Supporting Information

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Fig. S1. Study area. (A) Map of the cruise track (pink) during the ASCOS expedition (08-2008, at 87–88° N, 2–10° W), with ice-drift period highlighted (red, and inset area indicated) and (inset) shown in detail with the start of the drift marked by the circle. The ice edge (blue line) is shown for the start of the drift period on 12 August 2008. The drift area was within air with a minimal influence by man-made sources. (*B*) Aerial photograph of the ice floe used for the 3-wk drift taken from a helicopter where the blue circle indicates the location of the lcebreaker Oden, the yellow circle is the location of the meteorological sampling camp, and the red circle shows the location of lead sampled. (*C*) Air mass trajectories with an arrival height of 100 m at the position of the icebreaker.



Fig. 52. Partial characterization of DOM in SML and SSW waters. (A) Protein concentration (fraction $>0.7 \mu$ m) in SSW and SML waters. (B) Dissolved poly- and monosaccharide as well as particulate protein enrichment; proteins and polysaccharides were enriched in the SML immediately after cold periods.



Fig. S3. Hydrophobic moieties in microgels. (A) Cloud water microgels stained with Red Nile indicate the presence of hydrophobic moeities, with green corresponding to nonhydrophobic moeities. (B) Enrichment of dissolved hydrophobic amino acids (leucine, isoleucine, phenylalanine, and cysteine) in SML with respect to SSW waters.



Fig. S4. Microgels in fogs and aerosols. (A) Fog and (B) airborne aerosol nano and microgels immunostained with an antibody probe against material toward surface ocean waters from 87 to 88°N, 2–10°W. Relative frequency of Np [dNp/log(dDp): delta of Np/log delta Dp] in a specific size range for immunostained gels observed with confocal microscopy shows that 80% of the gels are in the smaller nanometer size range for both (C) fog and (D) airborne aerosol particles.



Fig. S5. Cleavage of microgels irradiated with environmental levels of UV at 87°N. Environmental levels of UV irradiation on spontaneously assembled microgels from the SSW DOM resulted in a factor of three reduction of the microgel yield indicating dispersion of microgels and cleavage of biopolymers.

Table S1. Cross reactivity of antibody to DOM from selected phytoplankton species kept in culture and field samples

No.*	Class	Genus	Species	Reactivity
327	Cryptophyceae	Guillardia	theta	_
334	Coscinodiscophyceae	Cyclotella	meneghiniana	_
336	Coscinodiscophyceae	Cyclotella	meneghiniana	_
338	Coscinodiscophyceae	Cyclotella	meneghiniana	_
419	Dinophyceae	Protodinium	sp.	_
425	Dinophyceae	Gymnodinium	sp.	_
440	Cryptophyceae	Hemiselmis	rufescens	_
452	Raphidophyceae	Heterosigma	akashiwo	—
525	Eustigmatophyceae	Nannochloropsis	oculata	_
689	Dinophyceae	Prorocentrum	micans	_
702	Dinophyceae	Prorocentrum	sp.	_
703	Dinophyceae	Prorocentrum	sp.	_
1178	Cryptophyceae	Rhodomonas	abbreviata_cf	_
1322	Dinophyceae	Heterocapsa	pygmaea	_
1577	Coscinodiscophyceae	Cyclotella	striata_cf	_
1594	Euglenophyceae	Eutreptiella	gymnastica_cf	_
1595	Raphidophyceae	Heterosigma	akashiwo	—
1647	Coscinodiscophyceae	Thalassiosira	rotula	_
2283	Prymnesiophyceae	Emiliania	huxleyi	_
2715	Cryptophyceae	Proteomonas	sp.	—
2948	Dinophyceae	Symbiodinium	sp.	_
3162	Bacillariophyceae	Surirella	sp.	—
3171	Prasinophyceae	unid	sp.	_
3194	Rhodophyceae	Bostrychia	calliptera	—
1013	Coscinodiscophyceae	Thalassiosira	pseudonana	—
ASCOS	Coscinodiscophyceae	Melosira	artica	+
	Phytoplankton from Puget Sound			—

The antibody was generated against seawater DOM collected during ASCOS (08-2008, 87° N, 2–10° W). A negative sign indicates no immuno reactivity, whereas a positive sign indicates reactivity. *Culture Center for Marine Phytoplankton, Bigelow Laboratory for Ocean Sciences

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