Table S1. Primers Used to Prepare ToMOH Variants.

Variant	Primer					
I100W	5'-CAA CTT CAC TTC GGA GCG <u>TGG</u> GCA CTT GAA GAA TAC G-3'					
	5'-C GTA TTC TTC AAG TGC <u>CCA</u> CGC TCC GAA GTG AAG TTG-3'					
I100A	5'-CAC TTC GGA GCG <u>GCT</u> GCA CTT GAA GAA TAC-3'					
	5'-GTA TTC TTC AAG TGC <u>AGC</u> CGC TCC GAA GTG-3'					
W167E	5'-CAT ACT AAC GAA GAA GCC GCA ATC GCG GCA CGG-3'					
	5'-CCG TGC CGC GAT TGC GGC TTC TTC GTT AGT ATG-3'					
F176A	5'-GCT GCA CGG TCT TTC <u>GCT</u> GAC GAC ATG ATG-3'					
	5'-CAT CAT GTC GTC <u>AGC</u> GAA AGA CCG TGC AGC-3'					
F176W	5'-GCA CGG TCT TTC <u>TGG</u> GAC GAC ATG ATG ATG -3'					
	5'-CAT CAT CAT GTC GTC <u>CCA</u> GAA AGA CCG TGC -3'					
F205A	5'-C <u>AGC</u> AAT ATG CAG <u>GCT</u> CTC GGT TTG GCC G-3'					
	5'-C GGC CAA ACC GAG <u>AGC</u> CTG CAT ATT <u>GCT</u> G-3'					
F205W	5'-GGC TTC <u>AGC</u> AAT ATG CAG <u>TGG</u> CTC GG T TTG GCC GCT GAC-3'					
	5'- GTC AGC GGC CAA ACC GAG <u>CCA</u> CTG CAT ATT <u>GCT</u> GAA GCC-3'					
L208W	5'-CAG TTT CTC GGT <u>TGG</u> GCC GCT GAC GCT G-3'					
	5'-C AGC GTC AGC GGC <u>CCA</u> ACC GAG AAA CTG-3'					
L208W/D211W	5'-G TTT CTC GGT <u>TGG</u> GCC GCT <u>TGG</u> GCT GCT G-3'					
	5'-C AGC AGC <u>CCA</u> AGC GGC <u>CCA</u> ACC GAG AAA C-3'					
V271A	5'-G AAG CTT TTC TCG <u>GCT</u> CTC ACC GGC CC-3'					
	5'-GG GCC GGT GAG <u>AGC</u> CGA GAA AAG CTT C-3'					
L272A	5'-G CTT TTC TCG GTA <u>GCT</u> ACC GGC CCC ATC-3'					

	5'-GAT GGG GCC GGT AGC TAC CGA GAA AAG C-3'
L272E	5'-G CTT TTC TCG GTA <u>GAA</u> ACC GGC CCC ATC-3'
	5'-GAT GGG GCC GGT <u>TTC</u> TAC CGA GAA AAG C-3'
1276E	5'-GTA CTC ACC GGC CCC <u>GAA</u> ATG GAT TAT TAC AC-3'
	5'-GT GTA ATA ATC CAT <u>TTC</u> GGG GCC GGT GAG TAC-3'

Location of	Fe Content,	Specific steady state	Relative steady state
the mutation	[Fe]/[ToMOH]	activity (mU/mg)	activity (%) ^b
-	3.7 ± 0.6	1200 ± 200	50 ± 10
-	4.15 ± 0.04	2400 ± 300	100
Cavity 1	4.14 ± 0.06	470 ± 130	20 ± 6
Cavity 1	3.43 ± 0.03	nd ^d	nd ^d
Cavity 3	5.97 ± 0.29 ^c	720 ± 50	30 ± 4
Cavity 1	2.72 ± 0.10	1290 ± 80	54 ± 8
Cavity 1	3.19 ± 0.14	nd ^d	nd ^d
Cavity 1	2.76 ± 0.08	230 ± 50	10 ± 2
Cavity 1	4.72 ± 0.13	nd ^d	nd ^d
Channel	4.74 ± 0.09	nd ^d	nd ^d
Channel	4.10 ± 0.06	nd ^d	nd ^d
Cavity 2	3.22 ± 0.07	1140 ± 100	48 ± 7
Cavity 2	4.78 ± 0.02	940 ± 140	39 ± 8
Cavity 2	6.8 ± 0.1 ^c	310 ± 40	13 ± 2
Channel	5.9 ± 0.1°	1400 ± 490	58 ± 22
	Location of the mutation - Cavity 1 Cavity 1 Cavity 3 Cavity 1 Cavity 1 Cavity 1 Cavity 1 Cavity 1 Cavity 1 Cavity 1 Cavity 2 Cavity 2 Cavity 2 Cavity 2 Cavity 2 Cavity 2 Cavity 2	Location of the mutationFe Content, [Fe]/[ToMOH] $ 3.7 \pm 0.6$ $ 4.15 \pm 0.04$ $ 4.15 \pm 0.04$ Cavity 1 4.14 ± 0.06 Cavity 1 3.43 ± 0.03 Cavity 3 $5.97 \pm 0.29^{\circ}$ Cavity 1 2.72 ± 0.10 Cavity 1 3.19 ± 0.14 Cavity 1 2.76 ± 0.08 Cavity 1 4.72 ± 0.13 Channel 4.74 ± 0.09 Channel 4.10 ± 0.06 Cavity 2 3.22 ± 0.07 Cavity 2 $6.8 \pm 0.1^{\circ}$ Channel $5.9 \pm 0.1^{\circ}$	Location of the mutationFe Content, [Fe]/[ToMOH]Specific steady state activity (mU/mg)- 3.7 ± 0.6 1200 ± 200 - 4.15 ± 0.04 2400 ± 300 Cavity 1 4.14 ± 0.06 470 ± 130 Cavity 1 3.43 ± 0.03 nd ^d Cavity 3 $5.97 \pm 0.29^{\circ}$ 720 ± 50 Cavity 1 2.72 ± 0.10 1290 ± 80 Cavity 1 3.19 ± 0.14 nd ^d Cavity 1 2.76 ± 0.08 230 ± 50 Cavity 1 4.72 ± 0.13 nd ^d Channel 4.10 ± 0.06 nd ^d Cavity 2 3.22 ± 0.07 1140 ± 100 Cavity 2 $6.8 \pm 0.1^{\circ}$ 310 ± 40 Channel $5.9 \pm 0.1^{\circ}$ 1400 ± 490

Table S2. Fe Content and Specific Steady State Activity of ToMOH Variants.^a

^aFe content and specific steady state activity were measured in triplicate and the average and standard deviations were calculated. ^bRelative steady state activity of the variants were measured relative to the value of T201S single variant. The errors for the relative steady state activity are determined by propagating the standard deviation values for the variants. ^cA few variants contained > 4 Fe atoms per protein, possibly due to binding of adventitious iron. In such cases, the protein was reduced with dithionite and dialyzed anaerobically to remove the excess iron, which led to approximately 4 Fe ions per protein. ^dNot detected.

	T201S/I100A	T201S/I100W	T201S/W167E	T201S/F176W	T201S/V271A	T201S/L272E	T201S/I276E
Data Collection							
Beamline	NE-CAT 24 ID-E	NE-CAT 24 ID-E	NE-CAT 24 ID-E	NE-CAT 24 ID-E	SSRL 12-2	NE-CAT 24 ID-E	NE-CAT 24 ID-E
Wavelength (Å)	0.979	0.979	0.979	0.979	1.034	0.979	0.979
Space group	P3 ₁ 21						
Unit cell dimensions (Å)	182.6 x 182.6 x 67.0	182.8 x 182.8 x 67.7	182.6 x 182.6 x 68.3	183.1 x 183.1 x 68.0	183.4 x 183.4 x 68.3	182.4 x 182.4 x 67.5	183.0 x 183.0 x 68.0
Resolution range (Å)	30.0 - 2.75	50.0 - 3.00	50.0 - 2.80	50.0 – 2.65	50.0 - 2.2	50.0 - 2.80	50.0 - 2.50
Total reflections	195303	149031	199829	230251	598335	244050	293744
Unique reflections	27980	24140	29390	36462	66756	30252	43135
Completeness (%) ^a	93.7 (95.3)	96.6 (97.4)	96.6 (97.9)	99.7 (100)	99.7 (100)	99.6 (100)	99.7 (99.8)
l/σ(l) ^a	21.8 (4.0)	30.1 (9.0)	26.2 (4.6)	33.1 (7.4)	16.4 (6.9)	25.3 (7.2)	26.9 (5.2)
R _{sym} (%) ^{a,b}	10.2 (42.1)	10.3 (26)	10.1 (42.3)	9.2 (36.6)	10.0 (31.7)	13.9 (40.8)	9.9 (39.3)
Refinement							
R _{cryst} (%) ^c	19.0	18.0	19.5	17.8	15.6	16.4	18.4

Table S3. X-ray Data Collection and Refinement Statistics for ToMOH Variants.

R_{free} (%) ^d	25.1	24.1	26.0	23.5	20.0	22.7	23.4
Average B-value (Å ²)	64.1	53.0	52.3	45.9	24.2	27.6	42.8
r.m.s deviation bond length (Å)	0.008	0.016	0.015	0.019	0.024	0.016	0.021
r.m.s deviation bond angles (°)	1.11	1.65	1.66	1.81	1.81	1.61	1.83
No. protein atoms	7331	7340	7329	7368	7355	7348	7335
No. non-protein atoms	53	57	51	169	586	239	298
Fe atoms	2	2	2	2	2	2	2
Water molecules	46	55	49	152	566	212	296
PEG-400 fragments ^e	1	0	0	0	2	2	0
RMSD to wild- type ToMOH	0.20	0.19	0.23	0.16	0.17	0.16	0.15
PDB Code	3RNC	3RNA	3RNG	3RNB	3RNF	3RN9	3RNE

^aValues in parentheses are for the highest resolution shell. ^bR_{sym} = $\Sigma_i \Sigma_{hkl} |I_i(hkl) - \langle I(hkl) \rangle |/ \Sigma_{hkl} \langle I(hkl) \rangle$, where $I_i(hkl)$ is the *i*th measured diffraction intensity and $\langle I(hkl) \rangle$ is the mean intensity for the Miller index (*hkl*). ^cR_{cryst} = $\Sigma_{hkl} ||F_o(hkl)| - |F_c(hkl)||/$ $\Sigma_{hkl} |F_o(hkl)|$. ^dR_{free} = R_{cryst} for a test set of reflections (5% in each case). ^eA second PEG-400 molecule is observed in the T201S/V271A and T201S/L272E variants. They are positioned in the cavity 3 as shown in Fig S5.

Figures Captions

Figure S1. (A) Superposition of residues gating access between cavities 1 and 2 in T4moH-T4moD (green sticks) and ToMOH (blue sticks) structures. Cavity 2 of the T4moH-T4moD structure is shown as red surfaces. Distances between L272 and L268, A172, and A99 in the T4moH-T4moD structure are 6.7 Å, 8.0 Å, and 4.5 Å, respectively. (B) Superposition of the T4moH-T4moD complex and ToMOH T201S/L272E (cyan sticks).

Figure S2. Plots of formation rate constants of $T201_{peroxo}$ vs dioxygen concentrations. Formation and decay rates were measured in the reaction of reduced ToMOH variants and ToMOD with various concentrations of dioxygen. Figures are arranged from top to bottom as: (A) T201S, (B) T201S/I100W, (C) T201S/L272E, (D) T201S/V271A, (E) T201S/W167E, and (F) T201S/I276E ToMOH variants. Formation and decay rate constants are represented as black squares and red circles, respectively. Both formation and decay rates were fit to a linear function as depicted with black and red lines, respectively. Second-order formation rate constants were obtained from the slope of a linear fit. Linear fits of decay rates in all variants, shown in red, yielded zero or very small slopes indicating that decay of T201_{peroxo} is independent of O₂ concentration.

Figure S3. Structural changes in the T201S/F176W ToMOH variant. The F176W substitution contracts the entrance to cavity 1. The closest C···C distance between the I100 and F176 is 6.0 Å in the T201S variant is reduced to 4.2 Å by the F176W mutation.

Figure S4. Structural changes in the T201S/W167E ToMOH variant. (A) Change in distances between W167E and nearby residues between cavities 2 and 3. Wild-type and W167E ToMOH gating residues are depicted as gray and green sticks, respectively. The distance between the W167E δ 1 oxygen and V271 γ carbon atoms is 7.8 Å (not shown). (B) Newly formed hydrogen bonds between W167E and adjacent residues, E166 and T341. Oxygen and nitrogen atoms in W167E, E166, and T341 residues are depicted by red and blue sticks, respectively.

Figure S5. A second PEG 400 molecule, in addition to one observed in the channel of the wild-type and T201S/I100A proteins, is observed in cavity 3 of the (A) T201S/L272E and (B) T201S/V271A ToMOH variants. In (A) the cavity 3 surfaces are colored light yellow and a $2F_{o}$ - F_{c} omit map for PEG is contoured to 1.0 σ (blue mesh). Residues delineating the gate between cavities 2 and 3 are represented by green sticks. The oxygen and carbon atoms of PEG are depicted by red and gray sticks, respectively. Ribbon structures at the top right of the panel correspond to the ToMOH γ -subunit. In (B), cavity surfaces are depicted pink as in Fig 2D to denote connectivity between all three cavities. Atoms and molecules are depicted as in (A), with the exception of the γ -subunit, which is not shown.

Figure S1.



Figure S2A.



Figure S2B.



Figure S2C.



Figure S2D.



Figure S2E.



Figure S2F.



Figure S3.



Figure S4.



Figure S5.

