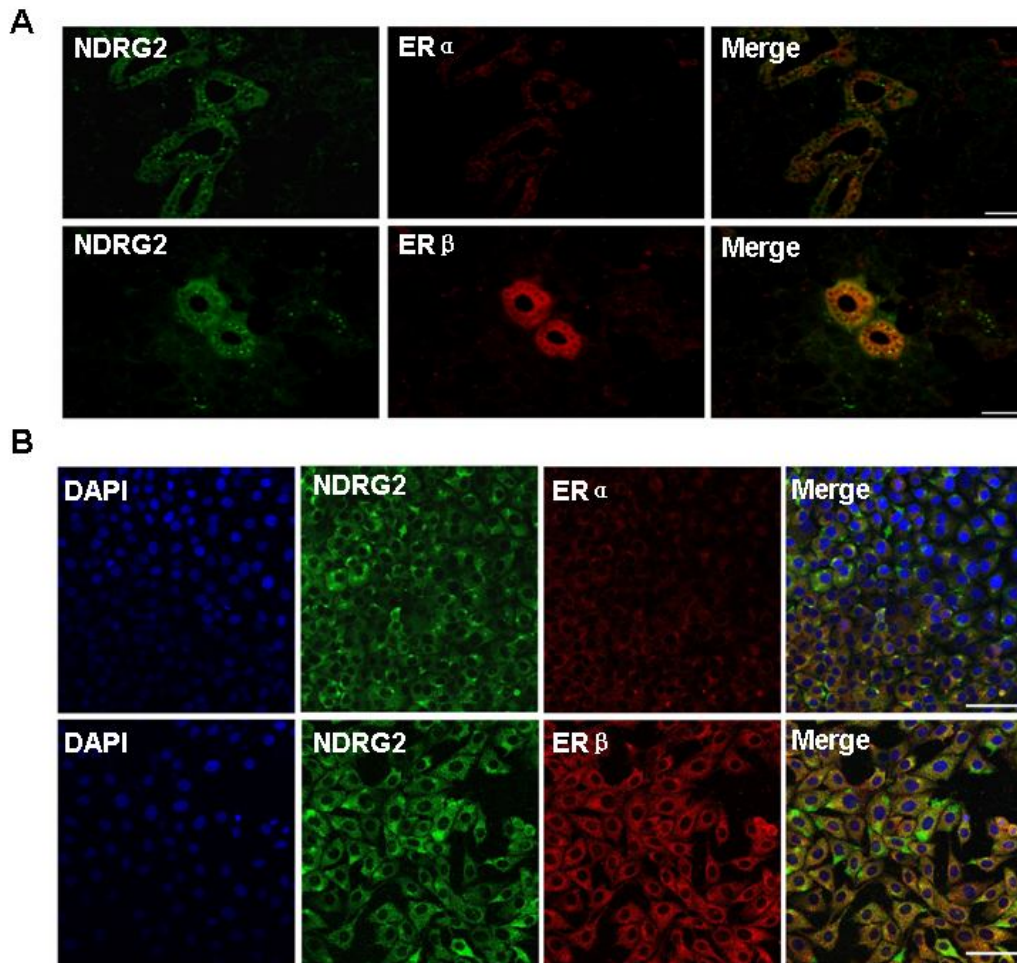
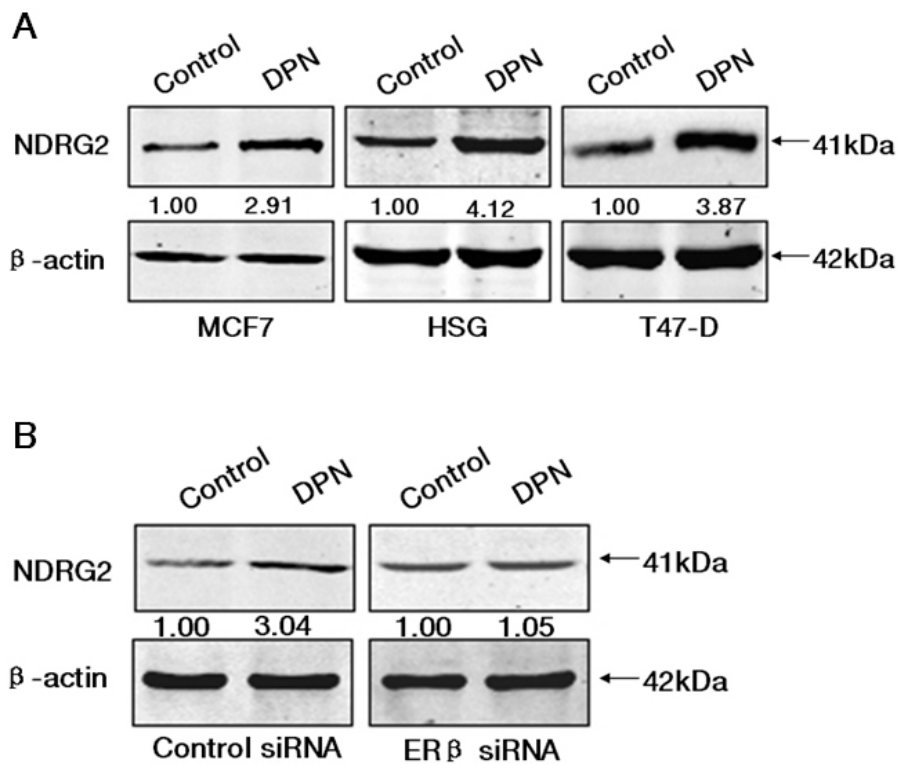


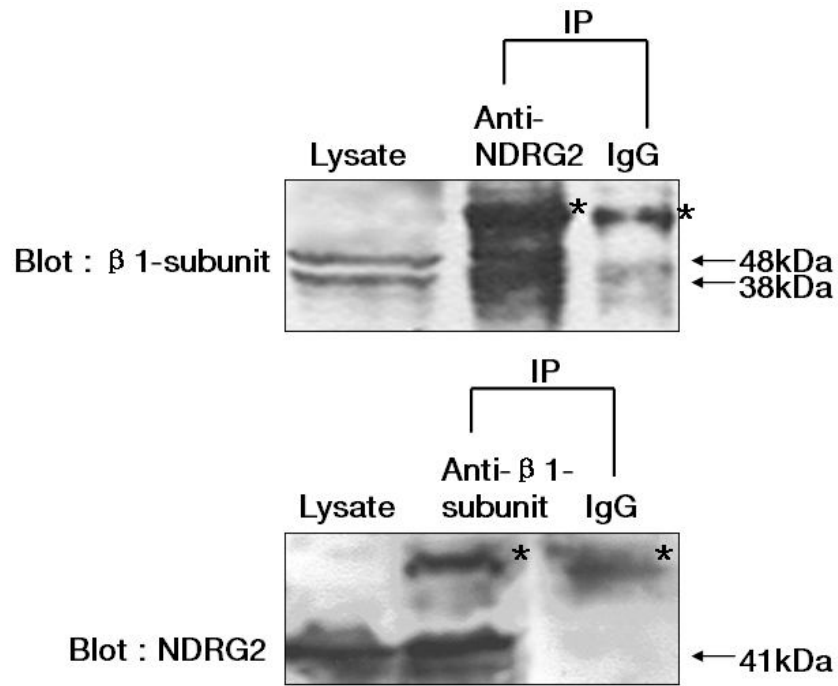
**Fig. S2. NDRG2 interacts with the Na<sup>+</sup>/K<sup>+</sup>-ATPase β1.** (A) Schematic of the full-length Na<sup>+</sup>/K<sup>+</sup>-ATPase β1 and its truncated fragments. The cDNAs encoding them were fused to Gal4-AD in pACT vector, and co-transformed into yeast *AH109* with full-length NDRG2-Gal4-BD vector. The grown clones on SD/-Trp-Leu-His-Ade/X-α-Gal dish were transferred onto nitrocellulose membrane and tested for β-galactosidase activity. The “+” stands for positive interaction, while “-” for negative clones. (B) Mammalian two-hybrid assay. HEK293 cells were co-transfected with 250 ng Na<sup>+</sup>/K<sup>+</sup>-ATPase β1-AD plasmid, 250 ng NDRG2-Gal4-BD plasmid and 250 ng pG5-*luc* reporter. The luciferase activity was measured and expressed as fold increase relative to the control cells from three experiments. 1. Background control: Gal4-AD vector, Gal4-BD vector, and pG5-*luc* vector; 2. Negative control: Gal4-BD vector, Na<sup>+</sup>/K<sup>+</sup>-ATPase β1-AD vector, and pG5-*luc* vector; 3. Negative control: NDRG2-Gal4-BD vector, Gal4-AD vector, and pG5-*luc* vector; and 4. Experiment group: NDRG2-Gal4-BD vector, Na<sup>+</sup>/K<sup>+</sup>-ATPase β1-AD vector, and pG5-*luc* vector. \*\* *p*<0.01 versus background control.



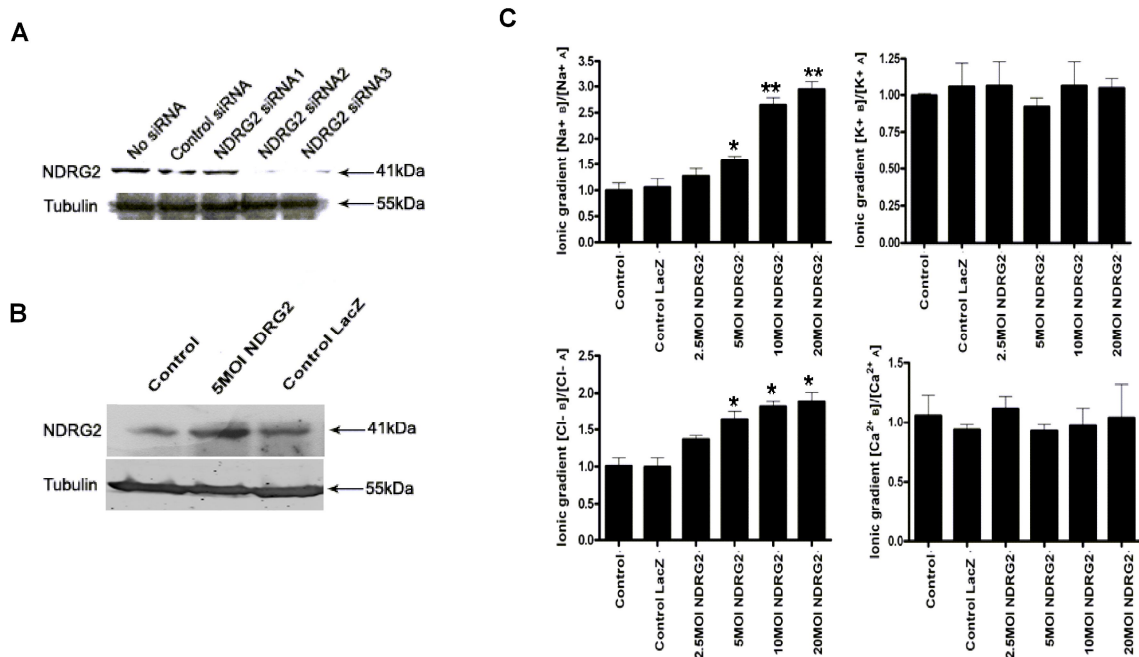
**Fig. S3. NDRG2 is co-localized with the ER $\beta$  in human salivary gland tissues and cells.** (A) The expression of NDRG2 and ER in salivary glands (Scale bars=50  $\mu$ m). (B) The expression of NDRG2 and ER in HSG cells (Scale bars=20  $\mu$ m). Salivary gland tissue sections and HSG cells were probed with mAb against NDRG2 antibody and rabbit anti-ER $\alpha$  or anti-ER $\beta$  antibodies and the bound antibodies were detected by FITC-goat anti-mouse IgG and Cy3-goat anti-rabbit IgG. The isotype mouse and rabbit IgG were used as negative controls (data not shown). The expression of NDRG2 and ER were examined under a fluorescent microscope. Data shown are representative images from five individual subjects in three independent experiments.



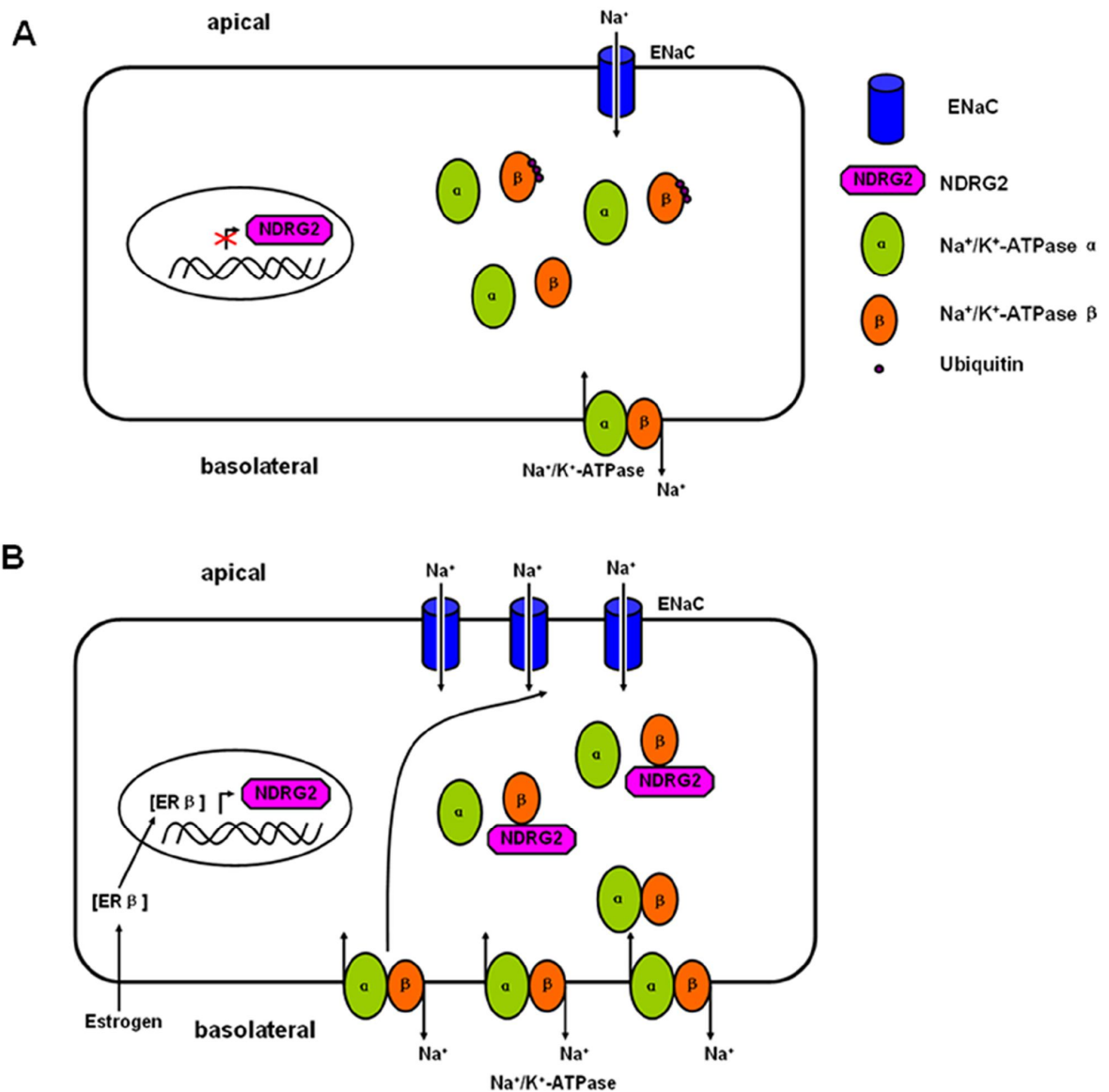
**Fig. S4. DPN enhances endogenous NDRG2 protein expression.** (A) MCF7, HSG and T-47D cells that express endogenous ER $\beta$  were treated with ER $\beta$  selective agonist, 10 nM DPN, or control ethanol for 24 h. The NDRG2 protein expression was determined by immunoblotting. (B) Silence of ER $\beta$  expression reduces their responses to DPN. HSG cells were transfected with 100 pmol ER $\beta$  specific siRNA or control siRNA for 48 h and treated with DPN for 24 h. The NDRG2 protein expression was determined by immunoblotting. Data are representative images of each group of cells from three separated experiments.



**Fig. S5. The interaction of endogenous NDRG2 and endogenous  $\text{Na}^+/\text{K}^+$ -ATPase  $\beta$ 1.** IPs were performed with whole cell lysates of HSG cells pretreated with protein A conjugated sepharose beads. Whole cell lysates were probed for input. The antibodies for immunoprecipitation and western blot are as indicated. The location of various proteins is indicated with arrowhead. Blot, western blot; IP, immunoprecipitation; \*, Ig heavy chain.



**Fig. S6. NDRG2 stimulate ionic transport in HSG cells.** (A) Silence of NDRG2 expression by siRNA. HSG cells were transfected with, or without, control siRNA or individual *NDRG2*-specific siRNAs, and the NDRG2 expression was determined by western blotting. The *NDRG2*-specific siRNA 2 was used for the following experiments. (B) Induction of NDRG2 expression by NDRG2 adenovirus. HSG cells were transfected with NDRG2 or LacZ adenovirus (5MOI) for 48 h, and the NDRG2 expression was determined by western blotting. (C) Induction of NDRG2 over-expression modulates the Na<sup>+</sup> and Cl<sup>-</sup> gradients. HSG cells were cultured on collagen I-coated Transwell-Col filters for monolayer and infected with NDRG2 or control LacZ adenovirus (2.5-20 MOI) for 48 h. The concentrations of Na<sup>+</sup>, K<sup>+</sup>, Cl<sup>-</sup> and Ca<sup>2+</sup> in the harvested media were measured. Data are expressed as the mean ± SD of the relative ratios of individual electrolyte in the basolateral compartment to that in the apical compartment from three independent experiments. A, apical; B, basolateral. \**p*<0.05; \*\* *p*<0.01 versus control.



**Fig. S7. Model for the mechanism of NDRG2 participates in estrogen-mediated Na<sup>+</sup>/K<sup>+</sup>-ATPase regulation in epithelial cells.** (A) In the absence of estrogen, NDRG2 transcripts at low levels. Na<sup>+</sup>/K<sup>+</sup>-ATPase  $\beta$ 1 is ubiquitinated and naturally degraded in cytosol. Na<sup>+</sup>/K<sup>+</sup>-ATPase and ENaC mediate Na<sup>+</sup> transport on basolateral and apical sides of plasma membrane at low levels, respectively. (B) In the presence of estrogen, estrogen binds to ER $\beta$ , forming transcription factor, which activates the *NDRG2* transcription. Increased NDRG2 protein binds and stabilizes Na<sup>+</sup>/K<sup>+</sup>-ATPase  $\beta$ 1 protein by inhibiting its ubiquitination and degradation. A greater accumulation of Na<sup>+</sup>/K<sup>+</sup>-ATPase promotes Na<sup>+</sup> pumping across the basolateral plasma membrane, leading to a low cytosolic concentrations of Na<sup>+</sup>, which stimulates the sensors of cytosolic Na<sup>+</sup> and activates the ENaC in the apical plasma membrane, facilitating the reabsorption of Na<sup>+</sup> in epithelial cells.

**Table S1. The sequences of primers.** The restricted endonuclease sites were underlined, and the mutant bases were framed. \* For vector construction.

pCMV-Myc-Na <sup>+</sup> /K <sup>+</sup> -ATPase β *	Forward: ACGC <u>GTCGACC</u> ATCGCCATGGCCCGCGGGAAAGCCAAG
	Reverse: CCGC <u>TGAGG</u> TGGGAAAGATTTGTGCTTGTG
NDRG2 siRNA1	ACATCCTGGCGAGATATGCTCTTAA
NDRG2 siRNA2	GCTCTCTGGAAATTCTGAGTTGATA
NDRG2 siRNA3	GCACCCAACCTGGATAACATTGAAT
Control siRNA	GGCCGCAAAGACCTTGTCTTAGAA
M1	Forward: CAGGTTCCCCCTGA <u>TTC</u> ATCCTGACCCAGAGACGAATTTAG
	Reverse: CTAAATTCGTCTCTGGGT <u>CAG</u> GATGA <u>ATC</u> AGGGGGAACCTG
M2	Forward: CAGGTTCCCCCTGAGTCATCCT <u>C</u> ACCCAGAGACGAATTTAG
	Reverse: CTAAATTCGTCTCTGGGT <u>G</u> AGGATGACTCAGGGGGAACCTG
M3	Forward: CAGGTTCCCCCTGA <u>TTC</u> ATCCT <u>C</u> ACCCAGAGACGAATTTAG
	Reverse: CTAAATTCGTCTCTGGGT <u>G</u> AGGATGA <u>ATC</u> AGGGGGAACCTG
ERβΔE (7-8) *	Forward: 5'-CCG <u>TGAG</u> ATAATGACCTTTGTGCCTC-3'
	Reverse: 5'-CG <u>GATCCT</u> CAGGAATTGAGCAGGATCATG-3'
ERβ siRNA	GCTACAAATCAGTGTACAATT
Control siRNA	GGCCGCAAAGACCTTGTAAATT
ChIP (-1378/-1240 NDRG2 promoter)	Forward: CCAAGGTCTAGGAAGCCAATG
	Reverse: CCTGAAGGTCCAGCATCTAA
ChIP (-75/+50 NDRG2 promoter)	Forward: GGCATTGACCCAGAGTCCCTG
	Reverse: GAAGTTGGACAACAAGGCGGGG
EMSA wt ERE NDRG2 promoter	Forward: CAGGTTCCCCCTGAGTCATCCTGACCCAGAGACGAATTTAG
	Reverse: CTAAATTCGTCTCTGGGT <u>CAG</u> GATGACTCAGGGGGAACCTG
EMSA mutant ERE NDRG2 promoter	Forward: CAGGTTCCCCCTGA <u>TTC</u> ATCCT <u>C</u> ACCCAGAGACGAATTTAG
	Reverse: CTAAATTCGTCTCTGGGT <u>G</u> AGGATGA <u>ATC</u> AGGGGGAACCTG
NDRG2-Gal4-BD *	Forward: ACGC <u>GTCGACC</u> ATGGCGGAGCTGCAGGAGG
	Reverse: CGGGT <u>TACCA</u> AGGGCCATTCAACAGGAGAC
Na <sup>+</sup> /K <sup>+</sup> -ATPase β1-AD *	Forward: CGCG <u>GATCCT</u> CGCCATGGCCCGCGGGCCC
	Reverse: CGGGT <u>TACCT</u> GTTGGGAAAGATTTGTGCTTGTG
Na <sup>+</sup> /K <sup>+</sup> -ATPase β1(real-time PCR)	Forward: TAAGGCGTACGGTGAGAACA
	Reverse: AATACTTAGGGCCAGAAACA
NDRG2 (real-time PCR)	Forward: GAGATATGCTCTTAACCACCCG
	Reverse: GCTGCCCAATCCATCCAA
β-actin (real-time PCR)	Forward: TGGCACCCAGCACAATGAA
	Reverse: CTAAGTCATAGTCCGCCTAGAAGCA