

FBF represses the Cip/Kip cell cycle inhibitor CKI-2 to
promote self-renewal of germline stem cells in *C. elegans*

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Supplemental Data

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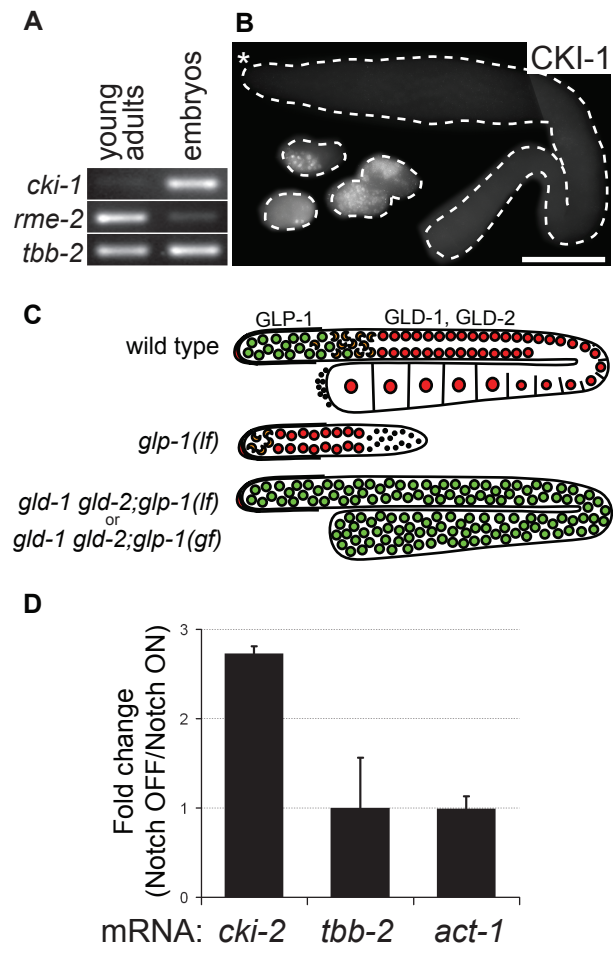


Figure S1

CKI-2, but not CKI-1, is predominantly expressed in the adult germ line.

(A) Semi-quantitative RT-PCR of indicated mRNAs. (B) Immuno-staining for CKI-1 protein on dissected gonads and embryos. Scale bar: 50 μ m. Both *cki-1* mRNA and CKI-1 protein can be detected in embryos, but are absent from non-gravid young adults. (C) A simplified view of GLP-1/Notch function in maintaining germline stem cells. In *glp-1(lf)* mutants, all germ cells enter meiosis, which depends on the pro-meiotic factors GLD-1 and GLD-2. In *gld-1 gld-2* mutants, germ cells proliferate independently of Notch signaling in either the presence of an additional loss-of-function mutation *glp-1(lf)* or a gain-of-function mutation *glp-1(gf)*. green, proliferating germline stem cells; yellow, nuclei entering meiosis; red, nuclei undergoing meiotic differentiation. (D) GLP-1/Notch signaling negatively regulates *cki-2* mRNA levels. The abundance of indicated mRNAs in gonads dissected from either *gld-1 gld-2; glp-1(lf)* (Notch OFF) or *gld-1 gld-2; glp-1(gf)* (Notch ON) gonads was measured by RT-qPCR. Control mRNAs: *tbb-2* (tubulin) and *act-1* (actin). Error bars = SEM, n = 3 (see Material and Methods).

Figure S2

Dissecting potential regulatory elements in the cki-2 3'UTR.

Alignment of *cki-2* 3'UTR sequences from *C.elegans*, *C.remanei*, *C.brenneri*, and *C.briggsae*, with the positions of both halvesites of MRE1 and MRE2 and the UGU trinucleotides of FBE1-4 indicated in colors.

The region required for repression, Δ FBE1-4, is boxed. Bold: TAA, the stop codon of the *cki-2* S ORF.

Nucleotides conserved between all four species are marked with an asterisk.

Kalchauer et al, Supplementary Fig.3

A

Oligo name	Kd, app (GS)	Kd, app (FP)
<i>cki-2</i> MRE2	9 ± 1 nM	15 ± 2 nM
<i>cki-2</i> MRE2 mut1	80 ± 20 nM	76 ± 4 nM
<i>cki-2</i> MRE2 mut2	>500 nM	>500 nM

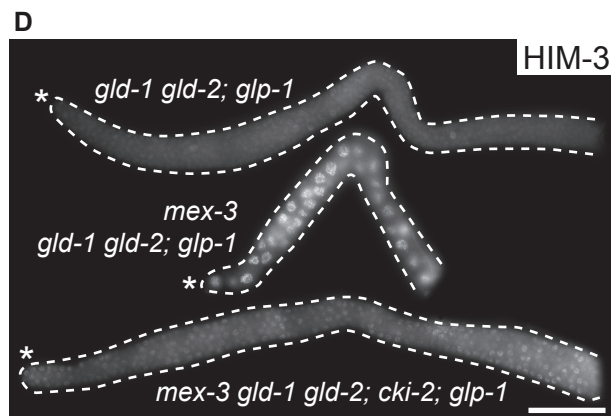
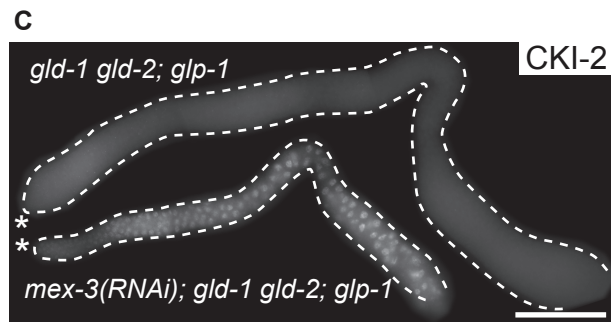
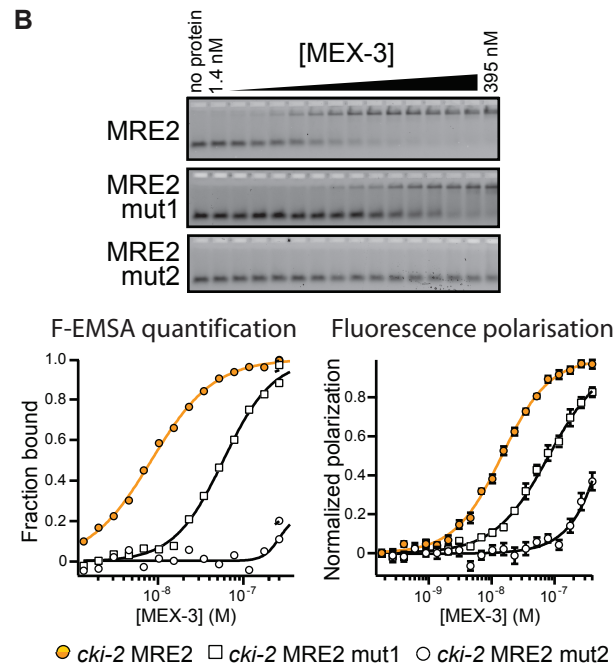


Figure S3

MEX-3 associates with MRE2 in the cki-2 3'UTR and regulates CKI-2 expression in certain genetic backgrounds.

(A) Affinities of MEX-3 to indicated RNA elements as determined by F-EMSA and FP assays. While MRE2 associates with MEX-3 tightly, MRE1 is predicted to be occluded by a stable hairpin (unpublished observation) and is not expected to associate with MEX-3. $K_{d,app}$, apparent equilibrium dissociation constant. For oligo sequences used see Table S1 in Supplemental Methods. (B) Top panel: F-EMSA gel pictures demonstrating association of MEX-3 to wild-type and mutated MRE2 from the *cki-2* 3'UTR. Bottom left: quantification of gel shifts shown above. The fraction of bound RNA was plotted as a function of protein concentration and fit to the Hill equation to determine the apparent equilibrium dissociation constant ($K_{d,app}$). Bottom right: affinity as determined by fluorescence polarization assays. Polarization is plotted as a function of protein concentration and fit to the Hill Equation to determine $K_{d,app}$. (C) CKI-2 immuno-staining on gonads (outlined) dissected from animals of the indicated genotype. MEX-3 prevents CKI-2 expression in *gld-1 gld-2; glp-1(lf)* gonads. (D) Immuno-staining for the synaptonemal complex component HIM-3 on dissected gonads (outlined) of indicated genotypes. Upregulation of CKI-2 in *mex-3 gld-1 gld-2; glp-1(lf)* gonads correlates with proliferation arrest and expression of the meiotic marker HIM-3. Removing *cki-2* from the *mex-3 gld-1 gld-2; glp-1(lf)* gonad rescues the proliferation defect and prevents accumulation of HIM-3. Scale bars: 50 μ m. Although MEX-3 is non-essential in the wild-type gonad, these experiments demonstrate that, in the presence of additional mutations, MEX-3 prevents meiosis by repressing CKI-2 expression.

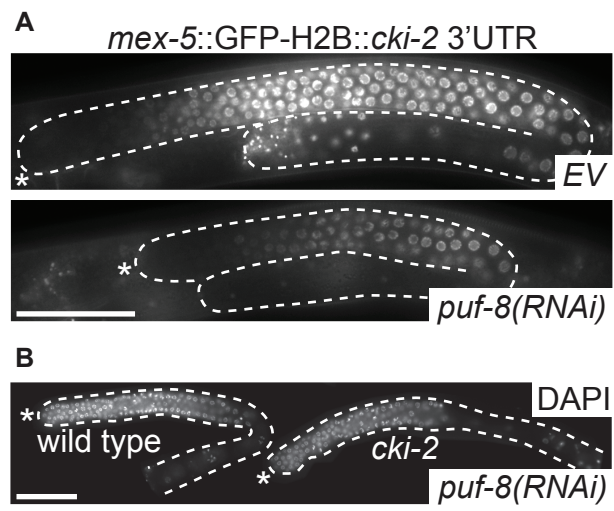


Figure S4

PUF-8 does not repress CKI-2.

(A) Live worms (gonads outlined) expressing the GFP::H2B reporter under the control of the wild type *cki-2* 3'UTR upon mock (EV: empty vector) or *puf-8* RNAi. Depletion of *puf-8* did not de-repress the reporter, though it did reduce germ line size as previously reported (Subramaniam and Seydoux 2003). (B) Dissected DAPI-stained gonads (outlined) of the indicated genotype upon RNAi of *puf-8*. *puf-8*(RNAi) caused germ line underproliferation in both wild-type and *cki-2* animals, indicating that this phenotype does not depend on CKI-2. Scale bars: 50µm

Supplementary Methods

Table S1

Oligonucleotide sequences used in in-vitro RNA binding assays.

MRE2 WT	ACUCAAAUCAUAGACAUUCUAGUUAUAAAU
MRE2 mut1	ACUCAAAUCCCCACAUCCCCGUUAUAAAU
MRE2 mut2	ACUCAAAUCCCCACAUCCCCGUCCCCAAU
FBEa (<i>gld-1</i>)	AUAGAAUCAUGUGCCAUAUCAUGUUG
FBE1 WT	UUUAUCUGUGAAUUUGAAAU
FBE1 mut	UUUAUCACAGAAUUUGAAAU
FBE2 WT	CAUACCCUGUCCAUUUCUGU
FBE2 mut	CAUACCCACACCAUUUCUGU
FBE3 WT	CAUUUCUGUGUUCUACUCCU
FBE3 mut	CAUUUCACAGUUCUACUCCU
FBE4 WT	CUACUCCUGUAAAAAAAGUC
FBE4 mut	CUACUCCACAAAAAAAGUC

Table S2

*Coordinates of regions deleted in the *cki-2* 3'UTR reporters*

name	deleted nucleotides (of 520)
Δ M2	409-426
Δ 1 Δ M2	1-302, 409-426
Δ 2 Δ M2	106-235, 409-426
Δ 3 Δ M2	211-340, 409-426
Δ 4	314-463
Δ 5 Δ M2	409-426, 409-426
Δ FBE1-4	246-313

Supplementary References

Biedermann, B., J. Wright, et al. (2009). "Translational repression of cyclin E prevents precocious mitosis and embryonic gene activation during *C. elegans* meiosis." *Dev Cell* 17(3): 355-64.

Ciosk, R., M. DePalma, et al. (2004). "ATX-2, the *C. elegans* ortholog of ataxin 2, functions in translational regulation in the germline." *Development* 131(19): 4831-41.

Draper, B. W., C. C. Mello, et al. (1996). "MEX-3 is a KH domain protein that regulates blastomere identity in early *C. elegans* embryos." *Cell* 87(2): 205-16.

Subramaniam, K. and G. Seydoux (2003). "Dedifferentiation of primary spermatocytes into germ cell tumors in *C. elegans* lacking the pumilio-like protein PUF-8." *Curr Biol* 13(2): 134-9.