SUPPLEMENTAL INFORMATION

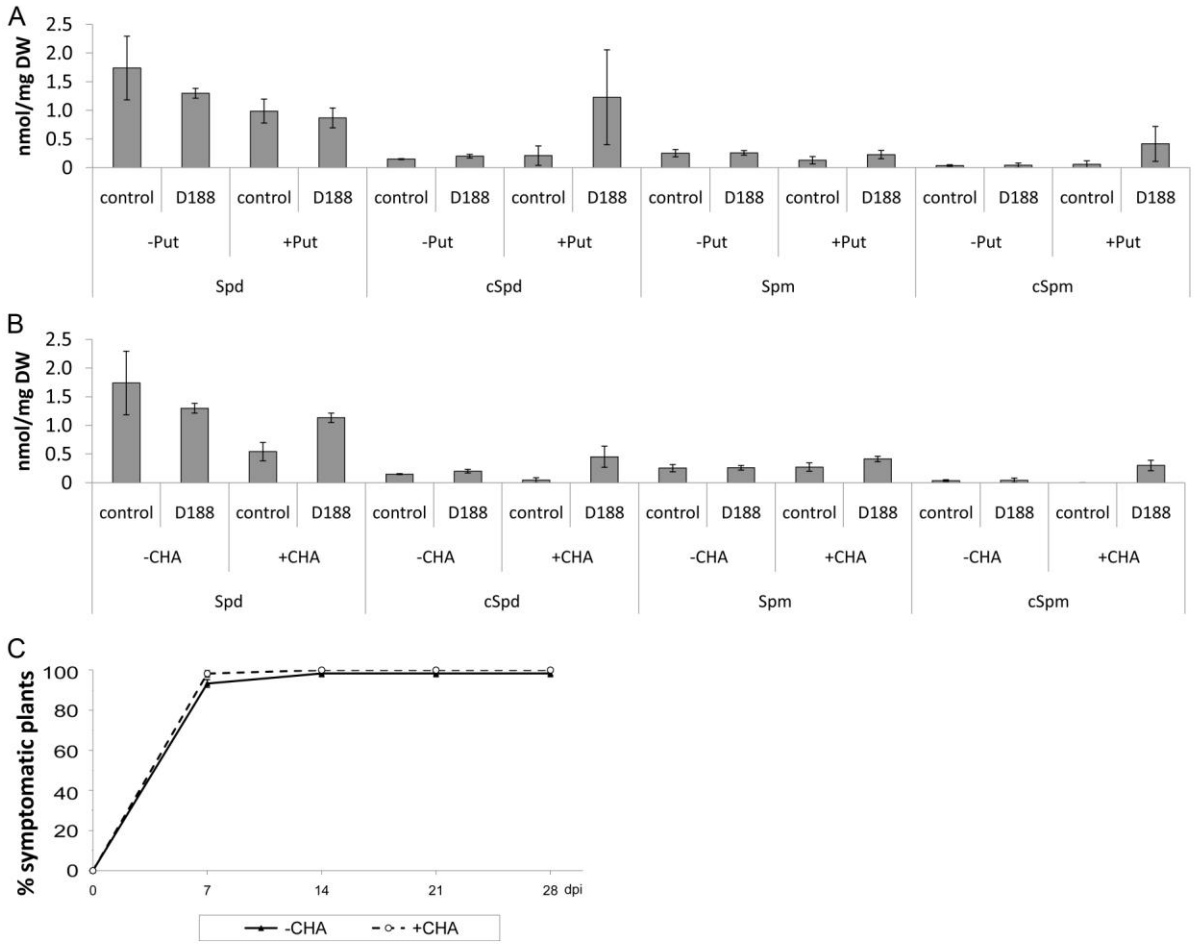

Figure S1. Effect of modulating the Put content on polyamine levels and symptom development in *R. fascians*-infected Arabidopsis Col-0.

Error bars represent standard errors (n=3). Student's *t*-tests did not reveal any statistical differences. A, Free and conjugated Spd and Spm levels measured at 14 dpi. Plants were placed on 1 mM Put 10 days before infection. B, Free and conjugated Spd and Spm levels measured at 14 dpi. Plants were placed on the SPDS inhibitor CHA (1 mM) just before infection. C, Percentage of symptomatic plants at different time points after infection.

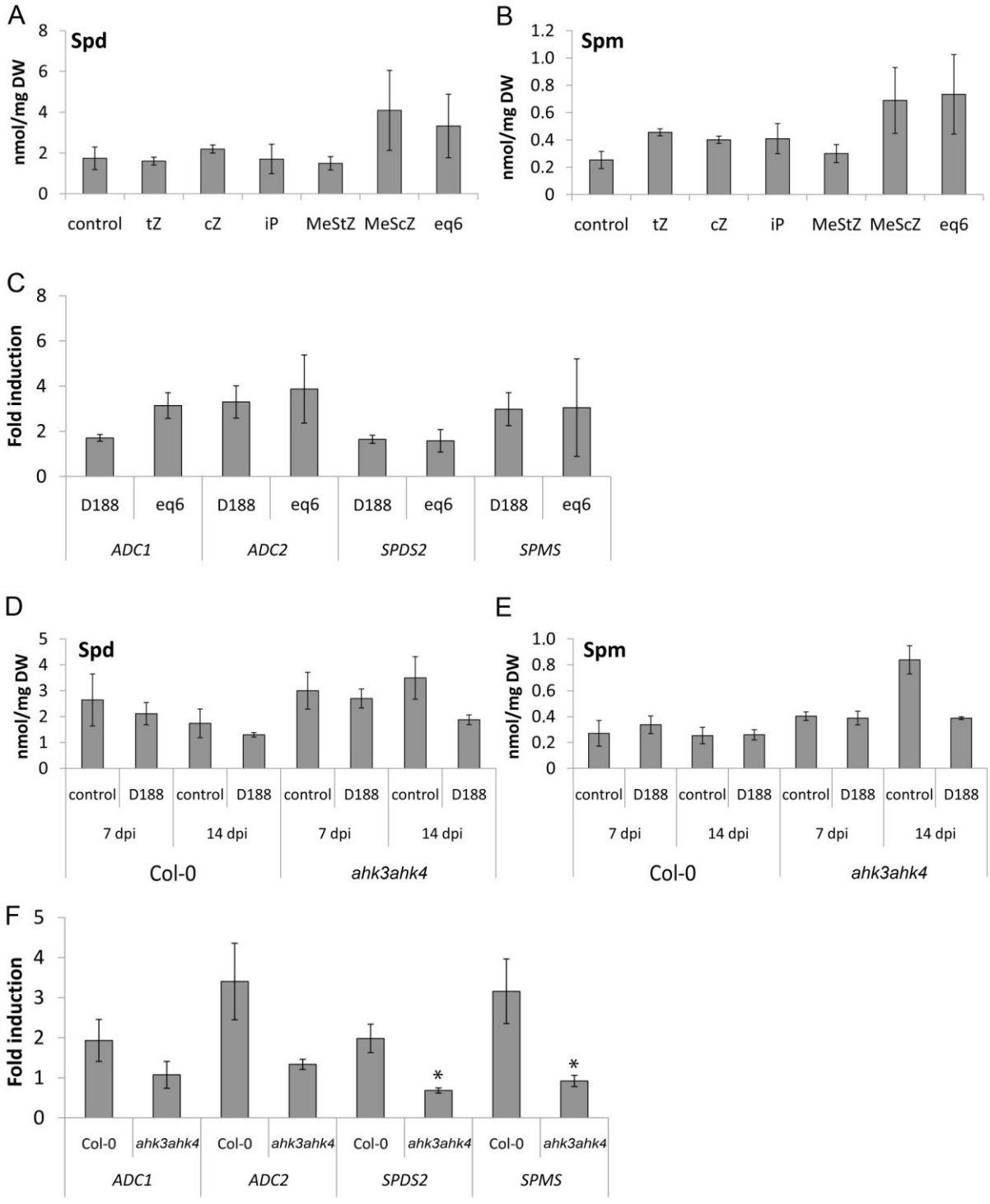
A
Spd
nmol/mg DW
8
6
4
2
0
control
tZ
cZ
iP
MeStZ
MeScZ
eq6
B
Spm
nmol/mg DW
1.2
1.0
0.8
0.6
0.4
0.2
0
control
tZ
cZ
iP
MeStZ
MeScZ
eq6
C
Fold induction
8
6
4
2
0
D188
eq6
D188
eq6
D188
eq6
D188
eq6
ADC1
ADC2
SPDS2
SPMS
D
Spd
nmol/mg DW
5
4
3
2
1
0
control
D188
control
D188
control
D188
control
D188
7 dpi
14 dpi
7 dpi
14 dpi
Col-0
ahk3ahk4
E
Spm
nmol/mg DW
1.0
0.8
0.6
0.4
0.2
0
control
D188
control
D188
control
D188
control
D188
7 dpi
14 dpi
7 dpi
14 dpi
Col-0
ahk3ahk4
F
Fold induction
5
4
3
2
1
0
*
*
Col-0
ahk3ahk4
Col-0
ahk3ahk4
Col-0
ahk3ahk4
Col-0
ahk3ahk4
ADC1
ADC2
SPDS2
SPMS

Figure S2. Importance of cytokinins on Spd and Spm production.

Error bars represent standard errors (n=3). Student's *t*-tests did not reveal any statistical differences (A-E). A-B, Free Spd (A) and (B) Spm levels in Col-0 plants treated with 10 μM of different cytokinins or with an equimolar mix of 1 μM each of iP, tZ, cZ, and their 2MeS derivatives (eq6) for 14 days. C, Fold-change in transcript levels of polyamine biosynthesis genes in D188-infected (D188 versus mock-infected) and eq6-treated (treated versus non-treated) Col-0 plants at 14 dpt. D-E, Free Spd (D) and Spm (E) levels in D188- and mock-infected (control) Col-0 and *ahk3ahk4* plants at 14 dpi. F, Fold-change in transcript levels of polyamine biosynthesis genes in D188 versus mock-infected Col-0 and *ahk3ahk4* plants at 14 dpi. Asterisks mark statistical differences ($p<0.05$) between wild-type and mutant plants according to a Student's *t*-test.

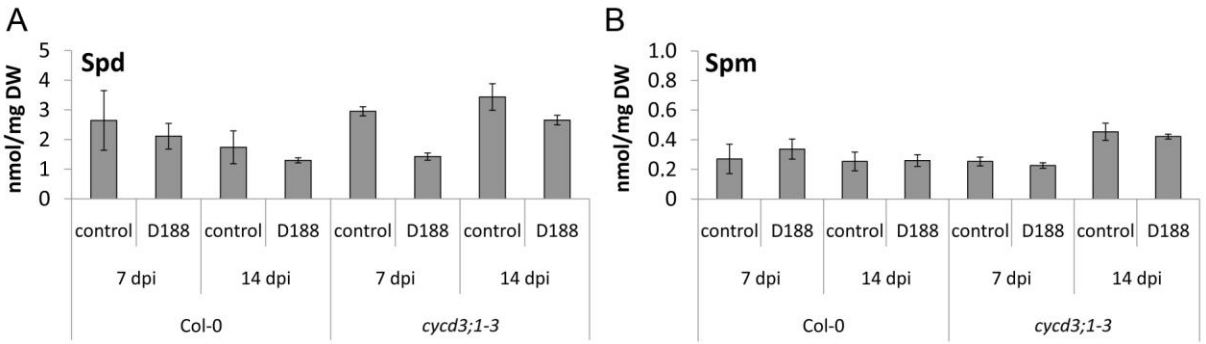

Figure S3. Free Spd (A) and Spm (B) levels in D188- and mock-infected (control) Col-0 and *cycd3;1-3* plants at 14 dpi.

Error bars represent standard errors (n=3). Student's *t*-tests did not reveal any statistical differences.

Table S1. *In vitro* and *in planta* bacterial growth and *fas*A gene expression in the presence and absence of Put.

To assess the effect of exogenous Put on bacterial growth, the turbidity of cultures (OD_{600}) and the colony forming units (CFU/mg fresh weight (FW)) present in infected Arabidopsis plants were determined, both upon growth in the presence or absence of 1 mM Put. We also monitored the expression of the *fas*A gene involved in cytokinin production in cultures of the GUS reporter strain D188-pSPIP*fasAgus* (Pertry et al., 2010) incubated under optimal conditions for *fas* gene expression (see "Material and Methods") and in bacterial cells present in plant tissues, again in the presence or absence of 1 mM Put. No significant differences were measured for either of these parameters using Student's *t*-tests.

Treatment	**OD_{600}**	**CFU/mg FW**	**GUS activity**
In vitro			
Pyr	0.837 ± 0.018		612.34 ± 124.36
His + Pyr	0.834 ± 0.011		7740.21 ± 517.57
Put + Pyr	0.893 ± 0.017		506.01 ± 111.50
Put + His + Pyr	0.997 ± 0.061		9404.64 ± 824.89
In planta			
Control		3615.22 ± 1401.66	40.06 ± 9.50
+Put		3477.66 ± 1177.51	58.08 ± 8.22

Table S2. Primers used for qRT-PCR amplifications.

Gene	AGI		Primer sequence	Reference
ACT2	At3g18780	Forward:	GGCTCCTCTTAACCCAAAGGC	Simón-Mateo et al. (2006)
		Reverse:	CACACCATCACCAGAATCCAGC	
ADC1	At2g16500	Forward:	CCAAGGTGTGTATCCTGTGAAAT	Jubault et al. (2008)
		Reverse:	AGCTTCTAAACCGAATCGAAAAC	
ADC2	At4g34710	Forward:	GCGATGGACCACACAGCTTT	Jubault et al. (2008)
		Reverse:	AGAACATCCGCTGAGGACTGA	
AIH	At5g08170	Forward:	TCGAGAATGCAAGAGAGATCGTT	Jubault et al. (2008)
		Reverse:	CATTTTCGGCGACGGAAGTA	
CPA	At2g27450	Forward:	GATCAAGTCGAAAAGGCAAAGCT	Jubault et al. (2008)
		Reverse:	CCATCCATAGTAAGAAGCACCTTGT	
SPDS1	At1g23820	Forward:	AATCACCACCTCTCACAAACCC	Jumtee et al. (2008)
		Reverse:	TCGGTGGCAGAGGTTTCTTTA	
SPDS2	At1g70310	Forward:	TTGCCCGTGAAGAGACCTAGA	Jubault et al. (2008)
		Reverse:	TCCACCGTTCTCTGTTTCCAT	
SPMS	At5g53120	Forward:	TGGCTCCATACTCATCTTATTGAA	Jubault et al. (2008)
		Reverse:	CGCATAGTGAACACTTTTGAATG	
ACL5	At5g19530	Forward:	CCATCATTTGCGGACACATG	Jubault et al. (2008)
		Reverse:	GAGACGAAAGAAGGAGCGTTTAGA	
SAMDC1	At3g02470	Forward:	TCTTTGAGCCAAGCATCTTTCA	Jubault et al. (2008)
		Reverse:	GCAGCAGGTGTAAGAATTTCATCA	
SAMDC2	At5g15950	Forward:	TCTCCGAGATCTACCTTGAAATG	Jubault et al. (2008)
		Reverse:	GATTCCCTATTCCTTCTCGTCCT	
SAMDC3	At3g25570	Forward:	GGTGGATAGGGTTCTGGTTTG	Cui et al. (2010)
		Reverse:	GGTGAGCAACATTCAACAGTC	
SAMDC4	At5g18930	Forward:	GGTGACCGTTACTCAACTATCCA	This work
		Reverse:	CGAAGCTCGCGTAGCTAAA	
ARR5	At3g48100	Forward:	TTTAAAAGCTCAAAGATTCACACACA	Jasinski et al. (2005)
		Reverse:	ATCAGCAAAAGAAGCCGTAATGT	
CYCD3;1	At4g34160	Forward:	CGTTCGTAGACCACATTATCAGGAG	Depuydt et al. (2009)
		Reverse:	CGGAGATTACAGAGAGGAGGAGAC	
CDKB1;1	At3g54180	Forward:	GGTGGTGACATGTGGTCTGTTGG	Boudolf et al. (2004)
		Reverse:	CGCAGTGTGGAAACACCCGG	

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