SUPPLEMENTARY DATA

Supplementary Research design and methods

Western Immunobloting

Total cell extracts were prepared using RIPA buffer. Briefly, ST-2 cells were washed twice with ice cold PBS and then the cells were lysed using ice cold modified RIPA buffer (50mM Tris-HCl pH=7.4, 1% NP-40, 0.25% Na deoxycholate, 150 mM Nacl, 1mM EDTA, 1mM PMSF, 1µg/ml of aprotinin, leupeptin and pepstatin, 1mM Na3VO4, 1mM NaF). Protein electrophor esis was performed in a 12% acrylamide gel and immunoblotting was performed as previously described(1). The primary antibodies used were anti-Nrf2 C-20 (Santa Cruz, Santa Cruz, CA), anti-α-tubulin T8203 (Sigma, St Louis, MO) and the secondary HRP conjugated antibodies (mouse #7076 and rabbit #7074S) were from Cell Signaling Technology (Beverly, MA).

Fluorescence staining of ST-2 cells

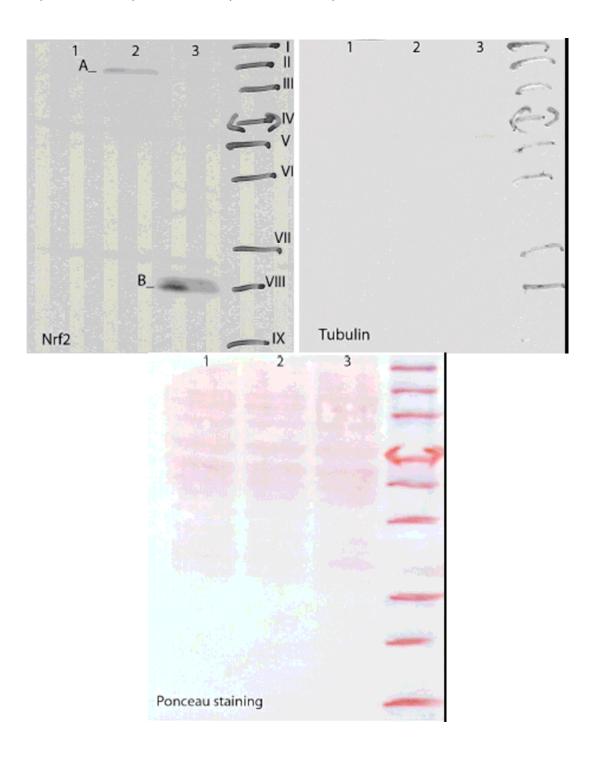
Cells were grown and transfected on sterile glass coverslips. Before the staining procedure, they were washed briefly with PBS 1x and then they were fixed with cold 4% paraformaldehyde (in PBS pH=7.4) for 10 minutes. After three 5-minute washing steps with PBS 1x (the first washing step with PBS 1x 1% glycine), they were incubated for 10 minutes with 1% Triton-x 100 in PBS 1x. Then, after three 5-minute washing steps with PBS 1x, cells were mounted in Vectashield Mounting Medium with DAPI (Vector labs, Burlingame, CA). Cells were observed and photographed using the Nikon Eclipse TE2000-U microscope of the Advanced Light Microscopy facility of the Medical School, University of Patras.

Transfection of ARE-ST2 cells

ARE-ST2 cells are ST-2 cells that stably express a plasmid (pGL2-basic-luc) that bears the Antioxidant Response Element (ARE) sequence upstream of the luciferase gene. Generation of these cells has been described previously (2). The ARE sequence is found in the promoter of Nrf2 target genes such as NQO1 (NADPH quinine oxidoreductase, HO-1 (Heme Oxygenase 1) etc. Thus, any increase in the transcriptional activity of Nrf2 is translated into increased luciferase activity. In this study, we transfected ARE-ST2 cells with Nrf2, DN-Nrf2 or empty vector under the conditions the transfection experiments with ST-2 cells were performed (in figure 6 of the main text).

Supplementary Figure 1. Nrf2 and DN-Nrf2 protein expression after transfection of their corresponding plasmids in ST-2 cells.

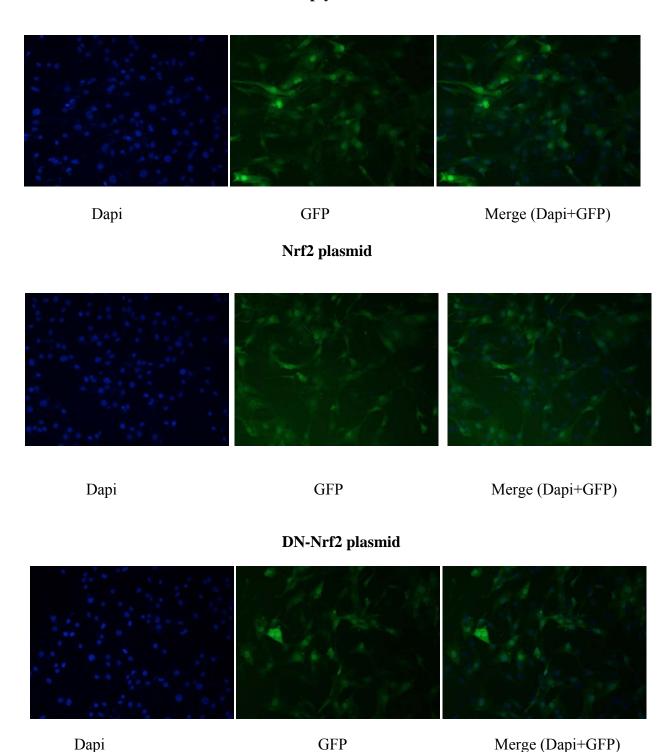
Total protein extracts from ST-2 cells transfected with Nrf2 (lane 2), DN-Nrf2 (lane 3) or empty plasmid (lane 1). Nrf2 expression was assayed by Western Immunoblotting. Tubulin was employed as a loading control. Ponceau staining of the nitrocellulose membrane was employed as a loading control as well. Due to low exposure time (1 min) endogenous Nrf2 is not detected in lane 1. Band A: Nrf2 band, Band B: DN-Nrf2 band. Bands of the protein ladder, I: 181.8 kDa, II: 115.5 kDa, III: 82.2 kDa, IV: 64.2 kDa, V: 48.5 kDa, VI: 37.1 kDa, VII: 25.9 kDa, VIII: 19.4 kDa, IX: 14.8 kDa.



Supplementary Figure 2. Transfection efficiency of ST-2 cells.

To check the efficiency of the delivery of plasmids in our experimental settings, ST-2 cells were transfected with pcDNA3-EGFP plasmid along with the plasmids used in the transfection experiments described in figure 6 of the main text. Then, the cells were stained with Dapi and were observed and photographed under UV microscope. The percentage of cells transfected is about the same (70%) in each experiment, consistent with the expected transfection efficiency of lipofectamine.

Empty vector

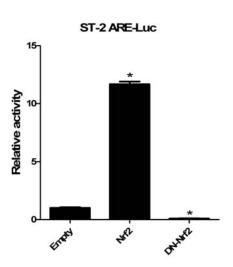


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SUPPLEMENTARY DATA

Supplementary Figure 3. Transcriptional activity of Nrf2 in ARE-ST2 cells after transfection with Nrf2, DN-Nrf2 or empty plasmid. Data show means±SEM. *p<0.001 compared to empty vector. The results are indicative of three different experiments performed on three different days. The statistical analysis was performed using ANOVA with Tukey's post-test.

Transfection of Nrf2 increases Nrf2 transcriptional activity more than 10-fold while transfection of DN-Nrf2 almost eliminates Nrf2 transcriptional activity.



Supplementary References

- 1. Ziros PG, Gil AP, Georgakopoulos T, Habeos I, Kletsas D, Basdra EK, Papavassiliou AG: The bone-specific transcriptional regulator Cbfa1 is a target of mechanical signals in osteoblastic cells. *J Biol Chem* 277:23934-23941, 2002
- 2. Habeos IG, Ziros PG, Chartoumpekis D, Psyrogiannis A, Kyriazopoulou V, Papavassiliou AG: Simvastatin activates Keap1/Nrf2 signaling in rat liver. *J Mol Med* 86:1279-1285, 2008