

Telomeric sequence detection using FISH

The products obtained using the previously published protocol (Ijdo *et al.*, 1991) were too large (>10Kb) and resulted in heavy precipitation on FISH slides. Size of products was reduced to 200-500bp by increasing the primer concentration from 0.1 μ M to 10 μ M. Labeling was carried out in the same reaction by substituting 0.1mM of dTTP for fluorescein-12-dUTP or biotin-16-dUTP (Roche).

Eigenmannia sp.1 and sp.2 slides were prepared following Henegariu *et al.* (2001) and treated with 100 μ L of RNase A (100 μ g/mL in 2xSSC) for 1h at 37°C, washed in PBS (2x5min) and dehydrated in ethanol. Pepsin treatment (0.005% in 10mM HCl) was carried out for 10min at 37°C. The slides were then washed (2x5min in PBS) and fixed with 1% formaldehyde (in PBS, 50mM MgCl₂) for 15min and washed with PBS (2x5min). Denaturing was performed at 65 °C for 2min in 70% formamide (in 0.66xSSC), after which the slides were dehydrated in ethanol and air dried.

Slides were covered with 12 μ L of either a low or high stringency hybridization solution consisting of 1-2 μ L (50-100ng) labeled probe, 70% (high stringency) or 50% (low stringency) of formamide, 10% dextran sulfate and 2xSSC. The hybridization solution was denatured for 10min at 75°C. Hybridization took place overnight at 37°C, and the slides were then washed at 42°C in 2xSSC containing 50% formamide (2x5min) and 2xSSC. When biotin labeled probes were used, slides were incubated at 37°C in 100mM Tris-HCl (pH 7.5), 150mM NaCl, 0.5% m/v dry milk for 30min, followed by incubation in the same conditions but with the addition of 10 μ L Cy3. Slides

were then washed 3 times for 5min in 4xSSC containing 0.005% tween 20 and then dehydrated in ethanol and air-dried. Slides prepared with directly labeled probes were washed 3 times for 5min with 4xSSC containing 0.005% Tween 20, dehydrated and air-dried following overnight hybridization. Slides were counter-stained with DAPI using Vectashield[®] Mounting Medium with DAPI (Vector Labs., CA, USA) and analyzed with the ImagePro Plus[®] software. The results are displayed in the figure below (Figure S3.1).

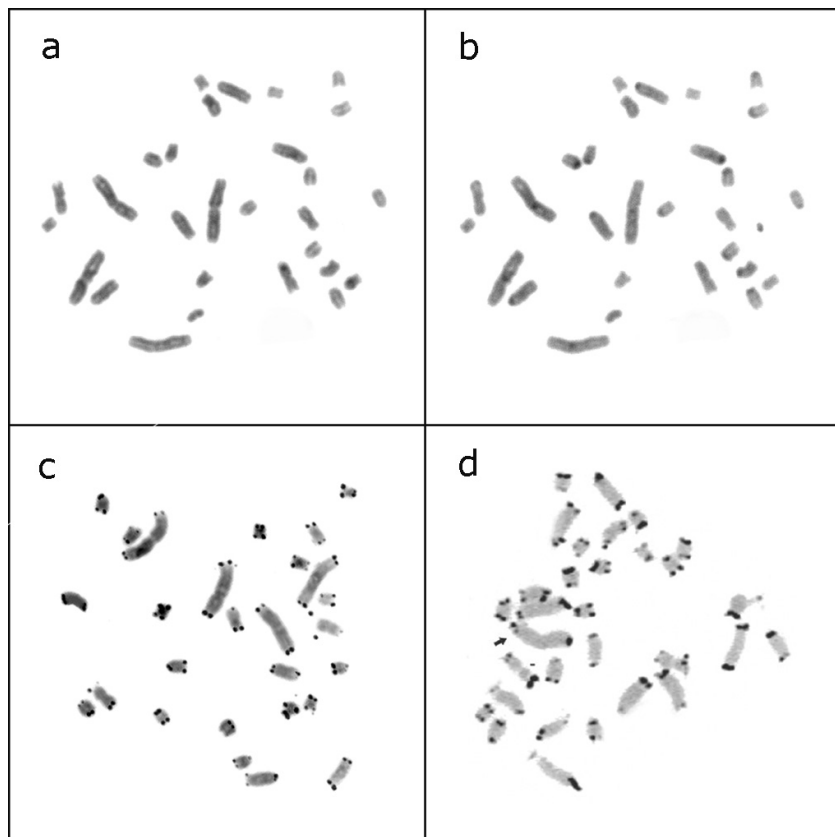


Figure S3.1. Chromomycin-DAPI double staining of *Eigenmannia sp.1*. using **a)** DAPI and **b)** chromomycin. Results of telomeric sequence detection using FISH on **c)** *Eigenmannia sp.1* and; **d)** *Eigenmannia sp.2*. The Y chromosome is shown with an arrow in **d)**.

Cited References

Henegariu O, Heerema NA, Lowe Wright L, Bray-Ward P, Ward DC, Vance GH (2001). Improvements in cytogenetic slide preparation: controlled chromosome spreading, chemical aging and gradual denaturing. *Cytometry* **43**(2): 101-109.

Ijdo JW, Wells RA, Baldini A, Reeders ST (1991). Improved telomere detection using a telomere repeat probe (TTAGGG)_n generated by PCR. *Nucleic Acids Res* **19**(17): 4780.