

# HETEROGENEOUS CATALYTIC HYDROGENATION OF UNPROTECTED INDOLES IN WATER: A GREEN SOLUTION TO A LONG-STANDING CHALLENGE

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**MATERIALS:** All the indoles and acid additives were purchased from Aldrich and used without any purification. Pt/C was purchased from Acros Organics and Pt/Al<sub>2</sub>O<sub>3</sub> was purchased from Engelhard. CDCl<sub>3</sub> used as a solvent (99.8%) for NMR studies was an Aldrich product. Other solvents used in synthesis with minimum purity of 99.5% were Fisher products. Water used as a solvent in the synthesis was deionized water.

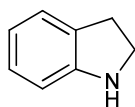
**NMR ANALYSIS:** The <sup>1</sup>H and <sup>13</sup>C spectra were obtained on a 300 MHz Varian NMR spectrometer, in CDCl<sub>3</sub> with tetramethylsilane as internal standards or the residual solvent signals. The temperature was 25 °C (accuracy ±1 °C) and controlled by the Varian control unit.

**GC-MS ANALYSIS:** The mass spectrometric identification of the products have been carried out by an Agilent 6850 gas chromatograph- 5973 mass spectrometer system (70 eV electron impact ionization) using a 30m long DB-5 type column (J&W Scientific).

**MELTING POINTS:** All the melting points are uncorrected and recorded on a MEL-TEMP apparatus.

**Representative procedure for the hydrogenation of indoles to indolines:** The hydrogenations were performed in a Berghof HR-100 autoclave using a Teflon liner at room temperature (25°C). Pt/C (30 mg), *p*-TSA·H<sub>2</sub>O (228 mg, 1.2 mmol), indole (117 mg, 1 mmol) and deionized water (5 mL) were charged to the Teflon liner. Then the autoclave was flushed with hydrogen three times, filled to a pressure of 30 bar and stirred (1000 rpm) for 3 h. After the completion of the reaction, the catalyst was removed by filtration. The filtrate was quenched with 1 mol % aqueous NaOH solution by adjusting the pH to 8. The aqueous layer was extracted with two portions of 15 mL CH<sub>2</sub>Cl<sub>2</sub>. The CH<sub>2</sub>Cl<sub>2</sub> extracts were combined and dried over Na<sub>2</sub>SO<sub>4</sub>. The solvent was removed *in vacuo* and the crude product was purified by flash chromatography.

#### 4. Indoline



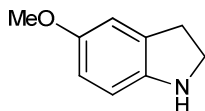
Colorless oil,  $R_f = 0.30$  (20 % EtOAc in hexane)

<sup>1</sup>H NMR (300.128 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.21 (d,  $J = 7.2$  Hz, 1H), 7.11 (t,  $J = 7.8$  Hz, 1H), 6.80 (t,  $J = 7.5$  Hz, 1H), 6.72 (d,  $J = 7.8$  Hz, 1H), 3.59 (t,  $J = 8.4$  Hz, 2H), 3.10 (t,  $J = 8.1$  Hz, 2H).

<sup>13</sup>C NMR (75.474 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 151.4, 129.1, 126.9, 124.4, 118.4, 109.2, 47.1, 29.6.

MS-C<sub>8</sub>H<sub>9</sub>N (119)  $m/z$  (%): 119 (M<sup>+</sup>, 100), 91 (21), 77 (2), 65 (6), 58 (7).

#### 10. 5-Methoxyindoline



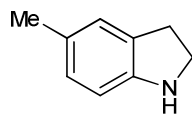
Light brown oil,  $R_f = 0.25$  (30 % EtOAc in hexane)

<sup>1</sup>H NMR (300.128 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 6.78 (s, 1H), 6.60 (m, 2H), 3.75 (s, 3H), 3.50 (t,  $J = 8.4$  Hz, 2H), 3.00 (t,  $J = 8.1$  Hz, 2H).

<sup>13</sup>C NMR (75.474 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 153.6, 145.5, 131.3, 112.2, 111.6, 110.2, 56.0, 47.9, 30.6.

MS-C<sub>9</sub>H<sub>11</sub>NO (149)  $m/z$  (%): 149 (M<sup>+</sup>, 54), 134 (100), 117 (3), 104 (9), 77 (7).

### 11. 5-Methylindoline



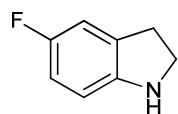
Light brown oil,  $R_f = 0.25$  (20 % EtOAc in hexane)

$^1\text{H NMR}$  (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 6.93 (s, 1H), 6.80 (d,  $J = 7.8$  Hz, 1H), 6.52 (d,  $J = 7.5$  Hz, 1H), 3.47 (t,  $J = 8.1$  Hz, 2H), 2.95 (t,  $J = 8.1$  Hz, 2H), 2.23 (s, 3H).

$^{13}\text{C NMR}$  (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 149.4, 129.9, 128.2, 127.7, 125.6, 109.6, 47.8, 30.2, 21.0.

$\text{MS-C}_9\text{H}_{11}\text{N}$  (133)  $m/z$  (%): 133 ( $\text{M}^+$ , 100), 117 (28), 103 (5), 77 (9), 65 (7).

### 12. 5-Fluoroindoline



Colorless oil,  $R_f = 0.25$  (20 % EtOAc in hexane)

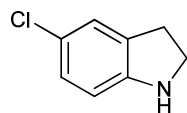
$^1\text{H NMR}$  (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 6.84 (dd,  $J = 8.7, 2.4$  Hz, 1H), 6.70 (dt,  $J = 9.0, 2.7$  Hz, 1H), 6.54 (dd,  $J = 8.1, 4.2$  Hz, 1H), 3.64 (bs, 1H), 3.55 (t,  $J = 8.4$  Hz, 2H), 3.01 (t,  $J = 8.4$  Hz, 2H).

$^{13}\text{C NMR}$  (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 158.5, 155.3, 147.4, 113.1, 112.8, 112.1, 111.8, 109.5, 109.4, 47.8, 30.1.

$^{19}\text{F NMR}$  (282.4 MHz,  $\text{CDCl}_3$ ),  $\delta$ (ppm)  $-126.4$

$\text{MS-C}_8\text{H}_8\text{FN}$  (137)  $m/z$  (%): 137 ( $\text{M}^+$ , 100), 116 (3), 109 (33), 83 (8), 67 (7).

### 13. 5-Chloroindoline



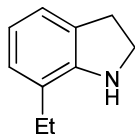
Light brown oil,  $R_f = 0.27$  (20 % EtOAc in hexane)

$^1\text{H NMR}$  (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 7.04 (s, 1H), 6.94 (d,  $J = 8.4$  Hz, 1H), 6.51 (d,  $J = 8.1$  Hz, 1H), 3.72 (bs, 1H), 3.54 (t,  $J = 8.4$  Hz, 2H), 2.99 (t,  $J = 8.1$  Hz, 2H).

$^{13}\text{C NMR}$  (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 150.1, 131.2, 126.8, 124.7, 122.9, 109.8, 47.5, 29.6.

$\text{MS-C}_8\text{H}_8\text{ClN}$  (153)  $m/z$  (%): 153 ( $\text{M}^+$ , 100), 125 (7), 117 (77), 89 (23), 58 (9).

#### 14. 7-Ethylindoline



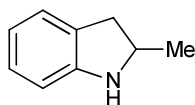
Brown oil,  $R_f = 0.33$  (20 % EtOAc in hexane)

$^1\text{H NMR}$  (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 7.90 (d,  $J = 7.2$  Hz, 1H), 6.98 (d,  $J = 7.2$  Hz, 1H), 6.79 (t,  $J = 7.5$  Hz, 1H), 3.63 (t,  $J = 8.1$  Hz, 2H), 3.55 (bs, 1H), 3.12 (t,  $J = 8.4$  Hz, 2H), 2.56 (q,  $J = 7.5$  Hz, 2H), 1.31 (t,  $J = 7.5$  Hz, 3H).

$^{13}\text{C NMR}$  (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 149.3, 128.7, 125.9, 124.8, 122.0, 118.8, 47.1, 29.9, 24.0, 13.1.

$\text{MS-C}_{10}\text{H}_{13}\text{N}$  (147)  $m/z$  (%): 147 ( $\text{M}^+$ , 63), 132 (100), 117 (24), 105 (7), 91 (6).

#### 15. 2-Methylindoline



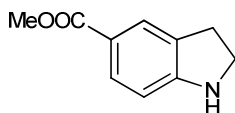
Light brown oil,  $R_f = 0.40$  (20 % EtOAc in hexane)

$^1\text{H NMR}$  (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 7.12 (d,  $J = 7.2$  Hz, 1H), 7.06 (t,  $J = 7.8$  Hz, 1H), 6.74 (t,  $J = 7.8$  Hz, 1H), 6.64 (d,  $J = 8.1$  Hz, 1H), 4.02 (m, 1H), 3.60 (bs, 1H), 3.18 (dd,  $J = 15.6, 8.4$  Hz, 1H), 2.67 (dd,  $J = 15.3, 7.8$  Hz, 1H), 1.32 (d,  $J = 6.0$  Hz, 3H).

$^{13}\text{C NMR}$  (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 151.1, 129.1, 127.4, 124.9, 118.7, 109.3, 55.4, 37.9, 22.4.

$\text{MS-C}_9\text{H}_{11}\text{N}$  (133)  $m/z$  (%): 133 ( $\text{M}^+$ , 33), 118 (100), 91 (17), 77 (4), 65 (3).

#### 16. Methyl indoline-5-carboxylate



Light orange solid,  $R_f = 0.20$  (20 % EtOAc in hexane)

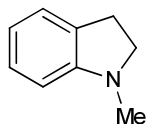
**M.P.:** 67-69 °C

$^1\text{H NMR}$  (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 7.76 (m, 2H), 6.55 (d,  $J = 8.4$  Hz, 1H), 3.84 (s, 3H), 3.65 (t,  $J = 8.4$  Hz, 2H), 3.07 (t,  $J = 8.4$  Hz, 2H).

$^{13}\text{C}$  NMR (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 199.6, 157.4, 130.8, 129.1, 126.3, 107.5, 100.2, 51.7, 47.4, 28.9.

MS- $\text{C}_{10}\text{H}_{11}\text{NO}_2$  (177)  $m/z$  (%): 177 ( $\text{M}^+$ , 82), 146 (100), 118 (27), 89 (12), 72 (6).

### 17. 1-methylindoline



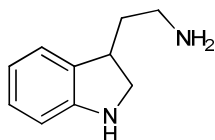
Brown oil,  $R_f = 0.25$  (10 % EtOAc in hexane)

$^1\text{H}$  NMR (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 7.07 (m, 2H), 6.66 (t,  $J = 7.5$  Hz, 1H), 6.48 (d,  $J = 8.1$  Hz, 1H), 3.27 (t,  $J = 8.4$  Hz, 2H), 2.92 (t,  $J = 8.4$  Hz, 2H), 2.74 (s, 3H).

$^{13}\text{C}$  NMR (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 153.3, 130.2, 127.2, 124.1, 117.6, 107.1, 56.0, 36.2, 28.6.

MS- $\text{C}_9\text{H}_{11}\text{N}$  (133)  $m/z$  (%): 133 ( $\text{M}^+$ , 100), 117 (41), 103 (4), 91 (10), 77 (6).

### 18. 2-(Indolin-3-yl)ethanamine



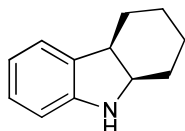
Light brown oil,  $R_f = 0.50$  (5 % MeOH in DCM)

$^1\text{H}$  NMR (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 7.08 (d,  $J = 7.2$  Hz, 1H), 7.03 (t,  $J = 7.5$  Hz, 1H), 6.72 (t,  $J = 7.2$  Hz, 1H), 6.64 (d,  $J = 7.8$  Hz, 1H), 3.67 (t,  $J = 8.7$  Hz, 1H), 3.32 (m, 1H), 3.20 (t,  $J = 8.4$  Hz, 1H), 2.80 (bs, 2H), 1.96 (m, 2H), 1.70 (m, 2H).

$^{13}\text{C}$  NMR (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 151.4, 132.8, 127.6, 123.9, 118.8, 109.7, 53.6, 40.4, 39.9, 38.4.

MS- $\text{C}_{10}\text{H}_{14}\text{N}_2$  (162)  $m/z$  (%): 162 ( $\text{M}^+$ , 30), 145 (19), 130 (50), 117 (100), 91 (20).

### 19. ( $\pm$ )-cis-2,3,4,4a,9,9a-hexahydro-1H-carbazole



White solid,  $R_f = 0.38$  (20 % EtOAc in hexane)

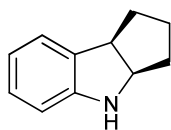
**M.P.** : 93-94°C

**$^1\text{H}$  NMR** (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 7.09 (d,  $J = 7.2$  Hz, 1H), 7.03 (t,  $J = 7.8$  Hz, 1H), 6.74 (t,  $J = 7.2$  Hz, 1H), 6.68 (d,  $J = 7.8$  Hz, 1H), 3.73 (q,  $J = 6.6$  Hz, 1H), 3.65 (bs, 1H), 3.10 (q,  $J = 6.6$  Hz, 1H), 1.76 (m, 2H), 1.56 (m, 3H), 1.39 (m, 3H).

**$^{13}\text{C}$  NMR** (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 150.7, 133.4, 126.9, 123.1, 118.7, 110.1, 59.6, 40.8, 29.1, 26.9, 22.4, 21.6.

**MS**- $\text{C}_{12}\text{H}_{15}\text{N}$  (173)  $m/z$  (%): 173 ( $\text{M}^+$ , 30), 144 (9), 130 (100), 117 (14), 90 (5).

## 20. ( $\pm$ )-*cis*-1,2,3,3a,4,8b-hexahydrocyclopenta[*b*]indole



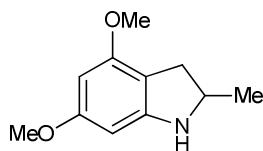
Colorless oil,  $R_f = 0.25$  (10 % EtOAc in hexane)

**$^1\text{H}$  NMR** (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 7.03 (d,  $J = 7.2$  Hz, 1H), 6.97 (t,  $J = 7.8$  Hz, 1H), 6.66 (dt,  $J = 7.5, 1.2$  Hz, 1H), 6.51 (d,  $J = 8.1$  Hz, 1H), 4.34 (m, 1H), 3.76 (dt,  $J = 6.6, 2.1$  Hz, 1H), 1.94 (m, 2H), 1.70 (m, 4H).

**$^{13}\text{C}$  NMR** (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 151.3, 133.3, 127.2, 124.4, 118.2, 108.3, 63.2, 47.1, 36.8, 34.8, 24.3.

**MS**- $\text{C}_{11}\text{H}_{13}\text{N}$  (159)  $m/z$  (%): 159 ( $\text{M}^+$ , 28), 130 (100), 117 (10), 89 (5), 77 (4).

## 21. 4,6-dimethoxy-2-methylindoline



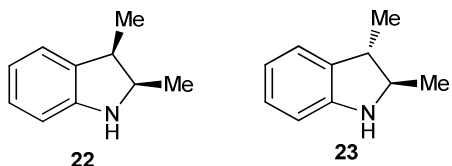
Light brown oil,  $R_f = 0.25$  (20 % EtOAc in hexane)

**$^1\text{H}$  NMR** (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 5.90 (m, 2H), 3.77 (s, 3H), 3.75 (s, 3H), 3.38 (m, 1H), 3.14 (dd,  $J = 8.7, 5.1$ , 1H), 1.26 (d,  $J = 6.6$  Hz, 3H).

**$^{13}\text{C}$  NMR** (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 161.2, 157.0, 153.1, 112.8, 89.3, 88.6, 55.5, 55.3, 55.0, 34.5, 19.0.

**MS**- $\text{C}_{11}\text{H}_{15}\text{NO}_2$  (193)  $m/z$  (%): 193 ( $\text{M}^+$ , 38), 178 (100), 163 (18), 147 (20), 132 (7).

**22, 23.** (±)-2,3-dimethylindoline



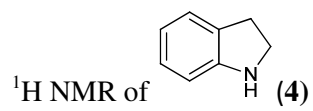
22:23 = 6:1

Brown oil,  $R_f = 0.40$  (20 % EtOAc in hexane)

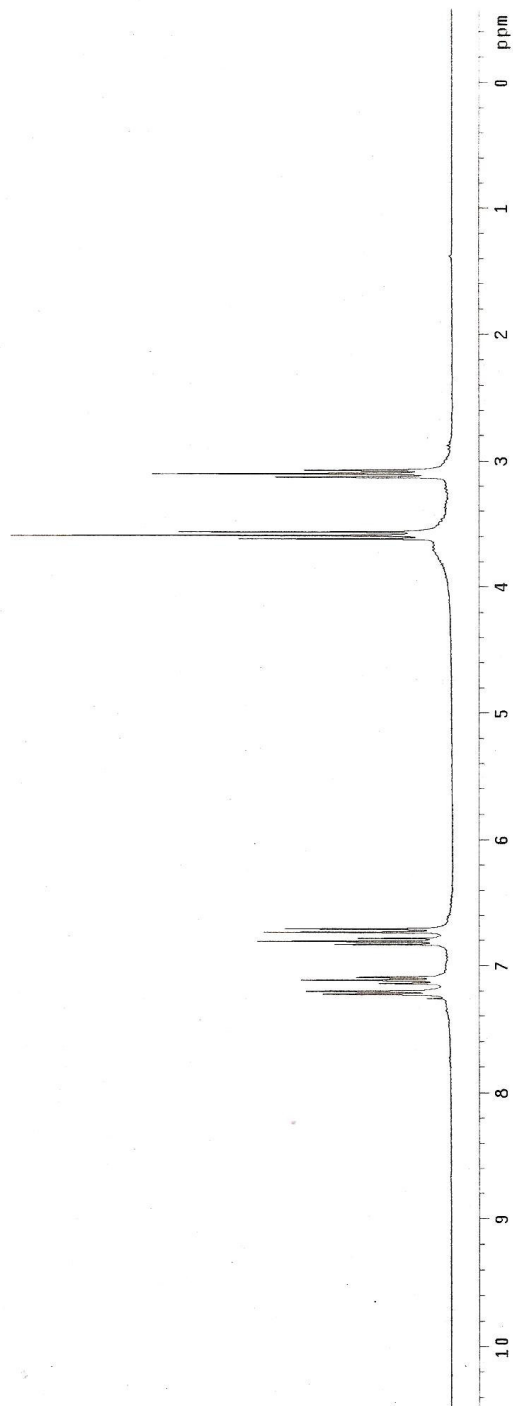
**$^1\text{H}$  NMR** (300.128 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 7.12 (d,  $J = 7.5$  Hz, 1H), 7.06 (d,  $J = 7.8$  Hz, 1H), 6.78 (t,  $J = 7.5$  Hz, 1H), 6.65 (d,  $J = 7.8$  Hz, 1H), 3.97 (m, 1H), 3.66 (bs, 1H), 3.50 (m, 1H, minor isomer), 3.31 (quin,  $J = 7.5$  Hz, 1H), 2.86 (quin,  $J = 6.9$  Hz, minor isomer), 1.36 (d,  $J = 2.1$  Hz, 3H, minor isomer), 1.34 (d,  $J = 3.0$  Hz, 3H, minor isomer), 1.22 (d,  $J = 7.2$  Hz, 3H), 1.17 (d,  $J = 6.6$  Hz, 1H).

**$^{13}\text{C}$  NMR** (75.474 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm) 149.9, 134.1, 127.1, 127.1 (minor isomer), 123.6, 123.1 (minor isomer), 118.5, 118.4 (minor isomer), 109.2, 109.0 (minor isomer), 63.7 (minor isomer), 58.2, 44.1, (minor isomer), 39.3, 20.3 (minor isomer), 17.0 (minor isomer), 16.1, 13.5.

**MS**- $\text{C}_{10}\text{H}_{13}\text{N}$  (147)  $m/z$  (%): 147 ( $\text{M}^+$ , 38), 132 (100), 117 (39), 103 (4), 77 (8).

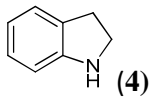


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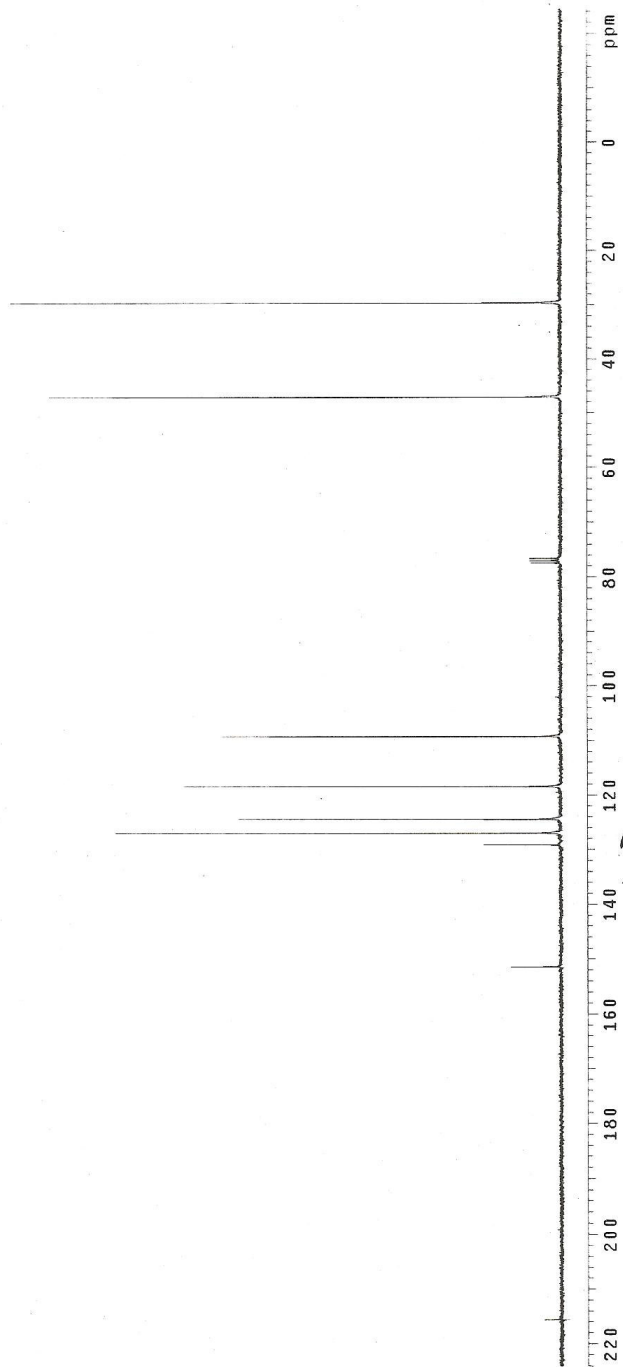




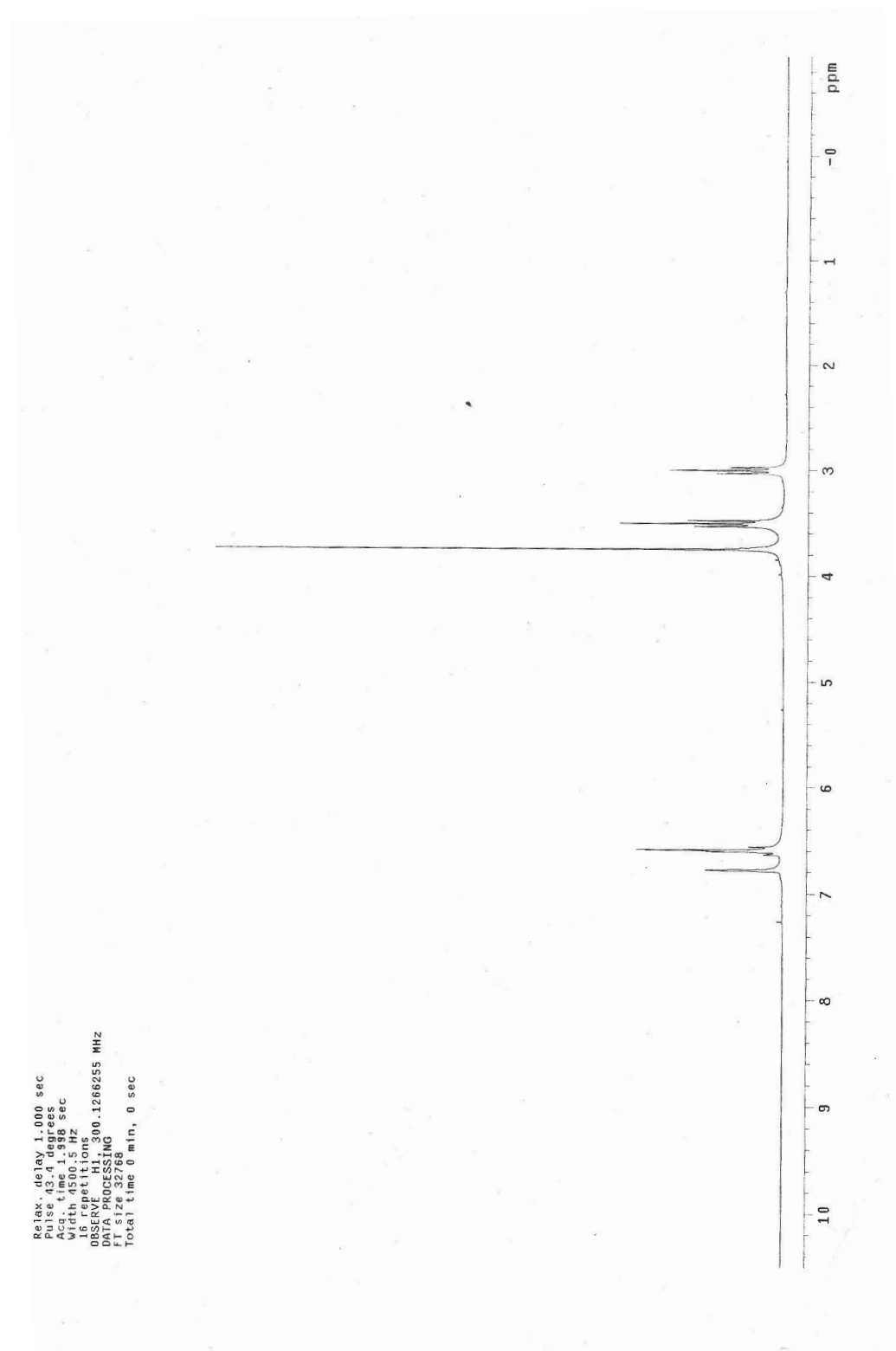
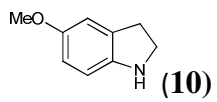
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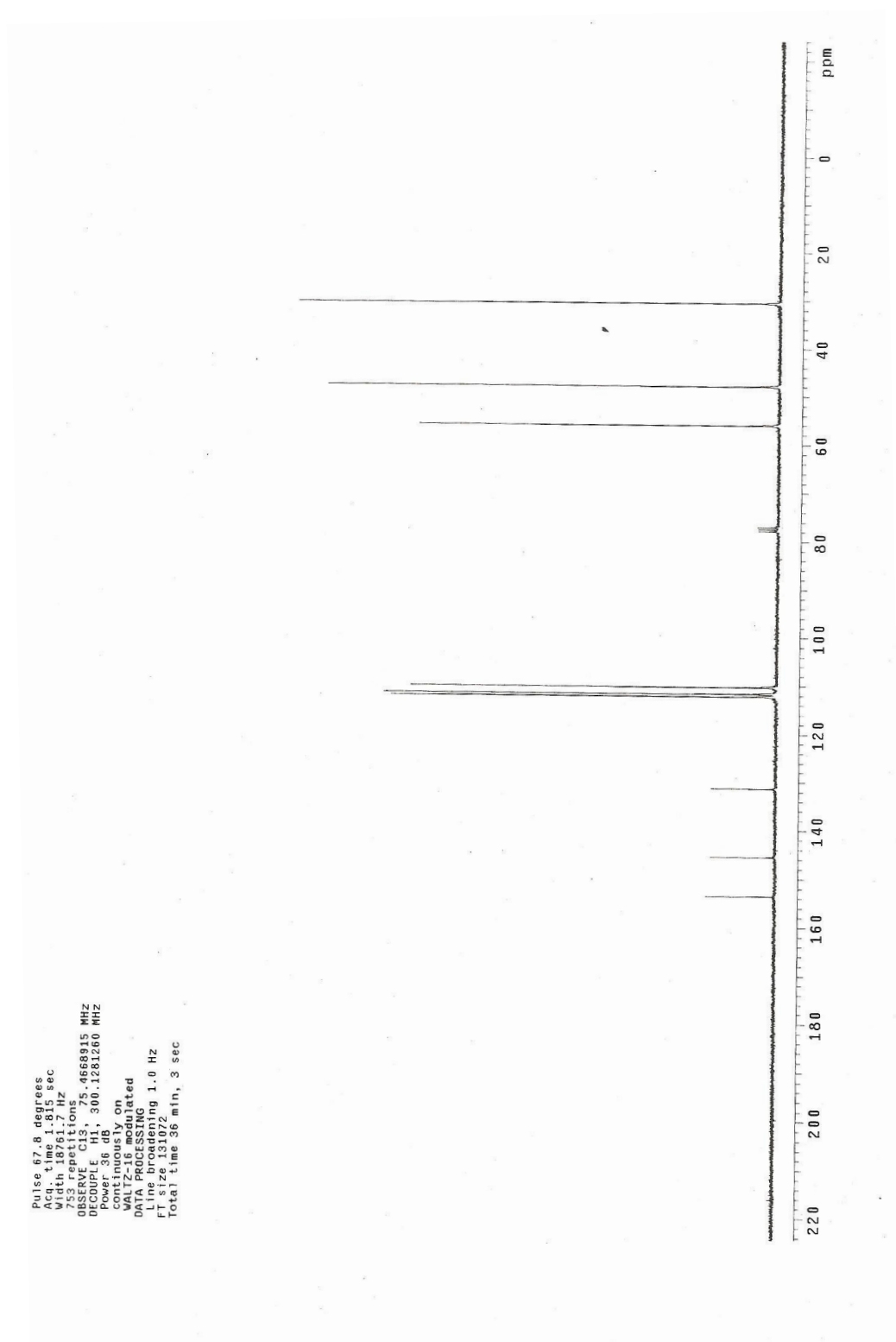
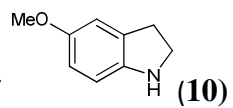
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continuously on  
with decoupling  
Data Processing  
Line broadening 1.0 Hz  
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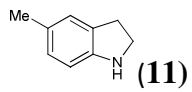
<sup>1</sup>H NMR of



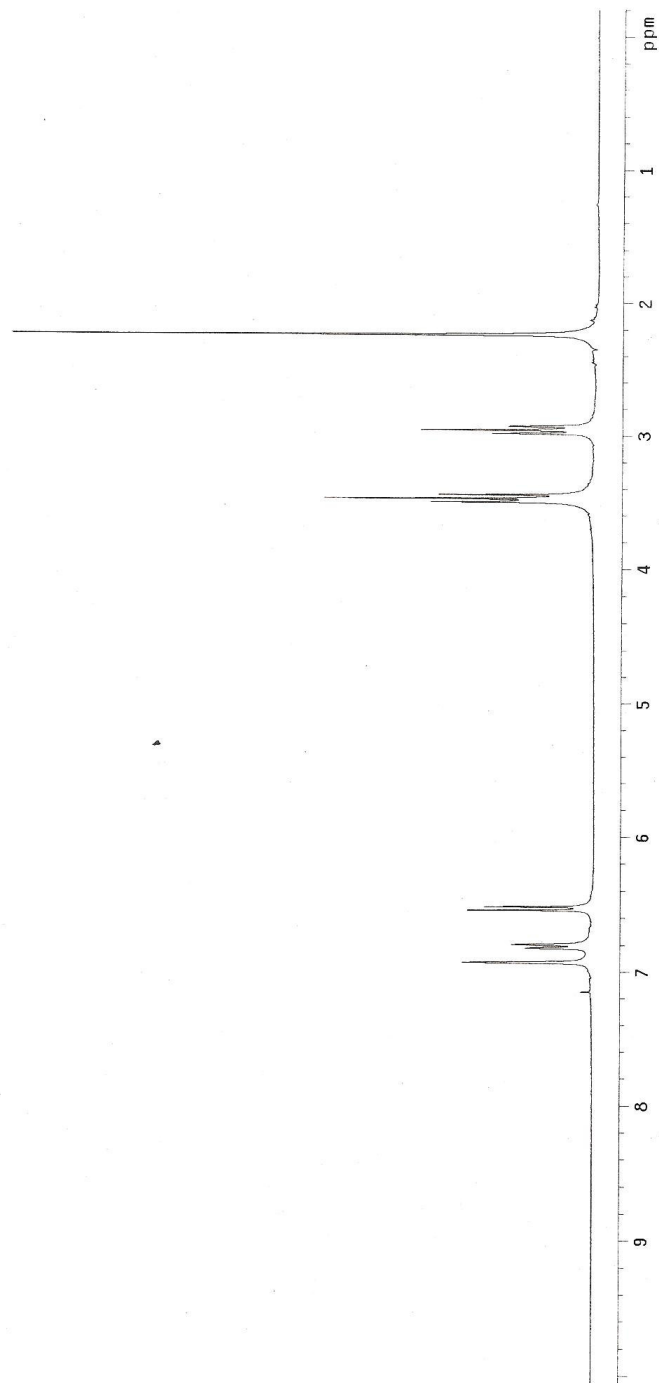
<sup>13</sup>C NMR of



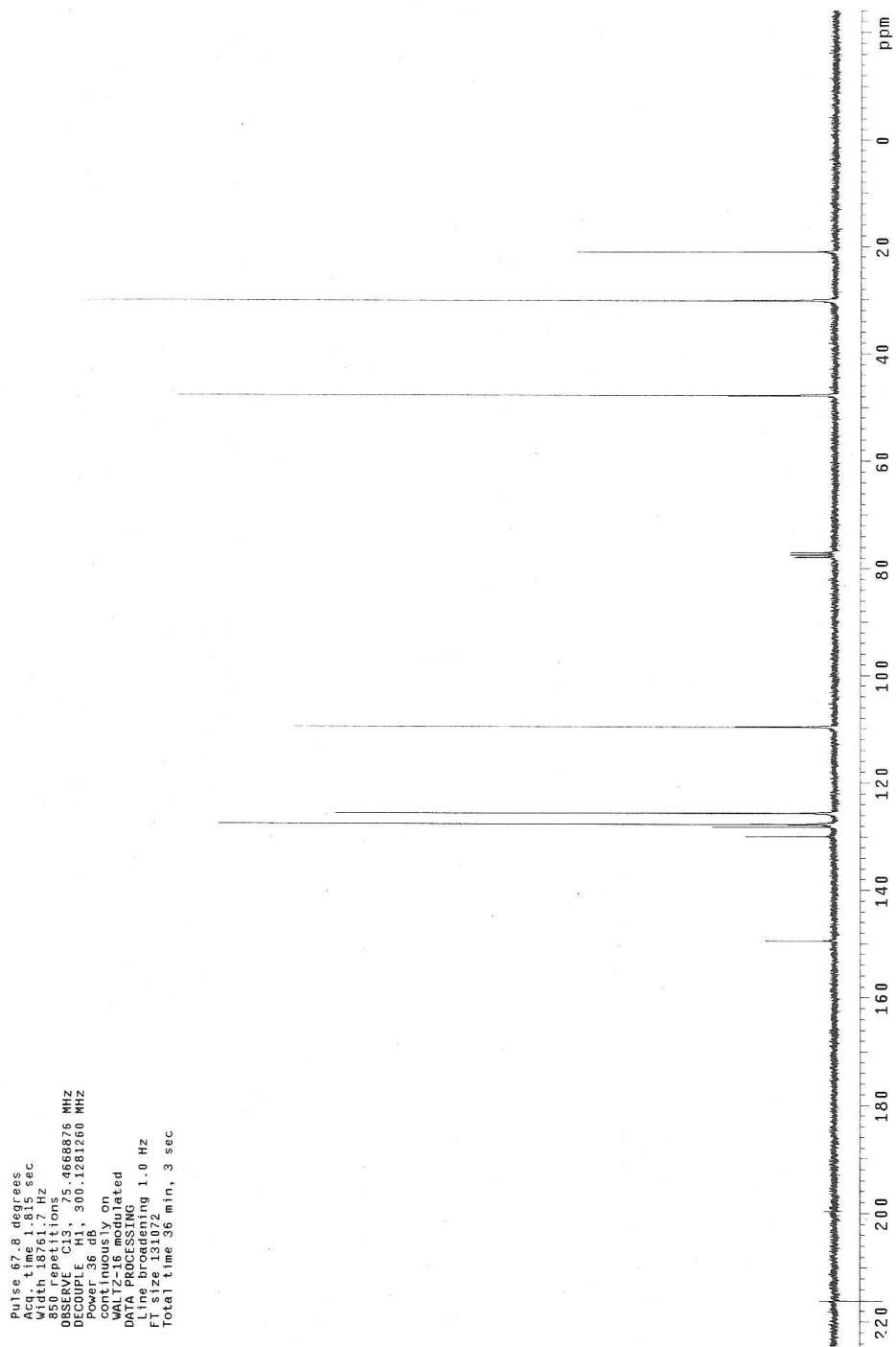
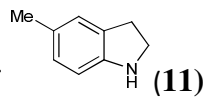
<sup>1</sup>H NMR of



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Total time 0 min, 49 sec

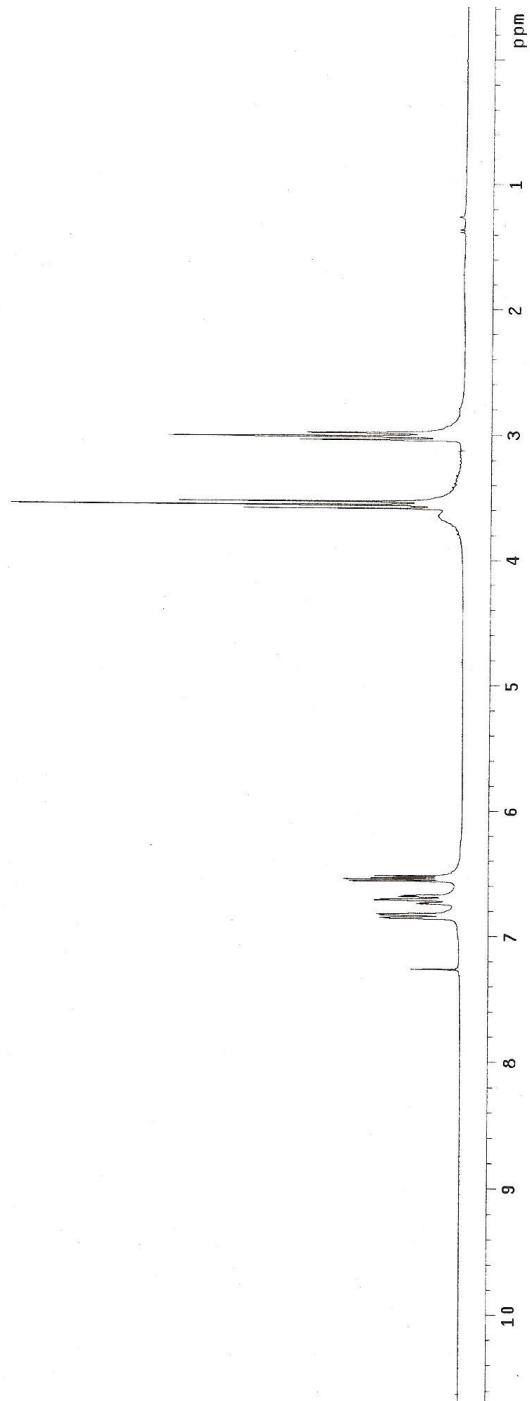


<sup>13</sup>C NMR of



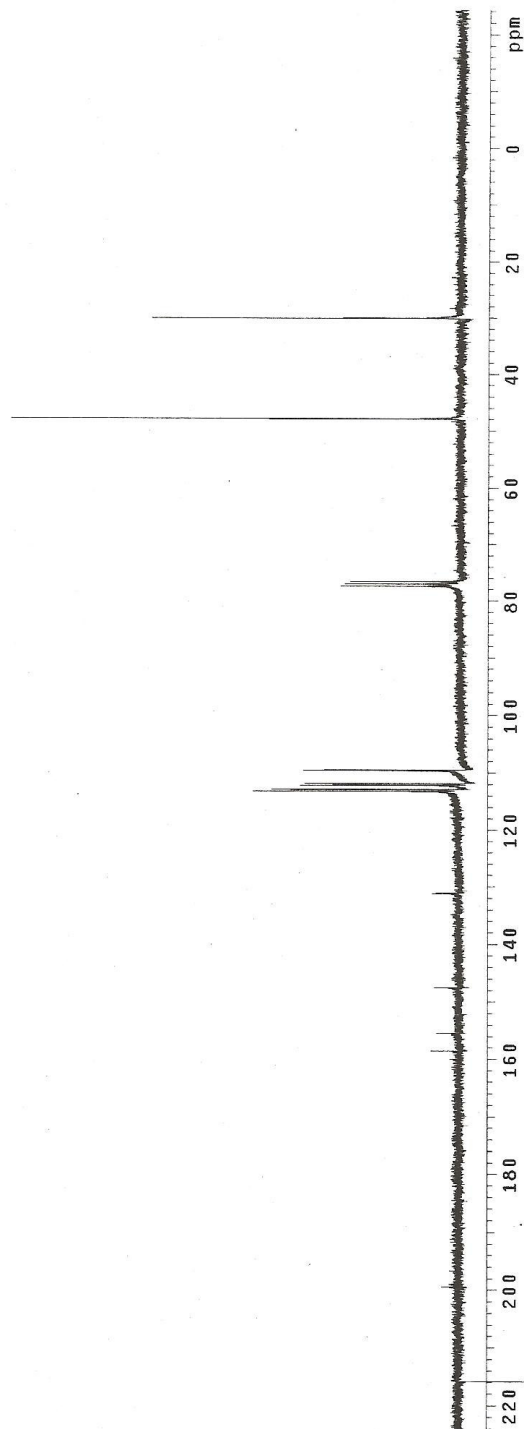
<sup>1</sup>H NMR of Fc1ccc2c(c1)CCN2 (12)

Relax. delay 1.000 sec  
Pulse prog zgpg30  
Acq time 1.186 sec  
Width 4500.5 Hz  
32 repetitions  
OBSERVE H1, 300.1266255 MHz  
F1 127.80  
DATA PROCESSING  
Total time 0 min, 0 sec

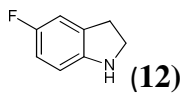


<sup>13</sup>C NMR of Fc1ccc2c(c1)ncn2 (12)

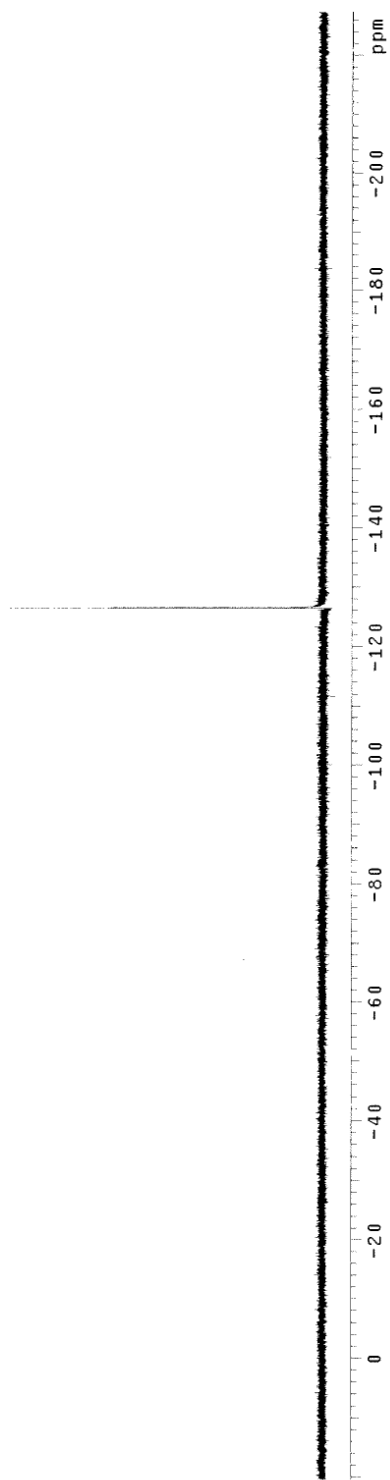
Pulse 67.8 degrees  
Time 1.91 sec  
Width 18761.5 Hz  
1024 repetitions  
OBSERVE C13, 75.4669044 MHz  
DECOUPLE H1, 300.1281260 MHz  
Power 36 dB  
SOLVENT CDCl3  
WALTZ16 selected  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 131072  
Total time 36 min, 3 sec



<sup>19</sup>F NMR of



Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient temperature  
GEMINI-300BB "Mighty300"  
Relax. delay 4.000 sec  
Pulse 5.0 usec  
Acq. time 0.300 sec  
Width 7000 Hz  
16 repetitions  
OBSERVE F19, 282.4011453 MHZ  
DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 65536  
Total time 1 min, 11 sec

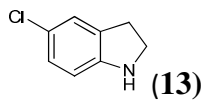




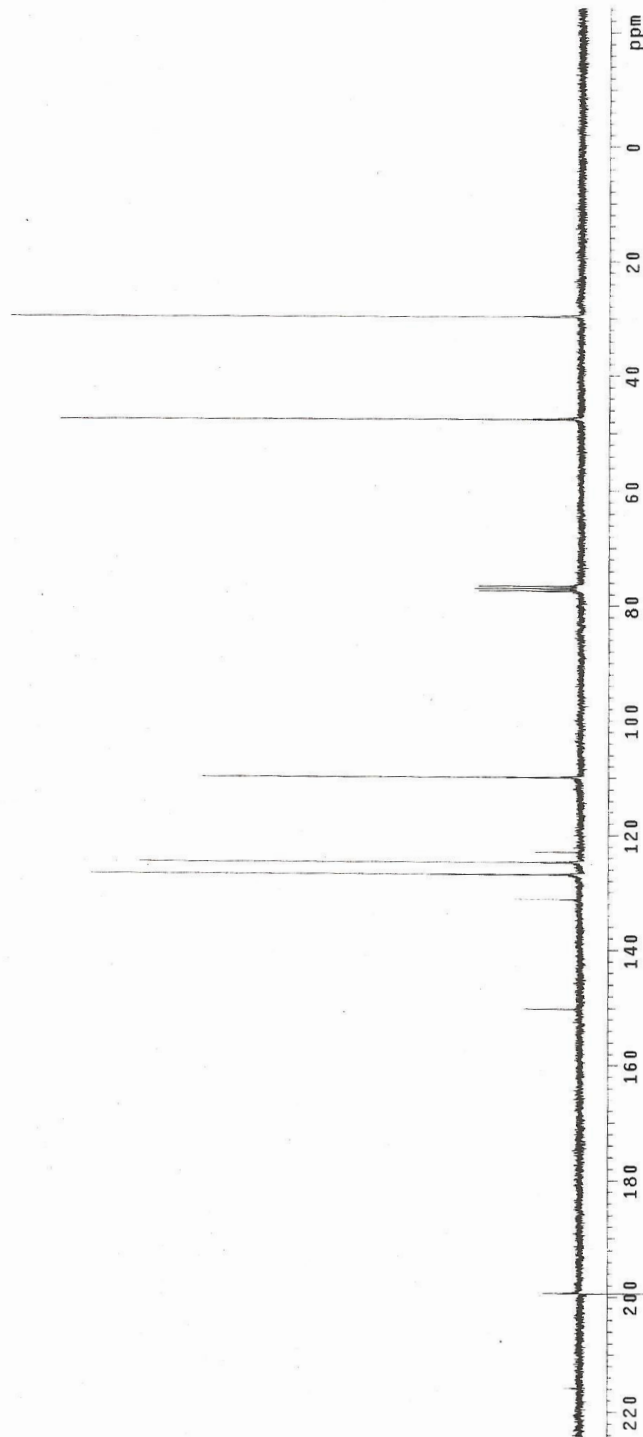
<sup>1</sup>H NMR of Clc1ccc2c(c1)ncn2 (13)

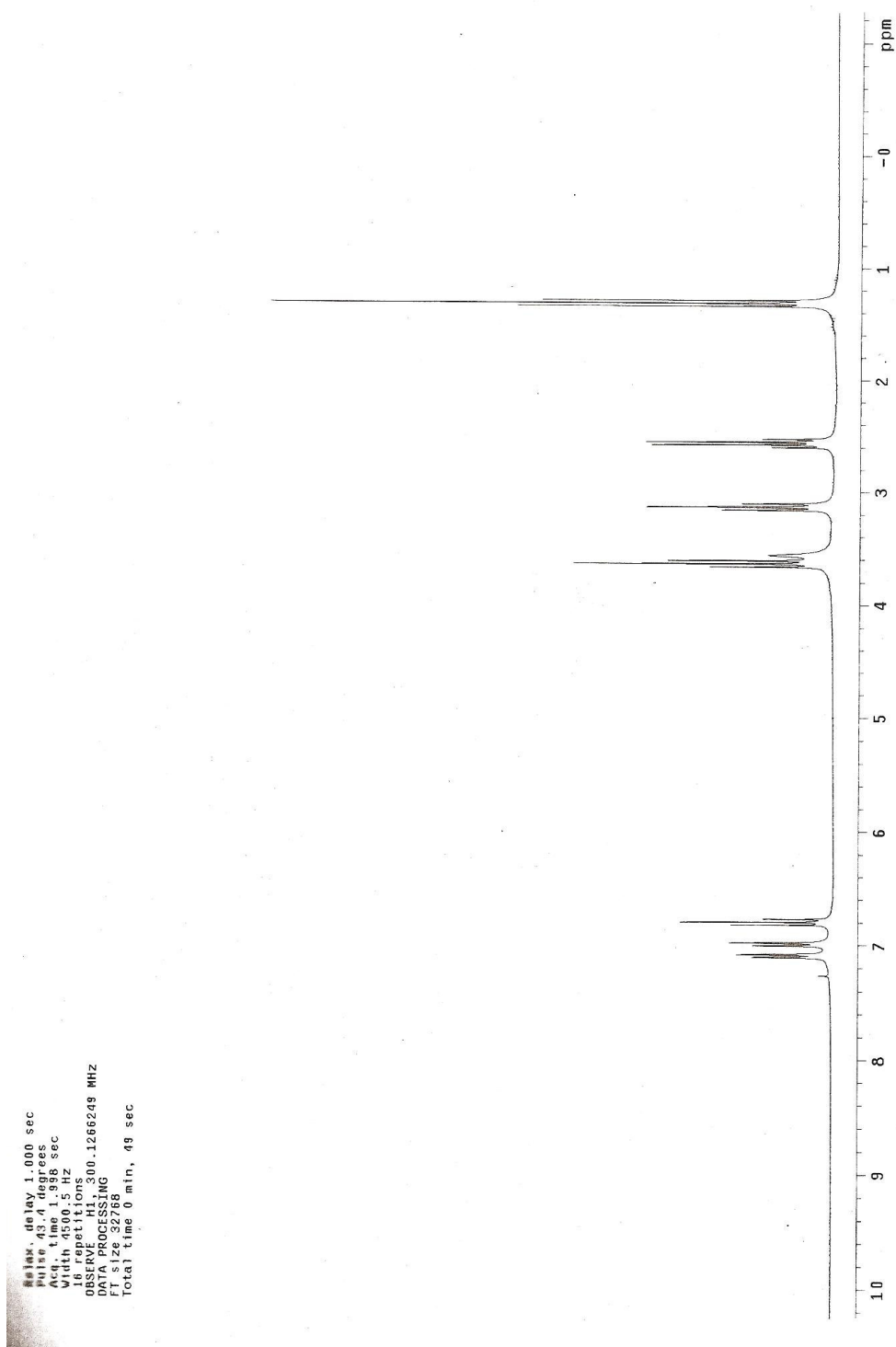
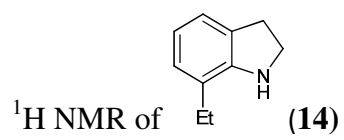


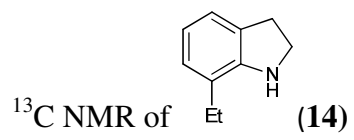
<sup>13</sup>C NMR of



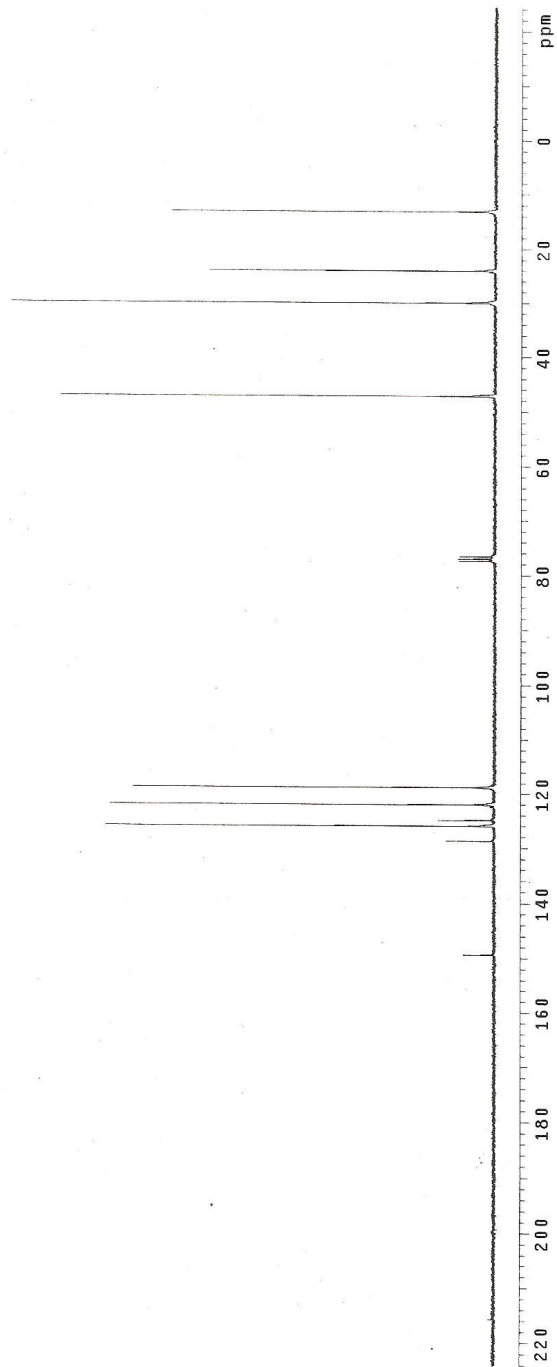
Pulse 67.8 degrees  
Acq. time 1.815 sec  
Width 18761.7 Hz  
861 repetitions  
OBSERVE C13, 75.4669061 MHz  
PULSE P1, 300.1281260 MHz  
P2, 36  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 131072  
Total time 36 min, 3 sec

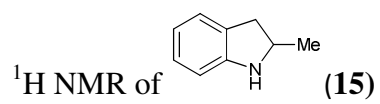




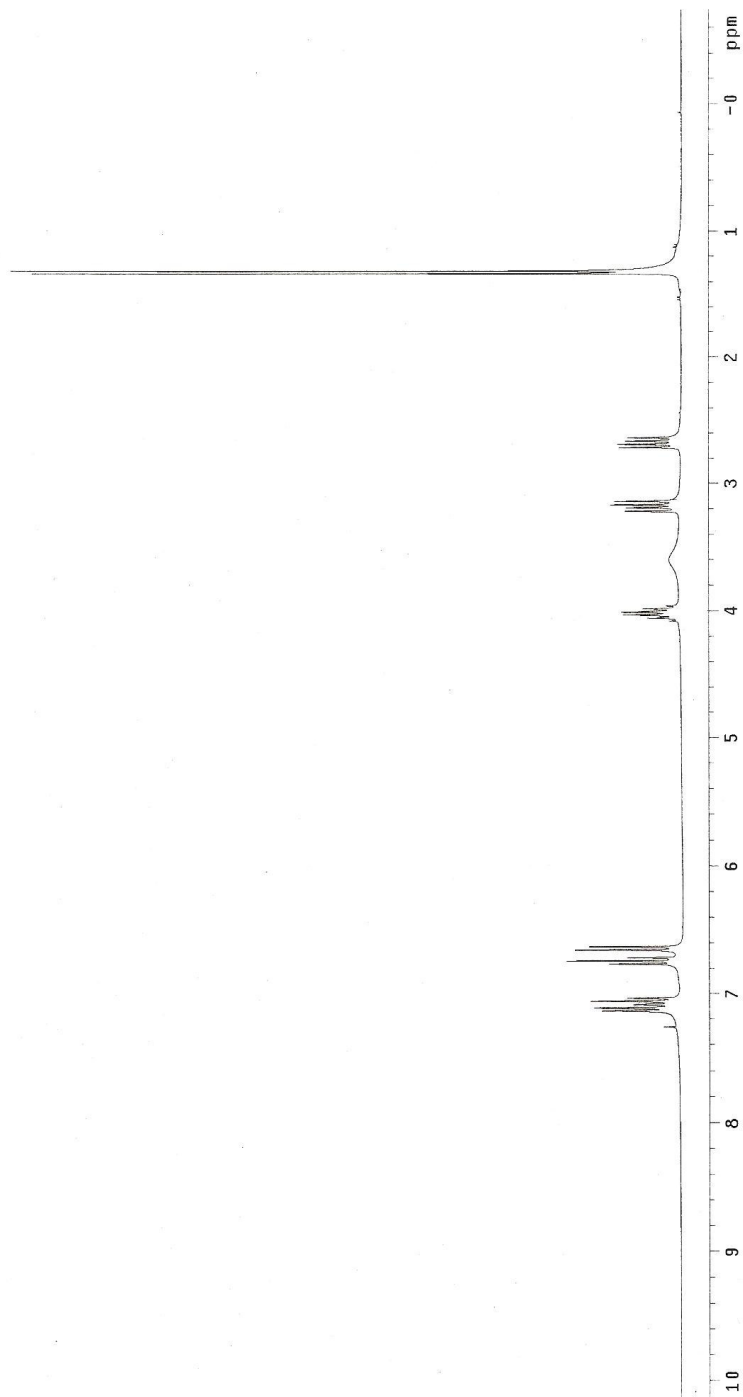


Pulse 67.8 degrees  
Acq. time 1.815 sec  
Width 18761.7 Hz  
Observed repetitions  
DECOUPLE CH1, 300.1281280 MHz  
Power 36 dB, continuously on  
WALTZ-16 modulated  
D1 1.00000000 sec  
Line broadening 1.0 Hz  
FT size 131072  
Total time 36 min, 3 sec

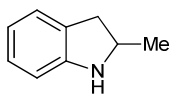




Relax. delay 1.000 sec  
Pulse prog zgpg30  
Acq time 1.998 sec  
Width 4500.5 Hz  
16 repetitions  
OBSERVE H1, 300.1266257 MHz  
DATA PROCESSING  
P1 1.20 sec  
Total time 0 min, 0 sec

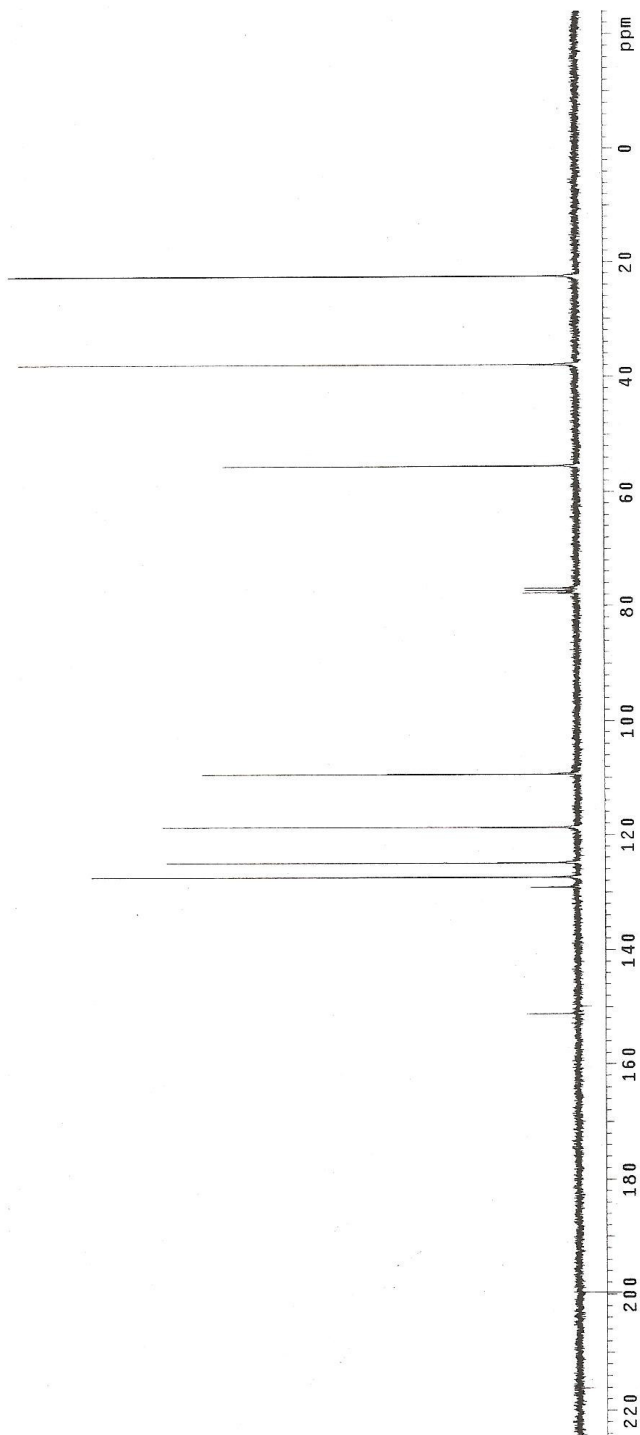


<sup>13</sup>C NMR of

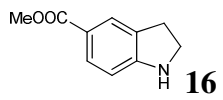


(15)

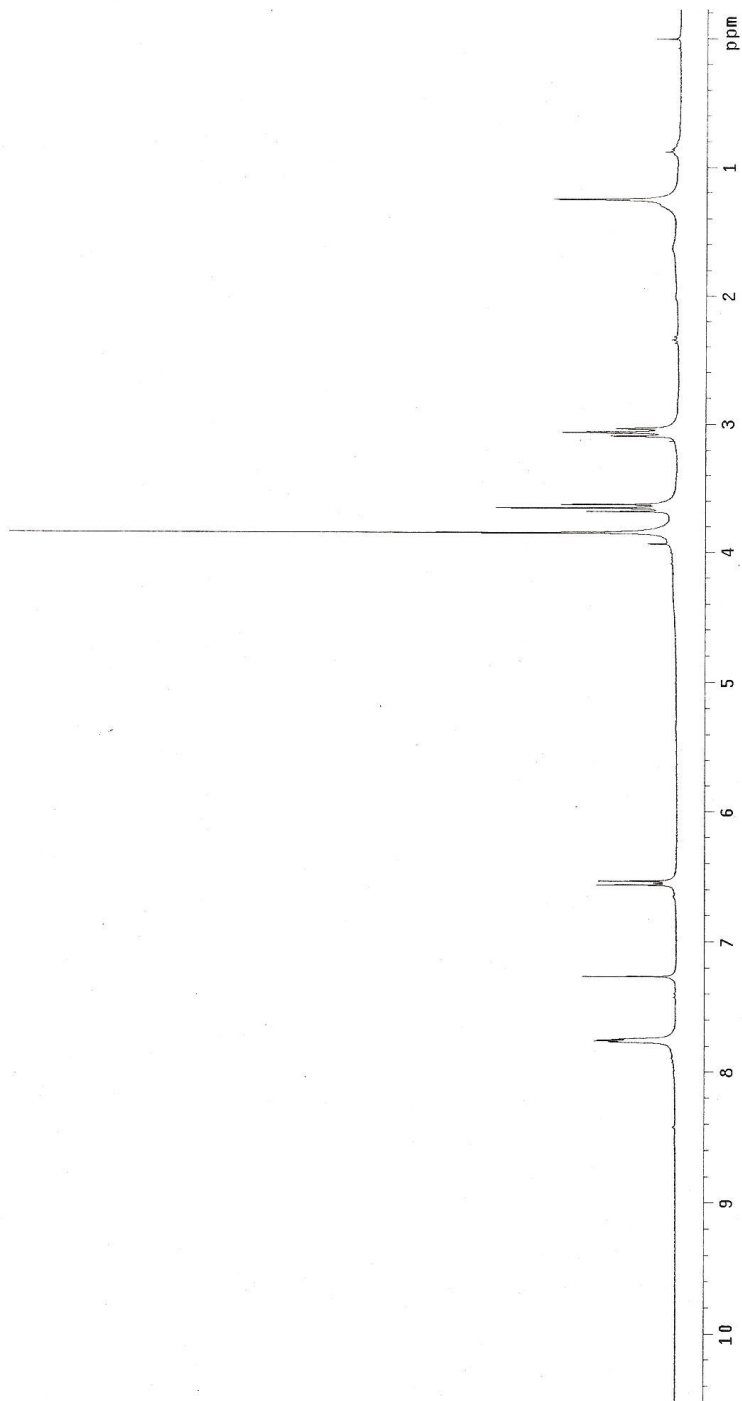
Pulse 67.8 degrees  
Acq time 1.815  
Width 18761.7 Hz  
733 repetitions  
OBSERVE C13, 75.4668876 MHz  
DECOUPLE H1, 300.1281260 MHz  
Power 36.000 W  
Locking On  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 131072  
Total time 36 min, 3 sec



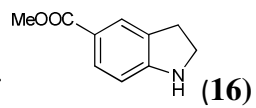
<sup>1</sup>H NMR of



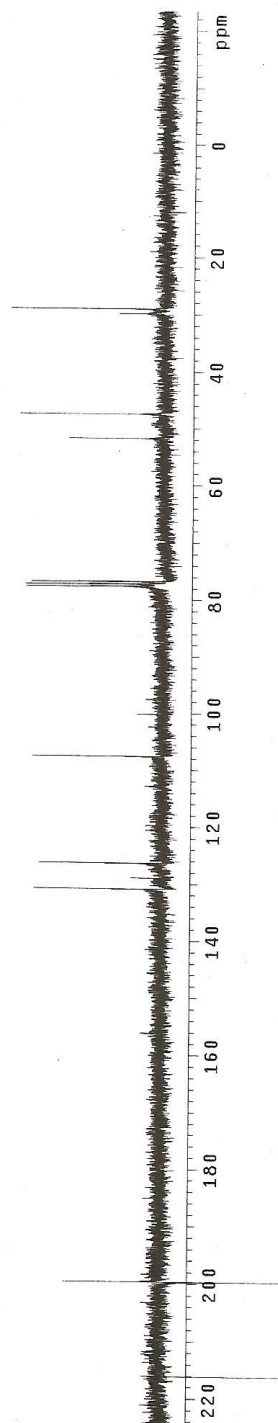
Relax. delay 1.000 sec  
Pulse 43.4 degrees  
Acq. time 1.998 sec  
Width 4500.5 Hz  
16 Repetitions  
05/16/05 300.1266241 MHz  
QNP-1H  
PCP00551MG  
FT size 32768  
Total time 0 min, 0 sec



<sup>13</sup>C NMR of

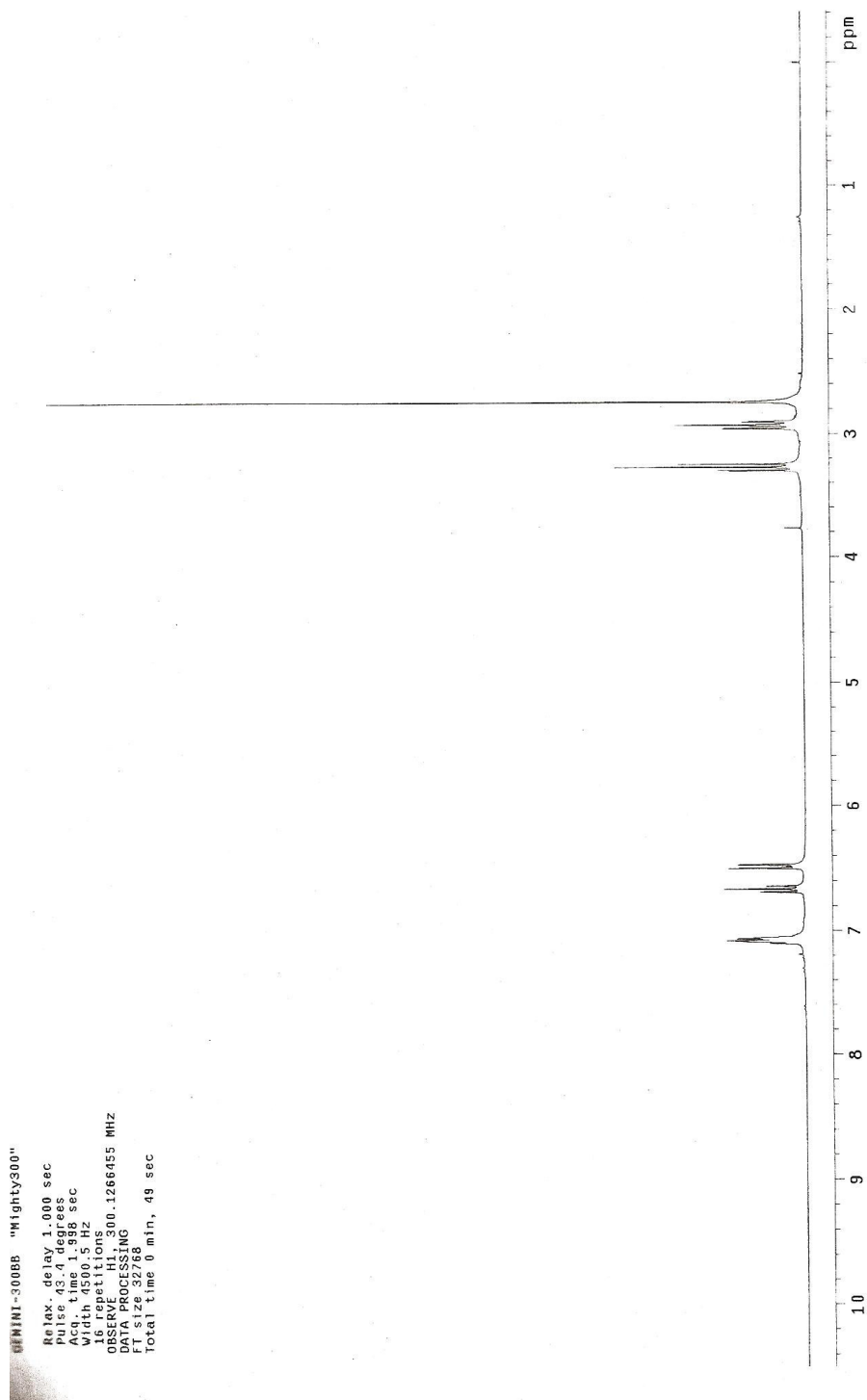
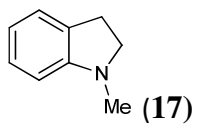


Pulse 67.8 degrees  
Acq. time 1.815 sec  
Date\_ 10/01/77  
1024 F<sub>2</sub> 177.7 Hz  
OBSERVE C13, 75.4668876 MHz  
DECOUPLE H1, 300.1281260 MHz  
Power 36 dB  
continuously on  
with F2C modulated  
DATA F2C modulated  
SOLVENT CDCl<sub>3</sub>  
Line broadening 1.0 Hz  
FT size 131072  
Total time 36 min, 3 sec

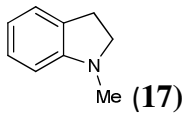




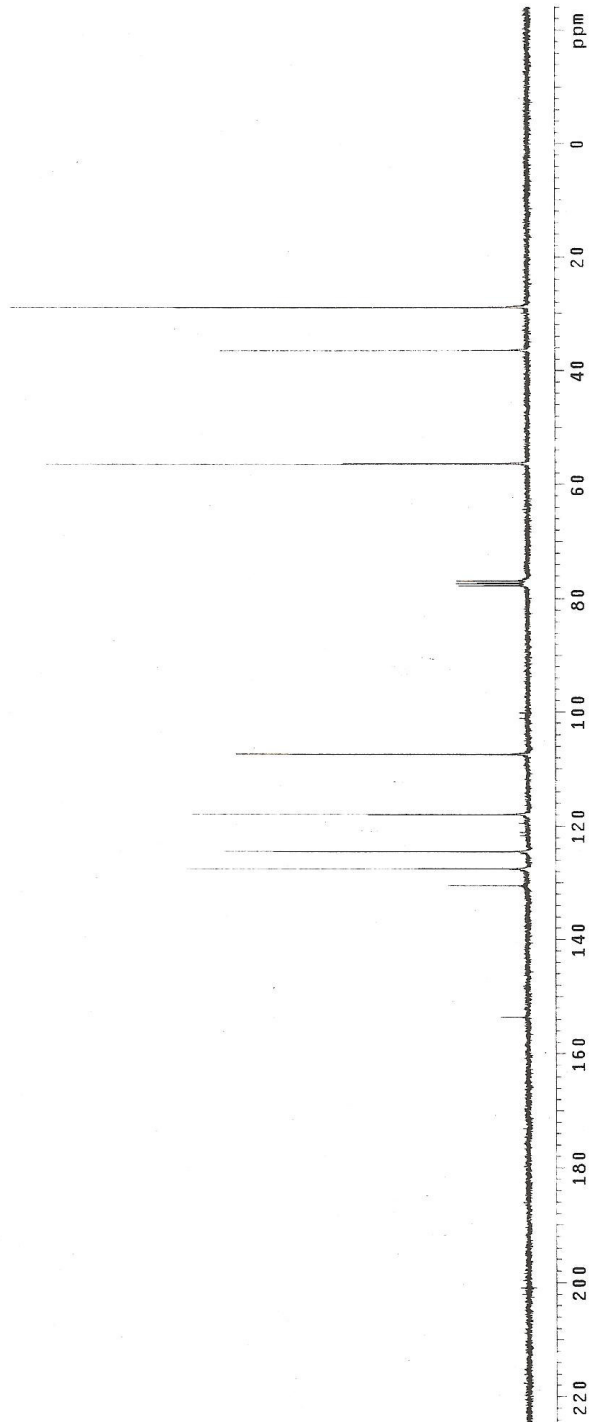
<sup>1</sup>H NMR of



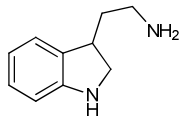
<sup>13</sup>C NMR of



Pulse 67.8 degrees  
Acq. time 1.11 sec  
Width 18761.7 Hz  
1024 repetitions  
OBSERVE C13, 75.4668876 MHz  
DECUPLE H1, 300.1281260 MHz  
Power 36 dB  
C13 pulse on  
C13 frequency 75.4668876 MHz  
WALTZ16 pulsed  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 131072  
Total time 36 min, 3 sec

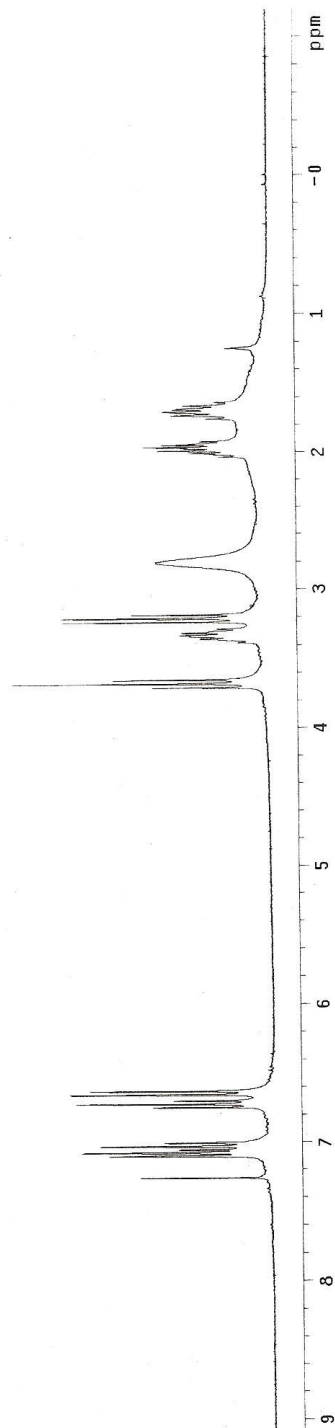


<sup>1</sup>H NMR of

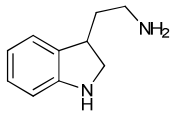


(18)

Relax. delay 1.000 sec  
Pulse 43.4 degrees  
Acq. time 1.988 sec  
Width 4500.5 Hz  
16 repetitions  
AQ 0.1266255 MHz  
DATA PROCESSING  
FT size 32768  
Total time 0 min, 49 sec

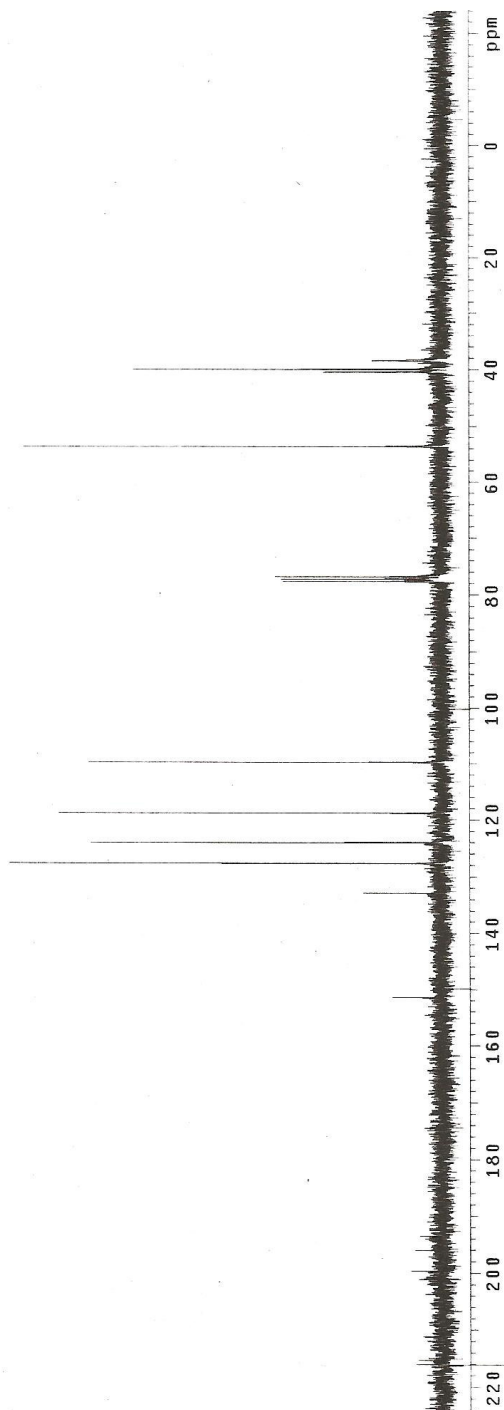


<sup>13</sup>C NMR of

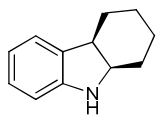


(18)

Pulse 67.8 degrees  
Acq. time 1.15 sec  
Width 18761.7 Hz  
1024 repetitions  
OBSERVE C13, 75.4666876 MHz  
DECOUPLE H1, 300.1281260 MHz  
Power 36 dB  
Coupling on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 size 131072  
Total time 36 min, 3 sec

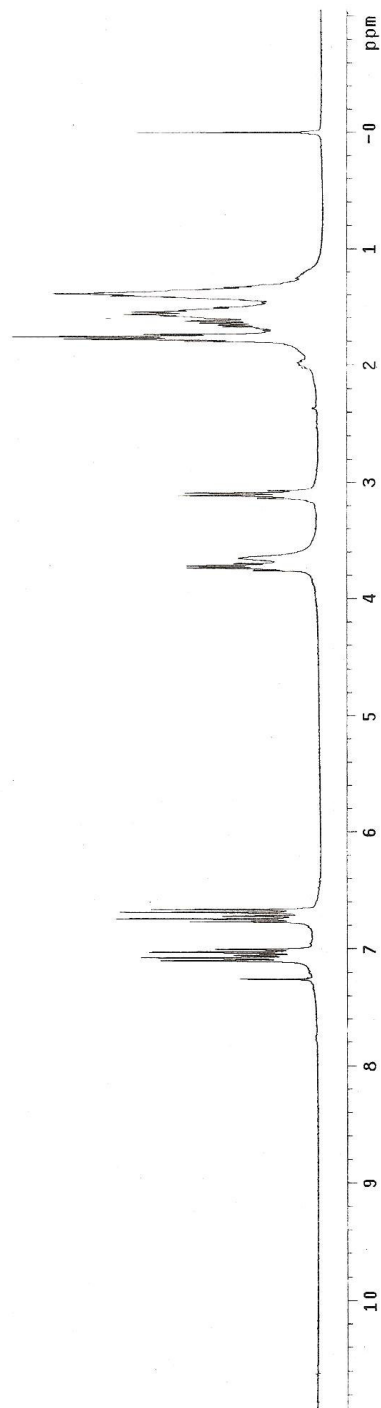


<sup>1</sup>H NMR of

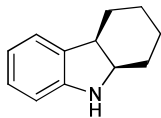


(19)

Relax. delay 1.000 sec  
Pulse 43.4 degrees  
Acq. time 1.988 sec  
Width 4500.5 Hz  
64 repetitions  
SFO 400.126250 MHz  
DATA PROCESSING  
FT size 32768  
Total time 3 min, 19 sec

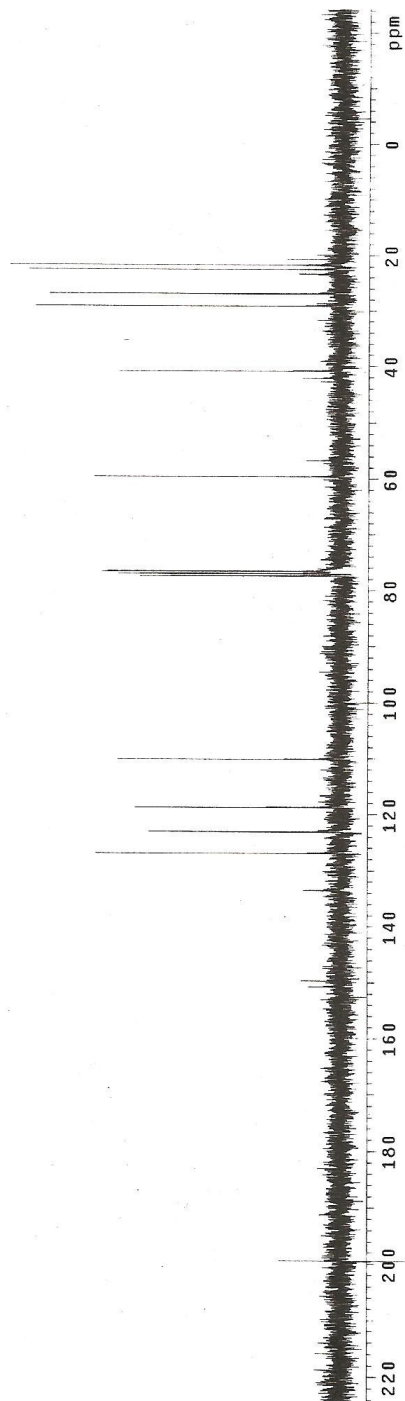


<sup>13</sup>C NMR of

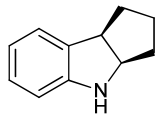


(19)

Pulse 67.8 degrees  
Acq. time 1.815 sec  
Width 18761.7 Hz  
F2 101.253 MHz  
REPEATS 4559018  
DECOUPLE H1, 300.1281260 MHz  
Power 36 dB  
continuously on  
WALTZ-16 modulated  
LINE SCANNING  
Line width 1.0 Hz  
FI size 131072  
Total time 36 min, 3 sec

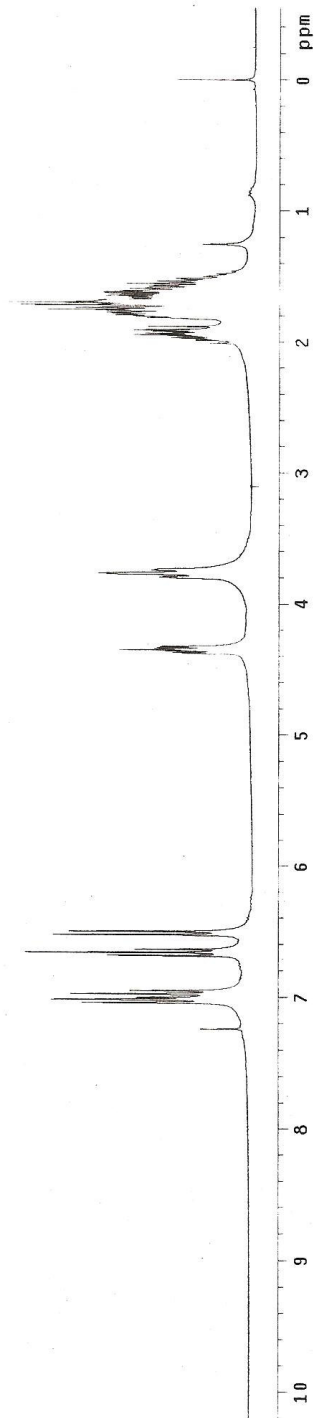


<sup>1</sup>H NMR of

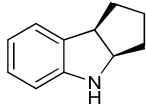


20

Relax. delay 1.000 sec  
Pulse 43.4 degrees  
Acq. time 1.398 sec  
Width 60.00 Hz  
32 repetitions  
OBSERVE H1, 300.1266318 MHz  
DATA PROCESSING  
FT size 32768  
Total time 1 min, 39 sec

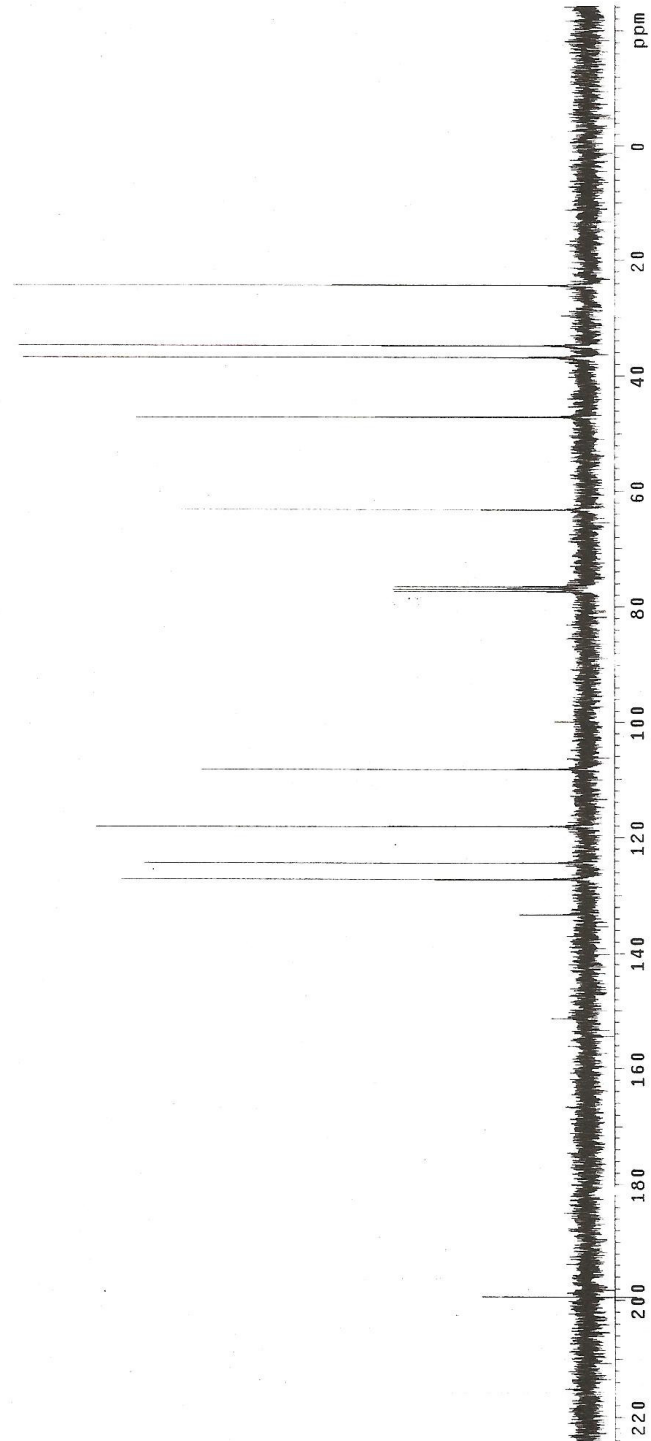


<sup>13</sup>C NMR of

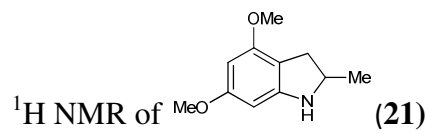


(20)

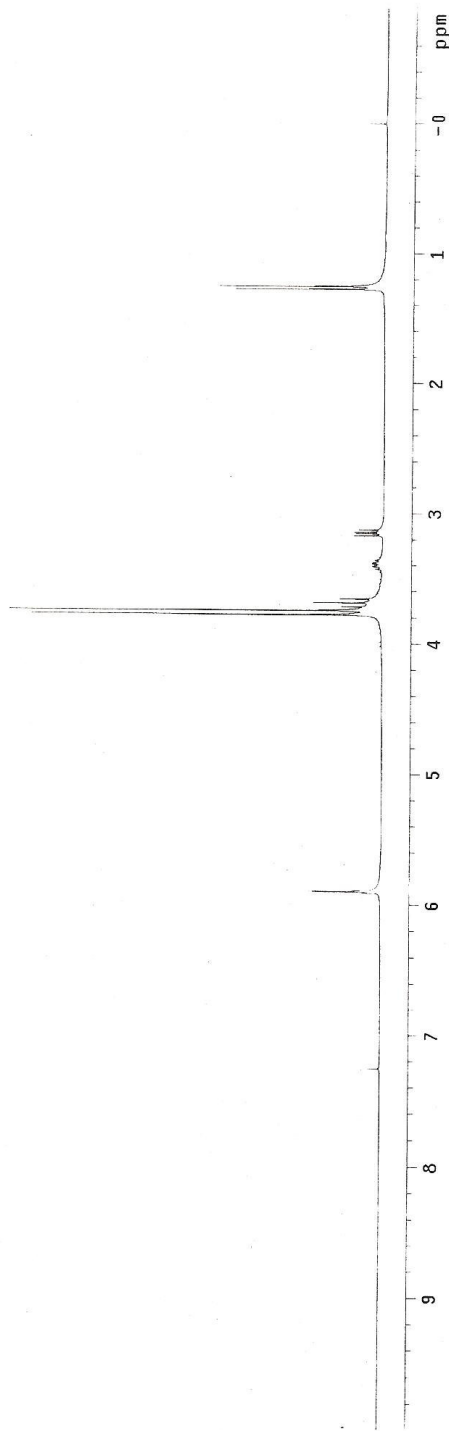
Pulse 67.8 degrees  
Acq. time 1.915 sec  
F1 125.761 MHz  
1024 repetitions  
OBSERVE C13, 75.4669044 MHz  
DECOUPLE H1, 300.1281260 MHz  
Power 36 dB  
continuously on  
WALTZ16 modulated  
NUC1  
Line broadening 1.0 Hz  
FT size 131072  
Total time 36 min, 3 sec

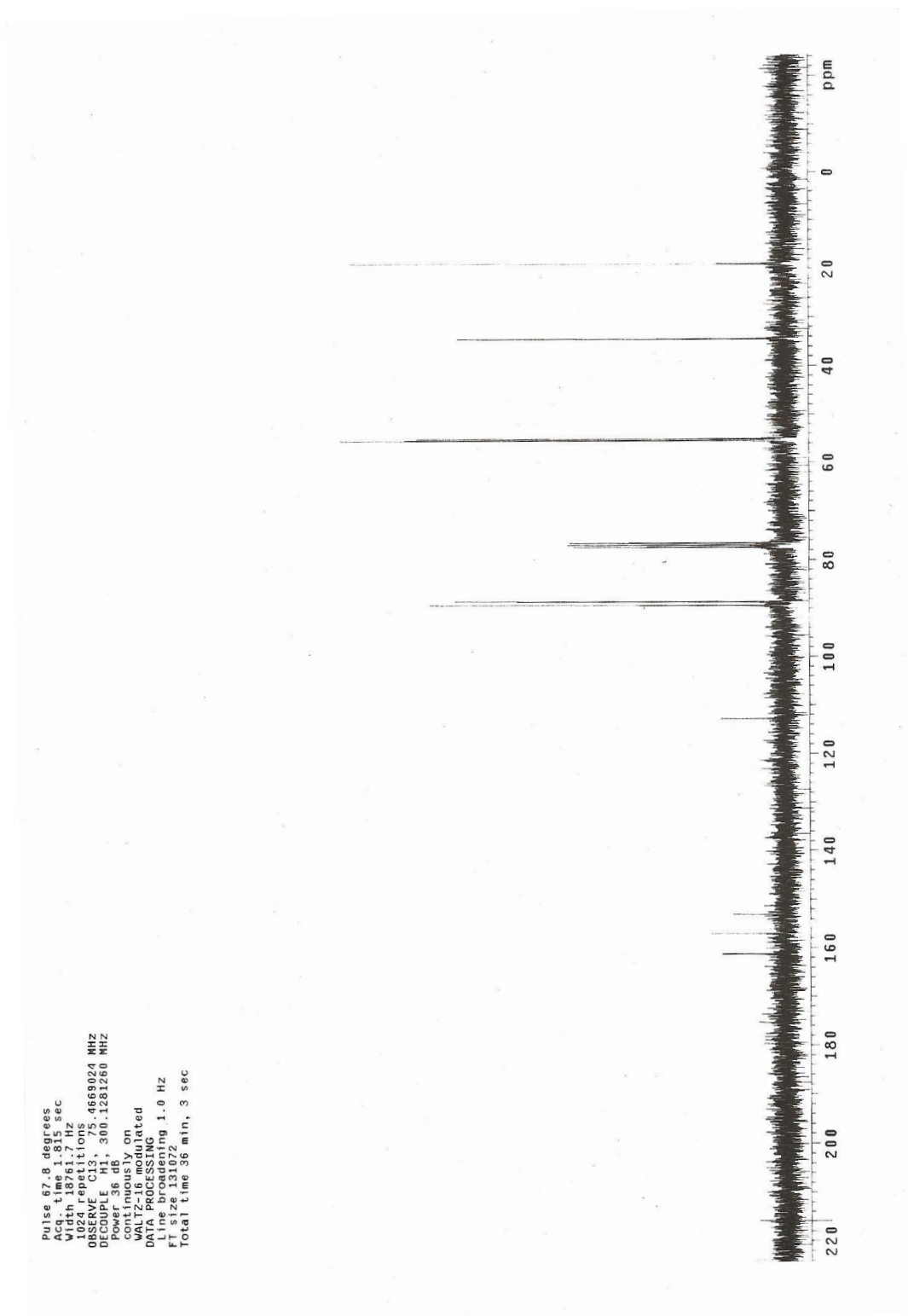
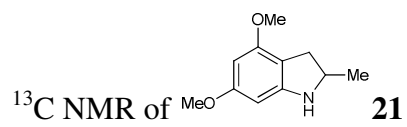


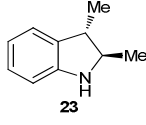
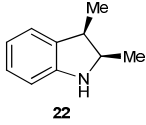




Relax. delay 1.000 sec  
Pulse 45.4 degrees  
Gain 1.000000  
Width 4500 Hz  
16 repetitions  
OBSERVE H1, 300.1266257 MHz  
DATA PROCESSING  
F1 size 32768  
Total time 0 min, 0 sec





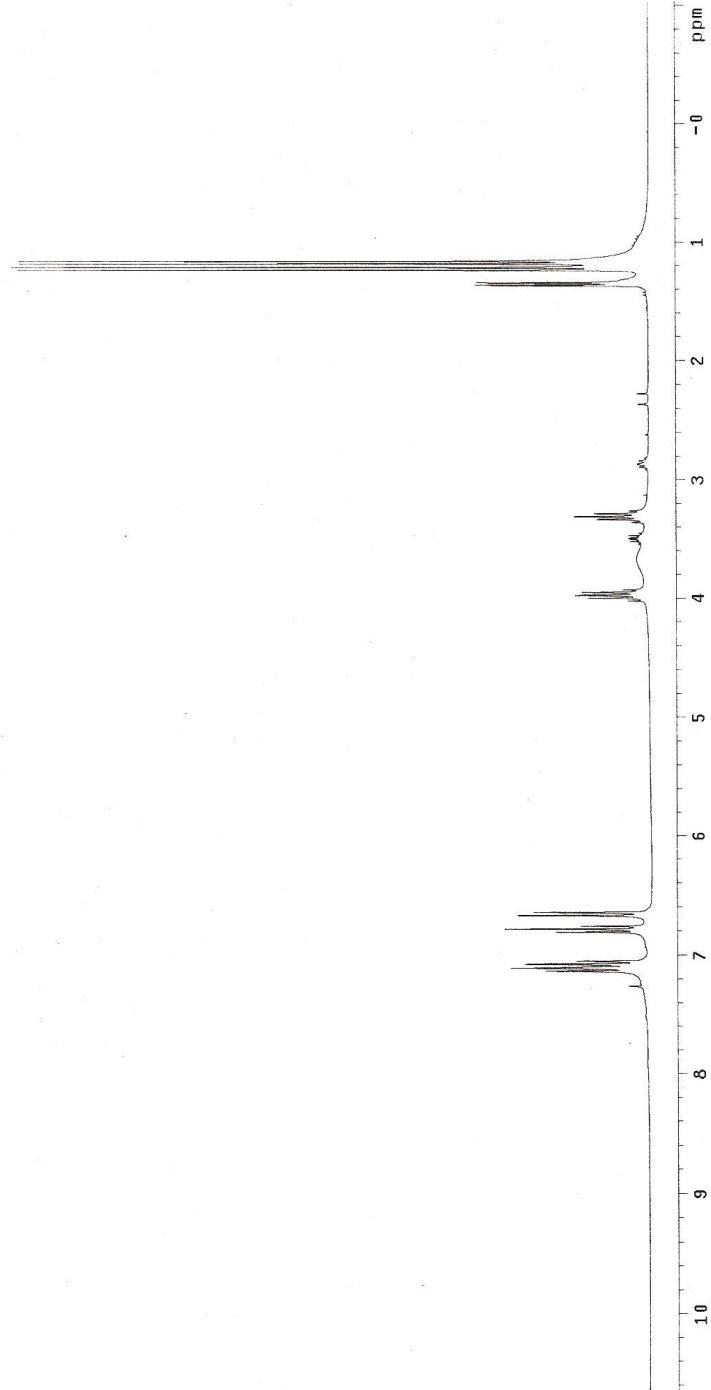


22:23 = 6:1

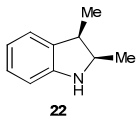
(22 and 23)

<sup>1</sup>H NMR of

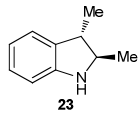
Relax. delay 1.000 sec  
Pulse 43.4 degrees  
Acq. time 1.988 sec  
Width 4500.5 Hz  
Sweep 11.000 MHz  
F1 300.1266252 MHz  
DATA PROCESSING  
FT size 32768  
Total time 1 min, 39 sec



$^{13}\text{C}$  NMR of



22:23 = 6:1



(22 and 23)

Pulse 67.8 degrees  
Acq. time 1.515 sec  
In 15761.7 Hz  
1024  
OBSERVE C13, 75.4669121 MHz  
DECOUPLE H1, 300.1281260 MHz  
Power 36 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
FT size 131072  
Total time 36 min, 3 sec

