

## SUPPLEMENTAL METHODS

RNA was isolated using the RNeasy isolation system (QIAGEN, Valencia, CA) and reverse transcribed using the iScript™ cDNA synthesis kit (Bio-Rad, Hercules, CA). Equal amounts of cDNA reactions were amplified with FastStart Universal SYBR Green Master (Rox) (Roche) using the primer sets listed in Supplemental Table IV. Target sequences were amplified using 95°C/30 sec, 59°C/60 sec, 72°C/30 sec conditions. Fold changes in gene expression between TM-treated and control animals were calculated using the comparative  $2^{-\Delta\Delta Ct}$  method. Samples were internally normalized to 18S ribosomal RNA,  $\Delta Ct = Ct_{\text{sample}} - Ct_{18S}$ . Samples were then normalized to untreated controls,  $\Delta\Delta Ct = \Delta Ct_{\text{TM-treated}} - \Delta Ct_{\text{untreated}}$ . The inverse log of the  $\Delta\Delta Ct$  was then calculated to give the relative fold change. Error bars represent the S.E.M.

**SUPPLEMENTAL TABLE I**  
**GENOTYPING AND CRE-INDUCED RECOMBINATION PRIMER SETS**

Mice were genotyped using the following primer sets:

Xbp1<sup>fl/+</sup> 5'-actgcaccaacacttgccatttc-3', 5'-caaggtgggtcactgcct-3'  
Mist1<sup>CreER/+</sup> 5'-ggtaaagcaaattgtcaagtacgg-3', 5'- atagtaagtatgtgcgctcagcg-3',  
5'-gaagcattttccaggtatgctcag-3'  
R26<sup>LacZ/+</sup> 5'-gcgaagagtttgcctcaacc-3', 5'- ggagcgggagaaatggatatg-3',  
5'-aaagtcgctctgagttgtat-3'

Xbp1<sup>fl/+</sup> recombination was detected using the following primer sets:

Xbp1<sup>WT</sup> 5'-ttgggactctctcgtgtg-3', 5'-caaggtgggtcactgcct-3'  
Xbp1<sup>ΔEx2</sup> 5'-tggccacgctacaaatgaa-3', 5'-caaggtgggtcactgcct-3'

**SUPPLEMENTAL TABLE II**  
**PRIMARY ANTIBODIES FOR IMMUNOHISTOCHEMISTRY**

anti-Amylase 171534, 1:100, Calbiochem, San Diego, CA  
anti-β-gal NB100-65209, 1:300, Novus Biologicals, Littleton, CO  
anti-Chop 2895S, 1:500, Cell Signaling, Boston, MA  
anti-E-Cadherin ab53033, 1:1000, Abcam, Cambridge, MA  
anti-Glucagon A0565, Dako, Carpinteria, CA  
anti-Ki67 M7249, 1:500, Dako, Carpinteria, CA  
anti-K19 Troma3, 1:100, Dev. Studies Hybridoma Bank, Iowa City, IA  
anti-Mist1 C175, 1:500, Pin et al, 2000  
anti-pH3 06-570, 1:100, Upstate (Millipore), Billerica, MA  
anti-LC3B 3868S, 1:200, Cell Signaling, Boston, MA  
anti-Sox9 AB5535, 1:500, Millipore, Billerica, MA  
anti-Hes1 Gift of Tetsuo Sudo, 1:1000  
anti-F4/80 MF48000, 1:50, Invitrogen, Camarillo, CA  
anti-CD3 A0452, 1:200, Dako, Carpinteria, CA

**SUPPLEMENTAL TABLE III**  
**PRIMARY ANTIBODIES FOR IMMUNOBLOTS**

anti-nATF6 $\alpha$	sc-22799, 1:1000, Santa Cruz Biotechnology, Santa Cruz, CA
anti-p-eIF2 $\alpha$	3597S, 1:1000, Cell Signaling, Boston, MA
anti-Amylase	sc-12821, 1:1000, Santa Cruz Biotechnology, Santa Cruz, CA
anti-CPA	1810-0006, 1:1000, AbD Serotec, Raleigh, NC
anti-Erk1/2	9102S, 1:1000, Cell Signaling, Boston, MA
anti-p-Erk1/2	9101S, 1:1000, Cell Signaling, Boston, MA
anti-p38	9212S, 1:1000, Cell Signaling, Boston, MA
anti-S6	sc-74459, 1:1000, Santa Cruz Biotechnology, Santa Cruz, CA

**SUPPLEMENTAL TABLE IV**  
**RT-QPCR PRIMER SETS**

Xbp1 <sup><math>\Delta</math>Ex2</sup>	5'-agaaagcgctgaggagaac-3', 5'-cctccacctctggaacctc-3'
Bip	5'-gtgtcctctctgggtatcagg-3', 5'-tgtctttgttaggggtcggtt-3'
Chop	5'-cctgaggagagagtgtccag-3', 5'-cagatcctcataccaggcttc-3'
Xbp1u	5'-tcagactatgtgcacctctgc-3', 5'-agagaaagggaggctggtaag-3'
Xbp1s	5'-tgagtccgcagcaggt-3', 5'-agagaaagggaggctggtaag-3'
Sec61a	5'-ctatttccagggtccgagt-3', 5'-aggtgtgtactggcctcggt-3'
Pdi1	5'-caagatcaagccccacctgat-3', 5'-agttcgcccccaaccagtactt-3'
Amylase	5'-cagagacatggtgacaagggtg-3', 5'-atcgtaaagtccaagcaga-3'
Elastase	5'-actatgtccagctgggtgttc-3', 5'-cagtaagaggagctggagcag-3'
Nestin	5'-gagagtcgcttagagggtcag-3', 5'-gatctgagcgatctgactctgt-3'
Hes1	5'-agagaaggcagacattctgga-3', 5'-gtcacctcgttcactgcactc-3'
Gli1	5'-tttcttgaggtgggatgaag-3', 5'-ggtaggtcattggattgaac-3'
Mist1	5'-tggtggctaaagctacgtgt-3', 5'-catagctccaggctggtttt-3'
Sox9	5'-cttctgtgggagcgacaactt-3', 5'-agggagggaaaacagagaacg-3'
Reg1	5'-atggctaggaacgcctacttc-3', 5'-cccaagttaacggcttctcagt-3'
18S	5'-tgtctcaaagattaagccatgc-3', 5'-gcgaccaaaggaaccataac-3'

**SUPPLEMENTAL TABLE V**  
**MARKER EXPRESSION PROFILE**

<b>COMPARTMENT</b>	<b>MARKERS EXPRESSED</b>
Acinar Cells	Mist1, Amylase, Carboxypeptidase (CPA)
Islets	Insulin, Glucagon
Ductal/centroacinar	Cytokeratin 19 (K19), Sox9
Centroacinar exclusive	Hes1, Nestin
All epithelial cells	E-cadherin

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<b>CONTEXT</b>	<b>MARKERS EXPRESSED</b>
General ER stress	BIP, CHOP
Xbp1 Pathway	Xbp1s, Sec61a, PDI
PERK Pathway	p-eIF2 $\alpha$
ATF6a Pathway	nATF6 $\alpha$
Apoptosis	TUNEL
Actively proliferating	Ki67, phospho-histone 3 (pH3)
Cre-recombination (ROSA)	$\beta$ -galactosidase
Autophagy	Autophagy marker light chain 3 - isoform B (LC3B)
Regeneration	Reg1

## SUPPLEMENTAL FIGURE LEGENDS

**FIGURE S1:** *The Xbp1<sup>fl/fl</sup> locus has no effect on acinar cell morphology. (A,B)* H&E staining of pancreata sections from control wild type and Xbp1<sup>ΔEx2/+</sup>; Mist1<sup>CreER/+</sup> mice treated with corn oil. All genotypes exhibit normal appearing pancreata. Scale = 40 μm. **(C,D)** Relative transcript levels of the common ER stress indicators BiP and Chop from 2 wk post-TM treated mice reveals that pancreata heterozygous for Xbp1 do not undergo an ER stress response. Only homozygous null Xbp1<sup>ΔEx2</sup> cells show ER stress responses. \*p<0.05

**FIGURE S2:** *Islet and duct populations are normal following acinar-specific Xbp1 ablation. (A)* Insulin/glucagon costaining of Xbp1<sup>ΔEx2</sup> pancreata 4 wk post-TM reveal normal islet morphology. **(B)** K19 staining of Xbp1<sup>ΔEx2</sup> pancreata also indicates normal ductal development in 4 wk post-TM pancreata. Scale = 20 μm.

**FIGURE S3:** *Xbp1<sup>fl/+</sup>;Mist1<sup>CreER/+</sup> and Xbp1<sup>fl/fl</sup>;Mist1<sup>CreER/+</sup> zymogen-bearing acini have normal zymogen accumulation, localization and development of rER. (A,B)* Xbp1<sup>fl/+</sup>;Mist1<sup>CreER/+</sup> pancreata show no alterations in zymogen granule accumulation, abortive zymogens, or ER disorganization. **(C,D)** Zymogen-containing, acinar cell populations in 4 wk post-TM Xbp1<sup>fl/fl</sup>;Mist1<sup>CreER/+</sup> pancreata are similar in ultrastructure to heterozygous and wild-type controls. **(E,F)** Non-zymogenic exocrine cells in 4 wk post-TM Xbp1<sup>fl/fl</sup>; Mist1<sup>CreER/+</sup> pancreata show the previously described cell abnormalities including few and mislocalized zymogens, self-destructing organelles, and disrupted ER organization. **(G,H)** Eight wk post-TM Xbp1<sup>fl/fl</sup>; Mist1<sup>CreER/+</sup> pancreata

exhibit an almost full recovery from Xbp1 deletion with an increase in ZG numbers and a well developed rER. Scale = 1  $\mu\text{m}$  (A,C,E,G); 0.5  $\mu\text{m}$  (B,D,F,H).

**FIGURE S4:** *Anti- $\beta$ -galactosidase staining indicates zymogenic cells fail to ablate Xbp1.*

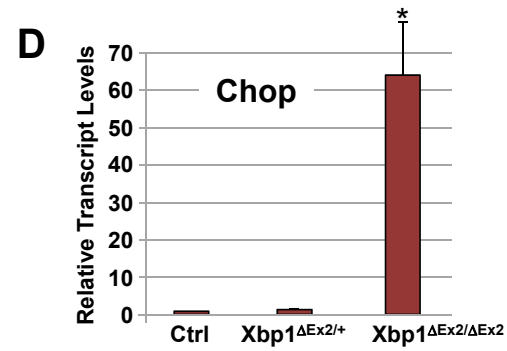
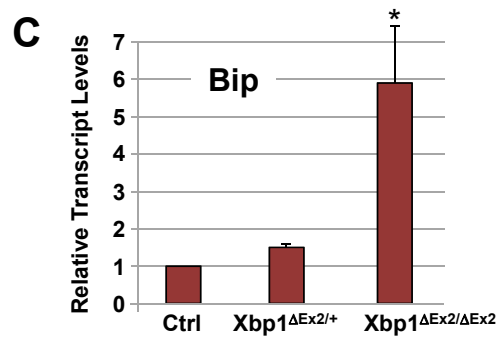
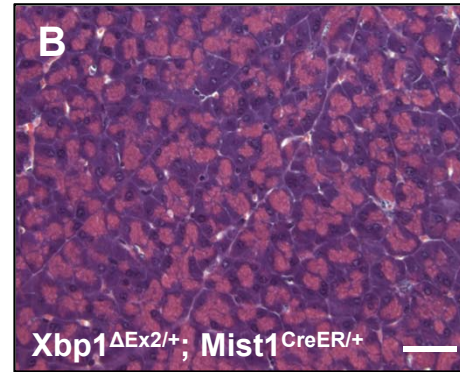
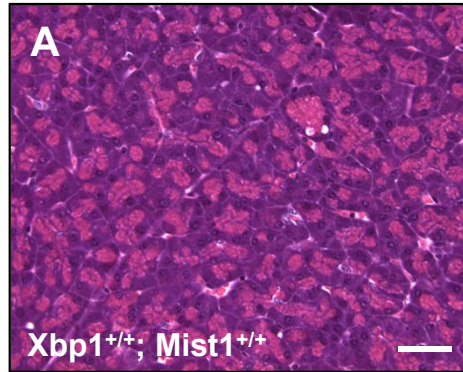
Zymogenic cells (outlines - identified under phase contrast by extensive ZG accumulation) in Xbp1<sup>fl/fl</sup>;Mist1<sup>CreER/+</sup>;R26R<sup>LacZ</sup> pancreata 4 wk post-TM do not express  $\beta$ -gal (green), indicating no Cre-dependent recombination. Scale = 20  $\mu\text{m}$ .

**FIGURE S5:** *Xbp1 <sup>$\Delta$ Ex2</sup> cells undergo autophagy.* **(A)** LC3B staining (green) in 4 wk post-TM pancreata reveals the presence of extensive autophagic cells within the non-zymogenic cell compartment. In contrast, zymogenic cells (red outlines) and islets (yellow outline) exhibit no signs of autophagy. **(B)** Electron micrograph of a non-zymogenic acinar cell showing a zymogen granule within an autolysosome (red box). Scale = 20  $\mu\text{m}$  (A); 1  $\mu\text{m}$  (B). **(C)** Immunoblots for additional stress pathway components (pERK1/2 and p38) show that they are elevated in Xbp1 <sup>$\Delta$ Ex2</sup> pancreata over the indicated post-TM time course. As predicted, maximum MAPK stress is observed at 4 wk post-TM.

**FIGURE S6:** *Xbp1 <sup>$\Delta$ Ex2</sup> pancreata undergo extensive cell proliferation of Xbp1<sup>fl/fl</sup> acinar cells.* **(A)** Quantification of Ki67 positive Xbp1 <sup>$\Delta$ Ex2</sup> acinar cells over the indicated post-TM time course. **(B)** H&E staining reveals mitotic figures (arrow) in pancreata 6 wk post-TM. Scale = 10  $\mu\text{m}$ . **(C)** Relative transcript levels of Reg1 over the indicated post-TM time points. \*p<0.05

**FIGURE S7:** *Xbp1<sup>ΔEx2</sup> pancreata reveal rare β-gal+ acinar cells 8 wk post-TM.* Anti-β-gal staining of 8 wk post-TM *Xbp1<sup>ΔEx2</sup>* pancreata shows a rare intact acinus (white outline) that is β-gal positive, presumably reflecting cells that recombined the R26<sup>LacZ</sup> locus but failed to delete both copies of the *Xbp1<sup>fl/fl</sup>* allele. Scale = 10 μm.

**FIGURE S8:** *Recovered Xbp1<sup>ΔEx2</sup> pancreata contain areas of tubular duct-like structures surrounded by stromal cells.* **(A)** High-magnification image of tubular duct complexes (arrows) in 8 wk *Xbp1<sup>ΔEx2</sup>* pancreata. **(B,C)** Surrounding the tubular complexes (TC) are CD3+ T cells and F4/80+ macrophages (arrows). Ac - acinar cells; BV - blood vessel. Scale = 20 μm.



**Figure S1**



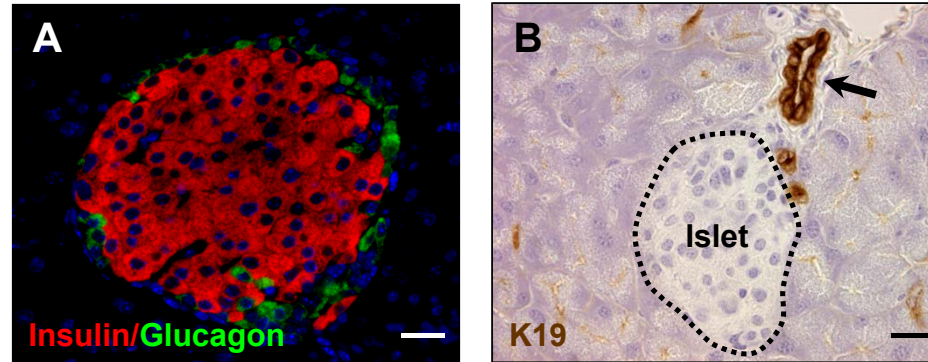
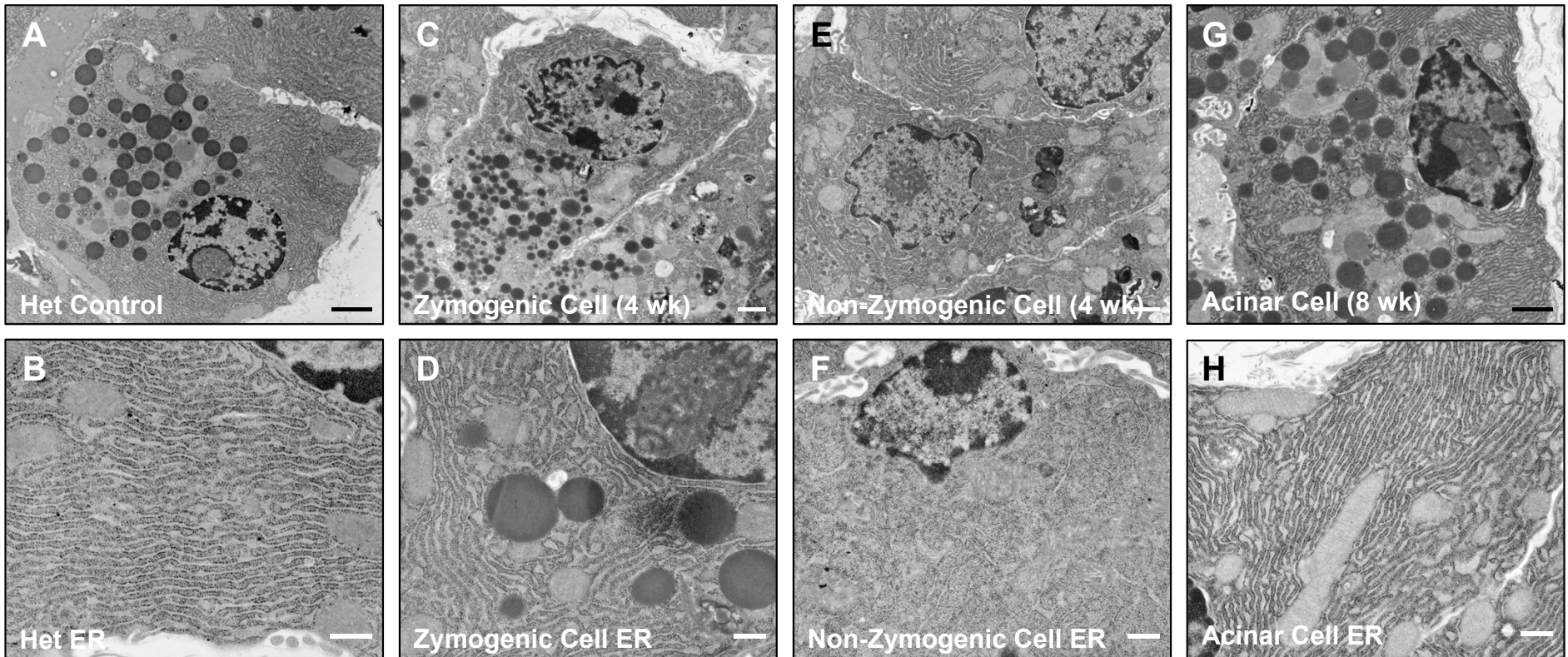


Figure S2



**Figure S3**

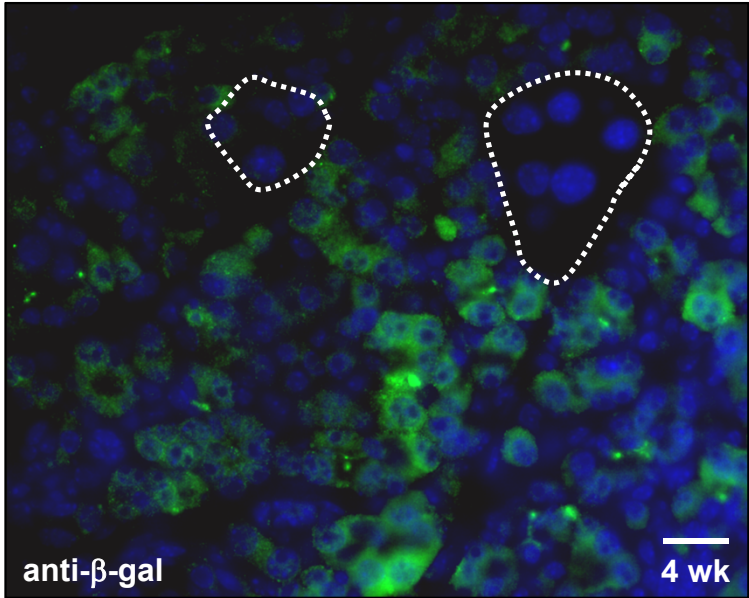


Figure S4

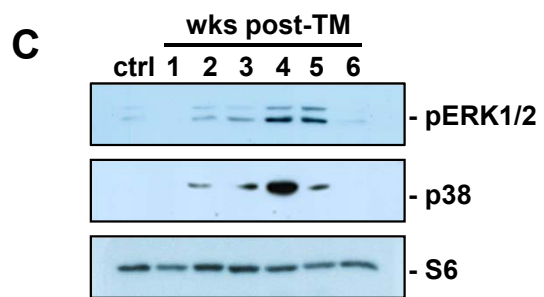
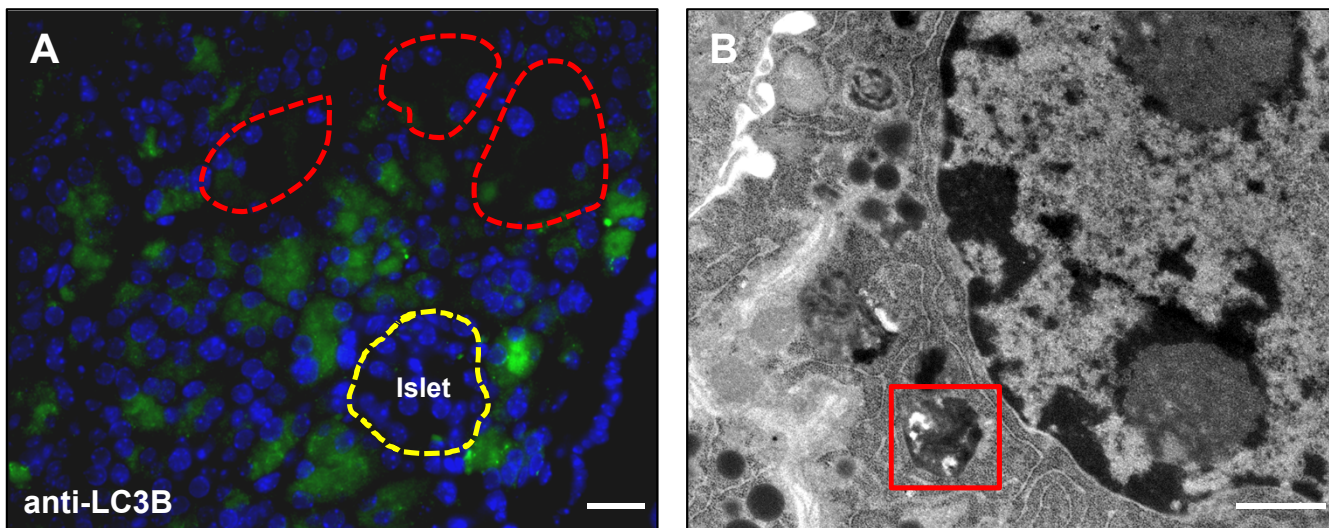
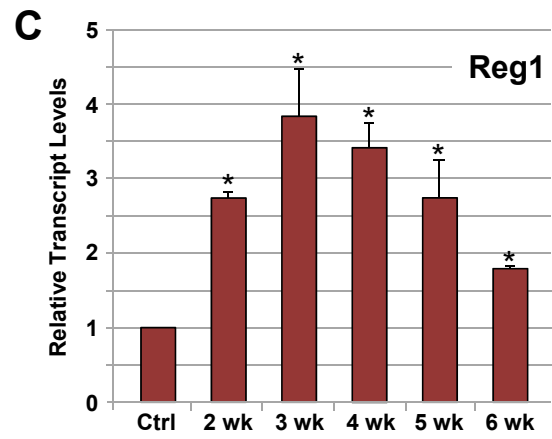
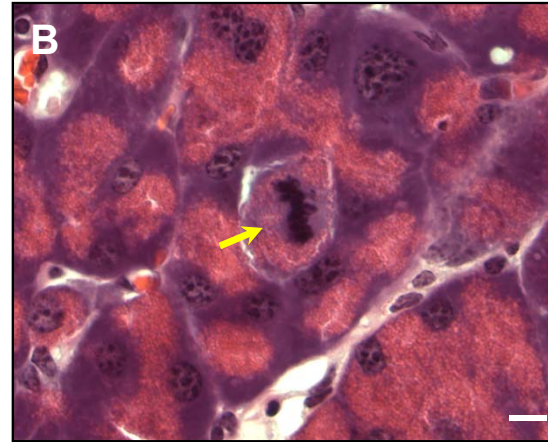
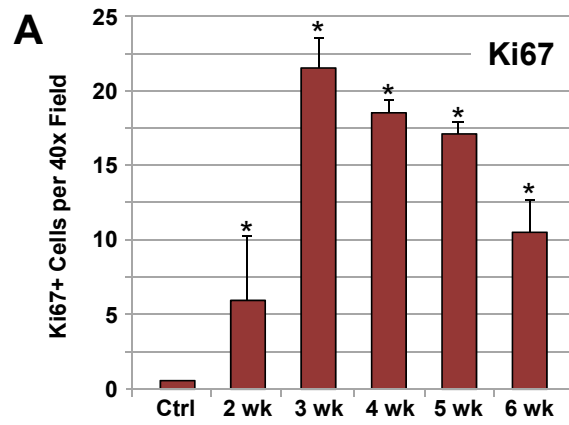


Figure S5



**Figure S6**

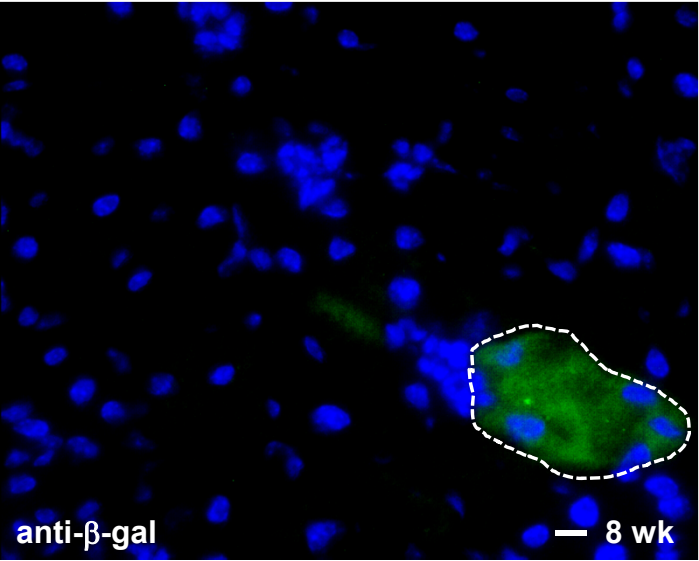
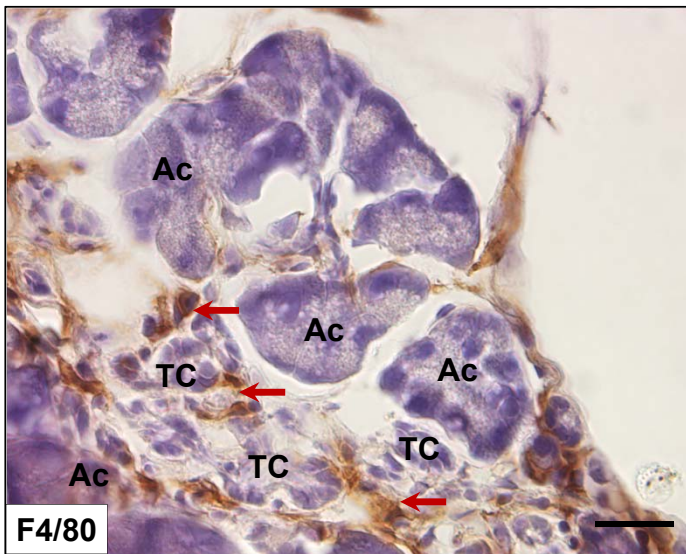
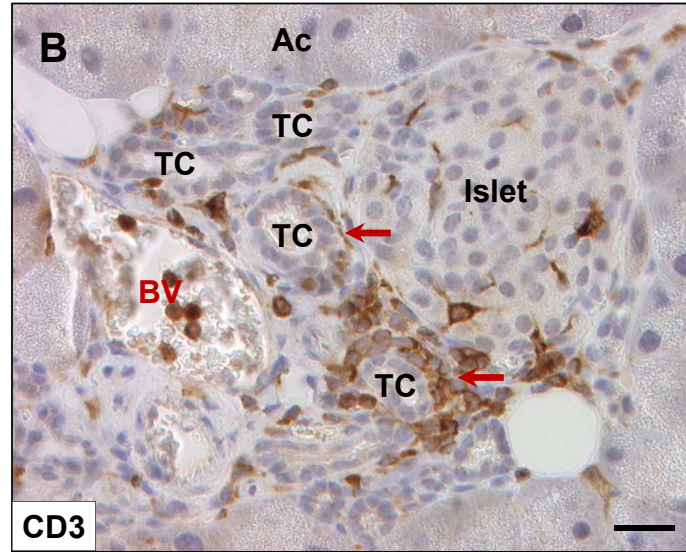
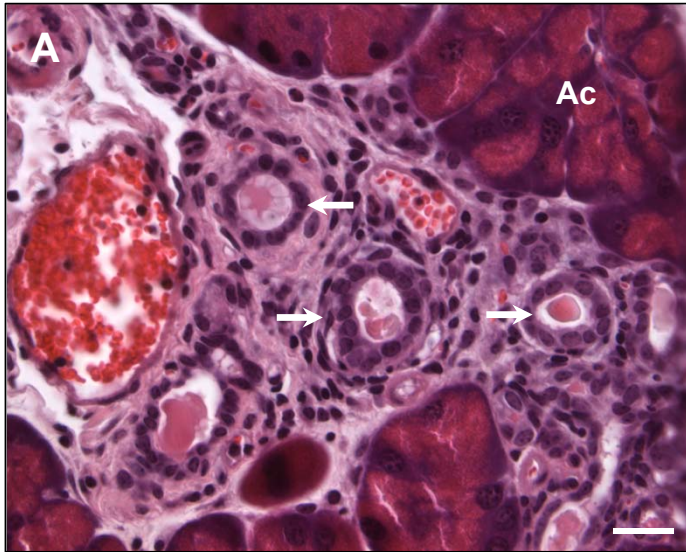


Figure S7



**Figure S8**