

# "N-O Chemistry for Antibiotics: Discovery of N-Alkyl-N-(pyridin-2-yl)hydroxylamine Scaffolds as Selective Antibacterial Agents Using Nitroso Diels-Alder and Ene Chemistry"

## Supporting Information

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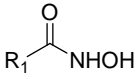


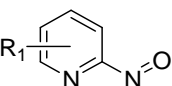
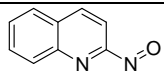

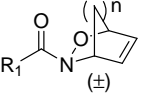
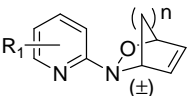
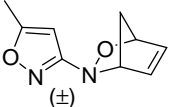
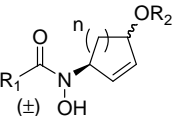
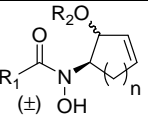
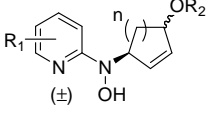
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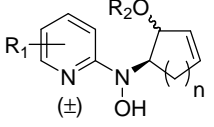
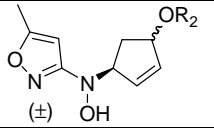
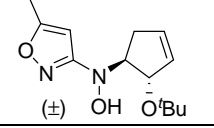
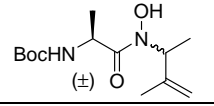
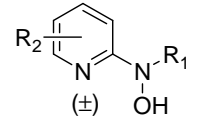

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## I. Table of All Structures From This Work

Table S1. Structures and antibacterial activity against <i>Micrococcus luteus</i> ATCC 10240 in the agar diffusion assay.								
Library <sup>a</sup>	Structure <sup>b</sup>	Comp. <sup>c</sup>	n	syn/anti <sup>d</sup>	R <sub>1</sub>	R <sub>2</sub>	Ref. <sup>e</sup>	Active? <sup>f</sup>
A Nitrosos & Precursors		S1	-	-	Bn	-	4	No
		S2	-	-	Ph	-	4	No
		S3	-	-	<i>t</i> -BuO	-	4	No
		S4	-	-	BocHN- 	-	4	No
		S5	-	-	BocHN- 	-	4	No
		8	-	-	6-Me	-	4	No
		10	-	-	H	-	4	nt
		17a	-	-	6-Et	-	4	No
		17b	-	-	4-Me	-	4	No
		17c	-	-	3-Me	-	4	Yes <sup>g</sup>
		17d	-	-	5-Cl	-	4	No
		17e	-	-	5-Br	-	4	No
		17f	-	-	5-I	-	4	No
		17g	-	-	3-Cl-5-Cl	-	4	No
		17h	-	-	5-Br-6-Me	-	4	No
	17i	-	-	3-Cl-5-CF <sub>3</sub>	-	4	No	
		13	-	-	-	-	4	No
		15	-	-	-	-	4	No
	B Diels-Alder Cycloadducts		S6	1	-	Bn	-	4
S7			2	-	Bn	-	4	No
S8			-	-	Ph	-	4	No
S9			1	-	<i>t</i> -BuO	-	4	No
		6	2	-	5-Br	-	1,4	No
		S10	1	-	5-Cl	-	1,4	No
		S11	1	-	5-Br	-	1,4	No
		S12	1	-	6-Me	-	1,4	No
		S13	2	-	6-Me	-	1,4	No
		S14	-	-	-	-	1,4	No
C Ring Opened Compounds		S15	1	anti	Bn	Me	3,4	No
		S16	1	anti	Bn	<i>t</i> -Bu	3,4	No
		S17	1	anti	<i>t</i> -BuO	Me	3,4	No
		S18	2	anti	Bn	Me	3,4	No
		S19	1	anti	<i>t</i> -BuO	Me	3,4	No
		S20	2	anti	Bn	Me	3,4	No
		7a	2	anti	5-Br	Me	1,4	Yes
		S21	1	syn/anti	5-Cl	<i>i</i> -Pr	1,4	Yes
		S22	1	syn/anti	5-Cl	<i>t</i> -Bu	1,4	Yes
		S23	1	syn	5-Br	H	1,4	Yes

		<b>S24</b>	1	anti	5-Br	H	1,4	Yes
		<b>S25</b>	1	syn/anti	5-Br	Me	1,4	Yes
		<b>S26</b>	1	syn/anti	5-Br	<i>i</i> -Pr	1,4	Yes
		<b>S27</b>	2	anti	5-Br	<i>i</i> -Pr	1,4	Yes
		<b>S28</b>	1	syn/anti	5-Br	<i>t</i> -Bu	1,4	Yes
		<b>S29</b>	2	anti	5-Br	<i>t</i> -Bu	1,4	Yes
		<b>S30</b>	2	anti	5-Br	allyl	1,4	Yes
		<b>S31</b>	1	syn/anti	6-Me	Me	1,4	Yes
		<b>S32</b>	1	syn/anti	6-Me	<i>i</i> -Pr	1,4	Yes
		<b>S33</b>	2	anti	6-Me	<i>i</i> -Pr	1,4	Yes
		<b>S34</b>	1	anti	6-Me	<i>t</i> -Bu	1,4	Yes
		<b>S35</b>	2	anti	6-Me	<i>t</i> -Bu	1,4	Yes
		<b>7b</b>	2	anti	5-Br	Me	1,4	Yes
		<b>S36</b>	1	anti	5-Cl	Me	1,4	Yes
		<b>S37</b>	1	anti	5-Cl	<i>t</i> -Bu	1,4	Yes
		<b>S38</b>	1	anti	5-Br	H	1,4	Yes
		<b>S39</b>	2	anti	5-Br	H	1,4	Yes
		<b>S40</b>	1	anti	5-Br	Me	1,4	Yes
		<b>S41</b>	1	anti	5-Br	<i>i</i> -Pr	1,4	Yes
		<b>S42</b>	2	anti	5-Br	<i>i</i> -Pr	1,4	Yes
		<b>S43</b>	2	anti	5-Br	<i>t</i> -Bu	1,4	Yes
		<b>S44</b>	2	anti	5-Br	Bn	1,4	Yes
		<b>S45</b>	2	anti	5-Br	allyl	1,4	Yes
		<b>S46</b>	1	anti	6-Me	Me	1,4	Yes
		<b>S47</b>	2	anti + 1,4- anti isomer	6-Me	Me	1,4	Yes
		<b>S48</b>	1	anti	6-Me	<i>i</i> -Pr	1,4	Yes
		<b>S49</b>	2	anti	6-Me	<i>i</i> -Pr	1,4	Yes
		<b>S50</b>	2	anti	6-Me	<i>t</i> -Bu	1,4	Yes
		<b>S51</b>	-	syn/anti	-	Me	1,4	No
		<b>S52</b>	-	syn/anti	-	<sup>t</sup> Bu	1,4	No
		<b>S53</b>	-	anti	-	-	1,4	No
<b>D</b> Ene Products		<b>S54</b>	-	-	-	-	-	No
		<b>9</b>	-	-		6-Me	2,4, This Work	Yes
		<b>12</b>	-	-		H	2,4	Yes
		<b>18a</b>	-	-		6-Et	This Work	Yes
		<b>18b</b>	-	-		4-Me	This Work	Yes
		<b>18c</b>	-	-		3-Me	This Work	Yes
		<b>18d</b>	-	-		5-Cl	4, This Work	Yes
		<b>18e</b>	-	-		5-Br	2,4, This Work	Yes
		<b>18f</b>	-	-		5-I	This Work	Yes

		<b>18g</b>	-	-		3-Cl-5-Cl	This Work	Yes
		<b>18h</b>	-	-		5-Br-6-Me	This Work	Yes
		<b>18i</b>	-	-		3-Cl-5-CF <sub>3</sub>	This Work	No
		<b>S55</b>	-	-		5-Br	2,4	Yes
		<b>11a</b>	-	-		H	2,4, This Work	Yes
		<b>S56</b>	-	-		5-Br	2,4	Yes
		<b>40a</b>	-	-		6-Et	This Work	Yes
		<b>19a</b>	-	-		H	This Work	Yes
		<b>19b</b>	-	-		H	This Work	Yes
		<b>19c</b>	-	-		H	This Work	Yes
		<b>14</b>	-	-	-	-	This Work	Yes
		<b>16</b>	-	-		-	2,4	Yes
		<b>S57</b>	-	-		-	2,4	Yes
		<b>S58</b>	-	-		-	2,4	Yes
		<b>S59</b>	-	-		-	2,4	Yes
<b>E</b> O-Alkyl-Hydroxyl-Amines		<b>30a</b>	-	-	Me	-	This Work	No
		<b>30b</b>	-	-	Pr	-	This Work	No
		<b>30c</b>	-	-	<i>n</i> -hexyl	-	This Work	No
		<b>30d</b>	-	-	Bn	-	This Work	No
		<b>30e</b>	-	-	(CH <sub>2</sub> ) <sub>2</sub> Ph	-	This Work	No
		<b>30f</b>	-	-	allyl	-	This Work	No
		<b>30g</b>	-	-	3-(PhO)Bn	-	This Work	No
		<b>30h</b>	-	-	4-(CF <sub>3</sub> )Bn	-	This Work	No
		<b>30i</b>	-	-	4-(CF <sub>3</sub> O)Bn	-	This Work	No
			<b>36</b>	-	-	-	-	This Work
		<b>38</b>	-	-	-	-	This Work	No
<b>F</b> Buchwald Hydroxyl-Amines		<b>31a</b>	-	-	Me	-	This Work	Yes
		<b>31b</b>	-	-	Pr	-	This Work	No
		<b>31c</b>	-	-	<i>n</i> -hexyl	-	This Work	Yes
		<b>31d</b>	-	-	Bn	-	This Work	Yes

		<b>31e</b>	-	-	(CH <sub>2</sub> ) <sub>2</sub> Ph	-	This Work	Yes
		<b>31f</b>	-	-	allyl	-	This Work	Yes
		<b>31g</b>	-	-	3-(PhO)Bn	-	This Work	No
		<b>31h</b>	-	-	4-(CF <sub>3</sub> )Bn	-	This Work	Yes
		<b>31i</b>	-	-	4-(CF <sub>3</sub> O)Bn	-	This Work	Yes
<b>G</b> O-Acyl- Hydroxyl- Amines		<b>33</b>	-	-	Bn	H	This Work	Yes
<b>35</b>		-	-		Br	This Work	Yes	
<b>H</b> N-O Reduced Compounds		<b>32</b>	-	-	Bn	H	This Work	No
		<b>34</b>	-	-		Br	This Work	No
<b>I</b> Homologated Hydroxyl- Amines		<b>39</b>	-	-	-	-	This Work	No

<sup>a</sup>Compound library is organized by types of structures (**A–I**) rather than compound numbers.

<sup>b</sup>(±) denotes a racemic compound (all compounds were tested in racemic form).

<sup>c</sup>Compounds only appearing in the supporting information are denoted with a capital **S** before the number and are numbered based on chronological appearance in Table S1. If a compound was used in main text, the original compound number assigned in the main text was retained throughout the supporting information.

<sup>d</sup>Some compounds, as indicated in the table, were isolated, characterized, and tested as a mixture of syn/anti diastereomeric mixtures of isomers.

<sup>e</sup>Reference contains disclosure of the original synthesis of the compound and spectral characterization.

<sup>f</sup>Indicates antibacterial activity against *Micrococcus luteus* ATCC 10240 in the agar diffusion antibacterial susceptibility assay. “Yes” denotes that the compound gave a growth inhibition zone diameter of >15 mm; “No” denotes that the compound gave a growth inhibition zone diameter of ≤15 mm; “nt” denotes that the compound was not tested. All of the tabulated biological data is given in Table S2.

<sup>g</sup>This was the only nitroso compound with moderate activity. See Table S2.

## II. Table of All Biological Data From This Work

**Table S2.** Complete list of biological data for compounds used in this study. Diameters of growth inhibition zones (mm) in the agar diffusion antibacterial susceptibility assay.<sup>a</sup> Minimum inhibitory concentrations (MIC<sub>90</sub>;  $\mu\text{M}$ ) in the broth microdilution antibacterial susceptibility assay.<sup>b</sup> Inhibitory concentrations (IC<sub>50</sub>;  $\mu\text{M}$ ) and percent growth inhibition values (compound concentration of 20  $\mu\text{M}$ ) against cancer cell lines.<sup>c</sup>

Comp <sup>d</sup>	Gram-positive Bacteria (diameter of growth inhibition zone in mm; MIC <sub>90</sub> in $\mu\text{M}$ given in parentheses)				Gram-negative Bacteria (diameter of growth inhibition zone in mm)			Yeast (diameter of zone in mm)	Cancer Cell Lines (% Inhibition @ 20 $\mu\text{M}$ ; IC <sub>50</sub> in $\mu\text{M}$ given in parentheses)		
	<i>M. luteus</i> ATCC 10240	<i>S. aureus</i> SG511	<i>E. faecalis</i> ATCC 49532	<i>M. vaccae</i> IMET 10670	<i>P. aeruginosa</i> 799/WT	<i>P. aeruginosa</i> 799/61	<i>E. coli</i> X580	<i>S. salmonicolor</i> 549	MCF7	PC3	HeLa
<b>6</b>	0	0	0	0	nt	0	0	nt	nt	nt	nt
<b>7a</b>	34 (4.0 $\mu\text{M}$ )	12 (>128 $\mu\text{M}$ )	16P (>128 $\mu\text{M}$ )	17p	nt	0	0	nt	33%	32%	25%
<b>7b</b>	34 (4.0 $\mu\text{M}$ )	18p ( $\geq$ 128 $\mu\text{M}$ )	20P (>128 $\mu\text{M}$ )	22s	nt	0	0	nt	nt	nt	nt
<b>8</b>	0	0	0	0	0	0	0	nt	nt	nt	nt
<b>9</b>	48 (4.0 $\mu\text{M}$ )	12 (>128 $\mu\text{M}$ )	15P (>128 $\mu\text{M}$ )	20P	nt	0	0	nt	11%	24%	0%
<b>10</b>	nt	nt	nt	nt	nt	nt	nt	nt	nt	nt	nt
<b>11a</b>	35 (8.0 $\mu\text{M}$ )	19 (64 $\mu\text{M}$ )	20P (>128 $\mu\text{M}$ )	16p	nt	0	18h	nt	57%	35%	31%
<b>12</b>	31 (8.0 $\mu\text{M}$ )	18 (128 $\mu\text{M}$ )	22P (>128 $\mu\text{M}$ )	nt	nt	nt	nt	nt	50%	14%	25%
<b>13</b>	9.5	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>14</b>	31 (8.0 $\mu\text{M}$ )	13 (128 $\mu\text{M}$ )	13P/h (>128 $\mu\text{M}$ )	nt	nt	nt	nt	nt	76% (15 $\mu\text{M}$ )	17%	41%

<b>15</b>	0	0	0	0	0	0	0	nt	nt	nt	nt
<b>16</b>	21 (64 $\mu$ M)	0 ( $>128 \mu$ M)	13P ( $>128 \mu$ M)	0	nt	nt	nt	nt	10%	28%	34%
<b>17a</b>	14	11	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>17b</b>	14	9.5	12h	nt	nt	nt	nt	nt	nt	nt	nt
<b>17c</b>	17	15	116p	nt	nt	nt	nt	nt	nt	nt	nt
<b>17d</b>	11.5p	0	nt	9.5	nt	nt	nt	nt	nt	nt	nt
<b>17e</b>	11/13p	9.5/12P	nt	10	nt	nt	nt	nt	nt	nt	nt
<b>17f</b>	14	11	9.5	nt	nt	nt	nt	nt	nt	nt	nt
<b>17g</b>	9.5	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>17h</b>	15	12	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>17i</b>	0	0	17	nt	nt	nt	nt	nt	nt	nt	nt
<b>18a</b>	52 (2.0 $\mu$ M)	11/20h ( $>128 \mu$ M)	22P/h ( $>128 \mu$ M)	nt	nt	nt	nt	nt	22%	11%	13%
<b>18b</b>	29 (4.0 $\mu$ M)	20 (128 $\mu$ M)	22P ( $>128 \mu$ M)	nt	nt	nt	nt	nt	74% (12 $\mu$ M)	17%	37%
<b>18c</b>	25 (16 $\mu$ M)	10 ( $>128 \mu$ M)	19P/h ( $>128 \mu$ M)	nt	nt	nt	nt	nt	nt	nt	nt
<b>18d</b>	40 (2.0 $\mu$ M)	18 (128 $\mu$ M)	15/21P ( $\geq 128 \mu$ M)	nt	nt	nt	nt	nt	68% (14 $\mu$ M)	0%	18%
<b>18e</b>	35 (4.0 $\mu$ M)	17P ( $>128 \mu$ M)	19P ( $>128 \mu$ M)	26s	nt	nt	nt	nt	70% (10 $\mu$ M)	12%	19%
<b>18f</b>	38 (2.0 $\mu$ M)	12 ( $>128 \mu$ M)	19P/h ( $>128 \mu$ M)	nt	nt	nt	nt	nt	75% (8 $\mu$ M)	17%	15%
<b>18g</b>	27 (16 $\mu$ M)	0 ( $>128 \mu$ M)	14P ( $>128 \mu$ M)	nt	nt	nt	nt	nt	29%	20%	24%

<b>18h</b>	35 (4.0 $\mu$ M)	13 ( $>128 \mu$ M)	12P ( $>128 \mu$ M)	nt	nt	nt	nt	nt	19%	0%	21%
<b>18i</b>	15 (128 $\mu$ M)	0 ( $>128 \mu$ M)	13P ( $>128 \mu$ M)	nt	nt	nt	nt	nt	20%	13%	30%
<b>19a</b>	33 (4.0 $\mu$ M)	15 (128 $\mu$ M)	18P ( $>128 \mu$ M)	nt	nt	nt	nt	nt	63%	20%	3%
<b>19b</b>	20 (64 $\mu$ M)	19 (32 $\mu$ M)	18P ( $>128 \mu$ M)	nt	nt	nt	nt	nt	nt	nt	nt
<b>19c</b>	30 (16 $\mu$ M)	15p (128 $\mu$ M)	16P ( $>128 \mu$ M)	nt	nt	nt	nt	nt	49%	24%	37%
<b>30a</b>	0	0	0	0	nt	nt	nt	nt	nt	nt	nt
<b>30b</b>	0	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>30c</b>	13P/h	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>30d</b>	0 ( $>128 \mu$ M)	0 ( $>128 \mu$ M)	0 ( $>128 \mu$ M)	nt	nt	nt	nt	nt	nt	nt	nt
<b>30e</b>	0	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>30f</b>	0	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>30g</b>	0	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>30h</b>	14P	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>30i</b>	14P	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>31a</b>	18p/24P ( $\geq 128 \mu$ M)	14h ( $>128 \mu$ M)	11P/h ( $>128 \mu$ M)	nt	nt	nt	nt	nt	90% (17 $\mu$ M)	63% (15 $\mu$ M)	97% (17 $\mu$ M)
<b>31b</b>	13p ( $>128 \mu$ M)	0 ( $>128 \mu$ M)	0 ( $>128 \mu$ M)	nt	nt	nt	nt	nt	nt	nt	nt
<b>31c</b>	15 (128 $\mu$ M)	17p ( $>128 \mu$ M)	16P ( $>128 \mu$ M)	nt	nt	nt	nt	nt	nt	nt	nt



<b>31d</b>	24 (64 $\mu$ M)	18 (128 $\mu$ M)	20P (>128 $\mu$ M)	15	0	0	0	nt	0%	0%	10%
<b>31e</b>	16 (128 $\mu$ M)	14h (>128 $\mu$ M)	15 (>128 $\mu$ M)	nt	nt	nt	nt	nt	nt	nt	nt
<b>31f</b>	14 (>128 $\mu$ M)	0 (>128 $\mu$ M)	12P (>128 $\mu$ M)	11P	0	0	0	nt	nt	nt	nt
<b>31g</b>	12p/P (>128 $\mu$ M)	13 (>128 $\mu$ M)	0 (>128 $\mu$ M)	14	0	0	12	nt	nt	nt	nt
<b>31h</b>	16 (128 $\mu$ M)	15s (>128 $\mu$ M)	15 (>128 $\mu$ M)	nt	nt	nt	nt	nt	nt	nt	nt
<b>31i</b>	17/21h (128 $\mu$ M)	14h (>128 $\mu$ M)	15P (>128 $\mu$ M)	nt	nt	nt	nt	nt	nt	nt	nt
<b>32</b>	0 (>128 $\mu$ M)	0 (>128 $\mu$ M)	0 (>128 $\mu$ M)	0	0	0	0	nt	nt	nt	nt
<b>33</b>	20 (128 $\mu$ M)	12 (>128 $\mu$ M)	11P (>128 $\mu$ M)	0	0	0	0	nt	nt	nt	nt
<b>34</b>	0 (>128 $\mu$ M)	0 (>128 $\mu$ M)	0 (>128 $\mu$ M)	0	nt	nt	nt	nt	nt	nt	nt
<b>35</b>	22 (32 $\mu$ M)	0 (>128 $\mu$ M)	0 (>128 $\mu$ M)	0	nt	0	0	nt	nt	nt	nt
<b>36</b>	24	15	0	23	0	11P	0	nt	nt	nt	nt
<b>37</b>	15p ( $\geq$ 128 $\mu$ M)	16p (>128 $\mu$ M)	0 (>128 $\mu$ M)	12P/h	0	13P	0	nt	nt	nt	nt
<b>38</b>	14P	13	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>39</b>	0 (>128 $\mu$ M)	0 (>128 $\mu$ M)	0 (>128 $\mu$ M)	0	0	0	0	nt	nt	nt	nt

<b>40a</b>	31 (2.0 $\mu$ M)	18P ( $>128 \mu$ M)	19.5P ( $>128 \mu$ M)	nt	nt	nt	nt	nt	nt	nt	nt
<b>S1</b>	0	0	0	0	0	0	0	0	nt	nt	nt
<b>S2</b>	0	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>S3</b>	0	0	0	0	0	0	0	0	nt	nt	nt
<b>S4</b>	0	0	0	0	0	0	0	0	nt	nt	nt
<b>S5</b>	0	0	0	0	0	0	0	0	nt	nt	nt
<b>S6</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S7</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S8</b>	0	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>S9</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S10</b>	11/15p/17P	10/15P	nt	12h	nt	nt	nt	nt	nt	nt	nt
<b>S11</b>	15.5	0	nt	0	nt	0	0	0	nt	nt	nt
<b>S12</b>	17P	9.5/11P	nt	0	nt	0	0	0	nt	nt	nt
<b>S13</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S14</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S15</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S16</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S17</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S18</b>	12h	0	nt	15P/h	nt	nt	nt	nt	nt	nt	nt
<b>S19</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S20</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S21</b>	29	10P	nt	14p	nt	nt	nt	nt	nt	nt	nt
<b>S22</b>	28	12p	nt	14p	nt	nt	nt	nt	nt	nt	nt
<b>S23</b>	25.5	9.5p	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S24</b>	26.5	0	nt	0	nt	nt	nt	nt	nt	nt	nt

<b>S25</b>	30	11/12p	nt	15p/18P	nt	0	0	nt	nt	nt	nt
<b>S26</b>	27	12.5p	nt	15p	nt	nt	nt	nt	nt	nt	nt
<b>S27</b>	23	0	nt	15P	nt	nt	nt	nt	nt	nt	nt
<b>S28</b>	25	12p	nt	14p	nt	nt	nt	nt	nt	nt	nt
<b>S29</b>	25	12h	nt	15P/h	nt	nt	nt	nt	nt	nt	nt
<b>S30</b>	20	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S31</b>	26	19h	nt	0	nt	0	0	nt	nt	nt	nt
<b>S32</b>	27.5	0	nt	12h	nt	nt	nt	nt	nt	nt	nt
<b>S33</b>	19	12P	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S34</b>	26.5	11p	nt	12P	nt	nt	nt	nt	nt	nt	nt
<b>S35</b>	26.5	15P	nt	13h	nt	nt	nt	nt	nt	nt	nt
<b>S36</b>	29	13p	nt	15p	nt	nt	nt	nt	nt	nt	nt
<b>S37</b>	30	11	nt	14p	nt	nt	nt	nt	nt	nt	nt
<b>S38</b>	28	0	nt	9.5	nt	0	0	nt	nt	nt	nt
<b>S39</b>	30	18	nt	16p	nt	nt	nt	nt	nt	nt	nt
<b>S40</b>	31	13	nt	15p/18P	nt	0	0	nt	nt	nt	nt
<b>S41</b>	30	11P	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S42</b>	27.5	10	nt	16p	nt	nt	nt	nt	nt	nt	nt
<b>S43</b>	27	13/18P	nt	18p	nt	nt	nt	nt	nt	nt	nt
<b>S44</b>	22	11	nt	12P	nt	nt	nt	nt	nt	nt	nt
<b>S45</b>	22	9.5	nt	16P	nt	nt	nt	nt	nt	nt	nt
<b>S46</b>	32	20P	nt	17p/P	nt	0	0	nt	nt	nt	nt
<b>S47</b>	29.5	20P	nt	13h	nt	nt	nt	nt	nt	nt	nt
<b>S48</b>	30	0	nt	12h	nt	nt	nt	nt	nt	nt	nt
<b>S49</b>	23	13P	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S50</b>	27	16P	nt	15P/h	nt	nt	nt	nt	nt	nt	nt

<b>S51</b>	0	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S52</b>	12p	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S53</b>	14	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S54</b>	0	0	0	nt	nt	nt	nt	nt	nt	nt	nt
<b>S55</b>	21	11.5	nt	15	nt	nt	nt	nt	nt	nt	nt
<b>S56</b>	29	11	nt	12/18p	nt	nt	nt	nt	nt	nt	nt
<b>S57</b>	23	0	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S58</b>	27	11	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>S59</b>	17	9.5	nt	0	nt	nt	nt	nt	nt	nt	nt
<b>cipro<sup>e</sup></b>	0 (8.0 $\mu$ M)	26 (0.5 $\mu$ M)	17 (2.0 $\mu$ M)	30	35	45	40	nt	nt	nt	nt
<b>TSA<sup>f</sup></b>	nt	nt	nt	nt	nt	nt	nt	nt	65% <sup>g</sup>	59% <sup>h</sup>	50% <sup>i</sup>

<sup>a</sup>A description of the agar diffusion antibacterial susceptibility assay is given in the main text along with a referenced literature protocol. Performed at UND.

<sup>b</sup>A description of the broth microdilution MIC<sub>90</sub> determination assay is given in the main text along with a referenced literature protocol. Performed at UND.

<sup>c</sup>A description of this assay has been reported by our group previously.<sup>5</sup> Performed at UND.

<sup>d</sup>Compounds are shown in chronological order beginning with compounds featured in the main article text followed by compounds only shown in the supporting information.

<sup>e</sup>Ciprofloxacin; used as a broad spectrum antibacterial standard at a concentration of 5  $\mu$ g/mL.

<sup>f</sup>Trichostatin A; used as a broad spectrum anticancer standard at concentrations indicated below.

<sup>g</sup>Percent inhibition at 25 nM TSA. <sup>h</sup>Percent inhibition at 200 nM TSA. <sup>i</sup>Percent inhibition at 150 nM TSA.

nt: indicates compound was not tested

h: indicates only a hint of growth inhibition detectable

P: indicates a very unclear growth inhibition zone

p: indicates unclear growth inhibition zone

s: indicates single colonies in the growth inhibition zone

### III. Table of All Cell Lines From This Work

<b>Table S3.</b> Origins and markers of cell lines used in this work.		
<b>Cell Line</b>	<b>Marker</b>	<b>Origin/Reference</b>
<b>Gram-positive bacteria</b>		
<i>Micrococcus luteus</i> ATCC 10240	wild type	American Type Culture Collection
<i>Staphylococcus aureus</i> SG511	wild type	Hans Knöll Institute, Jena, Germany
<i>Enterococcus faecalis</i> ATCC 49532	wild type	American Type Culture Collection
<i>Mycobacterium vaccae</i> IMET 10670	wild type	Hans Knöll Institute, Jena, Germany
<i>Bacillus subtilis</i> ATCC 6633	wild type	American Type Culture Collection
<i>Staphylococcus aureus</i> 134/94 (MRSA)	multidrug resistant	Witte et al. <b>1994</b> <sup>6</sup>
<i>Enterococcus faecalis</i> 1528 (VRE)	vancomycin resistant	Klare et al. <b>1995</b> <sup>7</sup>
<b>Gram-negative bacteria</b>		
<i>Pseudomonas aeruginosa</i> KW799/WT	wild Type	Zimmermann <b>1980</b> <sup>8</sup>
<i>Pseudomonas aeruginosa</i> KW799/61	antibiotic susceptible penetration mutant	Zimmermann <b>1980</b> <sup>8</sup>
<i>Escherichia coli</i> DC0	wild type	Richmond et al. <b>1976</b> <sup>9</sup>
<i>Escherichia coli</i> DC2	antibiotic susceptible penetration mutant	Richmond et al. <b>1976</b> <sup>9</sup>
<i>Escherichia coli</i> X580	$\beta$ -Lactam Hypersensitive	Eli Lilly & Co.
<b>Yeast</b>		
<i>Sporobolomyces</i> <i>salmonicolor</i> 549	wild type	Hans Knöll Institute, Jena, Germany
<b>Cancer cell lines</b>		
MCF-7	breast cancer	Soule et al. <b>1973</b> <sup>10</sup>
PC-3	prostate cancer	Alimerah et al. <b>2006</b> <sup>11</sup>
HeLa	cervical cancer	Rahbari et al. <b>2009</b> <sup>12</sup>

#### IV. Experimental Procedures and Compound Characterization Data

##### General Procedure A: Nitroso ene reaction.

This general procedure followed a method described previously.<sup>2,4</sup> The nitroso compound (1.0 equiv) was slowly added to a solution of the olefin (2.0 equiv) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (1.0 M with respect to olefin) at 0 °C (ice bath temp). Once complete (TLC analysis; hexanes/EtOAc), the CH<sub>2</sub>Cl<sub>2</sub> was evaporated under reduced pressure and the crude product was purified by silica gel column chromatography (hexanes/EtOAc) to give the desired nitroso ene adduct.

##### General Procedure B: Alkylation of BocNHOPMB (27).

Sodium hydride (60% in mineral oil; 1.2 equiv) solid was added to a solution of BocNHOPMB (27, 1.0 equiv) in anhydrous DMF (0.3–0.4 M with respect to 27) at 0 °C (ice bath temp). After stirring for 30 min, the alkyl halide (1.2 equiv) was added dropwise. The mixture was warmed to rt and stirred overnight under dry argon. The mixture was then cooled to 0 °C (ice bath temp) and quenched by slow addition of 10% aqueous NaHCO<sub>3</sub>. The DMF and H<sub>2</sub>O were removed by high vacuum rotary evaporation (~1 mm Hg). The resulting material was partitioned between EtOAc and H<sub>2</sub>O and the layers were separated. The H<sub>2</sub>O was extracted at least once with EtOAc. All the EtOAc layers were combined, washed with brine, dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated. The crude product was purified by silica gel column chromatography (hexanes/EtOAc) to give the desired *N*-alkylated product (28).

##### General Procedure C: *N*-Boc-deprotection of *N*-Boc-*N*-alkyl-*O*-(4-methoxybenzyl)hydroxylamines (28).

Anhydrous TFA was added to a solution of the *N*-Boc-*N*-alkyl-*O*-(4-methoxybenzyl)hydroxylamine (28, 1.0 equiv) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (20%–50% TFA v/v; ~0.3 M with respect to 28). After 15 min at rt, the TFA and CH<sub>2</sub>Cl<sub>2</sub> were removed under vacuum and the resulting material was dissolved in CHCl<sub>3</sub> and washed with saturated aqueous NaHCO<sub>3</sub>. The layers were separated and the CHCl<sub>3</sub> was dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated. The crude product was purified by silica gel column chromatography (hexanes/EtOAc) to give the desired *N*-alkyl-*O*-(4-methoxybenzyl)hydroxylamine (29).

##### General Procedure D: Buchwald-Hartwig amination cross-coupling reactions.

This procedure followed the general method described previously by Buchwald and coworkers.<sup>13</sup> The *N*-alkyl-*O*-(4-methoxybenzyl)hydroxylamine (29, 1.2 equiv), 2-bromopyridine (1.0 equiv), Pd<sub>2</sub>(dba)<sub>3</sub> (0.02 equiv), (±)-BINAP (0.04 equiv), and NaO<sup>t</sup>Bu (1.4 equiv) were dissolved in

anhydrous toluene (0.13 M with respect to the hydroxylamine), respectively, in an oven-dried, argon-flushed Schlenk tube. The mixture was heated to 70 °C (oil bath temp) over the course of the reaction. Once complete (TLC; hexanes/EtOAc), the mixture was cooled to rt, diluted with Et<sub>2</sub>O, and vacuum filtered through celite. The Et<sub>2</sub>O was washed once with brine, dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated. The crude product was purified by silica gel column chromatography (hexanes/EtOAc or hexanes/CH<sub>2</sub>Cl<sub>2</sub>) to give the desired *N*-alkyl-*O*-PMB-*N*-(pyridin-2-yl)hydroxylamine (**30**).

**General Procedure E: Removal of *O*-PMB protecting group.**

Anhydrous TFA was added slowly to a solution of the *N*-alkyl-*O*-PMB-*N*-(pyridin-2-yl)hydroxylamine (**30**, 1.0 equiv) and triethylsilane (2.0 equiv) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (30% TFA v/v; ~0.03 M with respect to **30**) and stirred at rt under dry argon. Once complete (TLC analysis; hexanes/EtOAc), the mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> and washed with saturated aqueous NaHCO<sub>3</sub>. The layers were separated and the CH<sub>2</sub>Cl<sub>2</sub> was dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated. The crude product was purified either by silica gel column chromatography (hexanes/EtOAc) or precipitation/trituration with cold pentanes to give the desired *N*-alkyl-*N*-(pyridin-2-yl)hydroxylamine (**31**).

(±) *N*-(6-Methylpyridin-2-yl)-*N*-(3-methylbut-3-en-2-yl)hydroxylamine (**9**). This compound was prepared according to general procedure **A** using 2-methyl-2-butene (62.4 mg, 0.89 mmol) and 5-methyl-2-nitrosopyridine **8** (53.7 mg, 0.44 mmol) in 1 mL of CH<sub>2</sub>Cl<sub>2</sub> for 20 min. Purification (0.5 x 6 in silica gel column; 17%–25% EtOAc in hexanes) provided the desired ene adduct (**9**) in 38% yield as a white solid (32.1 mg, 0.17 mmol). Mp 106–108 °C; <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.64 (s, N–OH, 1 H), 7.47 (dd, *J* = 8.2, 7.0 Hz, 1 H), 6.84 (d, *J* = 8.2 Hz, 1 H), 6.57 (d, *J* = 6.5 Hz, 1 H), 5.09 (dd, *J* = 13.2, 6.5 Hz, 1 H), 4.87 (m, 1 H), 4.84 (m, 1 H), 2.32 (s, 3 H), 1.72 (s, 3 H), 1.15 (d, *J* = 6.5 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 162.6, 155.3, 146.0, 137.8, 113.4, 111.6, 105.5, 58.1, 24.2, 21.5, 13.5; HRMS-ESI (*m/z*): [M+Na]<sup>+</sup> calcd. for C<sub>11</sub>H<sub>16</sub>N<sub>2</sub>NaO: 215.1155, found 215.1141; HPLC retention time 7.02 min.

(±) (*E*)-6-(*N*-Hydroxy-*N*-(pyridin-2-yl)amino)-3,7-dimethylocta-2,7-dien-1-ol (**11a**). This compound was prepared according to general procedure **A** using geraniol (137.0 mg, 0.89 mmol) and 2-nitrosopyridine **10** (47.6 mg, 0.44 mmol) in 1 mL of CH<sub>2</sub>Cl<sub>2</sub> for 25 min. Purification (0.5 x 6 in silica gel column; 50% EtOAc in hexanes) provided the desired ene adduct (**11a**) as a white solid (58.9 mg, 0.22 mmol) and a mixture of **11a** and **11b** as a yellow solid (23.1 mg, 0.09

mmol) in 71% overall yield. Mp 108–110 °C; <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.81 (s, N–OH, 1 H), 8.10 (ddd, *J* = 4.8, 1.9, 0.9 Hz, 1 H), 7.57 (ddd, *J* = 9.1, 7.0, 1.7 Hz, 1 H), 7.01 (dt, *J* = 8.3, 1.0, Hz, 1 H), 6.66 (ddd, *J* = 7.2, 4.8, 0.9 Hz, 1 H), 5.25–5.21 (m, 1 H), 4.94 (t, *J* = 6.9 Hz, 1 H), 4.86–4.84 (m, 1 H), 4.80 (t, *J* = 1.6 Hz, 1 H), 4.40 (t, *J* = 5.3 Hz, –OH, 1 H), 3.91 (t, *J* = 5.7 Hz, 2 H), 2.03–1.97 (m, 1 H), 1.91–1.74 (m, 3 H), 1.66 (s, 3 H), 1.56 (s, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 163.0, 146.9, 144.4, 137.4, 135.6, 125.1, 113.8, 112.6, 107.8, 62.6, 57.5, 27.2, 21.4, 16.1; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>23</sub>N<sub>2</sub>O<sub>2</sub>: 263.1754, found 263.1768; IR (neat): 3188.5 (br), 3113.2, 3070.2, 3022.4, 1664.7, 1500.9, 1470.1, 1296.4, 1017.0, 981.4 cm<sup>-1</sup>; HPLC retention time 6.18 min.

(±) ***N*-(3-Methylbut-3-en-2-yl)-*N*-(pyridin-2-yl)hydroxylamine (12)**. This compound was prepared according to general procedure **A** using 2-methyl-2-butene (62.4 mg, 0.89 mmol) and 2-nitrosopyridine **10** (47.6 mg, 0.44 mmol) in 1 mL of CH<sub>2</sub>Cl<sub>2</sub> for 15 min. Purification (0.5 x 6 in silica gel column; 17%–25% EtOAc in hexanes) provided the desired ene adduct (**12**) in 48% yield as a tan solid (37.6 mg, 0.21 mmol). Mp 67–70 °C; <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.73 (s, N–OH, 1 H), 8.13 (ddd, *J* = 4.8, 1.9, 0.9 Hz, 1 H), 7.59 (ddd, *J* = 8.5, 7.3, 2.0 Hz, 1 H), 7.04 (dt, *J* = 8.5, 0.9 Hz, 1 H), 6.70 (ddd, *J* = 7.2, 4.8, 1.2 Hz, 1 H), 5.07 (q, *J* = 6.7 Hz, 1 H), 4.88–4.87 (m, 2 H), 1.73 (s, 3 H), 1.16 (d, *J* = 6.7 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 162.9, 147.0, 145.8, 137.5, 114.2, 111.6, 108.5, 58.1, 21.4, 13.4; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>10</sub>H<sub>15</sub>N<sub>2</sub>O: 179.1179, found 179.1200; HPLC retention time 5.86 min.

(±) ***N*-(3-Methylbut-3-en-2-yl)-*N*-(quinolin-2-yl)hydroxylamine (14)**. This compound was prepared according to general procedure **A** using 2-methyl-2-butene (0.1 mL, 0.95 mmol) and 2-nitrosoquinoline **13** (50.0 mg, 0.32 mmol) in 2 mL of CH<sub>2</sub>Cl<sub>2</sub> for 1 h. Purification (0.75 x 6 in silica gel column; 10%–30% EtOAc in hexanes) provided the desired ene adduct (**14**) in 53% yield as an orange solid (38.0 mg, 0.17 mmol). Mp 83–85 °C; <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 9.12 (br s, N–OH, 1 H), 8.11 (d, *J* = 8.8 Hz, 1 H), 7.74 (dd, *J* = 7.9, 1.2 Hz, 1 H), 7.63–7.59 (m, 1 H), 7.57–7.53 (m, 1 H), 7.36 (d, *J* = 9.1 Hz, 1 H), 7.27 (ddd, *J* = 7.9, 6.8, 1.2 Hz, 1 H), 5.37 (q, *J* = 6.7 Hz, 1 H), 4.94 (s, 1 H), 4.90–4.88 (m, 1 H), 1.75 (s, 3 H), 1.27 (d, *J* = 6.7 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 160.8, 146.7, 145.6, 137.4, 129.4, 127.5, 126.3, 123.3, 122.4, 111.8, 110.7, 57.3, 21.4, 14.0; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>14</sub>H<sub>17</sub>N<sub>2</sub>O: 229.1335, found 229.1324; HPLC retention time 6.76 min.



(±) *N*-(5-Methylisoxazol-3-yl)-*N*-(3-methylbut-3-en-2-yl)hydroxylamine (**16**). This compound was prepared according to general procedure **A** using 2-methyl-2-butene (62.4 mg, 0.89 mmol) and 5-methyl-3-nitrosoisoxazole **15** (49.3 mg, 0.44 mmol) in 1 mL of CH<sub>2</sub>Cl<sub>2</sub> for 10 min. Purification (0.5 x 6 in silica gel column; 17%–25% EtOAc in hexanes) provided the desired ene adduct (**16**) in 94% yield as a white solid (75.4 mg, 0.41 mmol). Mp 80–82 °C; <sup>1</sup>H-NMR (500 MHz, DMSO-*d*<sub>6</sub>) δ 8.95 (s, N–OH, 1 H), 5.94 (m, 1 H), 4.91–4.88 (m, 1 H), 4.88–4.85 (m, 1 H), 4.09 (q, *J* = 6.5 Hz, 1 H), 2.28 (s, 3H), 1.75 (s, 3 H), 1.19 (d, *J* = 6.6 Hz, 1 H); <sup>13</sup>C-NMR (125 MHz, DMSO-*d*<sub>6</sub>) δ 169.6, 168.2, 144.5, 111.9, 94.6, 61.3, 20.7, 12.8, 11.7; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>9</sub>H<sub>15</sub>N<sub>2</sub>O<sub>2</sub>: 183.1128, found 183.1131; HPLC retention time 5.31 min.

(±) *N*-(3-Methylbut-3-en-2-yl)-*N*-(4-methylpyridin-2-yl)-hydroxylamine (**18b**). This compound was prepared according to general procedure **A** using 2-methyl-2-butene (0.42 mL, 3.97 mmol) and 4-methyl-2-nitrosopyridine **17b** (244.0 mg, 2.00 mmol) in 4 mL of CH<sub>2</sub>Cl<sub>2</sub> for 30 min. Purification (1.25 x 6 in silica gel column; 10%–20% EtOAc in hexanes) provided the desired ene adduct (**18b**) in 47% yield as a light brown solid (180.0 mg, 0.94 mmol). Mp 97.5–98.5 °C; <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.69 (br s, N–OH, 1 H), 7.99 (d, *J* = 5.0 Hz, 1 H), 6.88 (dt, *J* = 1.5, 0.8 Hz, 1 H), 6.56–6.54 (m, 1 H), 5.05 (q, *J* = 6.7 Hz, 1 H), 4.87–4.86 (m, 1 H), 4.86–4.84 (m, 1 H), 2.24 (s, 3 H), 1.74–1.71 (m, 3 H), 1.14 (d, *J* = 6.7 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 163.1, 147.9, 146.9, 146.0, 115.6, 111.6, 108.8, 58.3, 21.5, 20.9, 13.4; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>11</sub>H<sub>17</sub>N<sub>2</sub>O: 193.1335, found 193.1326; HPLC retention time 6.35 min.

(±) *N*-(3-Methylbut-3-en-2-yl)-*N*-(3-methylpyridin-2-yl)-hydroxylamine (**18c**). This compound was prepared according to general procedure **A** using 2-methyl-2-butene (0.42 mL, 3.97 mmol) and 3-methyl-2-nitrosopyridine **17c** (244.0 mg, 2.00 mmol) in 4 mL of CH<sub>2</sub>Cl<sub>2</sub> for 9 h. Purification (1.25 x 7 in silica gel column; 10%–30% EtOAc in hexanes) provided the desired ene adduct (**18c**) in 8% yield as a clear, colorless oil (30.0 mg, 0.16 mmol). <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.34 (br s, N–OH, 1 H), 8.09 (dd, *J* = 4.7, 1.5 Hz, 1 H), 7.48 (ddd, *J* = 7.3, 1.9, 0.7 Hz, 1 H), 6.94 (dd, *J* = 7.2, 4.8 Hz, 1 H), 4.89–4.87 (m, 1 H), 4.78–4.76 (m, 1 H), 4.43 (q, *J* = 6.5 Hz, 1 H), 2.31 (s, 3 H), 1.78 (s, 3 H), 1.16 (d, *J* = 6.5 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 160.4, 147.1, 144.1, 139.3, 126.1, 118.9, 111.1, 60.2, 20.5, 18.8, 15.0; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>11</sub>H<sub>17</sub>N<sub>2</sub>O: 193.1335, found 193.1335; HPLC retention time 6.75 min.

(±) ***N*-(5-Chloropyridin-2-yl)-*N*-(3-methylbut-3-en-2-yl)hydroxylamine (18d)**. This compound was prepared according to general procedure **A** using 2-methyl-2-butene (0.22 mL, 2.08 mmol) and 5-chloro-2-nitrosopyridine **17d** (141.0 mg, 0.99 mmol) in 2 mL of CH<sub>2</sub>Cl<sub>2</sub> for 30 min. Purification (0.75 x 6 in silica gel column; 5%–10% EtOAc in hexanes) provided the desired ene adduct (**18d**) in 59% yield as a tan solid (150.0 mg, 0.58 mmol). Mp 64.5–66 °C; <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.97 (s, N–OH, 1 H), 8.15 (dd, *J* = 2.6, 0.6 Hz, 1 H), 7.67 (dd, *J* = 8.8, 2.6 Hz, 1 H), 7.04 (dd, *J* = 8.9, 0.7 Hz, 1 H), 5.01 (q, *J* = 6.7 Hz, 1 H), 4.88–4.85 (m, 2 H), 1.71 (s, 3 H), 1.18 (d, *J* = 6.7 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 161.4, 145.4, 145.2, 137.3, 120.0, 111.9, 109.8, 58.3, 21.3, 13.7; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>10</sub>H<sub>14</sub>ClN<sub>2</sub>O: 213.0789, found 213.0806; HPLC retention time 7.99 min.

(±) ***N*-(5-Bromopyridin-2-yl)-*N*-(3-methylbut-3-en-2-yl)hydroxylamine (18e)**. This compound was prepared according to general procedure **A** using 2-methyl-2-butene (62.4 mg, 0.89 mmol) and 5-bromo-2-nitrosopyridine **17e** (82.3 mg, 0.44 mmol) in 1 mL of CH<sub>2</sub>Cl<sub>2</sub> for 10 min. Purification (0.5 x 6 in silica gel column; 17%–25% EtOAc in hexanes) provided the desired ene adduct (**18e**) in 64% yield as a faint yellow solid (72.4 mg, 0.28 mmol). Mp 65–67 °C; <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.98 (s, N–OH, 1 H), 8.20 (dd, *J* = 2.5, 0.7 Hz, 1 H), 7.76 (dd, *J* = 8.8, 2.6 Hz, 1 H), 6.99 (dd, *J* = 8.9, 0.7 Hz, 1 H), 5.01 (q, *J* = 6.7 Hz, 1 H), 4.87–4.85 (m, 2 H), 1.70 (s, 3 H), 1.18 (d, *J* = 6.7 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 161.5, 147.4, 145.3, 139.9, 111.9, 110.4, 108.0, 58.2, 21.3, 13.7; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>10</sub>H<sub>14</sub>BrN<sub>2</sub>O: 257.0284, found 257.0290; HPLC retention time 8.21 min.

(±) ***N*-(5-Iodopyridin-2-yl)-*N*-(3-methylbut-3-en-2-yl)hydroxylamine (18f)**. This compound was prepared according to general procedure **A** using 2-methyl-2-butene (0.10 mL, 0.95 mmol) and 5-iodo-2-nitrosopyridine **17f** (68.0 mg, 0.29 mmol) in 2 mL of CH<sub>2</sub>Cl<sub>2</sub> for 1 h. Purification (0.75 x 8 in silica gel column; 1%–15% EtOAc in hexanes) provided the desired ene adduct (**18f**) in 68% yield as a grey solid (60.0 mg, 0.20 mmol). Mp 88.5–90 °C; <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.95 (s, N–OH, 1 H), 8.29 (dd, *J* = 2.3, 0.6 Hz, 1 H), 7.86 (dd, *J* = 8.8, 2.3 Hz, 1 H), 6.91 (dd, *J* = 8.8, 0.6 Hz, 1 H), 5.00 (q, *J* = 6.7 Hz, 1 H), 4.87–4.85 (m, 2 H), 1.70 (s, 3 H), 1.17 (d, *J* = 6.7 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 161.7, 152.3, 145.4, 145.1, 111.9, 111.1, 79.3, 58.1, 21.3, 13.7; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>10</sub>H<sub>14</sub>IN<sub>2</sub>O: 305.0145, found 305.0145; HPLC retention time 8.56 min.

(±) *N*-(3,5-Dichloropyridin-2-yl)-*N*-(3-methylbut-3-en-2-yl)hydroxylamine (**18g**). This compound was prepared according to general procedure **A** using 2-methyl-2-butene (0.22 mL, 2.08 mmol) and 3,5-dichloro-2-nitrosopyridine **17g** (176.0 mg, 0.99 mmol) in 2 mL of CH<sub>2</sub>Cl<sub>2</sub> for 1 h. Purification (0.75 x 6 in silica gel column; 5%–10% EtOAc in hexanes) provided the desired ene adduct (**18g**) in 67% yield as a clear, colorless oil (165.0 mg, 0.67 mmol). <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.82 (s, N–OH, 1 H), 8.29 (d, *J* = 2.3 Hz, 1 H), 8.05 (d, *J* = 2.3 Hz, 1 H), 4.87–4.85 (m, 1 H), 4.80–4.78 (m, 1 H), 4.41 (q, *J* = 6.6 Hz, 1 H), 1.77 (s, 3 H), 1.21 (d, *J* = 6.5 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 157.0, 146.1, 143.6, 138.3, 124.7, 123.0, 111.7, 60.6, 20.6, 14.9; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>10</sub>H<sub>13</sub>Cl<sub>2</sub>N<sub>2</sub>O: 247.0399, found 247.0427; HPLC retention time 8.63 min.

(±) *N*-(5-Bromo-6-methylpyridin-2-yl)-*N*-(3-methylbut-3-en-2-yl)hydroxylamine (**18h**). This compound was prepared according to general procedure **A** using 2-methyl-2-butene (0.21 mL, 1.99 mmol) and 5-bromo-6-methyl-2-nitrosopyridine **17h** (201.0 mg, 1.00 mmol) in 2 mL of CH<sub>2</sub>Cl<sub>2</sub> for 30 min. Purification (1 x 6 in silica gel column; 5%–20% EtOAc in hexanes) provided the desired ene adduct (**18h**) in 62% yield as a faint orange solid (169.0 mg, 0.62 mmol). Mp 65–67 °C; <sup>1</sup>H-NMR (500 MHz, DMSO-*d*<sub>6</sub>) δ 8.90 (s, N–OH, 1 H), 7.71 (d, *J* = 8.8 Hz, 1 H), 6.81 (d, *J* = 8.8 Hz, 1 H), 5.04 (q, *J* = 6.5 Hz, 1 H), 4.88–4.86 (m, 1 H), 4.86–4.83 (m, 1 H), 2.42 (s, 3 H), 1.70 (s, 3 H), 1.17 (d, *J* = 6.8 Hz, 3 H); <sup>13</sup>C-NMR (125 MHz, DMSO-*d*<sub>6</sub>) δ 161.3, 153.2, 145.5, 141.0, 111.9, 108.3, 108.1, 58.2, 24.6, 21.4, 13.8; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>11</sub>H<sub>16</sub>BrN<sub>2</sub>O: 271.0441; found, 271.0432; HPLC retention time 9.27 min.

(±) *N*-(3-Chloro-5-(trifluoromethyl)pyridin-2-yl)-*N*-(3-methylbut-3-en-2-yl)hydroxylamine (**18i**). This compound was prepared according to general procedure **A** using 2-methyl-2-butene (0.11 mL, 1.00 mmol) and 3-chloro-5-(trifluoromethyl)-2-nitrosopyridine **17i** (105.0 mg, 0.50 mmol) in 2 mL of CH<sub>2</sub>Cl<sub>2</sub> for 45 min. Purification (0.75 x 8 in silica gel column; 5%–10% EtOAc in hexanes) provided the desired ene adduct (**18i**) in 75% yield as a faint purple solid (105.6 mg, 0.38 mmol). Mp 39–40 °C; <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 9.21 (s, 1 H), 8.52–8.50 (m, 1 H), 8.13–8.10 (m, 1 H), 4.92–4.90 (m, 1 H), 4.88–4.86 (m, 1 H), 4.84 (q, *J* = 6.5 Hz, 1 H), 1.76 (s, 3 H), 1.27 (d, *J* = 6.8 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 159.6, 145.2, 142.4 (q, *J*<sub>C-F</sub> = 4.3 Hz), 136.6 (q, *J*<sub>C-F</sub> = 3.2 Hz), 123.4 (q, *J*<sub>C-F</sub> = 271.5 Hz), 118.8, 118.3 (q, *J*<sub>C-F</sub> = 33.1 Hz), 112.1, 59.2, 21.1, 14.2; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>11</sub>H<sub>13</sub>ClF<sub>3</sub>N<sub>2</sub>O: 281.0663, found 281.0677; HPLC retention time 9.16 min.

***N*-(2,3-Dimethylbut-3-en-2-yl)-*N*-(pyridin-2-yl)hydroxylamine (19a).** This compound was prepared according to general procedure **A** using 2,3-dimethyl-2-butene (0.11 mL, 1.0 mmol) and 2-nitrosopyridine **10** (50.0 mg, 0.46 mmol) in 2 mL of CH<sub>2</sub>Cl<sub>2</sub> for 30 min. Purification (0.5 x 6 in silica gel column; 5%–10% EtOAc in hexanes) provided the desired ene adduct (**19a**) in 71% yield as a brown oil (63.2 mg, 0.33 mmol). <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.94 (s, N-OH, 1 H), 8.09–8.06 (m, 1 H), 7.57 (ddd, *J* = 8.5, 7.3, 2.1 Hz, 1 H), 7.12 (dt, *J* = 8.2, 0.9 Hz, 1 H), 6.74 (ddd, *J* = 7.2, 4.8, 1.2 Hz, 1 H), 4.74–4.73 (m, 1 H), 4.63–4.61 (m, 1 H), 1.79 (d, *J* = 0.6 Hz, 3 H), 1.35 (s, 6 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 163.1, 152.3, 146.3, 136.9, 115.3, 111.6, 107.4, 65.8, 24.7, 20.0; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>11</sub>H<sub>17</sub>N<sub>2</sub>O: 193.1335, found 193.1340; HPLC retention time 6.81 min.

***N*-(2-Phenylallyl)-*N*-(pyridin-2-yl)hydroxylamine (19b).** This compound was prepared according to general procedure **A** using 1-(prop-1-en-2-yl)benzene (109.0 mg, 0.92 mmol) and 2-nitrosopyridine **10** (50.0 mg, 0.46 mmol) in 2 mL of CH<sub>2</sub>Cl<sub>2</sub> for 3 h. Purification (0.5 x 6 in silica gel column; 5%–10% EtOAc in hexanes) provided the desired ene adduct (**19b**) in 6% yield as a yellow oil (6.3 mg, 0.03 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.24 (d, *J* = 4.7 Hz, 1 H), 7.60–7.54 (m, 3 H), 7.34 (t, *J* = 7.5 Hz, 2 H), 7.29 (d, *J* = 7.3 Hz, 1 H), 7.09 (d, *J* = 8.5 Hz, 1 H), 6.82–6.78 (m, 1 H), 5.59 (s, 1 H), 5.40 (s, 1 H), 4.75 (s, 2 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 161.6, 146.5, 143.3, 138.9, 137.9, 128.3, 127.8, 126.3, 115.8, 115.5, 109.3, 58.6; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>14</sub>H<sub>15</sub>N<sub>2</sub>O: 227.1179, found 227.1186; HPLC retention time, 7.39 min.

**5-(*N*-Hydroxy-*N*-(pyridin-2-yl)amino)-6-methylhept-6-en-2-one (19c).** This compound was prepared according to general procedure **A** using 6-methylhept-5-en-2-one (0.30 mL, 2.00 mmol) and 2-nitrosopyridine **10** (108.1 mg, 1.00 mmol) in 2 mL of CH<sub>2</sub>Cl<sub>2</sub> for 10 min. Purification (1 x 5 in silica gel column; 33% EtOAc in hexanes) provided the desired ene adduct (**19c**) in 94% yield as a light brown, crystalline solid (220.0 mg, 0.94 mmol). Mp 66–70 °C; <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.82 (s, N-OH, 1 H), 8.10–8.07 (m, 1 H), 7.61–7.54 (m, 1 H), 7.01 (d, *J* = 8.5 Hz, 1 H), 6.68–6.64 (m, 1 H), 4.94 (t, *J* = 7.5 Hz, 1 H), 4.85–4.78 (m, 2 H), 2.48–2.38 (m, 2 H), 2.00 (s, 3 H), 1.94–1.87 (m, 2 H), 1.66–1.60 (m, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ 208.2, 163.0, 147.0, 144.5, 137.6, 113.9, 112.6, 107.6, 61.8, 40.0, 29.8, 23.0, 21.4; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>13</sub>H<sub>19</sub>N<sub>2</sub>O<sub>2</sub>: 235.1441, found 235.1444; IR (neat): 3091.7 (br), 3052.8, 1713.6, 1593.3, 1430.0, 1163.5 cm<sup>-1</sup>; HPLC retention time 5.88 min.

***N*-Boc-*O*-(4-Methoxybenzyl)-*N*-methylhydroxylamine (28a).** This compound was prepared according to general procedure **B** using BocNHOPBM (2.0 g, 7.90 mmol), sodium hydride (60% in mineral oil; 415.0 mg, 10.38 mmol), and methyl iodide (0.59 mL, 9.46 mmol) in 20 mL of DMF for 26 h. Purification (1.5 x 6 in silica gel column; 100% hexanes–10% EtOAc in hexanes) provided the desired product (**28a**) in 91% yield as a clear, colorless liquid (1.91 g, 7.15 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.35–7.32 (m, 2 H), 6.90–6.87 (m, 2 H), 4.77 (s, 2 H), 3.81 (s, 3 H), 3.03 (s, 3 H), 1.50 (s, 9 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 159.8, 156.9, 131.0, 127.8, 113.7, 81.1, 76.0, 55.2, 36.7, 28.2; HRMS-ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>14</sub>H<sub>21</sub>NNaO<sub>4</sub>: 290.1363, found 290.1361.

***N*-Boc-*O*-(4-Methoxybenzyl)-*N*-(prop-1-yl)hydroxylamine (28b).** This compound was prepared according to general procedure **B** using BocNHOPBM (446.2 mg, 1.76 mmol), sodium hydride (60% in mineral oil; 102.0 mg, 2.55 mmol), and 1-bromopropane (0.19 mL, 2.09 mmol) in 6 mL of DMF for 5 h. Purification (1 x 6 in silica gel column; 10% EtOAc in hexanes) provided the desired product (**28b**) in 98% yield as a clear, colorless liquid (512.0 mg, 1.73 mmol). <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 7.33 (d, *J* = 8.6 Hz, 2 H), 6.88 (d, *J* = 8.8 Hz, 2 H), 4.76 (s, 2 H), 3.80 (s, 3 H), 3.41–3.33 (m, 2 H), 1.70–1.56 (m, 2 H), 1.51 (s, 9 H), 0.89 (t, *J* = 7.5 Hz, 3 H); <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 159.7, 156.5, 130.9, 127.8, 113.7, 80.9, 76.3, 55.2, 51.1, 28.2, 20.3, 11.2; HRMS-ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>25</sub>NNaO<sub>4</sub>: 318.1676, found 318.1687.

***N*-Boc-*N*-(hex-1-yl)-*O*-(4-Methoxybenzyl)hydroxylamine (28c).** This compound was prepared according to general procedure **B** using BocNHOPBM (550.0 mg, 2.17 mmol), sodium hydride (60% in mineral oil; 140.0 mg, 3.50 mmol), and 1-bromohexane (0.37 mL, 2.64 mmol) in 5 mL of DMF for 14 h. Purification (1 x 6 in silica gel column; 5%–20% EtOAc in hexanes) provided the desired product (**28c**) in 97% yield as a clear, colorless liquid (713.0 mg, 2.11 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.35–7.31 (m, 2 H), 6.90–6.86 (m, 2 H), 4.76 (s, 2 H), 3.80 (s, 3 H), 3.40–3.37 (m, 2 H), 1.62–1.55 (m, 2 H), 1.51 (s, 9 H), 1.33–1.24 (m, 6 H), 0.90–0.86 (m, 3 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 159.7, 156.5, 130.9, 127.8, 113.7, 80.9, 76.3, 55.2, 49.5, 31.4, 28.3, 26.9, 26.3, 22.5, 14.0; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>19</sub>H<sub>32</sub>NO<sub>4</sub>: 338.2326, found 338.2325.

***N*-Boc-*O*-(4-Methoxybenzyl)-*N*-(phenethyl)hydroxylamine (28e).** This compound was prepared according to general procedure **B** using BocNHOPBM (513.0 mg, 2.03 mmol), sodium

hydride (60% in mineral oil; 120.0 mg, 3.00 mmol), and (2-bromoethyl)benzene (0.33 mL, 2.42 mmol) in 5 mL of DMF for 14 h. Purification (1 x 6 in silica gel column; 5%–20% EtOAc in hexanes) provided the desired product (**28e**) in 98% yield as a clear, colorless liquid (712.0 mg, 1.99 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.38–7.35 (m, 2 H), 7.33–7.29 (m, 2 H), 7.25–7.21 (m, 3 H), 6.94–6.91 (m, 2 H), 4.79 (s, 2 H), 3.83 (s, 3 H), 3.66–3.62 (m, 2 H), 2.93–2.89 (m, 2 H), 1.49 (s, 9 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 159.7, 156.2, 138.9, 130.9, 128.8, 128.3, 127.8, 126.1, 113.7, 81.0, 76.4, 55.2, 51.2, 33.3, 28.2; HRMS-ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>21</sub>H<sub>27</sub>NNaO<sub>4</sub>: 380.1832, found 380.1833.

**N-Allyl-N-Boc-O-(4-methoxybenzyl)hydroxylamine (28f).** This compound was prepared according to general procedure **B** using BocNHOPBM (810.0 mg, 3.20 mmol), sodium hydride (60% in mineral oil; 180.0 mg, 4.50 mmol), and allyl bromide (0.33 mL, 3.90 mmol) in 8 mL of DMF for 10 h. Purification (1 x 6 in silica gel column; 10%–20% EtOAc in hexanes) provided the desired product (**28f**) in 87% yield as a clear, colorless liquid (815.0 mg, 2.78 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.34–7.31 (m, 2 H), 6.90–6.87 (m, 2 H), 5.91–5.83 (m, 1 H), 5.23 (dq, *J* = 17.3, 1.5 Hz, 1 H), 5.20–5.17 (m, 1 H), 4.78 (s, 2 H), 3.99 (dt, *J* = 6.2, 1.3 Hz, 2 H), 3.81 (s, 3 H), 1.51 (s, 9 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 159.8, 156.6, 132.8, 131.0, 127.8, 118.0, 113.7, 81.3, 76.7, 55.2, 52.8, 28.3; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>24</sub>NO<sub>4</sub>: 294.1700, found 294.1705.

**N-Boc-O-(4-Methoxybenzyl)-N-(3-phenoxybenzyl)hydroxylamine (28g).** This compound was prepared according general procedure **B** using BocNHOPBM (600.0 mg, 2.37 mmol), sodium hydride (60% in mineral oil; 125.0 mg, 3.13 mmol), and 3-phenoxybenzyl bromide (515.0 mg, 1.96 mmol) in 8 mL of DMF for 24 h. Purification (1.25 x 6 in silica gel column; 10% EtOAc in hexanes) provided the desired product (**28g**) in 50% yield as a clear, colorless liquid (518.1 mg, 1.19 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.35–7.31 (m, 2 H), 7.30 (t, *J* = 7.9 Hz, 1 H), 7.22–7.19 (m, 2 H), 7.11 (ddd, *J* = 7.5, 1.0, 0.9 Hz, 1 H), 7.10–7.07 (m, 1 H), 7.03–6.99 (m, 3 H), 6.95–6.93 (m, 1 H), 6.87–6.83 (m, 2 H), 4.66 (s, 2 H), 4.50 (s, 2 H), 3.81 (s, 3 H), 1.50 (s, 9 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 159.8, 157.3, 157.1, 156.5, 139.0, 131.0, 129.7, 129.6, 127.7, 123.4, 123.2, 119.0, 118.9, 117.9, 113.7, 81.6, 76.9, 55.2, 53.6, 28.3; HRMS-ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>26</sub>H<sub>29</sub>NNaO<sub>5</sub>: 458.1938, found 458.1942.

**N-Boc-O-(4-Methoxybenzyl)-N-(4-(trifluoromethyl)benzyl)hydroxylamine (28h).** This compound was prepared according to general procedure **B** using BocNHOPBM (798.0 mg, 3.15

mmol), sodium hydride (60% in mineral oil; 180.0 mg, 4.50 mmol), and 4-(trifluoromethyl)benzyl bromide (1.05 g, 4.39 mmol) in 10 mL of DMF for 22 h. Purification (1.5 x 5 in silica gel column; 5%–20% EtOAc in hexanes) provided the desired product (**28h**) in 98% yield as a clear, colorless liquid (1.26 g, 3.07 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.58 (d, *J* = 7.9 Hz, 2 H), 7.42 (d, *J* = 7.9 Hz, 2 H), 7.22–7.18 (m, 2 H), 6.87–6.84 (m, 2 H), 4.68 (s, 2 H), 4.57 (s, 2 H), 3.81 (s, 3 H), 1.51 (s, 9 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 159.9, 156.6, 141.0, 131.0, 129.8 (q, *J*<sub>C-F</sub> = 32.5 Hz), 128.7, 127.5, 125.3 (q, *J*<sub>C-F</sub> = 3.9 Hz), 123.2, 113.8, 81.9, 76.9, 55.2, 53.5, 28.3; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>21</sub>H<sub>25</sub>F<sub>3</sub>NO<sub>4</sub>: 412.1730, found 412.1738.

***N*-Boc-*O*-(4-Methoxybenzyl)-*N*-(4-(trifluoromethoxy)benzyl)hydroxylamine (28i).** This compound was prepared according to general procedure **B** using BocNHOPBM (816.0 mg, 3.22 mmol), sodium hydride (60% in mineral oil; 195.0 mg, 4.87 mmol), and 4-(trifluoromethoxy)benzyl bromide (0.72 mL, 4.50 mmol) in 10 mL of DMF for 22 h. Purification (1.5 x 5 in silica gel column; 5%–30% EtOAc in hexanes) provided the desired product (**28i**) in 86% yield as a clear, colorless liquid (1.18 g, 2.77 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.36–7.33 (m, 2 H), 7.21–7.18 (m, 2 H), 7.17 (dd, *J* = 8.8, 0.9 Hz, 2 H), 6.87–6.84 (m, 2 H), 4.66 (s, 2 H), 4.51 (s, 2 H), 3.81 (s, 3 H), 1.51 (s, 9 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 159.8, 156.6, 148.57 (q, *J*<sub>C-F</sub> = 1.7 Hz) 135.7, 131.0, 130.1, 127.5, 120.9, 120.4 (q, *J*<sub>C-F</sub> = 256.9 Hz), 113.8, 81.8, 76.9, 55.2, 53.1, 28.3; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>21</sub>H<sub>25</sub>F<sub>3</sub>NO<sub>5</sub>: 428.1679, found 428.1676.

***O*-(4-Methoxybenzyl)-*N*-methylhydroxylamine (29a).** This compound was prepared according to general procedure **C** using compound **28a** (754.0 mg, 2.82 mmol) in 8 mL of TFA:CH<sub>2</sub>Cl<sub>2</sub> (1:4) for 15 min. The desired compound (**29a**) was isolated in 99% crude yield as a clear, colorless liquid (465.0 mg, 2.78 mmol) and was used directly in the next reaction without purification or characterization.

***O*-(4-Methoxybenzyl)-*N*-(prop-1-yl)hydroxylamine (29b).** This compound was prepared according to general procedure **C** using compound **28b** (702 mg, 2.38 mmol) in 8 mL of TFA:CH<sub>2</sub>Cl<sub>2</sub> (1:1) for 15 min. The desired compound (**29b**) was isolated in 99% crude yield as a clear, colorless liquid (463.0 mg, 2.37 mmol) and was used directly in the next reaction without purification or characterization.

***N*-(Hex-1-yl)-*O*-(4-methoxybenzyl)hydroxylamine (29c).** This compound was prepared according to general procedure **C** using compound **28c** (1.00 g, 2.96 mmol) in 5 mL of

TFA:CH<sub>2</sub>Cl<sub>2</sub> (1:1) for 15 min. Purification (1 x 6 in silica gel column; 10%–25% EtOAc in hexanes) provided the desired product (**29c**) in 77% yield as a clear, colorless liquid (541.3 mg, 2.28 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.32–7.29 (m, 2 H), 6.91–6.87 (m, 2 H), 4.66 (s, 2 H), 3.81 (s, 3 H), 2.95–2.91 (m, 2 H), 1.54–1.48 (m, 2 H), 1.35–1.25 (m, 6 H), 0.91–0.87 (m, 3 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 159.3, 130.0, 129.9, 113.7, 75.8, 55.2, 52.1, 31.7, 27.2, 26.9, 22.6, 14.0; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>14</sub>H<sub>24</sub>NO<sub>2</sub>: 238.1802, found 238.1802.

**O-(4-Methoxybenzyl)-N-(phenethyl-2-yl)hydroxylamine (29e).** This compound was prepared according to general procedure C using compound **28e** (639.8 mg, 1.79 mmol) in 10 mL of TFA:CH<sub>2</sub>Cl<sub>2</sub> (3:7) for 15 min. Purification (0.75 x 5 in silica gel column; 5%–25% EtOAc in hexanes) provided the desired product (**29e**) in 73% yield as a clear, colorless liquid (336.1 mg, 1.31 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.33–7.28 (m, 4 H), 7.24–7.20 (m, 3 H), 6.92–6.88 (m, 2 H), 5.54 (br s, NH, 1 H), 4.67 (s, 2 H), 3.82 (s, 3 H), 3.19 (t, *J* = 7.2 Hz, 2 H), 2.86 (t, *J* = 7.2 Hz, 2 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 159.3, 139.5, 130.0, 129.9, 128.8, 128.5, 126.2, 113.7, 75.7, 55.3, 53.2, 33.7; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>20</sub>NO<sub>2</sub>: 258.1489, found 258.1489.

**N-Allyl-O-(4-methoxybenzyl)hydroxylamine (29f).** This compound was prepared according to general procedure C using compound **28f** (528.0 mg, 1.80 mmol) in 15 mL of TFA:CH<sub>2</sub>Cl<sub>2</sub> (1:5) for 15 min. Purification (0.75 x 6 in silica gel column; 10%–20% EtOAc in hexanes) provided the desired product (**29f**) in 58% yield as a clear, colorless liquid (200.0 mg, 1.04 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.32–7.28 (m, 2 H), 6.91–6.87 (m, 2 H), 5.96–5.88 (m, 1 H), 5.50 (br s, NH, 1 H), 5.24 (dq, *J* = 17.2, 1.6 Hz, 1 H), 5.19–5.15 (m, 1 H), 4.66 (s, 2 H), 3.82 (s, 3 H), 3.54 (ddd, *J* = 6.3, 1.5, 1.3 Hz, 2 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 159.3, 134.3, 130.0, 129.9, 117.8, 113.7, 75.8, 55.3, 55.0; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>11</sub>H<sub>16</sub>NO<sub>2</sub>: 194.1176, found 194.1177.

**O-(4-Methoxybenzyl)-N-(3-phenoxybenzyl)hydroxylamine (29g).** This compound was prepared according to general procedure C using compound **28g** (196.0 mg, 0.45 mmol) in 8 mL of TFA:CH<sub>2</sub>Cl<sub>2</sub> (1:7) for 15 min. Purification (1 x 6 in silica gel column; 10%–20% EtOAc in hexanes) provided the desired product (**29g**) in 70% yield as a clear, colorless liquid (105.0 mg, 0.31 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 7.37–7.33 (m, 2 H), 7.30 (t, *J* = 7.9 Hz, 1 H), 7.25–7.22 (m, 2 H), 7.14–7.10 (m, 1 H), 7.10–7.08 (m, 1 H), 7.04–7.03 (m, 2 H), 7.03–7.01 (m, 1 H), 6.94 (ddd, *J* = 8.1, 2.6, 1.0 Hz, 1 H), 6.88–6.85 (m, 2 H), 5.69 (br s, NH, 1 H), 4.58 (s, 2 H), 4.01



(s, 2 H), 3.81 (s, 3 H);  $^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  159.3, 157.3, 157.1, 139.8, 130.1, 129.8, 129.7, 129.6, 123.7, 123.2, 119.2, 118.9, 117.8, 113.7, 75.9, 56.1, 55.2; HRMS-ESI (m/z):  $[\text{M}+\text{H}]^+$  calcd. for  $\text{C}_{21}\text{H}_{22}\text{NO}_3$ : 336.1594, found 336.1599.

***O*-(4-Methoxybenzyl)-*N*-(4-(trifluoromethyl)benzyl)hydroxylamine (29h).** This compound was prepared according to general procedure **C** using compound **28h** (672.0 mg, 1.63 mmol) in 6 mL of TFA: $\text{CH}_2\text{Cl}_2$  (1:5) for 15 min. Purification (1.25 x 5 in silica gel column; 10%–20% EtOAc in hexanes) provided the desired product (**29h**) in 62% yield as an off-white solid (317.0 mg, 1.02 mmol). Mp 34.5–36.5 °C;  $^1\text{H}$ -NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.59 (d,  $J = 7.9$  Hz, 2 H), 7.46 (d,  $J = 7.9$  Hz, 2 H), 7.23–7.20 (m, 2 H), 6.88–6.85 (m, 2 H), 5.75 (br s, NH, 1 H), 4.57 (s, 2 H), 4.08 (s, 2 H), 3.81 (s, 3 H);  $^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  159.4, 142.0, 130.1, 129.6, 129.1, 125.2 (q,  $J_{\text{C-F}} = 3.9$  Hz), 113.7, 76.0, 55.9, 55.2; HRMS-ESI (m/z):  $[\text{M}+\text{H}]^+$  calcd. for  $\text{C}_{16}\text{H}_{17}\text{F}_3\text{NO}_2$ : 312.1206, found 312.1190.

***O*-(4-Methoxybenzyl)-*N*-(4-(trifluoromethoxy)benzyl)hydroxylamine (29i).** This compound was prepared according to general procedure **C** using compound **28i** (607.0 mg, 1.42 mmol) in 6 mL of TFA: $\text{CH}_2\text{Cl}_2$  (1:5) for 15 min. Purification (1.25 x 6 in silica gel column; 10%–20% EtOAc in hexanes) provided the desired product (**29i**) in 71% yield as an off-white solid (331.0 mg, 1.01 mmol). Mp 28–30 °C;  $^1\text{H}$ -NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.39–7.36 (m, 2 H), 7.23–7.20 (m, 2 H), 7.20–7.17 (m, 2 H), 6.88–6.85 (m, 2 H), 5.70 (br s, NH, 1 H), 4.57 (s, 2 H), 4.03 (s, 2 H), 3.81 (s, 3 H);  $^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  159.4, 136.6, 130.2, 130.1, 129.7, 120.8 (q,  $J_{\text{C-F}} = 1.12$  Hz), 113.7, 76.0, 55.6, 55.2; HRMS-ESI (m/z):  $[\text{M}+\text{H}]^+$  calcd. for  $\text{C}_{16}\text{H}_{17}\text{F}_3\text{NO}_3$ : 328.1155, found 328.1142.

***O*-(4-Methoxybenzyl)-*N*-methyl-*N*-(pyridin-2-yl)hydroxylamine (30a).** This compound was prepared according to general procedure **D** using crude compound **29a** (200.6 mg, 1.20 mmol), 2-bromopyridine (0.10 mL, 1.00 mmol),  $\text{Pd}_2(\text{dba})_3$  (18.0 mg, 0.02 mmol), ( $\pm$ ) BINAP (25.0 mg, 0.04 mmol), and  $\text{NaO}^t\text{Bu}$  (134.0 mg, 1.40 mmol) in 9 mL of toluene for 96 h. Purification (1.25 x 6 in silica gel column; 10%–20% EtOAc in hexanes) provided the desired product (**30a**) in 77% yield as a clear, colorless liquid (187.7 mg, 0.77 mmol).  $^1\text{H}$ -NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  8.25 (ddd,  $J = 4.8, 1.9, 0.9$  Hz, 1 H), 7.55 (ddd,  $J = 8.6, 7.0, 1.8$  Hz, 1 H), 7.41–7.38 (m, 2 H), 7.08 (dt,  $J = 8.3, 1.0$  Hz, 1 H), 6.94–6.91 (m, 2 H), 6.79 (ddd,  $J = 7.3, 4.9, 1.0$  Hz, 1 H), 4.85 (s, 3 H), 3.83 (s, 3 H), 3.26 (s, 3 H);  $^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  163.1, 159.7, 147.6, 137.6,

130.7, 128.4, 116.1, 113.9, 109.3, 75.4, 55.3, 40.0; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>14</sub>H<sub>17</sub>N<sub>2</sub>O<sub>2</sub>: 245.1285, found 245.1289; HPLC retention time 7.86 min.

***O*-(4-Methoxybenzyl)-*N*-(prop-1-yl)-*N*-(pyridin-2-yl)hydroxylamine (30b).** This compound was prepared according to general procedure **D** using compound **29b** (230.0 mg, 1.18 mmol), 2-bromopyridine (0.10 mL, 1.00 mmol), Pd<sub>2</sub>(dba)<sub>3</sub> (18.0 mg, 0.02 mmol), (±) BINAP (25.0 mg, 0.04 mmol), and NaO<sup>t</sup>Bu (134.0 mg, 1.40 mmol) in 9 mL of toluene for 72 h. Purification (1 x 6 in silica gel column; 5%–20% EtOAc in hexanes) provided the desired product (**30b**) in 51% yield as a clear, colorless liquid (140.0 mg, 0.51 mmol). <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 8.27 (ddd, *J* = 4.9, 1.7, 0.8 Hz, 1 H), 7.56 (ddd, *J* = 8.3, 7.2, 1.9 Hz, 1 H), 7.39 (d, *J* = 8.6 Hz, 2 H), 7.08 (d, *J* = 8.3 Hz, 1 H), 6.93 (d, *J* = 8.6 Hz, 2 H), 6.77 (ddd, *J* = 7.3, 4.9, 0.8 Hz, 1 H), 4.82 (s, 2 H), 3.83 (s, 3 H), 3.71–3.63 (m, 2 H), 1.80–1.66 (m, 2 H), 0.96 (t, *J* = 7.3 Hz, 3 H); <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 162.7, 159.7, 147.8, 137.7, 130.6, 128.3, 115.8, 113.9, 109.1, 75.8, 55.3, 55.2, 19.6, 11.6; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>21</sub>N<sub>2</sub>O<sub>2</sub>: 273.1598, found 273.1588; HPLC retention time 9.25 min.

***N*-(Hex-1-yl)-*O*-(4-methoxybenzyl)-*N*-(pyridin-2-yl)hydroxylamine (30c).** This compound was prepared according to general procedure **D** using crude compound **29c** (245.0 mg, 1.03 mmol), 2-bromopyridine (135.6 mg, 0.86 mmol), Pd<sub>2</sub>(dba)<sub>3</sub> (15.7 mg, 0.017 mmol), (±) BINAP (21.4 mg, 0.034 mmol), and NaO<sup>t</sup>Bu (115.5 mg, 1.20 mmol) in 8 mL of toluene for 72 h. Purification (1 x 8 in silica gel column; 5%–10% EtOAc in hexanes) provided the desired product (**30c**) in 40% yield as a clear, colorless liquid (107.3 mg, 0.34 mmol). <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>) δ 8.28–8.24 (m, 1 H), 7.56 (ddd, *J* = 8.6, 7.0, 1.9 Hz, 1 H), 7.38 (d, *J* = 8.4 Hz, 2 H), 7.09–7.04 (m, 1 H), 6.92 (d, *J* = 8.4 Hz, 2 H), 6.79–6.74 (m, 1 H), 4.81 (s, 2 H), 3.83 (s, 3 H), 3.71–3.66 (m, 2 H), 1.71–1.62 (m, 2 H), 1.39–1.23 (m, 8 H), 0.91–0.85 (m, 3 H); <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>) δ 162.7, 159.7, 147.8, 137.7, 130.6, 128.4, 115.8, 113.9, 109.1, 75.8, 55.3, 53.5, 31.8, 26.9, 26.2, 22.6, 14.1; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>19</sub>H<sub>27</sub>N<sub>2</sub>O<sub>2</sub>: 315.2067, found 315.2058; HPLC retention time 11.41 min.

***O*-(4-Methoxybenzyl)-*N*-(pheneth-2-yl)-*N*-(pyridin-2-yl)hydroxylamine (30e).** This compound was prepared according to general procedure **D** using compound **29e** (308.8 mg, 1.20 mmol), 2-bromopyridine (0.10 mL, 1.00 mmol), Pd<sub>2</sub>(dba)<sub>3</sub> (18.0 mg, 0.02 mmol), (±) BINAP (25.0 mg, 0.04 mmol), and NaO<sup>t</sup>Bu (134.0 mg, 1.40 mmol) in 9 mL of toluene for 60 h. Purification (1 x 6 in silica gel column; 5%–20% EtOAc in hexanes) provided the desired

product (**30e**) in 49% yield as a clear, colorless liquid (163.8 mg, 0.49 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.30–8.28 (m, 1H), 7.57 (ddd, *J* = 8.4, 7.2, 2.0 Hz, 1 H), 7.40–7.36 (m, 2 H), 7.31–7.24 (m, 5 H), 7.22–7.18 (m, 1 H), 7.11–7.08 (m, 1 H), 6.95–6.91 (m, 2 H), 6.81–6.78 (m, 1 H), 4.82 (s, 2 H), 3.99–3.93 (m, 2 H), 3.84 (s, 3 H), 2.99–2.92 (m, 2 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 162.2, 159.7, 147.9, 139.9, 137.7, 130.7, 128.9, 128.3, 126.0, 116.0, 113.9, 109.3, 75.8, 55.3, 54.5, 32.3; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>21</sub>H<sub>23</sub>N<sub>2</sub>O<sub>2</sub>: 335.1754, found 335.1755; HPLC retention time 10.40 min.

***N*-Allyl-*O*-(4-methoxybenzyl)-*N*-(pyridin-2-yl)hydroxylamine (30f).** This compound was prepared according to general procedure **D** using compound **29f** (198.8 mg, 1.03 mmol), 2-bromopyridine (130.7 mg, 0.83 mmol), Pd<sub>2</sub>(dba)<sub>3</sub> (15.2 mg, 0.017 mmol), (±) BINAP (20.7 mg, 0.033 mmol), and NaO<sup>*t*</sup>Bu (111.7 mg, 1.16 mmol) in 7.5 mL of toluene for 66 h. Purification (1 x 6 in silica gel column; 5%–20% EtOAc in hexanes) provided the desired product (**30f**) in 54% yield as a clear, colorless liquid (120.8 mg, 0.45 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.26 (ddd, *J* = 4.8, 1.9, 0.9 Hz, 1 H), 7.59–7.54 (m, 1 H), 7.39–7.36 (m, 2 H), 7.09 (dt, *J* = 8.5, 0.9 Hz, 1 H), 6.93–6.90 (m, 2 H), 6.79 (ddd, *J* = 7.3, 4.8, 1.0 Hz, 1 H), 6.05–5.97 (m, 1 H), 5.32–5.27 (m, 1 H), 5.20 (dddd, *J* = 10.3, 1.9, 1.2, 1.0 Hz, 1 H), 4.83 (s, 2 H), 4.34 (dt, *J* = 6.5, 1.2 Hz, 3 H), 3.83 (s, 3 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 162.2, 159.7, 147.7, 137.8, 133.7, 130.7, 128.3, 118.4, 116.2, 113.8, 109.3, 76.0, 56.0, 55.3; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>19</sub>N<sub>2</sub>O<sub>2</sub>: 271.1441, found 271.1436; HPLC retention time 8.82 min.

***O*-(4-Methoxybenzyl)-*N*-(3-phenoxybenzyl)-*N*-(pyridin-2-yl)hydroxylamine (30g).** This compound was prepared according to general procedure **D** using compound **29g** (138.7 mg, 0.41 mmol), 2-bromopyridine (54.4 mg, 0.34 mmol), Pd<sub>2</sub>(dba)<sub>3</sub> (6.3 mg, 0.007 mmol), (±) BINAP (8.58 mg, 0.014 mmol), and NaO<sup>*t*</sup>Bu (46.4 mg, 0.48 mmol) in 3.1 mL of toluene for 70 h. Purification (1 x 8 in silica gel column; 5%–30% EtOAc in hexanes) provided the desired product (**30g**) in 59% yield as a clear, colorless liquid (84.2 mg, 0.20 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.27 (ddd, *J* = 4.8, 1.9, 0.9 Hz, 1 H), 7.59–7.55 (m, 1 H), 7.33–7.27 (m, 3 H), 7.22–7.19 (m, 2 H), 7.18–7.16 (m, 1 H), 7.11 - 7.06 (m, 3 H), 6.99–6.96 (m, 2 H), 6.95 (ddd, *J* = 8.1, 2.5, 1.2 Hz, 1 H), 6.88–6.85 (m, 2 H), 6.81 (ddd, *J* = 7.2, 4.8, 0.9 Hz, 1 H), 4.83 (s, 2 H), 4.62 (s, 2 H), 3.82 (s, 3 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 162.2, 159.7, 157.2, 157.0, 147.7, 139.7, 137.9, 130.7, 129.7, 129.4, 128.0, 124.5, 123.1, 120.0, 118.8, 117.8, 116.4, 113.8, 109.2,

76.2, 57.1, 55.3; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>26</sub>H<sub>25</sub>N<sub>2</sub>O<sub>3</sub>: 413.1860, found 413.1865; HPLC retention time 11.18 min.

***O*-(4-Methoxybenzyl)-*N*-(4-(trifluoromethyl)benzyl)-*N*-(pyridin-2-yl)hydroxylamine (30h).**

This compound was prepared according to general procedure **D** using compound **29h** (310.0 mg, 1.00 mmol), 2-bromopyridine (131.1 mg, 0.83 mmol), Pd<sub>2</sub>(dba)<sub>3</sub> (15.2 mg, 0.017 mmol), (±) BINAP (20.7 mg, 0.033 mmol), and NaO<sup>t</sup>Bu (111.7 mg, 1.16 mmol) in 7.5 mL of toluene for 96 h. Purification (1 x 8 in silica gel column; 5%–30% EtOAc in hexanes) provided the desired product (**30h**) in 56% yield as a clear, colorless liquid (180.0 mg, 0.46 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.30 (ddd, *J* = 5.0, 1.8, 0.9 Hz, 1 H), 7.60 (ddd, *J* = 8.4, 7.2, 2.0 Hz, 1 H), 7.58–7.54 (m, 2 H), 7.52–7.49 (m, 2 H), 7.20–7.17 (m, 2 H), 7.12–7.09 (m, 1 H), 6.88–6.83 (m, 3 H), 4.91 (s, 2 H), 4.64 (s, 2 H), 3.81 (s, 3 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 162.0, 159.7, 147.8, 141.8, 138.0, 130.7, 129.6, 129.35 (q, *J*<sub>C-F</sub> = 32.5 Hz), 127.9, 125.0 (q, *J*<sub>C-F</sub> = 3.9 Hz), 123.3, 116.6, 113.8, 109.2, 76.2, 56.7, 55.2; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>21</sub>H<sub>20</sub>F<sub>3</sub>N<sub>2</sub>O<sub>2</sub>: 389.1471, found 389.1458; HPLC retention time 10.67 min.

***O*-(4-Methoxybenzyl)-*N*-(4-(trifluoromethoxy)benzyl)-*N*-(pyridin-2-yl)hydroxylamine (30i).**

This compound was prepared according to general procedure **D** using compound **29i** (290.0 mg, 0.89 mmol), 2-bromopyridine (116.7 mg, 0.74 mmol), Pd<sub>2</sub>(dba)<sub>3</sub> (13.5 mg, 0.015 mmol), (±) BINAP (18.4 mg, 0.029 mmol), and NaO<sup>t</sup>Bu (99.3 mg, 1.03 mmol) in 6 mL of toluene for 96 h. Purification (1 x 8 in silica gel column; 5%–10% EtOAc in hexanes) provided the desired product (**30i**) in 52% yield as a clear, colorless liquid (155.0 mg, 0.04 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.31 (ddd, *J* = 4.8, 1.9, 0.9 Hz, 1 H), 7.59 (ddd, *J* = 8.5, 7.2, 1.9 Hz, 1 H), 7.46–7.41 (m, 2 H), 7.19–7.15 (m, 4 H), 7.10 (dt, *J* = 8.3, 1.0 Hz, 1 H), 6.88–6.85 (m, 2 H), 6.84 (ddd, *J* = 7.3, 4.8, 1.0 Hz, 1 H), 4.85 (s, 2 H), 4.61 (s, 2 H), 3.82 (s, 3 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 162.1, 159.8, 148.4 (q, *J*<sub>C-F</sub> = 1.9 Hz), 147.8, 138.0, 136.4, 130.9, 130.7, 127.9, 120.6, 120.5 (q, *J*<sub>C-F</sub> = 256.9 Hz), 116.6, 113.8, 109.2, 76.2, 56.5, 55.2; HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>21</sub>H<sub>20</sub>F<sub>3</sub>N<sub>2</sub>O<sub>3</sub>: 405.1421, found 405.1425; HPLC retention time 10.86 min.

***N*-Methyl-*N*-(pyridin-2-yl)hydroxylamine (31a).** This compound was prepared according to general procedure **E** using compound **30a** (93.0 mg, 0.38 mmol) and triethylsilane (0.18 mL, 1.13 mmol) in 6 mL of TFA:CH<sub>2</sub>Cl<sub>2</sub> (30% TFA v/v) for 7 h. Purification (0.5 x 5 in silica gel column; 17%–33% EtOAc in hexanes) provided the desired product (**31a**) in 55% yield as a viscous faint yellow oil (26.0 mg, 0.21 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.24 (d, *J* = 4.1

Hz, 1 H), 7.60 (t,  $J = 8.2$  Hz, 1 H), 7.02 (d,  $J = 8.5$  Hz, 1 H), 6.84–6.82 (m, 1 H), 3.30 (s, 3 H);  $^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  162.9, 146.8, 137.7, 116.1, 109.3, 42.8; HRMS-ESI ( $m/z$ ):  $[\text{M}+\text{H}]^+$  calcd. for  $\text{C}_6\text{H}_9\text{N}_2\text{O}$ : 125.0709, found 125.0696; HPLC retention time 2.78 min.

***N*-(Prop-1-yl)-*N*-(pyridin-2-yl)hydroxylamine (31b).** This compound was prepared according to general procedure **E** using compound **30b** (100.0 mg, 0.37 mmol) and triethylsilane (0.15 mL, 0.94 mmol) in 10 mL of TFA: $\text{CH}_2\text{Cl}_2$  (30% TFA v/v) for 5 h. Purification (0.75 in x 6 in silica gel column; 20% EtOAc in hexanes) provided the desired product (**31b**) in 75% yield as a clear, colorless oil (42.0 mg, 0.28 mmol).  $^1\text{H}$ -NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  8.22 (d,  $J = 4.1$  Hz, 1 H), 7.57 (t,  $J = 7.2$  Hz, 1 H), 7.00 (d,  $J = 8.5$  Hz, 1 H), 6.81–6.76 (m, 1 H), 3.56 (t,  $J = 7.3$  Hz, 2 H), 1.76 (dq,  $J = 14.7, 7.3$  Hz, 2 H), 0.99 (t,  $J = 7.3$  Hz, 3 H);  $^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  162.1, 146.7, 137.6, 115.7, 109.3, 56.9, 19.6, 11.5; HRMS-ESI ( $m/z$ ):  $[\text{M}+\text{H}]^+$  calcd. for  $\text{C}_8\text{H}_{13}\text{N}_2\text{O}$ : 153.1022, found 153.1031. HPLC retention time 4.44 min.

***N*-(Hex-1-yl)-*N*-(pyridin-2-yl)hydroxylamine (31c).** This compound was prepared according to general procedure **E** using compound **30c** (35.9 mg, 0.11 mmol) and triethylsilane (0.12 mL, 0.75 mmol) in 6 mL of TFA: $\text{CH}_2\text{Cl}_2$  (30% TFA v/v) for 5 h. Purification (0.5 x 4 in silica gel column; 20% EtOAc in hexanes) provided the desired product (**31c**) in 69% yield as a clear, faint yellow oil (15.2 mg, 0.08 mmol).  $^1\text{H}$ -NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  8.20 (d,  $J = 3.2$  Hz, 1 H), 7.56 (t,  $J = 7.5$  Hz, 1 H), 6.99 (d,  $J = 8.2$  Hz, 1 H), 6.77 (t,  $J = 5.3$  Hz, 1 H), 3.58 (t,  $J = 7.1$  Hz, 2 H), 1.75–1.65 (m, 2 H), 1.41–1.24 (m, 6 H), 0.91–0.85 (m, 3 H);  $^{13}\text{C}$ -NMR (125 MHz,  $\text{CDCl}_3$ )  $\delta$  162.2, 146.7, 137.7, 115.6, 109.4, 55.2, 31.7, 26.7, 26.2, 22.6, 14.0; HRMS-ESI ( $m/z$ ):  $[\text{M}+\text{H}]^+$  calcd. for  $\text{C}_{11}\text{H}_{19}\text{N}_2\text{O}$ : 195.1492, found 195.1473; HPLC retention time 7.82 min.

***N*-(Pheneth-2-yl)-*N*-(pyridin-2-yl)hydroxylamine (31e).** This compound was prepared according to general procedure **E** using compound **30e** (105.0 mg, 0.31 mmol) and triethylsilane (0.10 mL, 0.63 mmol) in 10 mL of TFA: $\text{CH}_2\text{Cl}_2$  (30% TFA v/v) for 20 h. Purification (0.75 x 6 in silica gel column; 20%–25% EtOAc in hexanes) provided the desired product (**31e**) in 73% yield as a viscous yellow oil (49.0 mg, 0.23 mmol).  $^1\text{H}$ -NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  8.24 (d,  $J = 4.1$  Hz, 1 H), 7.60–7.54 (m, 1 H), 7.33–7.24 (m, 5 H), 7.24–7.19 (m, 1 H), 7.00 (d,  $J = 8.2$  Hz, 1 H), 6.83–6.77 (m, 1 H), 3.92–3.85 (m, 2 H), 3.06–2.99 (m, 2 H);  $^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  161.8, 146.8, 139.6, 137.7, 128.9, 128.4, 126.2, 115.9, 109.4, 56.5, 32.5; HRMS-ESI ( $m/z$ ):  $[\text{M}+\text{H}]^+$  calcd. for  $\text{C}_{13}\text{H}_{15}\text{N}_2\text{O}$ : 215.1179, found 215.1192; HPLC retention time 6.95 min.

***N*-Allyl-*N*-(pyridin-2-yl)hydroxylamine (31f).** This compound was prepared according to general procedure **E** using compound **30f** (80.0 mg, 0.30 mmol) and triethylsilane (0.10 mL, 0.63 mmol) in 10 mL of TFA:CH<sub>2</sub>Cl<sub>2</sub> (30% TFA v/v) for 5 h. Purification (0.75 x 6 in silica gel column; 20%–25% EtOAc in hexanes) provided the desired product (**31f**) in 45% yield as a faint yellow oil (20.0 mg, 0.13 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.25–8.23 (m, 1 H), 7.60 (ddd, *J* = 8.6, 7.0, 1.8 Hz, 1 H), 7.07 (dt, *J* = 8.5, 0.9 Hz, 1 H), 6.82 (ddd, *J* = 7.0, 5.0, 0.9 Hz, 1 H), 6.02–5.94 (m, 1 H), 5.34 (dq, *J* = 17.2, 1.5 Hz, 1 H), 5.27–5.23 (m, 1 H), 4.29 (dt, *J* = 6.2, 1.3 Hz, 3 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 161.8, 146.8, 137.8, 132.9, 118.9, 116.2, 109.7, 57.6; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>8</sub>H<sub>11</sub>N<sub>2</sub>O: 151.0866, found 151.0880; HPLC retention time 4.18 min.

***N*-(3-Phenoxybenzyl)-*N*-(pyridin-2-yl)hydroxylamine (31g).** This compound was prepared according to general procedure **E** using compound **30g** (35.6 mg, 0.086 mmol) and triethylsilane (0.05 mL, 0.31 mmol) in 6 mL of TFA:CH<sub>2</sub>Cl<sub>2</sub> (17% TFA v/v) for 7 h. Purification (0.5 x 4.5 in silica gel column; 10%–20% EtOAc in hexanes) provided the desired product (**31g**) in 60% yield as a white solid (15.1 mg, 0.052 mmol). Mp 81–83 °C; <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.22 (d, *J* = 4.7 Hz, 1 H), 7.62–7.57 (m, 1 H), 7.35–7.31 (m, 2 H), 7.29 (t, *J* = 7.8 Hz, 1 H), 7.16–7.09 (m, 3 H), 7.08–7.06 (m, 1 H), 7.00–6.97 (m, 2 H), 6.93 (dd, *J* = 8.2, 2.3 Hz, 1 H), 6.83 (dd, *J* = 7.0, 5.0 Hz, 1 H), 4.81 (s, 2 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 162.0, 157.3, 157.0, 147.0, 139.1, 137.8, 129.7, 129.6, 123.7, 123.2, 119.2, 118.9, 117.8, 116.4, 109.7, 58.5; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>18</sub>H<sub>17</sub>N<sub>2</sub>O<sub>2</sub>: 293.1285, found 293.1293; HPLC retention time 8.55 min.

***N*-(4-(Trifluoromethyl)benzyl)-*N*-(pyridin-2-yl)hydroxylamine (31h).** This compound was prepared according to general procedure **E** using compound **30h** (50.0 mg, 0.13 mmol) and triethylsilane (0.05 mL, 0.31 mmol) in 6 mL of TFA:CH<sub>2</sub>Cl<sub>2</sub> (30% TFA v/v) for 7 h. The desired product (**31h**) was precipitated by trituration with cold pentanes and isolated in 81% yield as an off-white solid (28.0 mg, 0.10 mmol). Mp 101–102 °C; <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.24 (dd, *J* = 4.8, 1.0 Hz, 1 H), 7.65–7.61 (m, 1 H), 7.59 (d, *J* = 8.2 Hz, 2 H), 7.52 (d, *J* = 8.2 Hz, 2 H), 7.14 (d, *J* = 8.5 Hz, 1 H), 6.86 (dd, *J* = 6.9, 5.1 Hz, 1 H), 4.88 (s, 2 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 161.9, 147.0, 141.3, 138.0, 129.7 (q, *J*<sub>C-F</sub> = 32.5 Hz), 129.1, 125.3 (q, *J*<sub>C-F</sub> = 3.9 Hz), 123.3, 116.7, 109.6, 58.3; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>13</sub>H<sub>12</sub>F<sub>3</sub>N<sub>2</sub>O: 269.0896, found 269.0892; HPLC retention time 8.19 min.

***N*-(4-(Trifluoromethoxy)benzyl)-*N*-(pyridin-2-yl)hydroxylamine (31i).** This compound was prepared according to general procedure **E** using compound **30i** (108.6 mg, 0.27 mmol) and triethylsilane (0.10 mL, 0.63 mmol) in 9 mL of TFA:CH<sub>2</sub>Cl<sub>2</sub> (30% TFA v/v) for 19 h. Purification (0.75 x 6 in silica gel column; 10%–20% EtOAc in hexanes) provided the desired product (**31i**) in 70% yield as an off-white, crystalline solid (53.0 mg, 0.19 mmol). Mp 101.5–103.5 °C; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>) δ ppm 8.20 (d, *J* = 4.8 Hz, 1 H), 7.64–7.57 (m, 1 H), 7.42 (d, *J* = 8.4 Hz, 2 H), 7.19–7.11 (m, 3 H), 6.84 (dd, *J* = 7.1, 5.1 Hz, 1 H), 4.80 (s, 2 H); <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>) δ ppm 162.1, 148.5 (q, *J*<sub>C-F</sub> = 1.6 Hz), 147.0, 138.0, 135.9, 130.3, 120.8, 120.4 (q, *J*<sub>C-F</sub> = 256.7 Hz), 116.5, 109.7, 58.0; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>13</sub>H<sub>12</sub>F<sub>3</sub>N<sub>2</sub>O<sub>2</sub>: 285.0845, found 285.0834; HPLC retention time 7.68 min.

(±) ***O*-Acetyl-*N*-(5-bromopyridin-2-yl)-*N*-((1*S*/*R*,4*S*/*R*)-2-methoxycyclohex-3-enyl)hydroxylamine (35).** Compound **7b** (22.0 mg, 0.074 mmol), Ac<sub>2</sub>O (9.0 mg, 0.09 mmol), *i*Pr<sub>2</sub>EtN (14.2 mg, 0.11 mmol), and catalytic DMAP (2.0 mg, 0.016 mmol) were dissolved in 4 mL of anhydrous CH<sub>2</sub>Cl<sub>2</sub>, respectively. After 9h, TLC (20% EtOAc in hexanes) showed complete consumption of the starting hydroxylamine compound. The CH<sub>2</sub>Cl<sub>2</sub> was evaporated and the crude product was purified by silica gel column chromatography (0.5 x 6 in silica gel; 10%–25% EtOAc in hexanes) to give the desired product (**35**) in 94% yield as a clear, colorless oil (23.5 mg, 0.07 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.33 (d, *J* = 2.0 Hz, 1 H), 7.66 (dd, *J* = 8.8, 2.3 Hz, 1 H), 6.68–6.64 (m, 1 H), 5.85–5.80 (m, 1 H), 5.78–5.70 (m, 1 H), 4.71–4.65 (m, 1 H), 4.12 (br s, 1 H), 3.39 (br s, 3 H), 2.28 (s, 3 H), 2.27–2.20 (m, 1 H), 2.19–2.12 (m, 1 H); <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 169.5, 159.3, 148.5, 140.1, 129.4, 126.5, 112.0, 110.5, 75.6, 62.1, 55.5, 29.7, 25.3, 18.7; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>14</sub>H<sub>18</sub>BrN<sub>2</sub>O<sub>3</sub>: 341.0495, found 341.0512; IR (neat): 3028.7, 1786.6, 1577.8, 1458.7, 1371.0, 1181.8, 1098.1, 1000.6 cm<sup>-1</sup>; HPLC retention time 7.93 min.

***N*-Benzyl-*O*-(4-methoxybenzyl)-*N*-(pyridin-3-yl)hydroxylamine (36).** This compound was prepared according to general procedure **D** using compound **29d** (279.0 mg, 1.15 mmol), 3-bromopyridine (158.0 mg, 1.00 mmol), Pd<sub>2</sub>(dba)<sub>3</sub> (18.0 mg, 0.02 mmol), (±) BINAP (25.0 mg, 0.04 mmol), and NaO<sup>t</sup>Bu (134.0 mg, 1.40 mmol) in 9 mL of toluene for 67 h. Purification (1.25 x 8 in silica gel column; 10%–50% EtOAc in hexanes) provided the desired product (**36**) in 36% yield as a clear, yellow liquid (115 mg, 0.36 mmol). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.49 (d, *J* = 2.6 Hz, 1 H), 8.25 (dd, *J* = 4.7, 1.5 Hz, 1 H), 7.45–7.41 (m, 3 H), 7.38–7.31 (m, 3 H), 7.20 (ddd,

$J = 8.4, 4.7, 0.7$  Hz, 1 H), 7.13–7.10 (m, 2 H), 6.85–6.82 (m, 2 H), 4.51 (s, 2 H), 4.42 (s, 2 H), 3.80 (s, 3 H);  $^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )  $\delta$  159.6, 147.4, 143.7, 139.6, 136.5, 130.7, 129.4, 128.3, 128.0, 127.7, 124.0, 123.3, 113.7, 75.8, 63.2, 55.2; HRMS-ESI ( $m/z$ ):  $[\text{M}+\text{H}]^+$  calcd. for  $\text{C}_{20}\text{H}_{21}\text{N}_2\text{O}_2$ : 321.1598, found 321.1618; IR (neat): 3060.7, 3031.9, 3005.0, 1611.5, 1572.5, 1513.1, 1246.8, 1030.2, 985.1  $\text{cm}^{-1}$ ; HPLC retention time 8.70 min.

***N*-Benzyl-*N*-(pyridin-3-yl)hydroxylamine (37).** This compound was prepared according to general procedure **E** using compound **36** (49.5 mg, 0.15 mmol) and triethylsilane (0.05 mL, 0.31 mmol) in 7 mL of TFA: $\text{CH}_2\text{Cl}_2$  (15% TFA v/v) for 30 h. Purification (0.5 x 4 in silica gel column; 30%–50% EtOAc in hexanes) provided the desired product (**37**) in 45% yield as an off-white solid (14.0 mg, 0.07 mmol). Mp 126–130 °C;  $^1\text{H-NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  8.49–8.45 (m, 1 H), 8.16–8.11 (m, 1 H), 7.52 (dt,  $J = 8.4, 1.2$  Hz, 1 H), 7.44–7.39 (m, 2 H), 7.36 (t,  $J = 7.2$  Hz, 2 H), 7.34–7.30 (m, 1 H), 7.21 (dd,  $J = 8.2, 4.7$  Hz, 1 H), 6.69 (br s, N–OH, 1 H), 4.45 (s, 2 H);  $^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )  $\delta$  148.7, 142.7, 138.5, 136.6, 129.0, 128.5, 127.7, 123.4, 123.3, 63.3; HRMS-ESI ( $m/z$ ):  $[\text{M}+\text{H}]^+$  calcd. for  $\text{C}_{12}\text{H}_{13}\text{N}_2\text{O}$ : 201.1022, found 201.1026; IR (neat) 3088.0 (br), 3062.3, 3033.1, 1572.5, 1487.0, 1422.5, 1232.0, 1189.8, 1062.9, 1002.8  $\text{cm}^{-1}$ ; HPLC retention time 5.32 min.

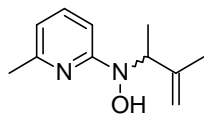
***N*-Benzyl-*N*-hydroxy(pyridin-2-yl)methanamine (39).** This compound was prepared according to general procedure **E** using compound **38** (15.0 mg, 0.045 mmol) and triethylsilane (0.025 mL, 0.16 mmol) in 3.5 mL of TFA: $\text{CH}_2\text{Cl}_2$  (15% TFA v/v) for 5 h. The desired product (**39**) was precipitated by trituration with cold pentanes and isolated in 73% yield as an off-white solid (7.0 mg, 0.033 mmol). Mp 80–82 °C;  $^1\text{H-NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  8.60–8.57 (m, 1 H), 7.69 (td,  $J = 7.6, 1.8$  Hz, 1 H), 7.45–7.41 (m, 2 H), 7.38–7.33 (m, 3 H), 7.32–7.28 (m, 1 H), 7.22 (ddd,  $J = 7.6, 4.9, 1.0$  Hz, 1 H), 4.09 (s, 2 H), 4.04 (s, 2 H);  $^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )  $\delta$  157.7, 148.9, 136.8, 129.6, 128.4, 127.5, 123.3, 122.4, 64.3, 64.0; HRMS-ESI ( $m/z$ ):  $[\text{M}+\text{H}]^+$  calcd. for  $\text{C}_{13}\text{H}_{15}\text{N}_2\text{O}$ : 215.1179, found 215.1161; IR (neat): 3144.6 (br), 3063.9, 3031.5, 1597.7, 1435.1, 1117.4, 1031.5  $\text{cm}^{-1}$ ; HPLC retention time 5.13 min.

**(±) (*E*)-6-(*N*-Hydroxy-*N*-(6-ethylpyridin-2-yl)amino)-3,7-dimethylocta-2,7-dien-1-ol (40a).** This compound was prepared according to general procedure **A** using geraniol (453.2 mg, 2.94 mmol) and 6-ethyl-2-nitrosopyridine **10** (200.0 mg, 1.46 mmol) in 5 mL of  $\text{CH}_2\text{Cl}_2$  for 25 min. Purification (1 x 6 in silica gel column; 10%–25% EtOAc in hexanes) provided a mixture of isomers (**40a:40b**; 70:30) in 75% yield as a yellow oil (321.0 mg, 1.10 mmol). Further



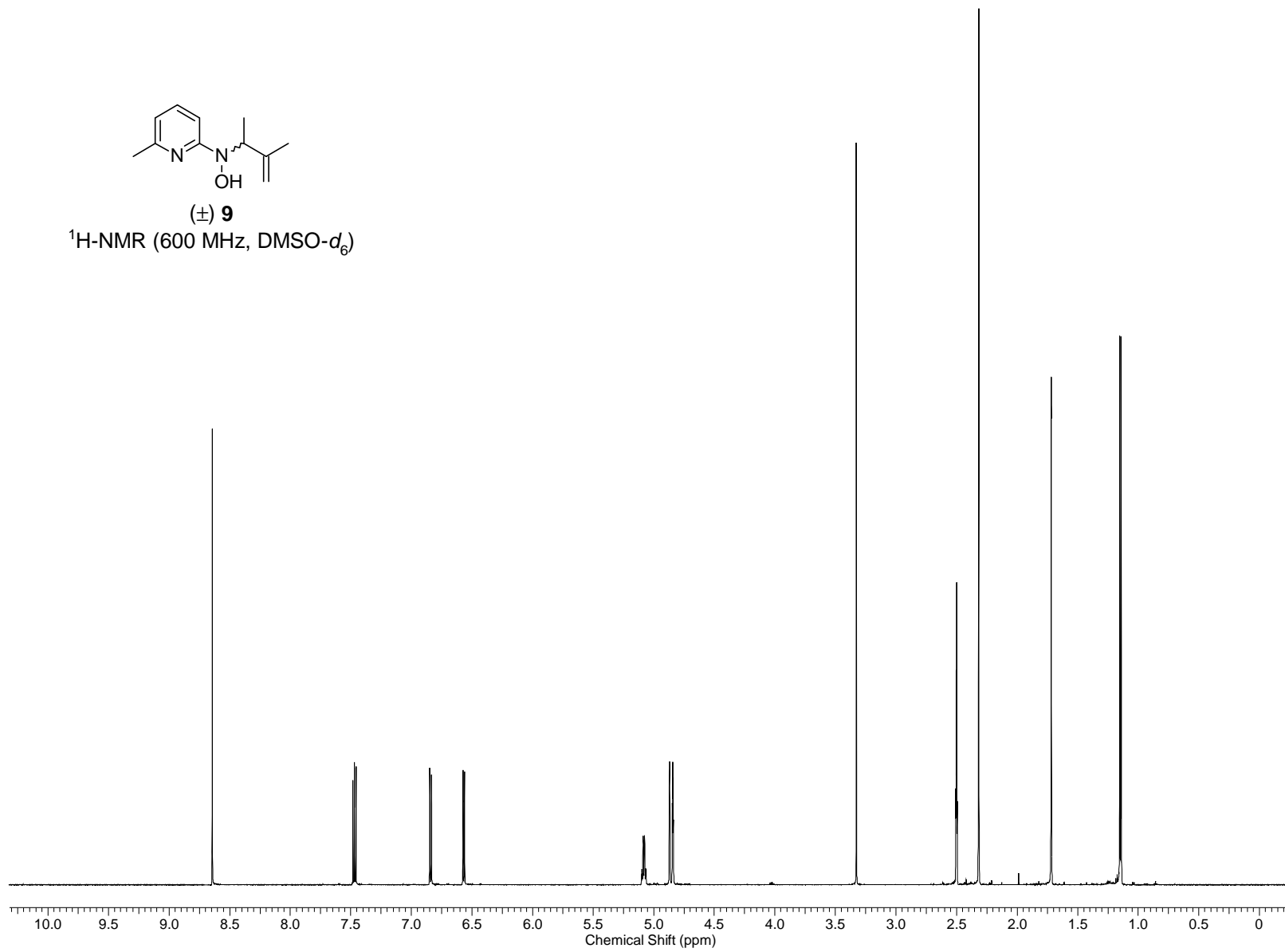
purification (1 x 6 in silica gel; 33%–50% EtOAc in hexanes) provided an analytically pure sample of isomer **40a** as a yellow oil (50.0 mg, 0.17 mmol). <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ ppm 8.73 (s, 1 H), 7.46 (dd, *J* = 8.2, 7.3 Hz, 1 H), 6.82 (d, *J* = 8.2 Hz, 1 H), 6.53 (d, *J* = 7.0 Hz, 1 H), 5.27–5.23 (m, 1 H), 4.99 (t, *J* = 7.0 Hz, 1 H), 4.87–4.84 (m, 1 H), 4.80–4.78 (m, 1 H), 4.40 (t, *J* = 5.3 Hz, 1 H), 3.92 (t, *J* = 5.7 Hz, 2 H), 2.57 (q, *J* = 7.4 Hz, 2 H), 2.09–1.96 (m, 2 H), 1.93–1.72 (m, 2 H), 1.66 (s, 3 H), 1.57 (s, 3 H), 1.19–1.14 (m, 3 H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>) δ ppm 162.7, 160.0, 144.6, 137.7, 135.7, 125.2, 112.7, 111.9, 105.2, 62.5, 57.6, 36.2, 30.5, 27.4, 21.4, 16.2, 13.5; HRMS-ESI (*m/z*): [M+H]<sup>+</sup> calcd. for C<sub>17</sub>H<sub>27</sub>N<sub>2</sub>O<sub>2</sub>: 291.2067, found 291.2075; HPLC retention time 7.82 min.

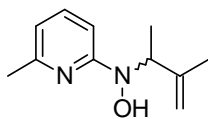
## V. Copies of $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR Spectra



(±) **9**

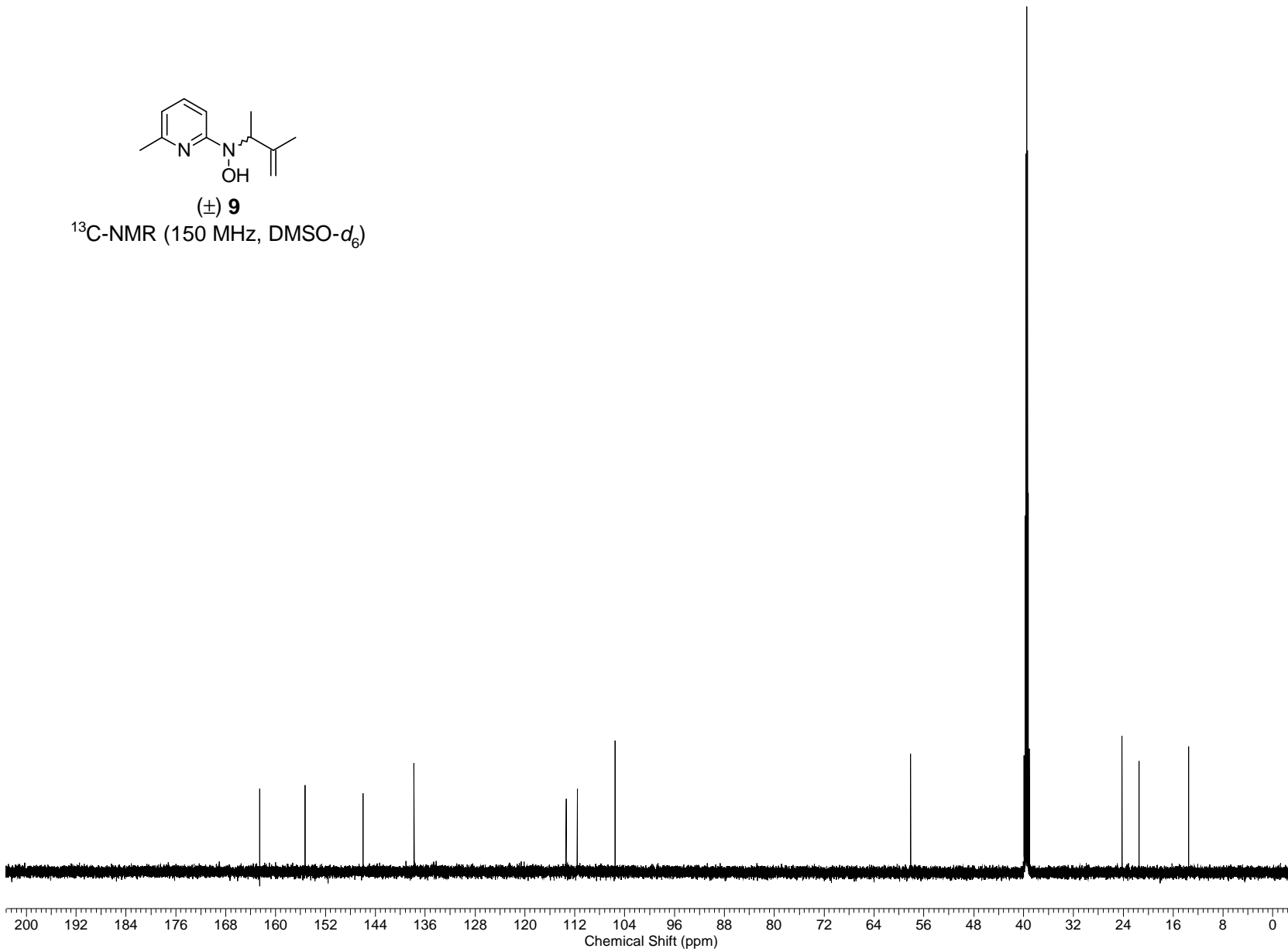
$^1\text{H}$ -NMR (600 MHz,  $\text{DMSO-}d_6$ )

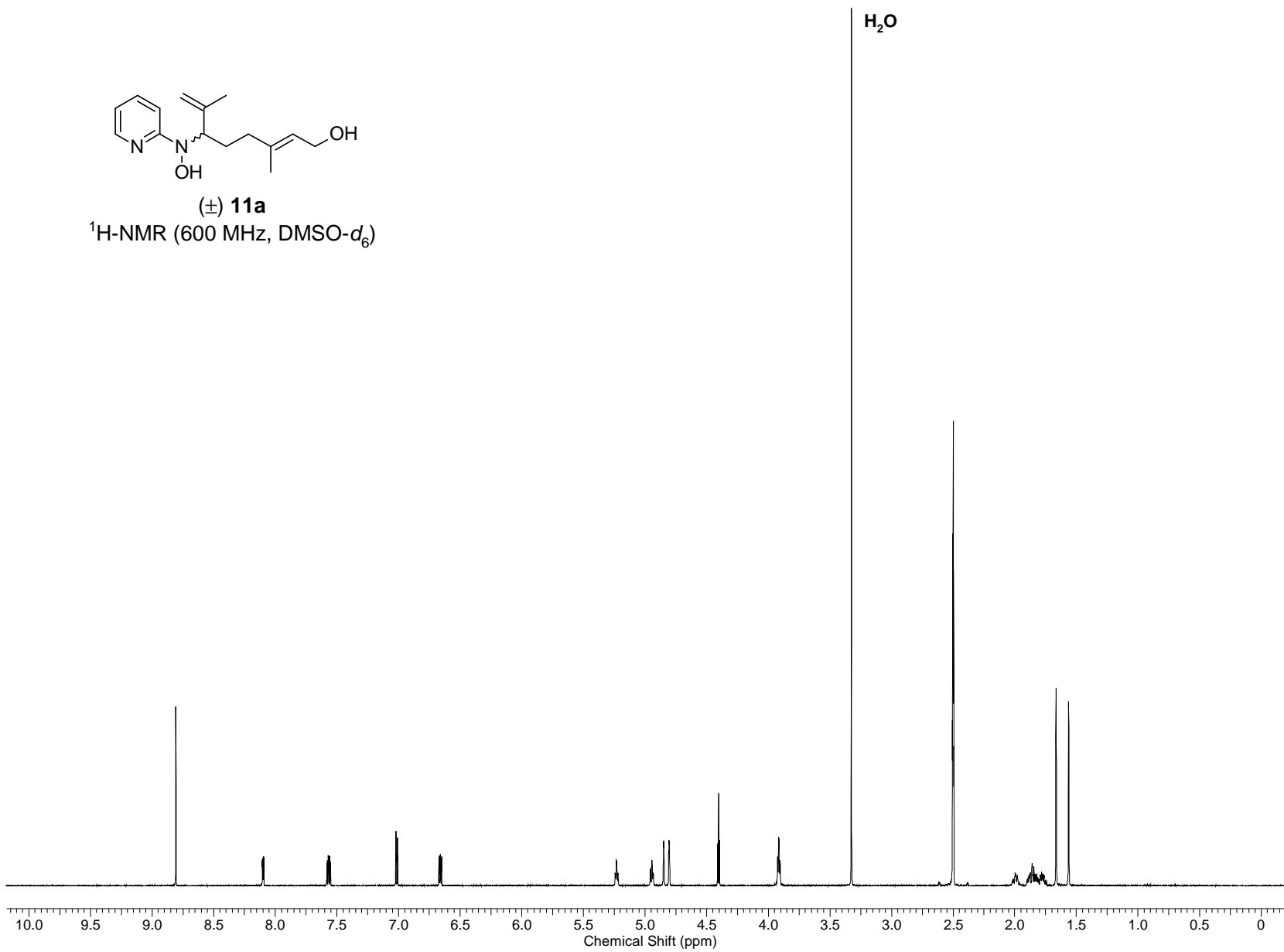
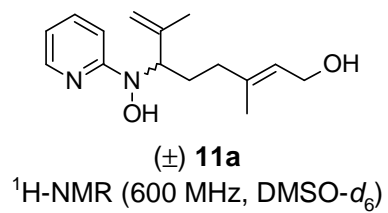


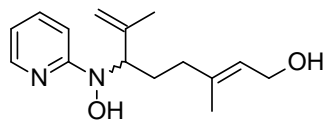


(±) **9**

$^{13}\text{C}$ -NMR (150 MHz,  $\text{DMSO-}d_6$ )

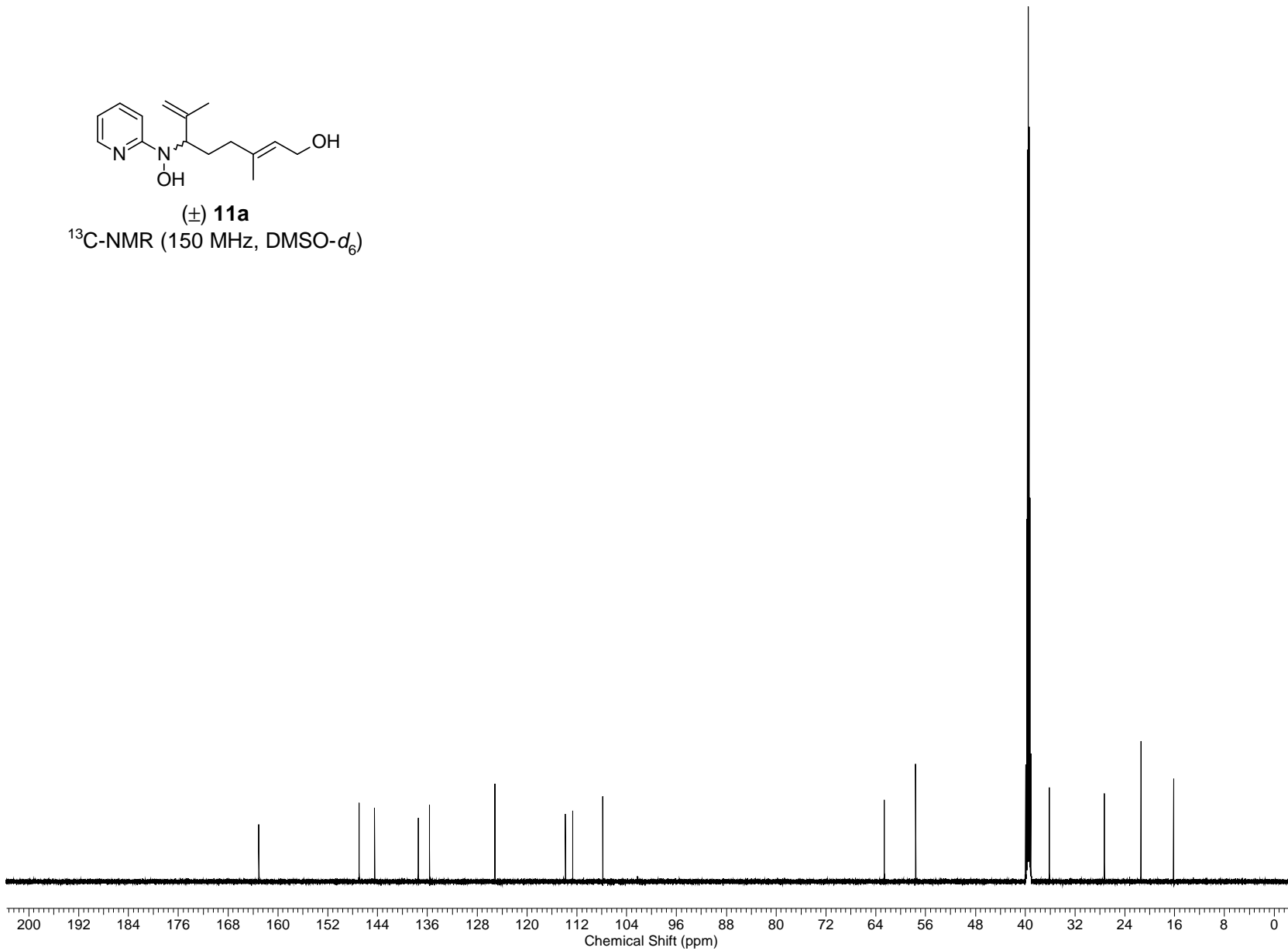


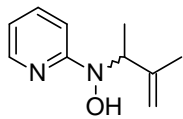




(±) **11a**

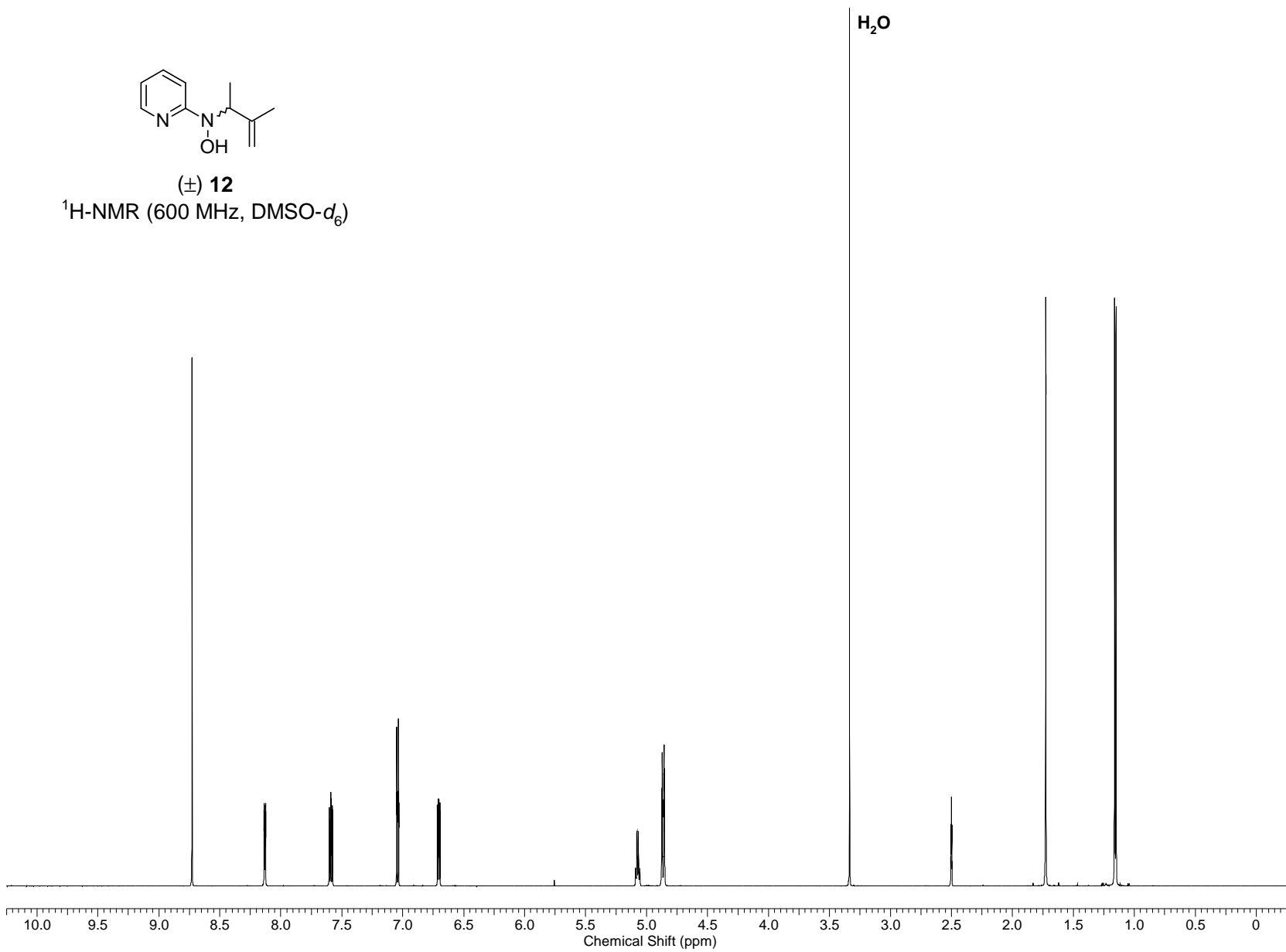
<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)

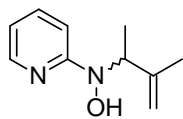




(±) **12**

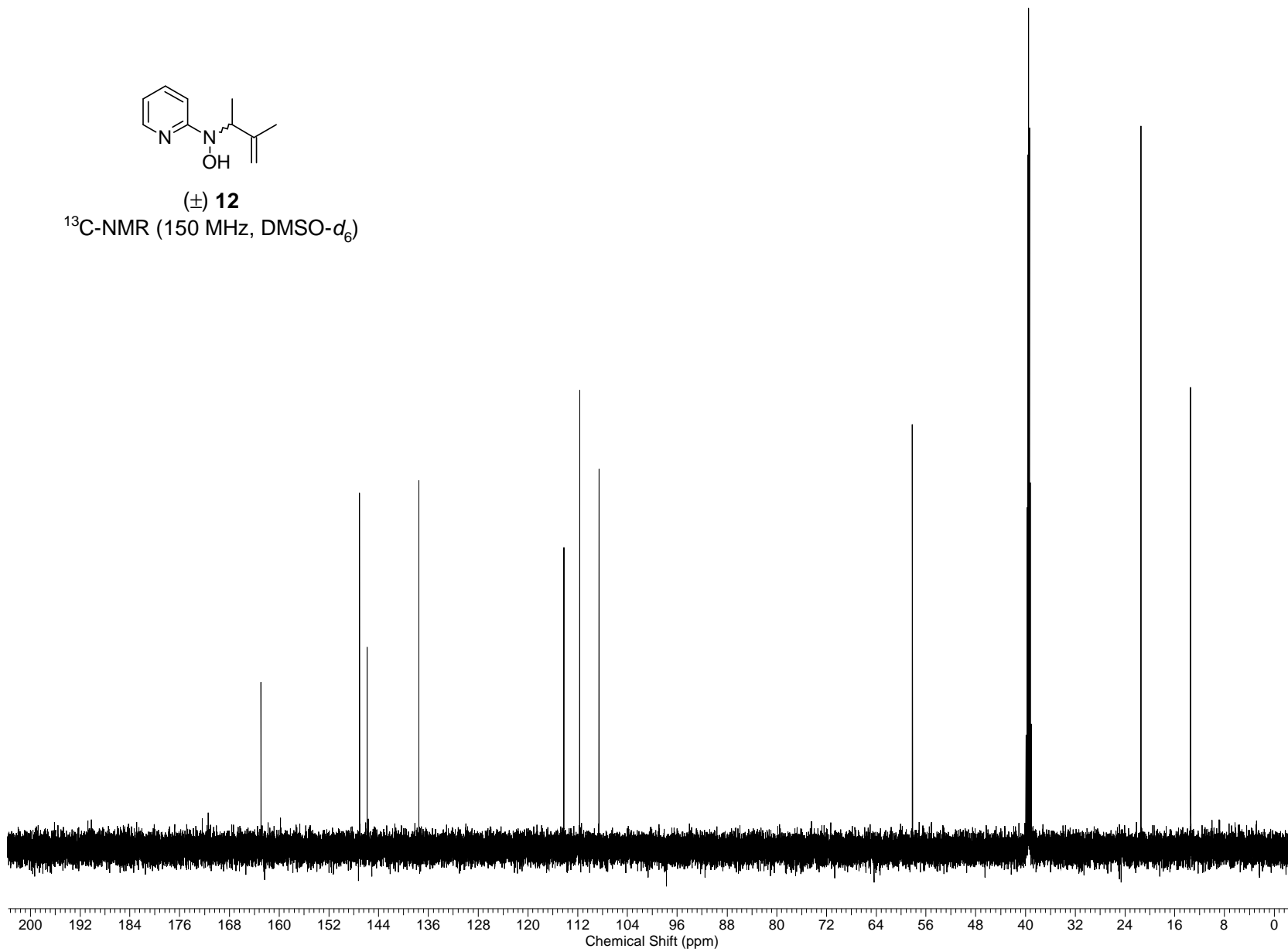
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

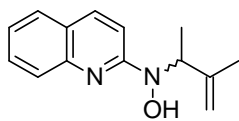




(±) **12**

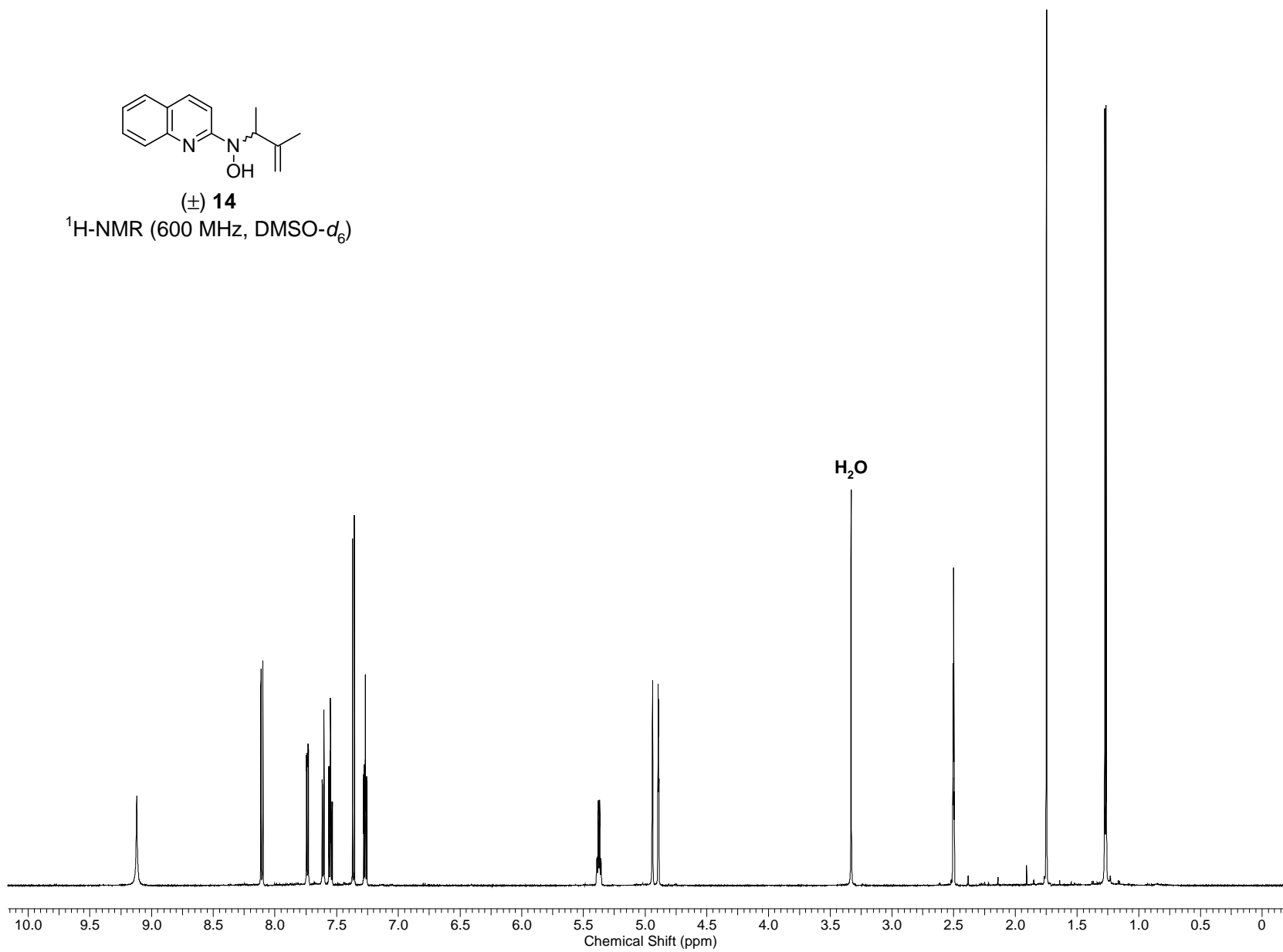
<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)



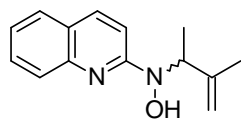


(±) **14**

<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

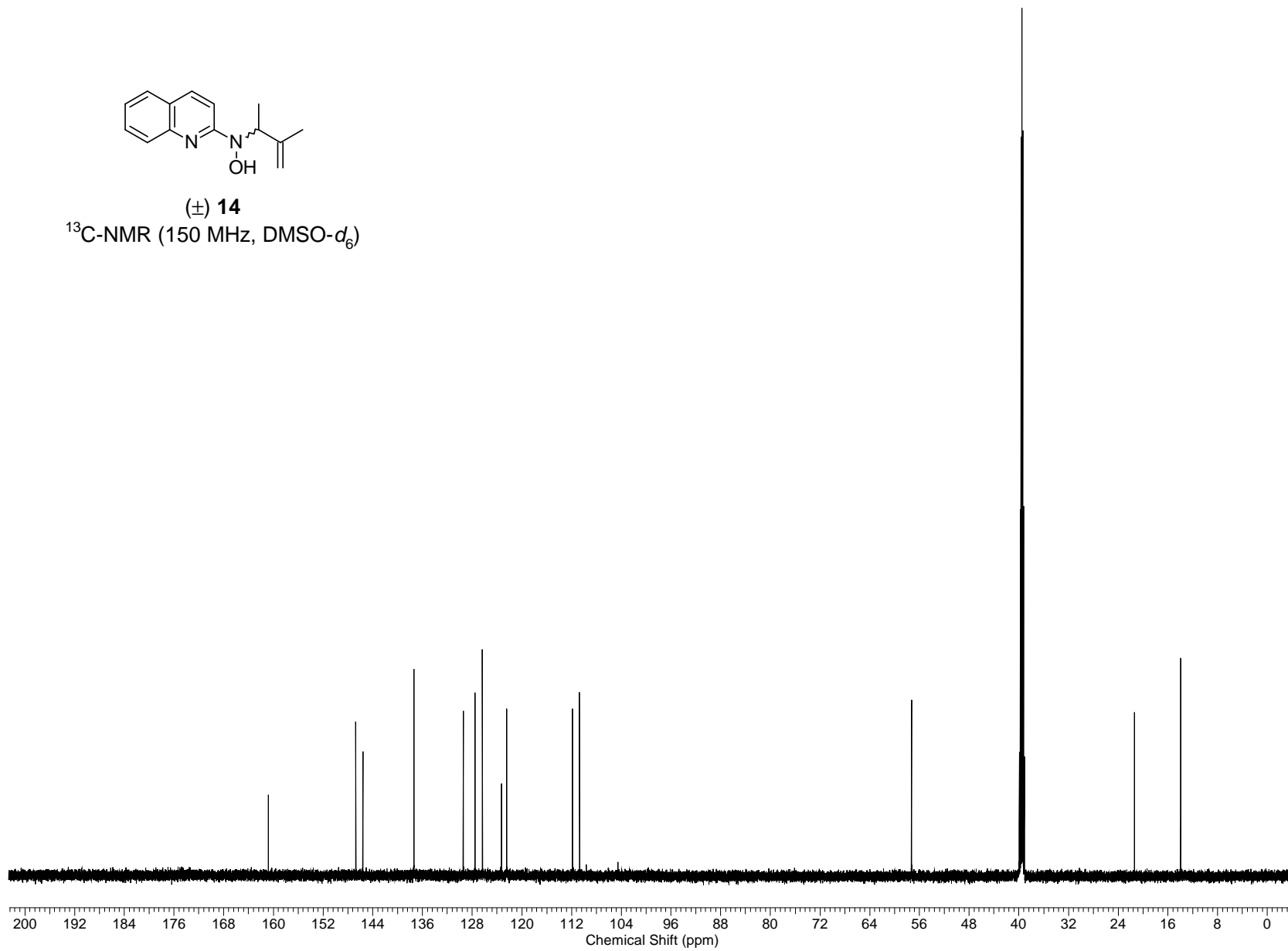


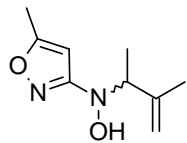




(±) **14**

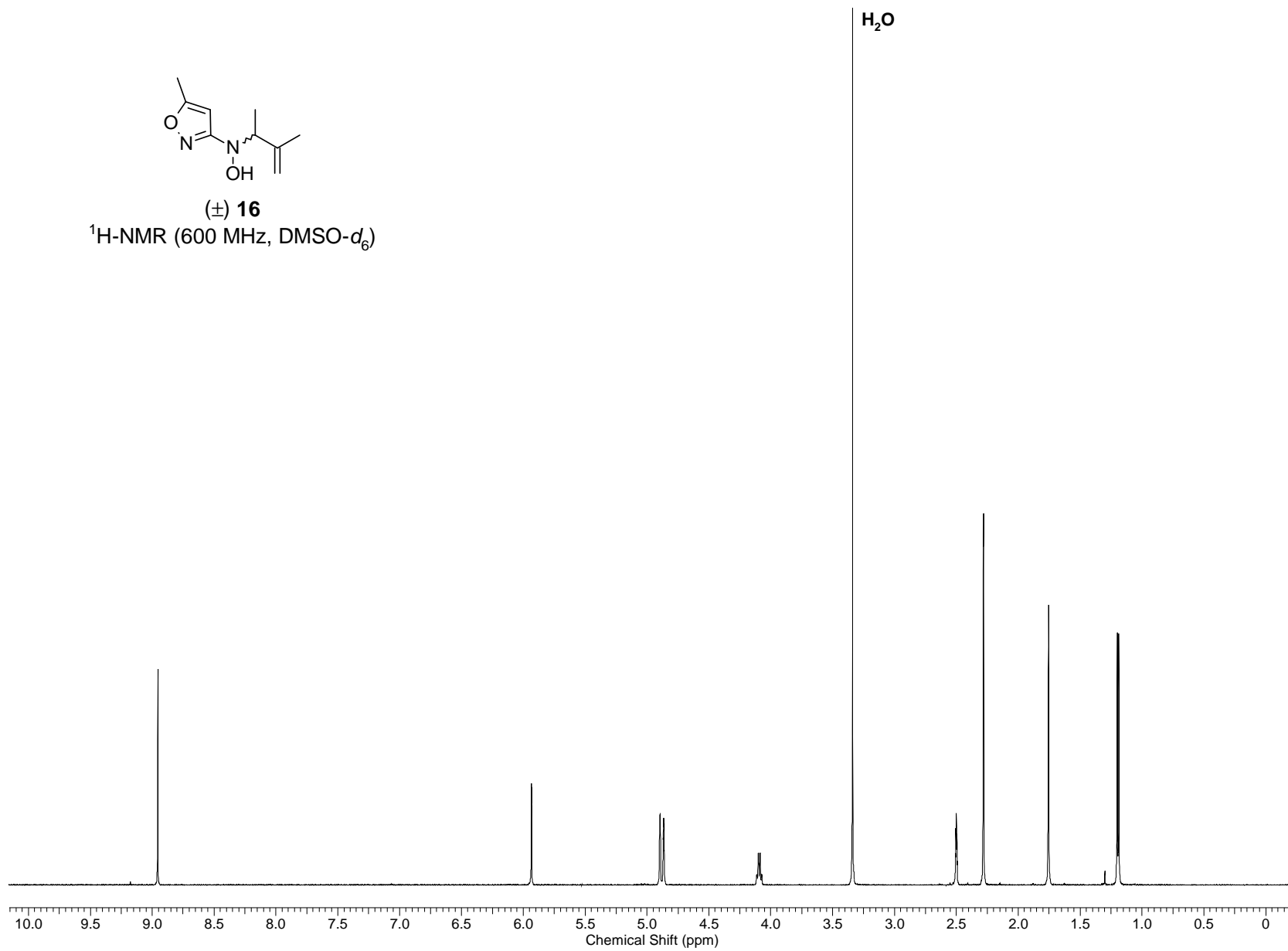
$^{13}\text{C}$ -NMR (150 MHz,  $\text{DMSO}-d_6$ )

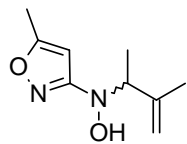




(±) **16**

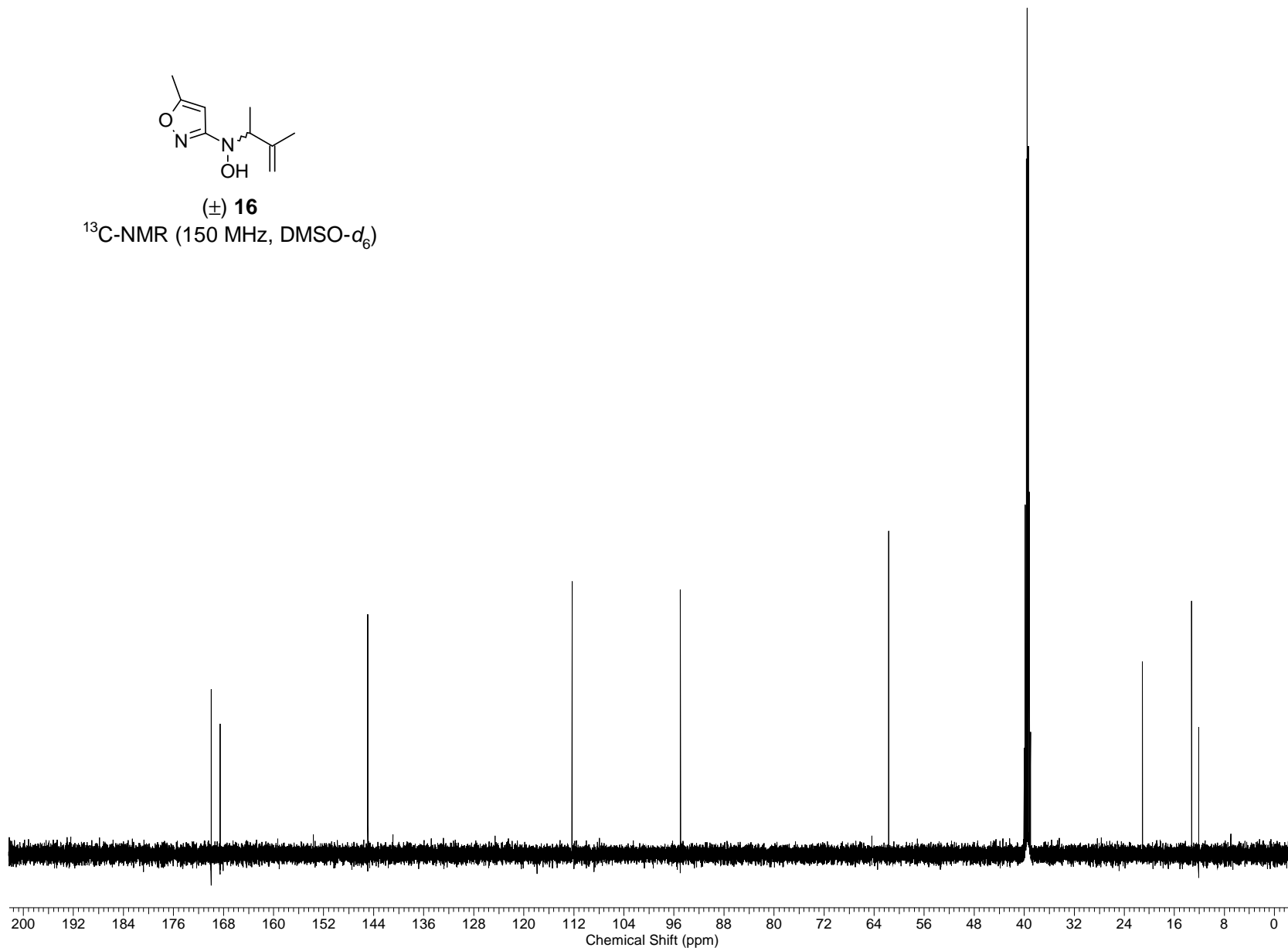
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

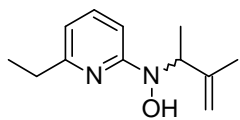




(±) **16**

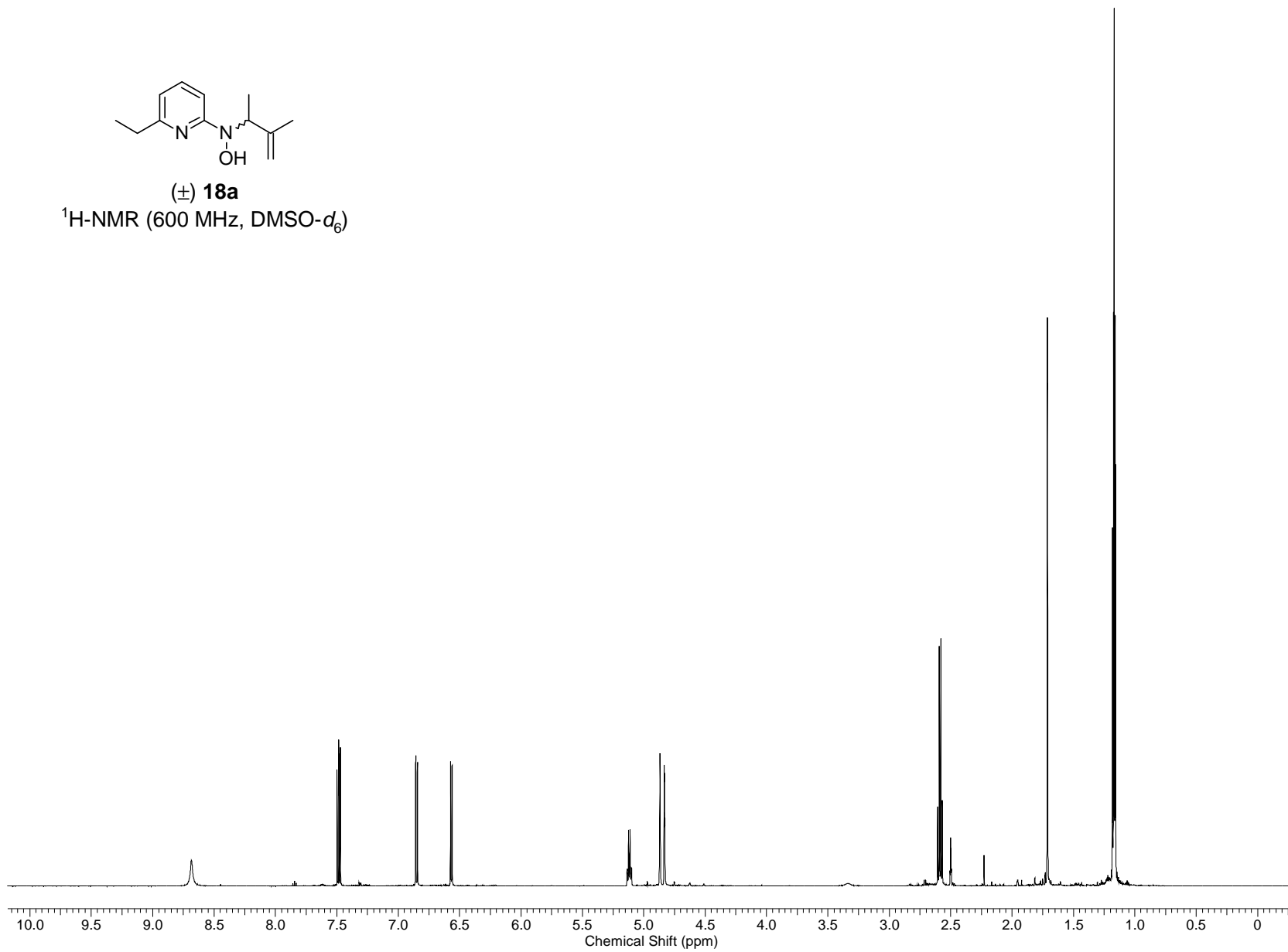
<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)

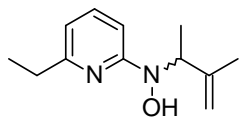




(±) **18a**

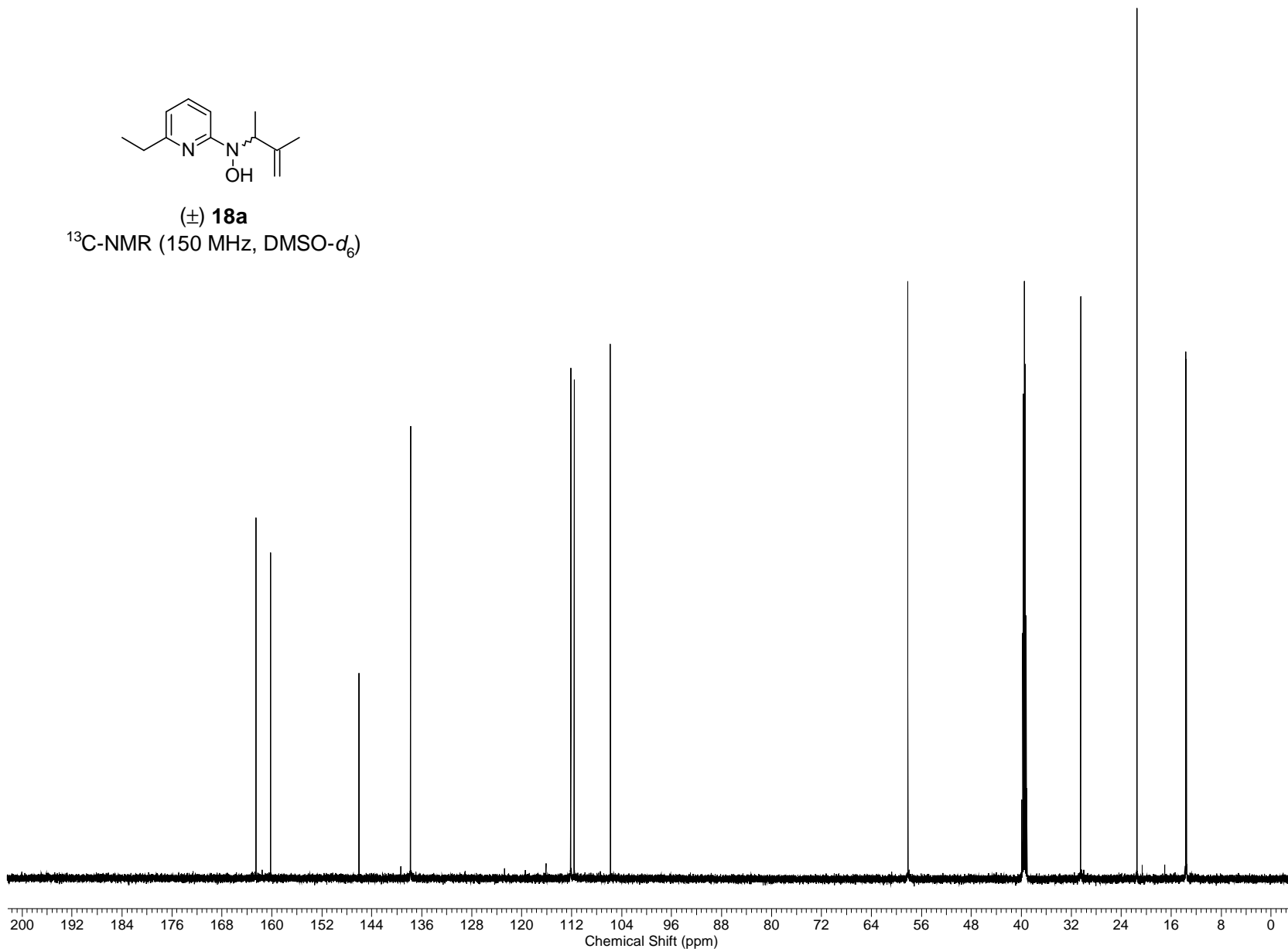
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

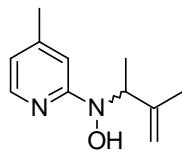




(±) **18a**

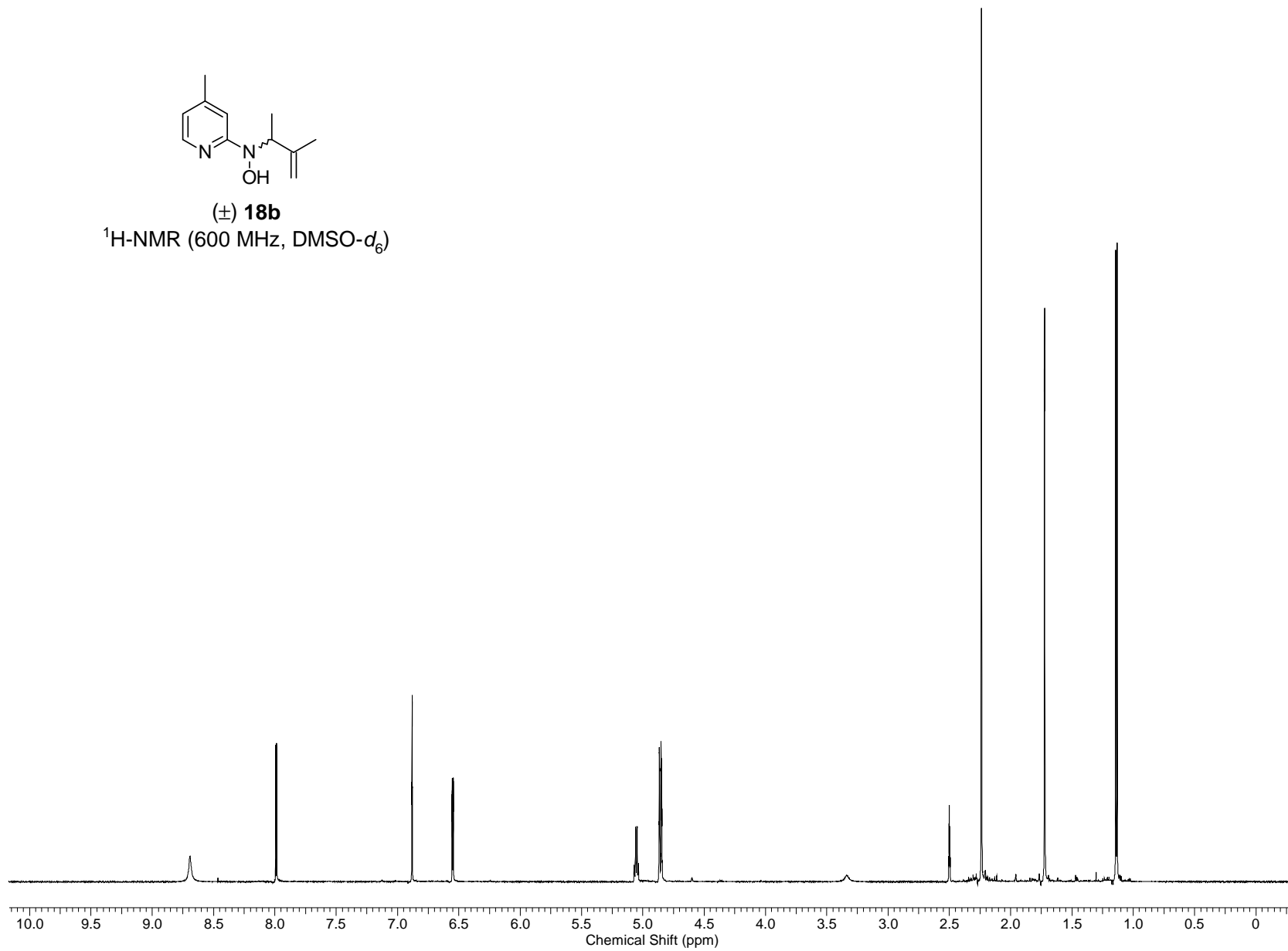
<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)

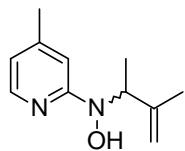




(±) **18b**

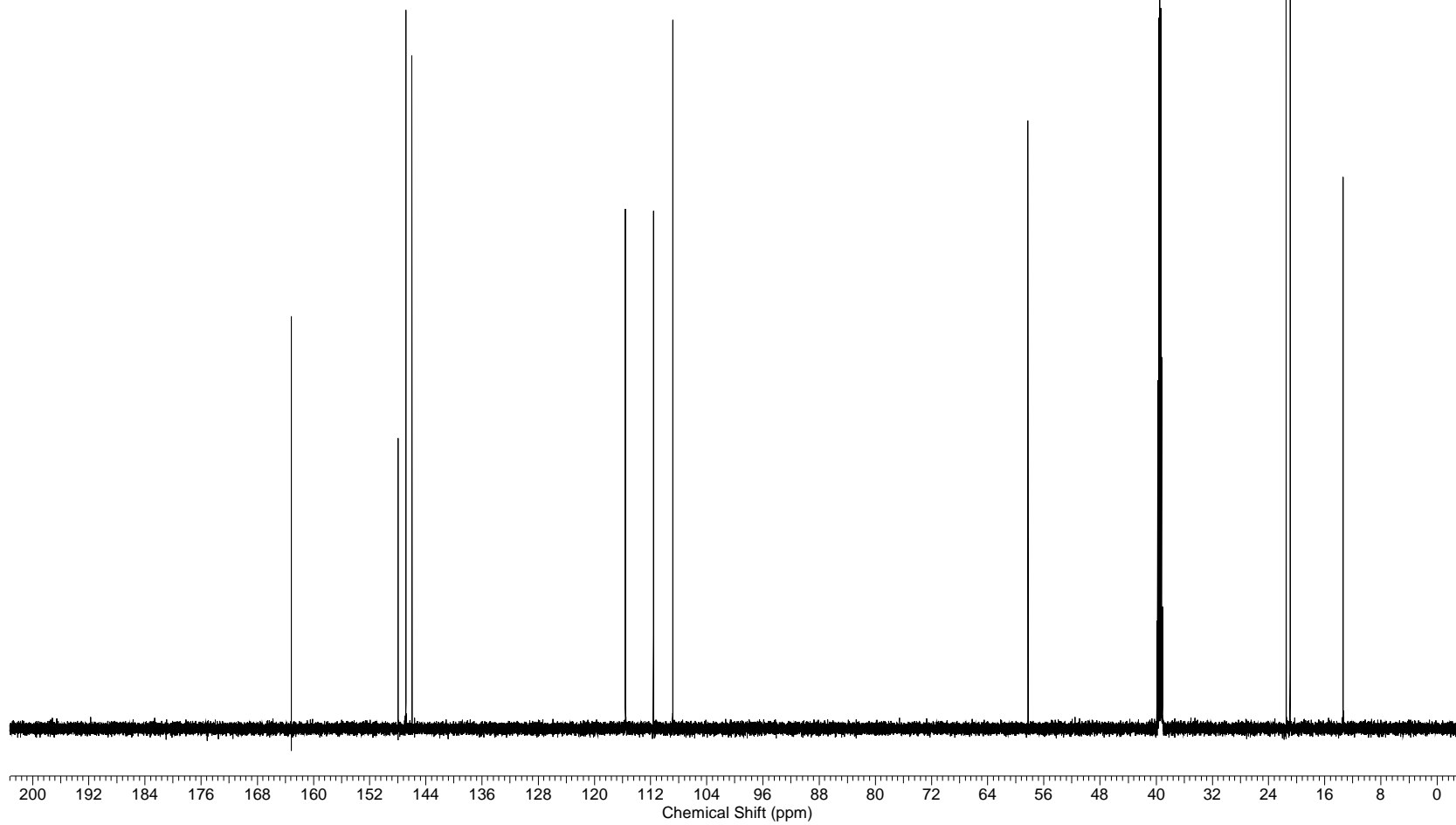
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)





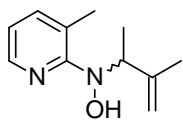
(±) **18b**

<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)



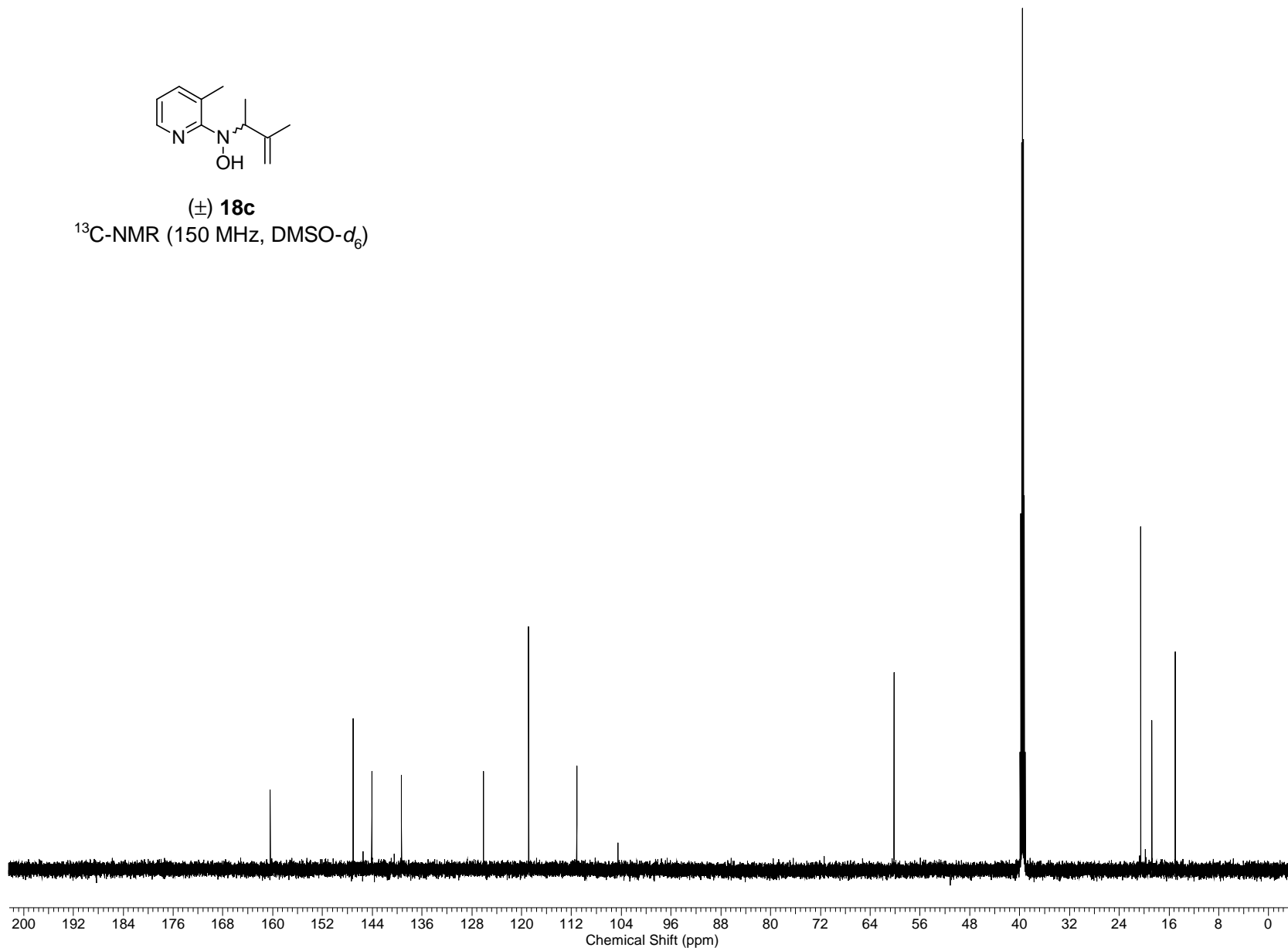


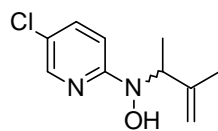




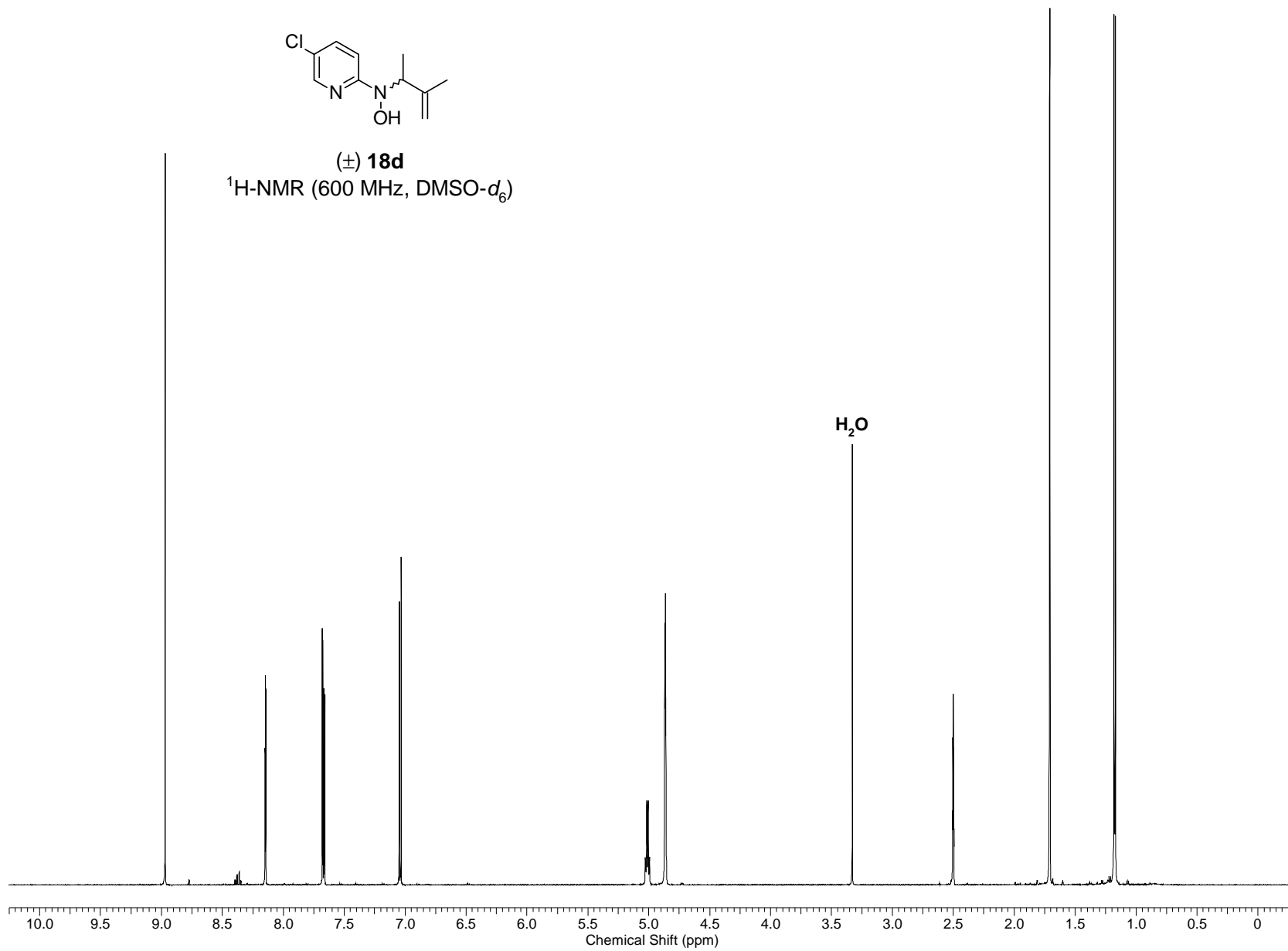
(±) **18c**

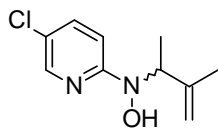
<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)





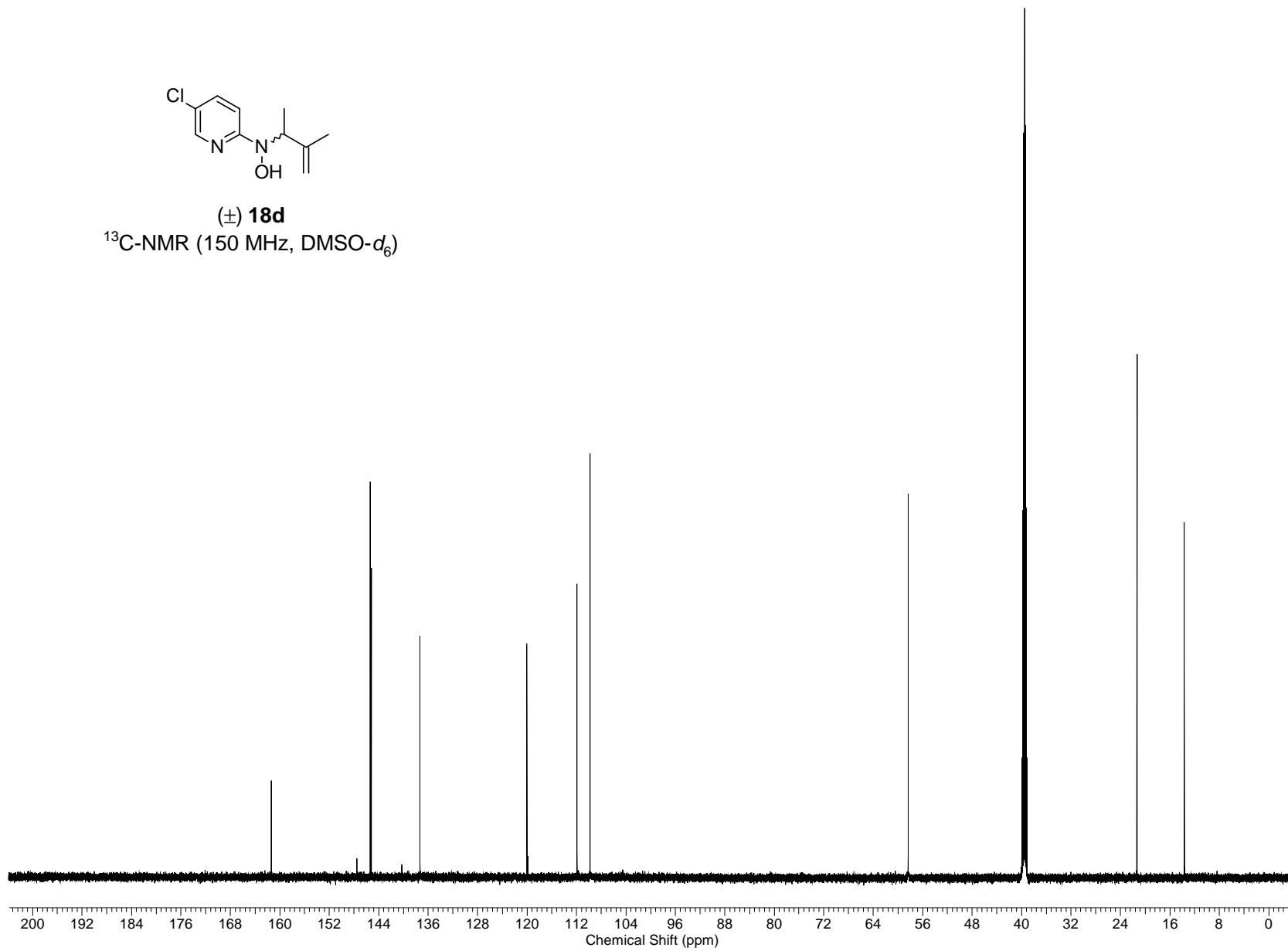
(±) **18d**  
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

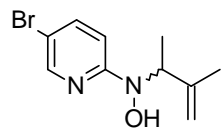




(±) **18d**

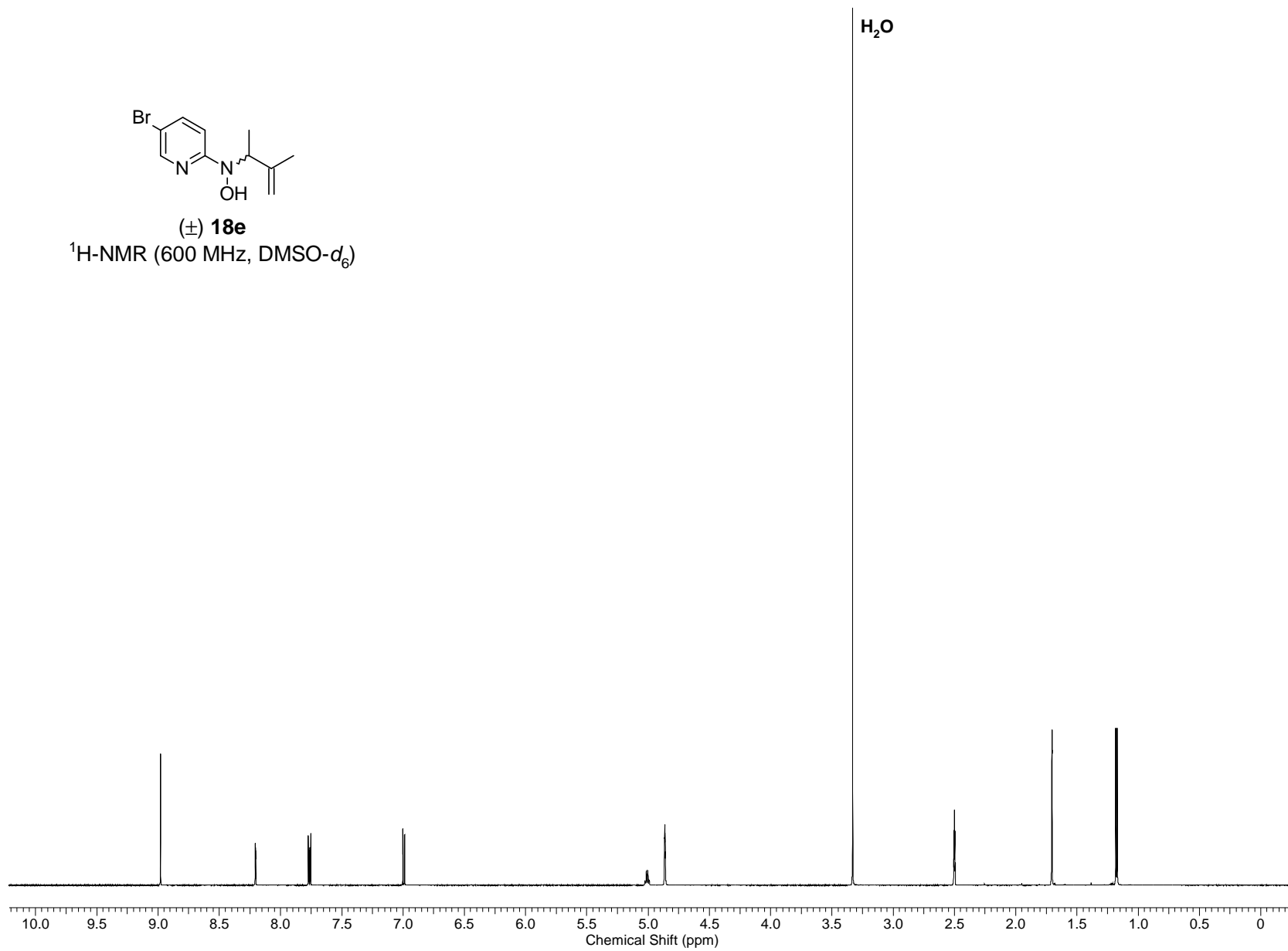
<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)

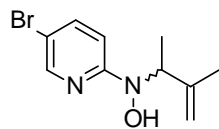




(±) **18e**

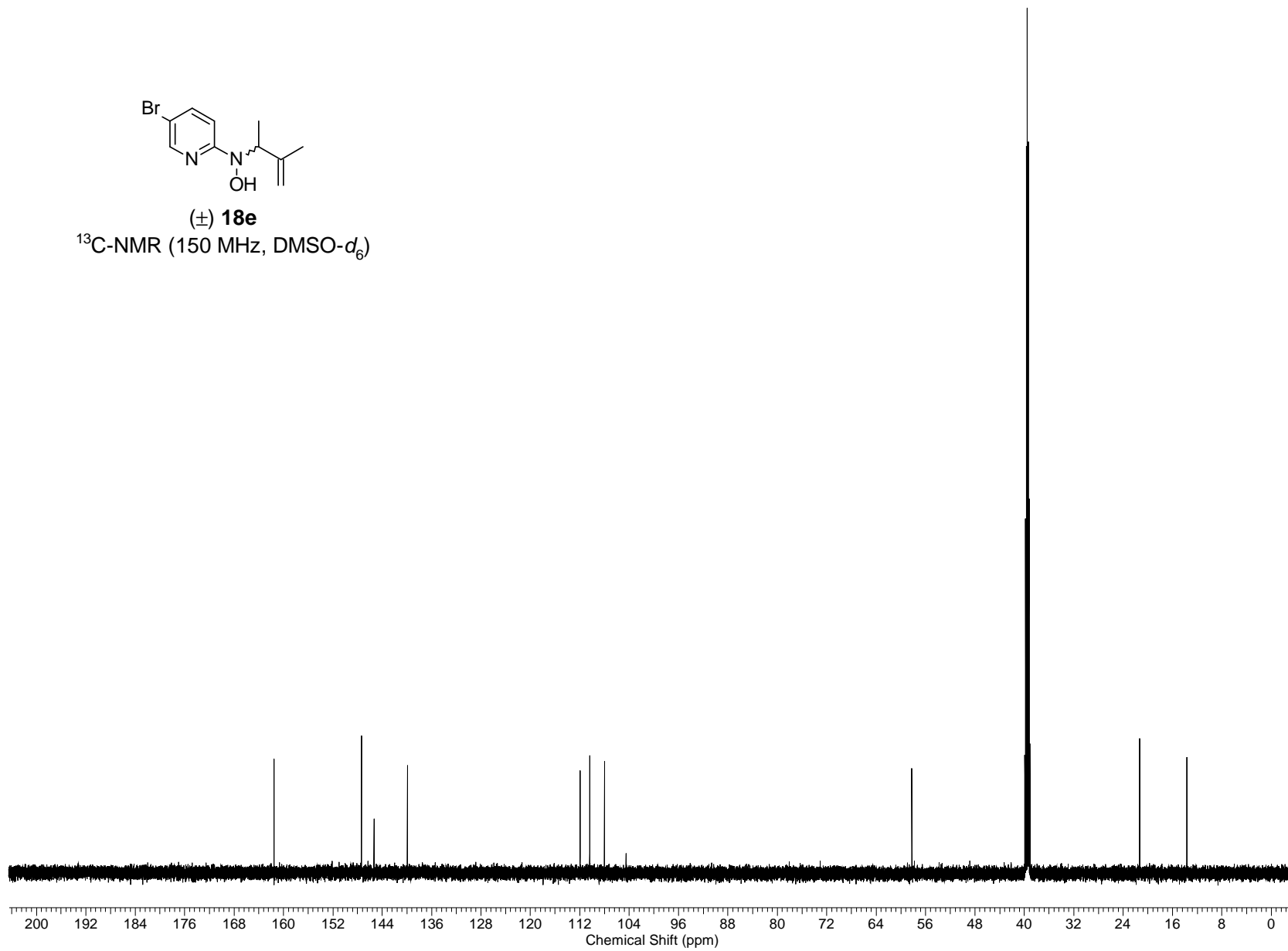
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

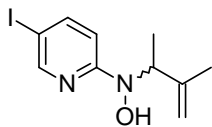




(±) **18e**

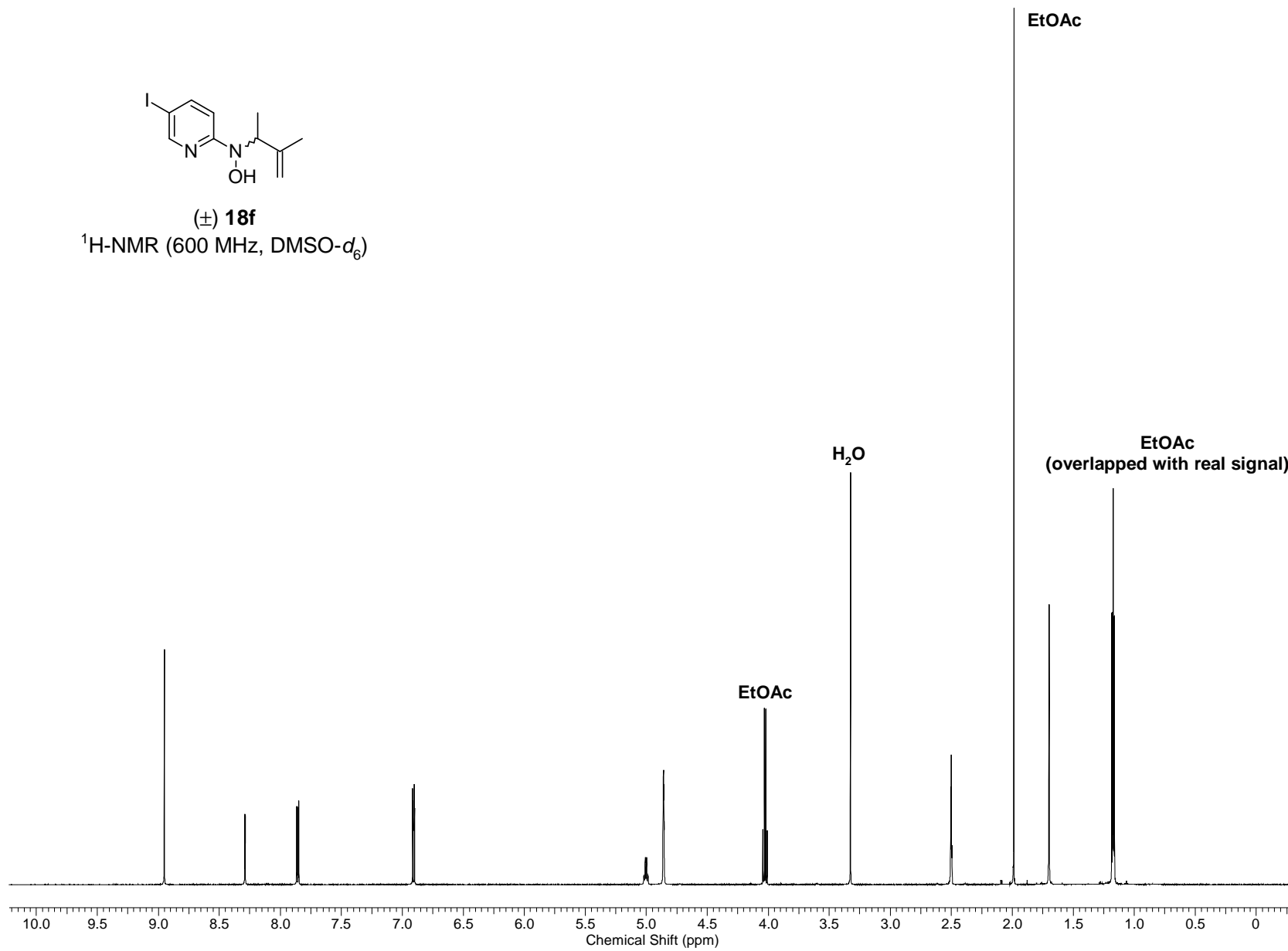
<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)

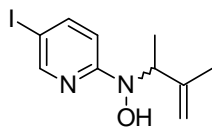




(±) **18f**

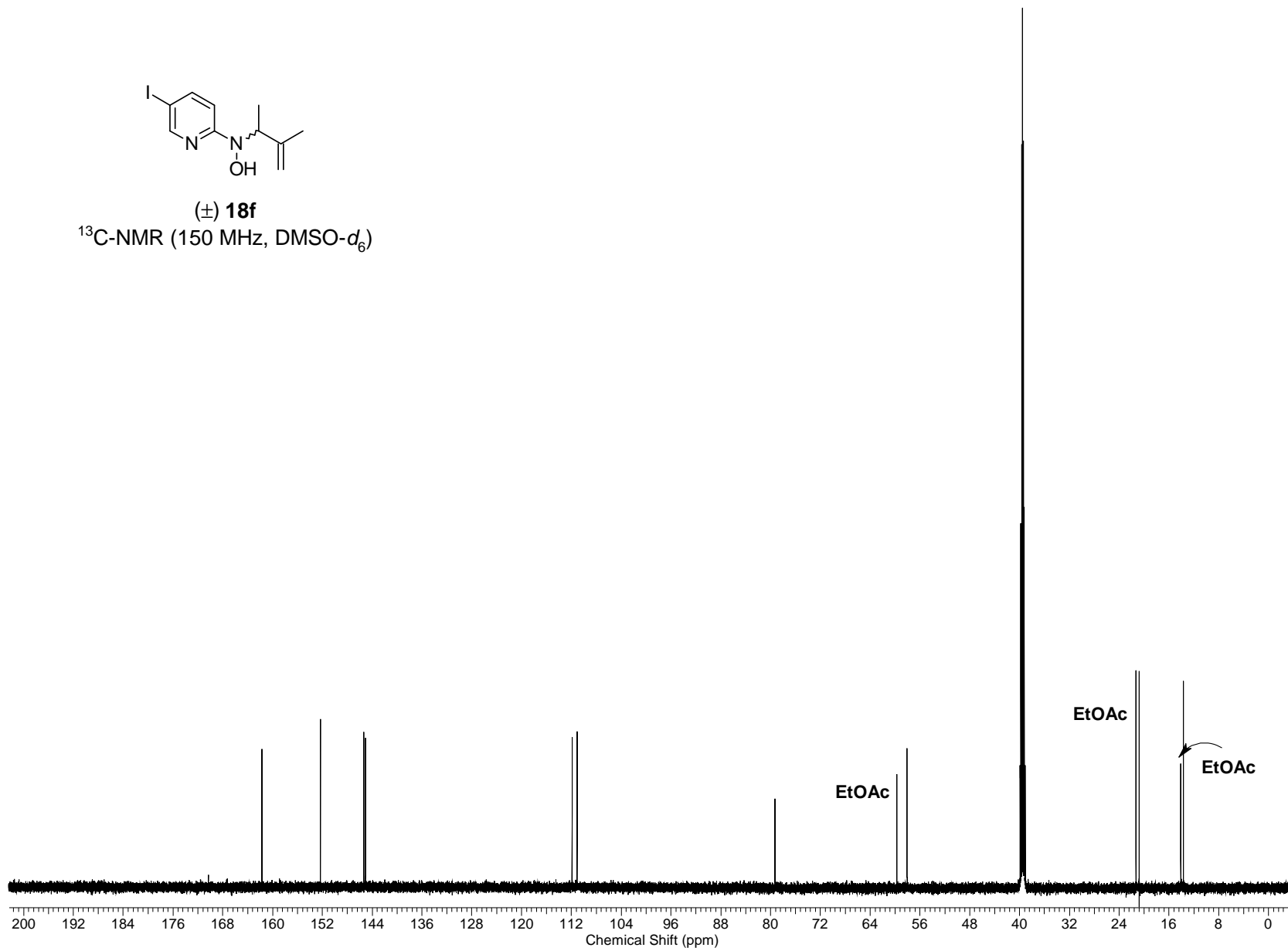
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

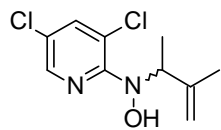




(±) **18f**

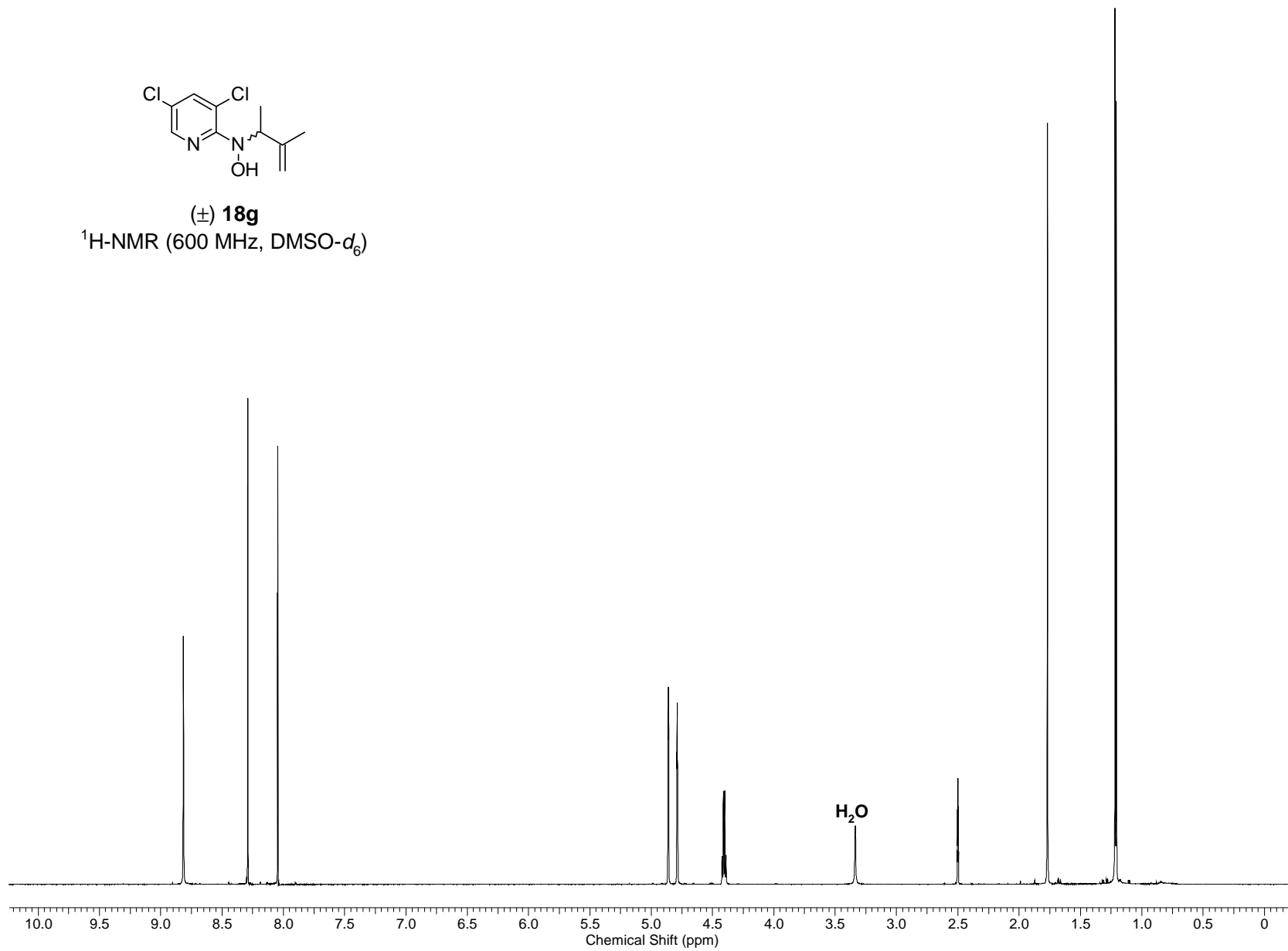
$^{13}\text{C}$ -NMR (150 MHz,  $\text{DMSO-}d_6$ )



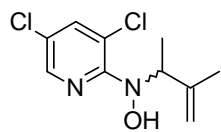


(±) **18g**

<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

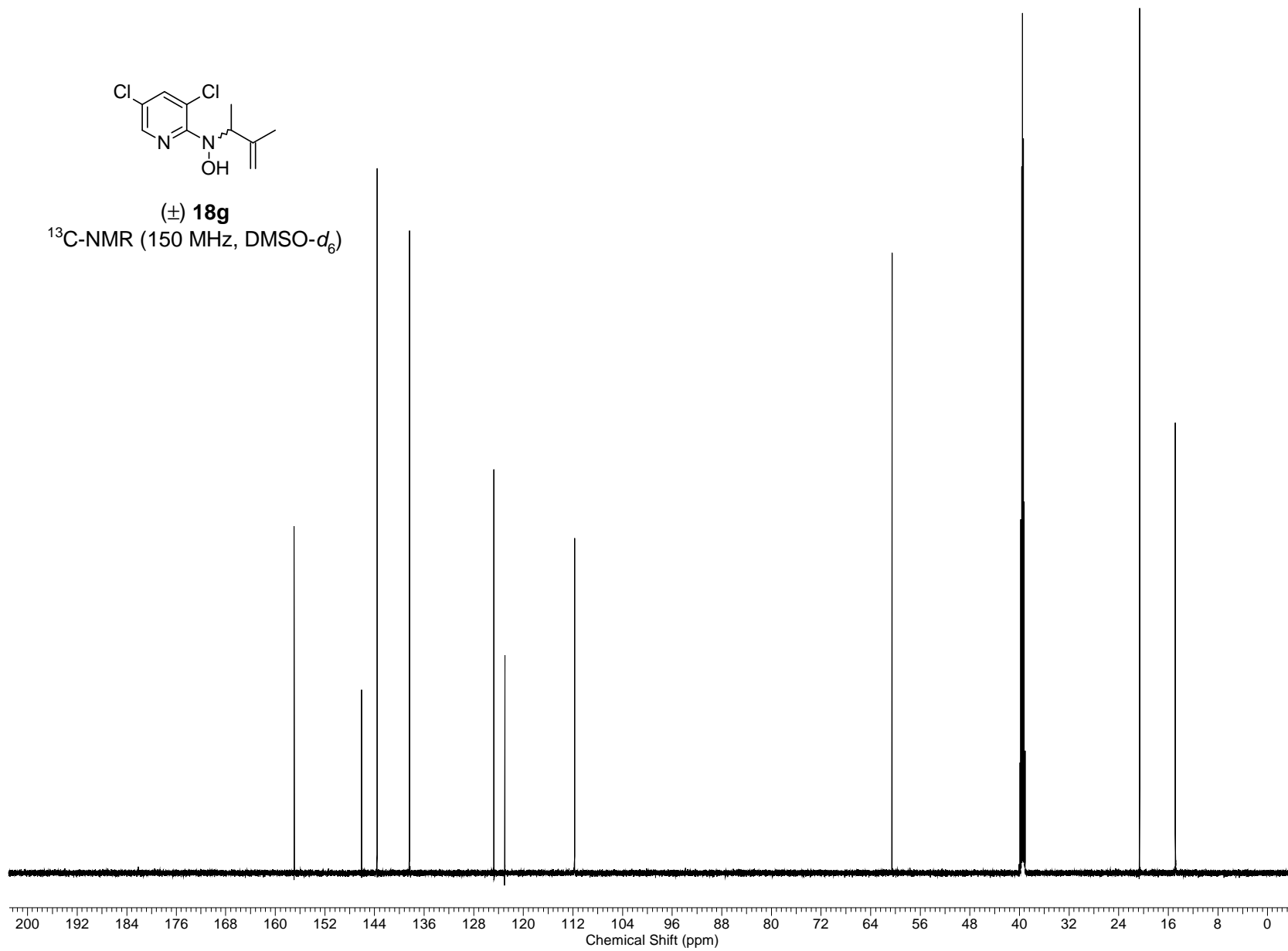


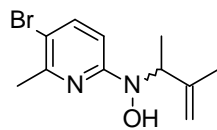




(±) **18g**

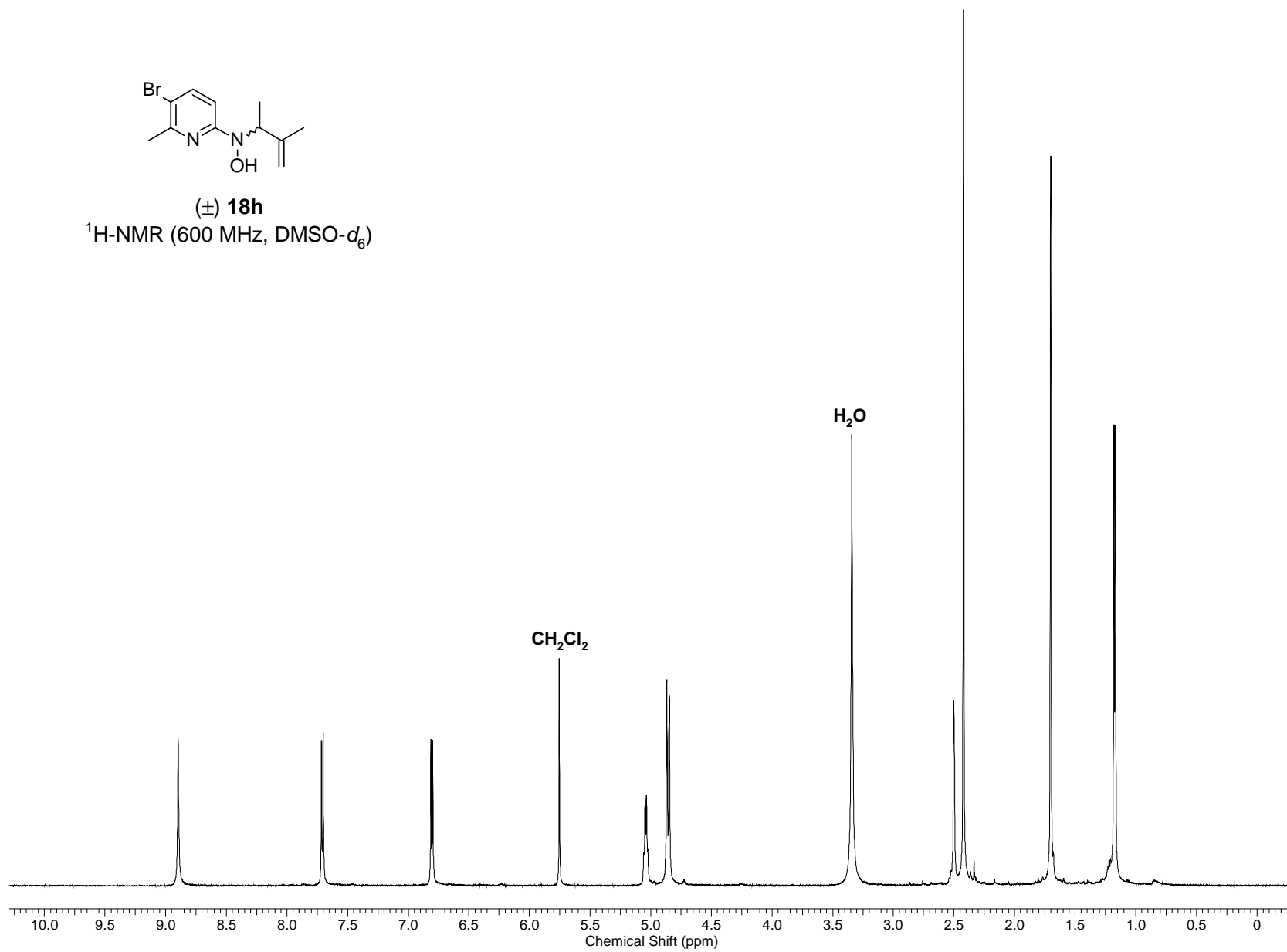
<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)

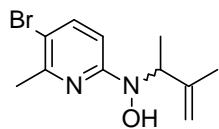




(±) **18h**

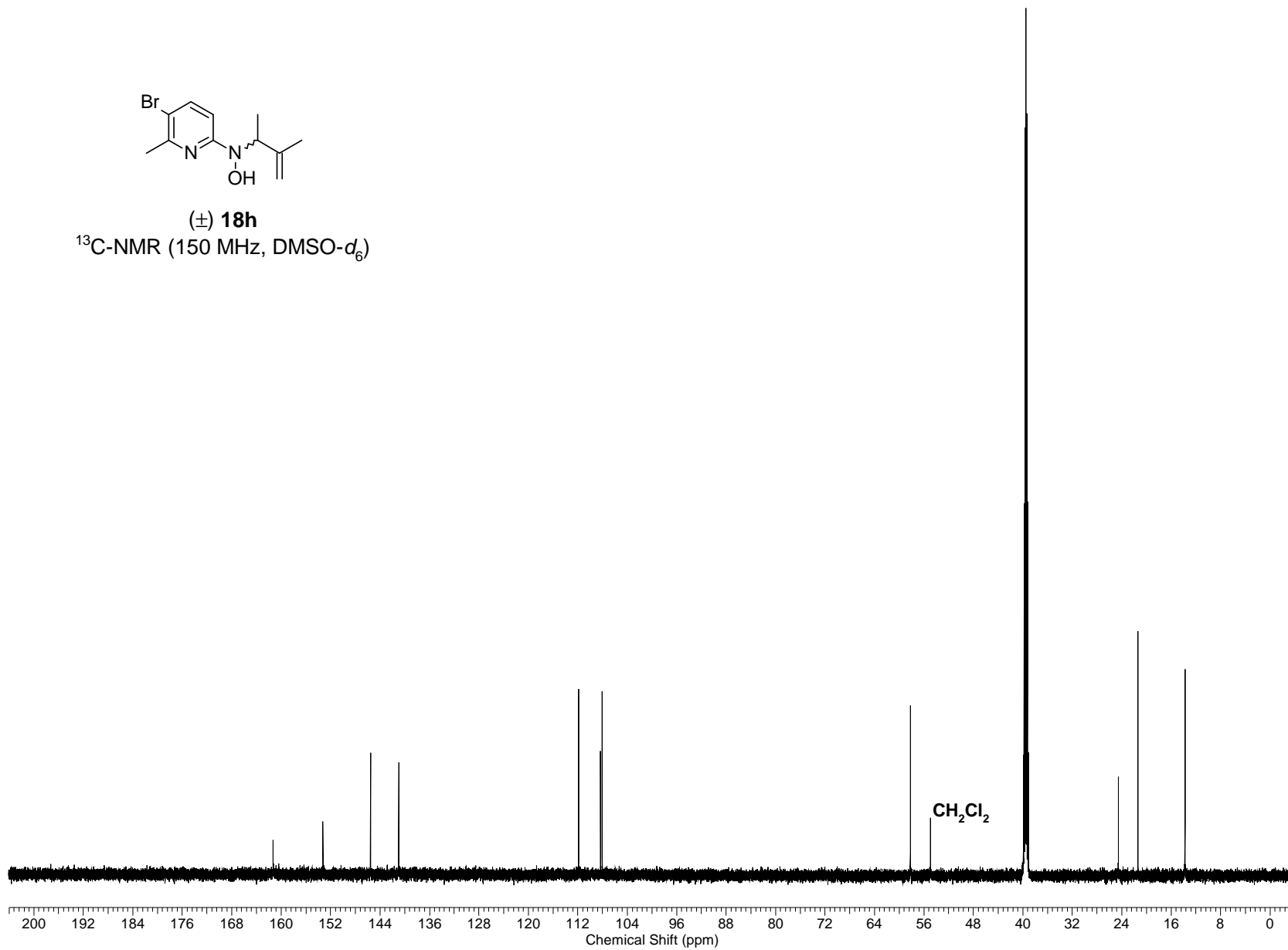
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)





(±) **18h**

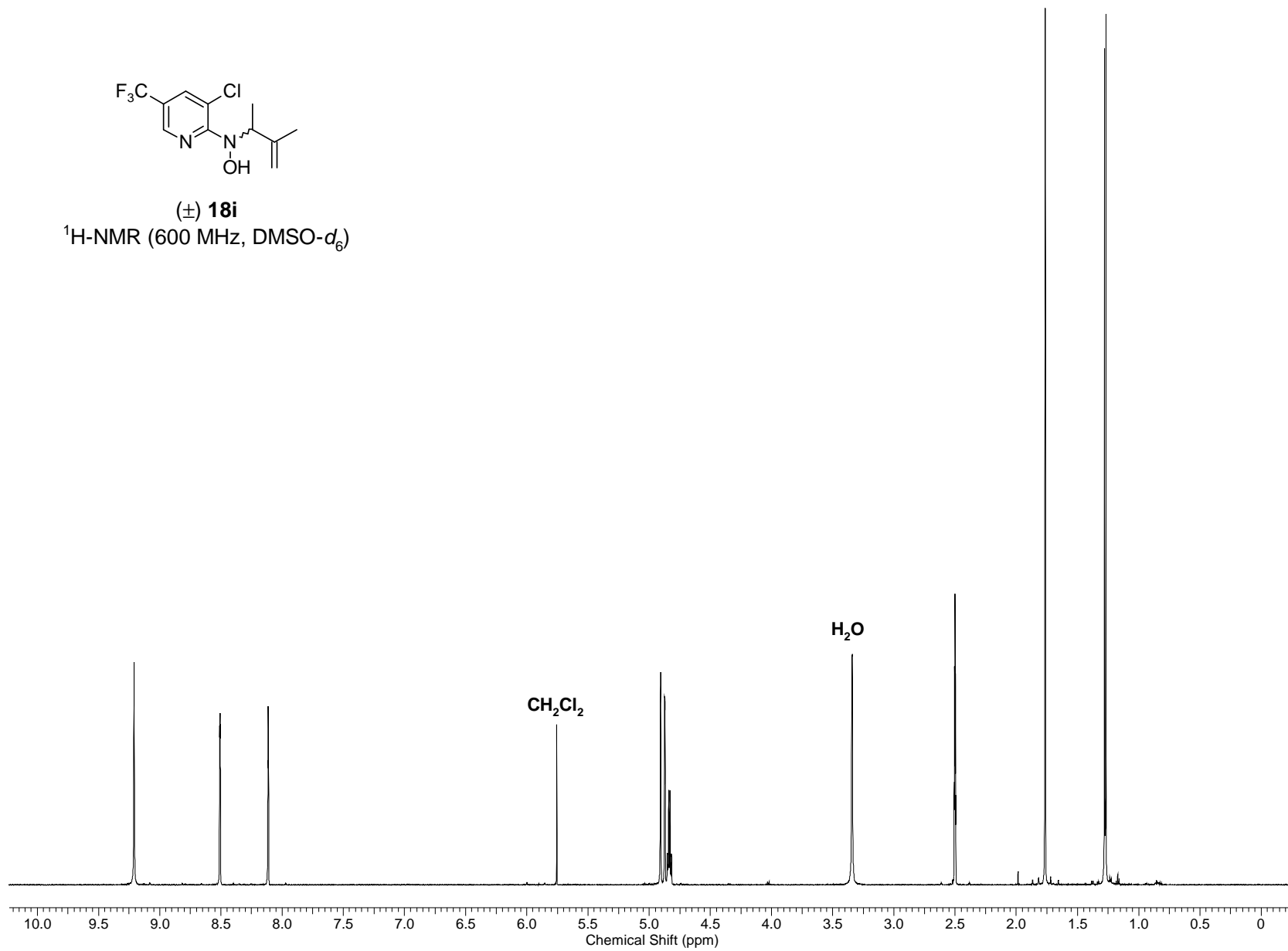
$^{13}\text{C}$ -NMR (150 MHz,  $\text{DMSO-}d_6$ )





(±) **18i**

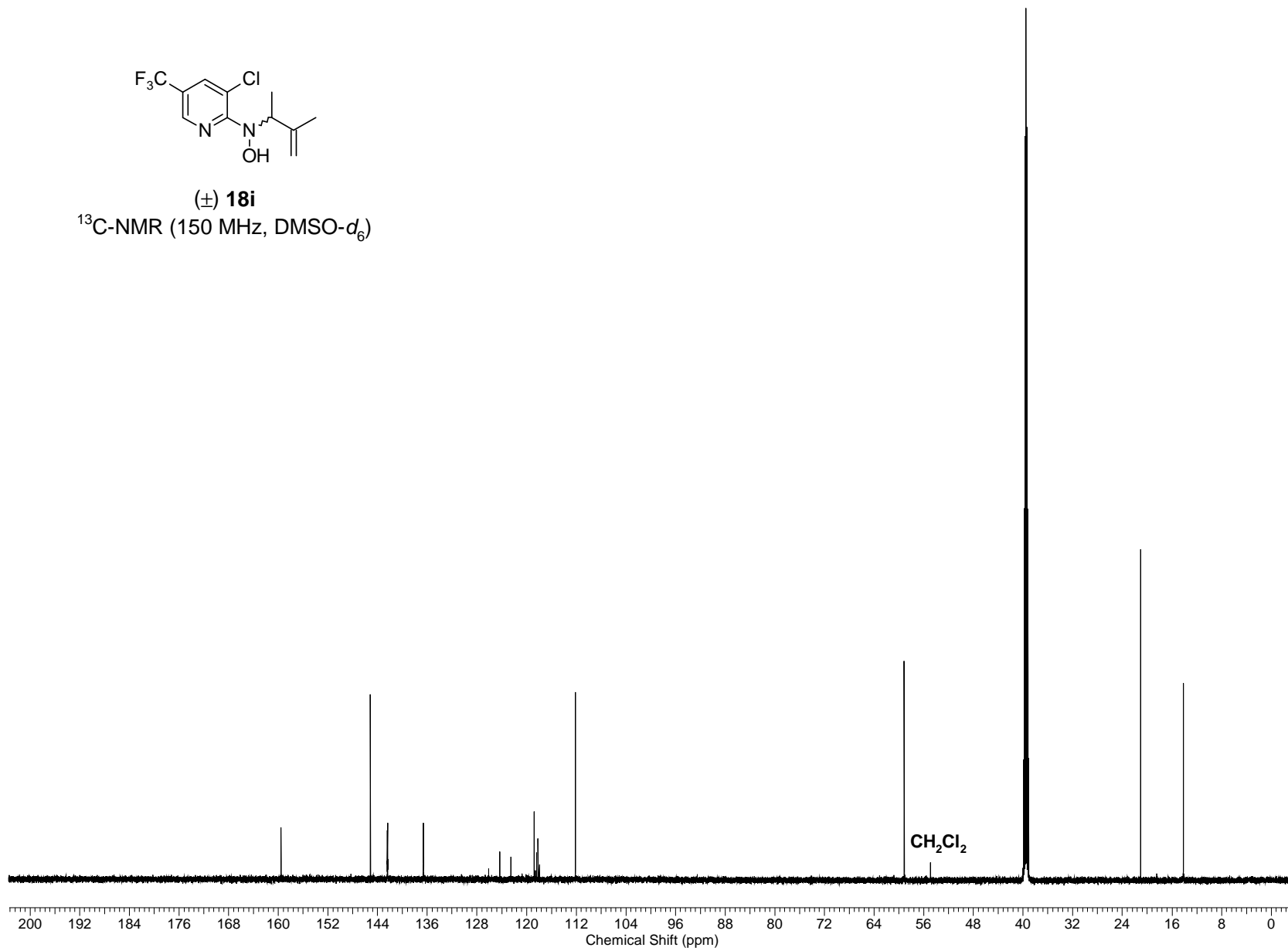
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

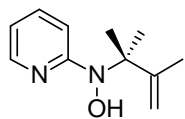




(±) **18i**

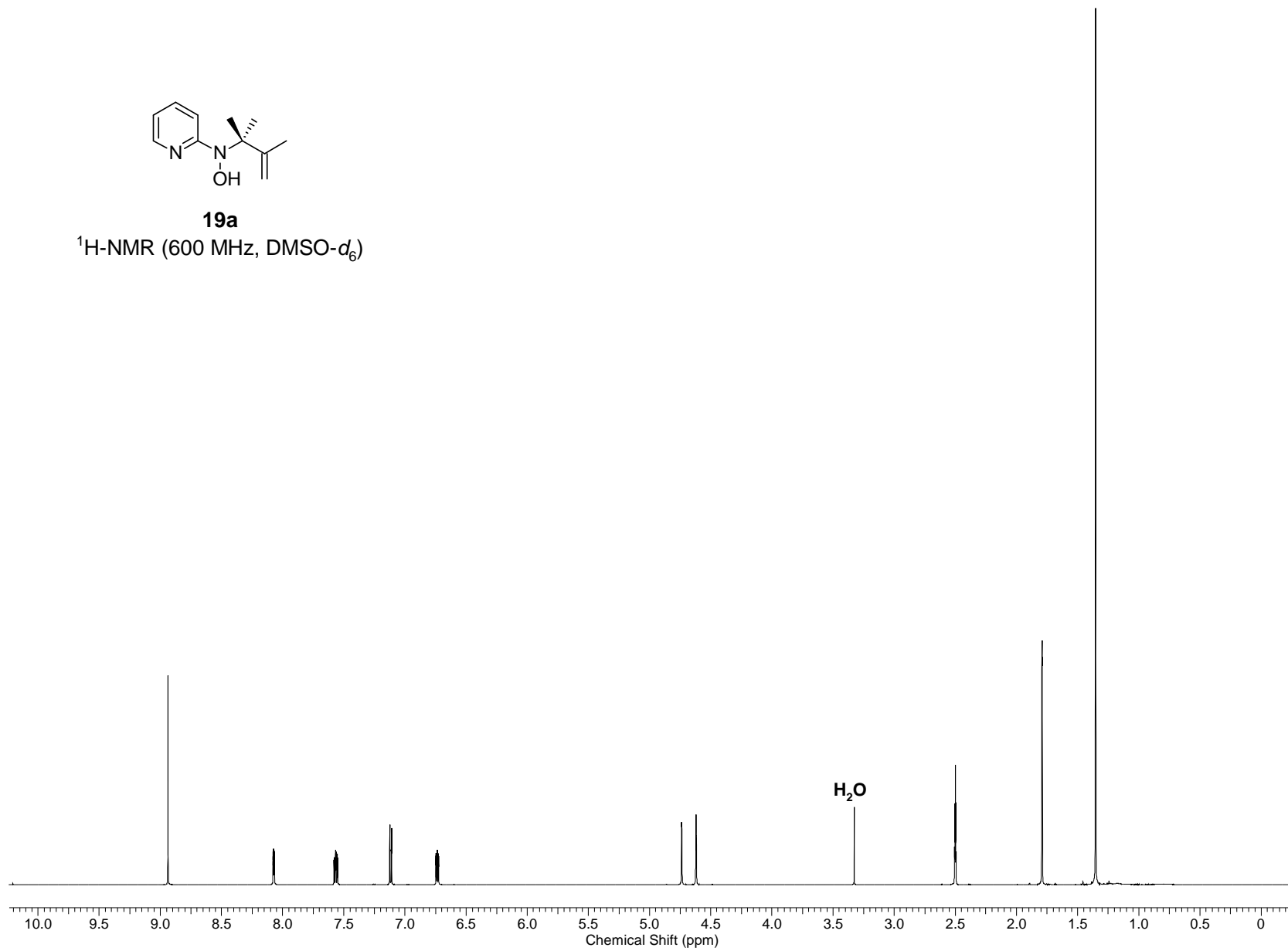
<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)

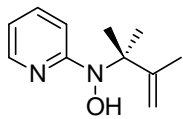




**19a**

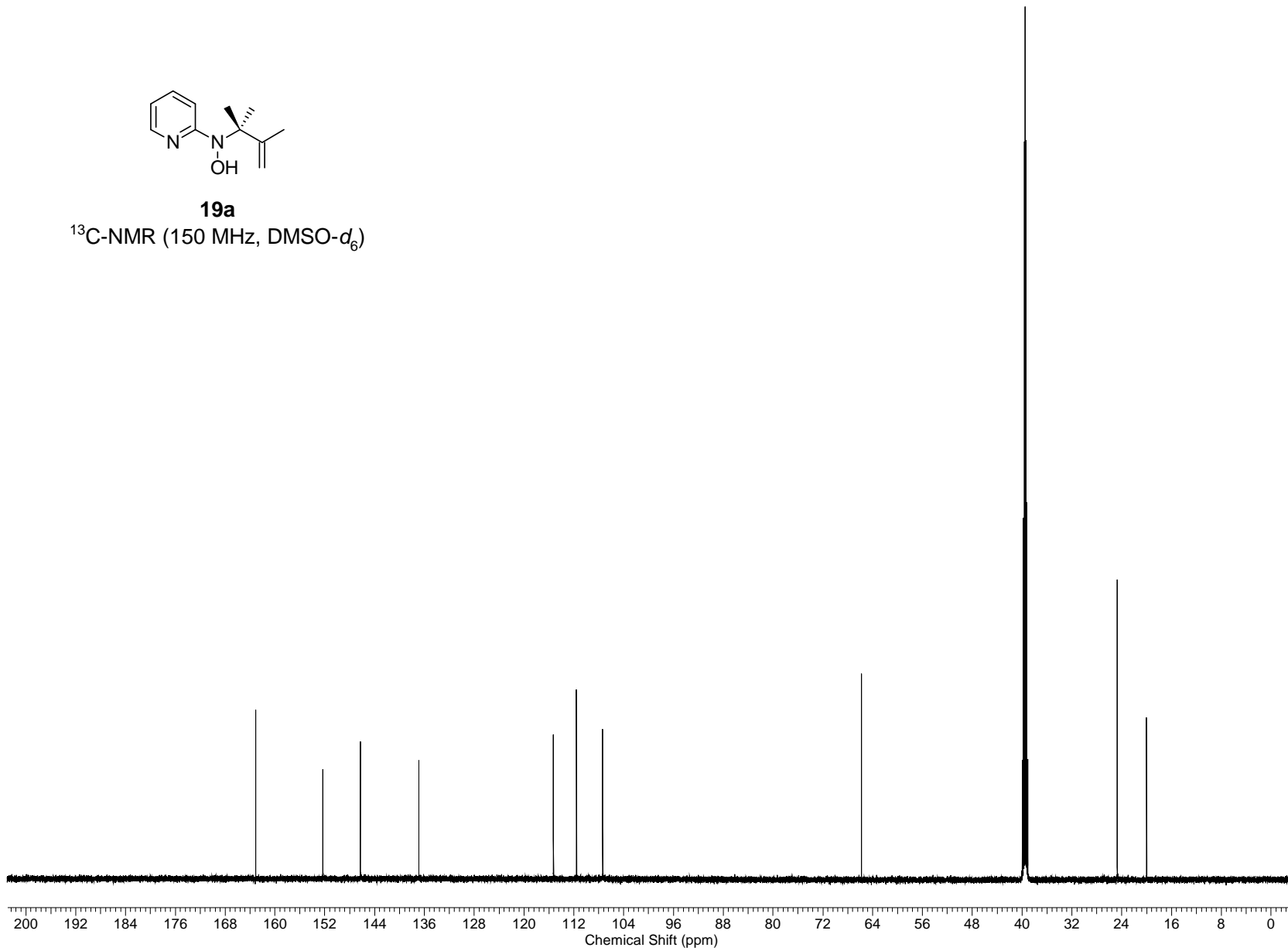
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

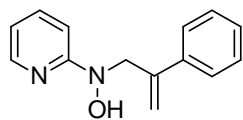




**19a**

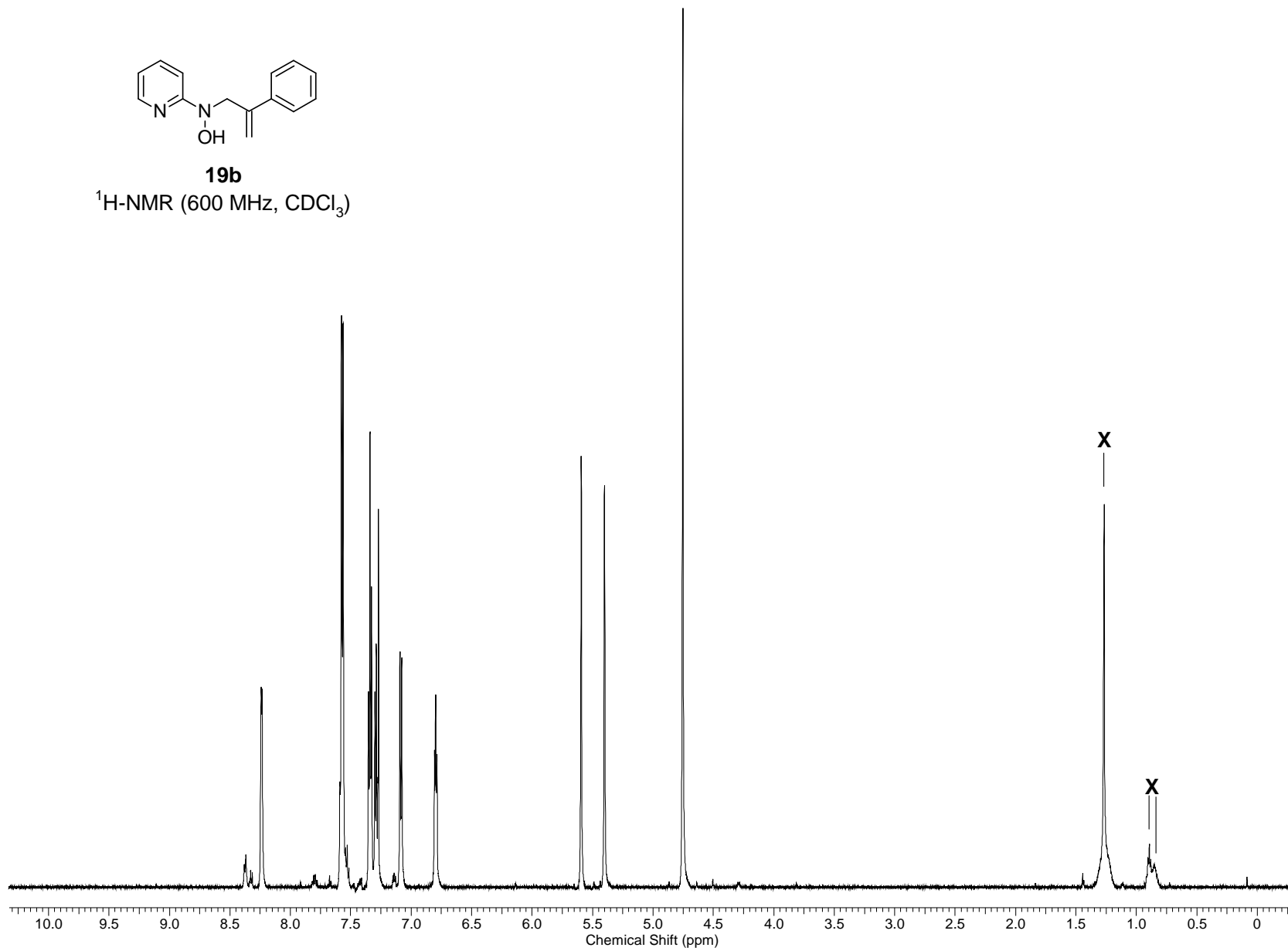
<sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)



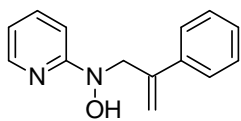


**19b**

<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

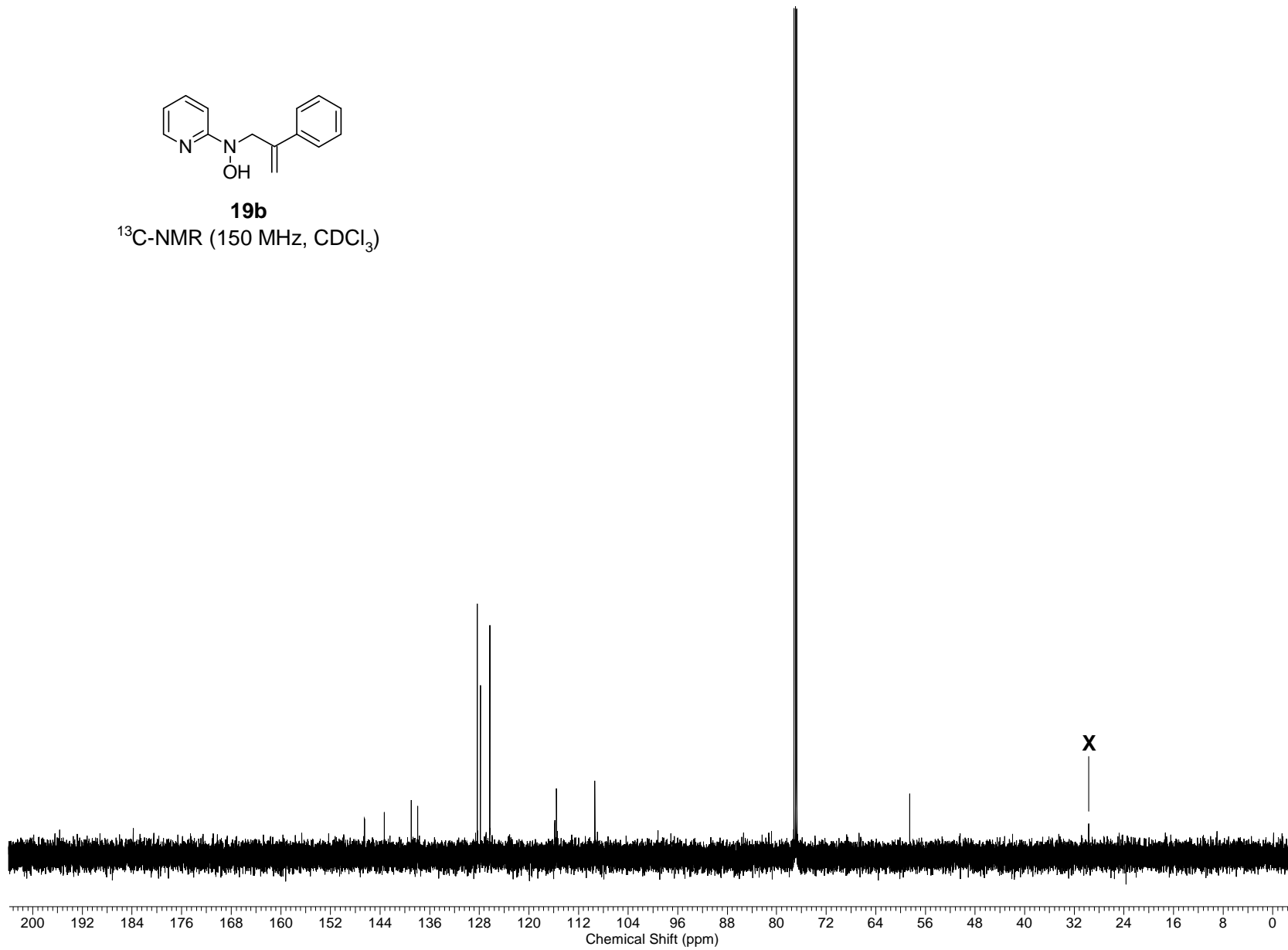


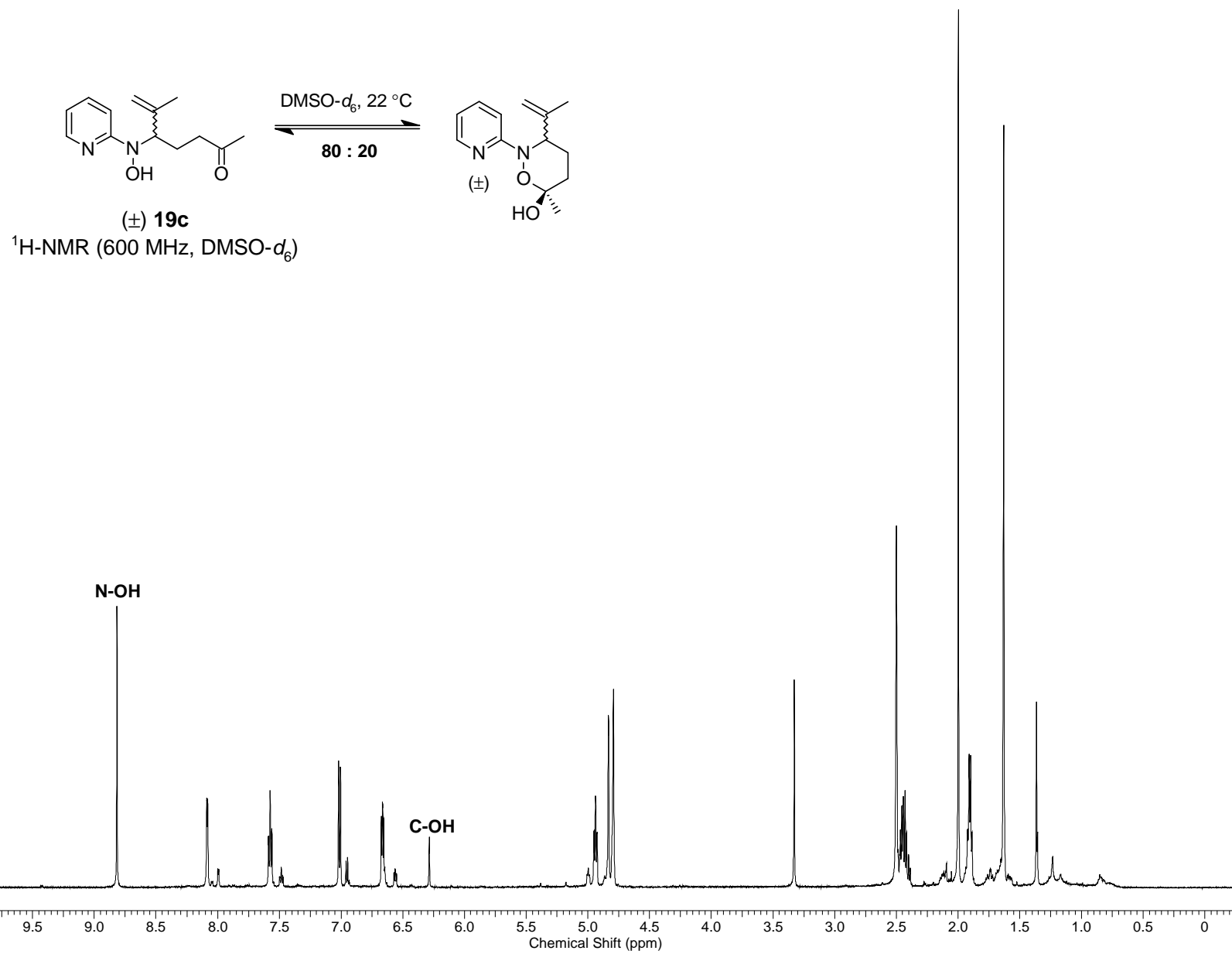


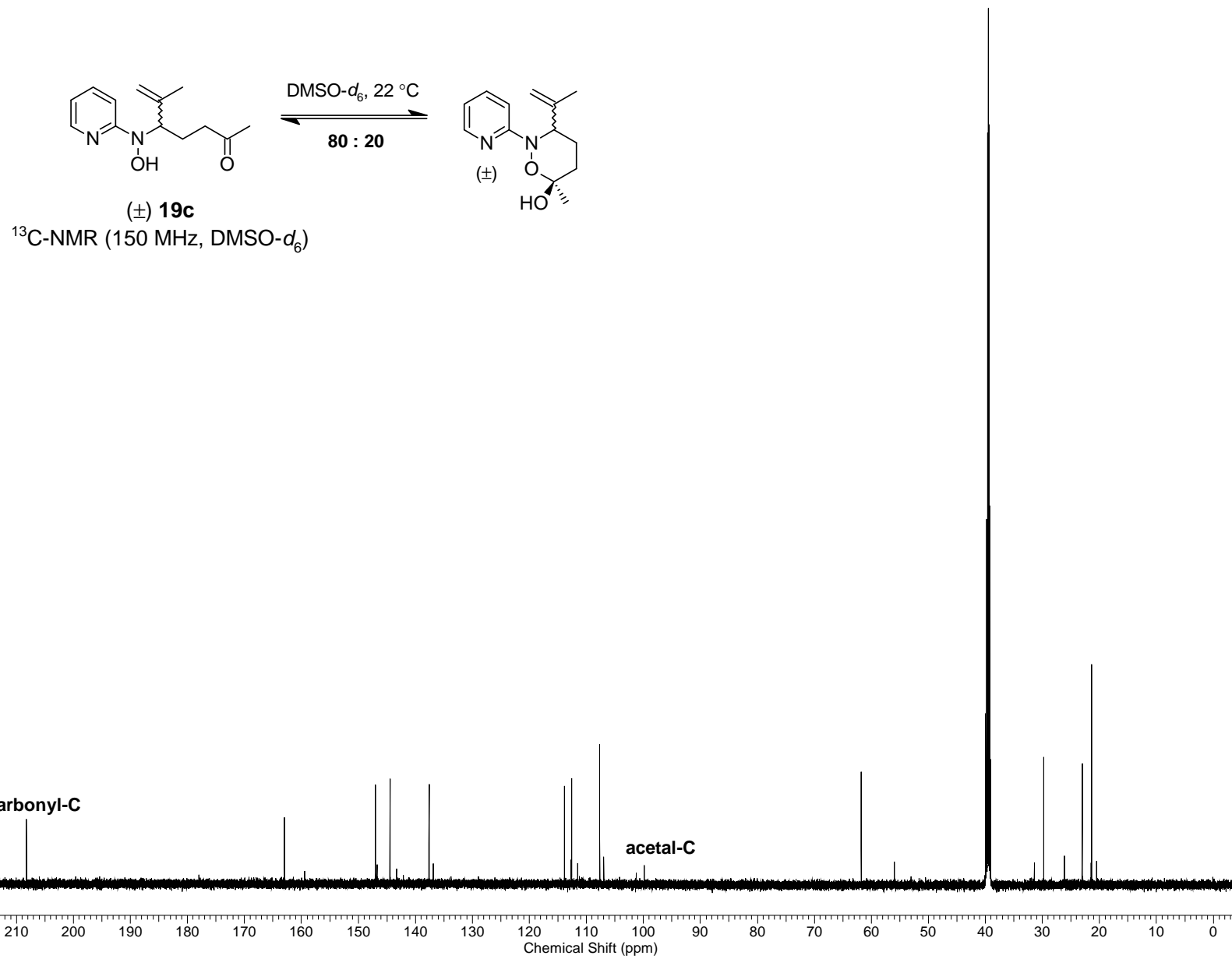


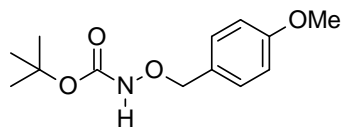
**19b**

$^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )



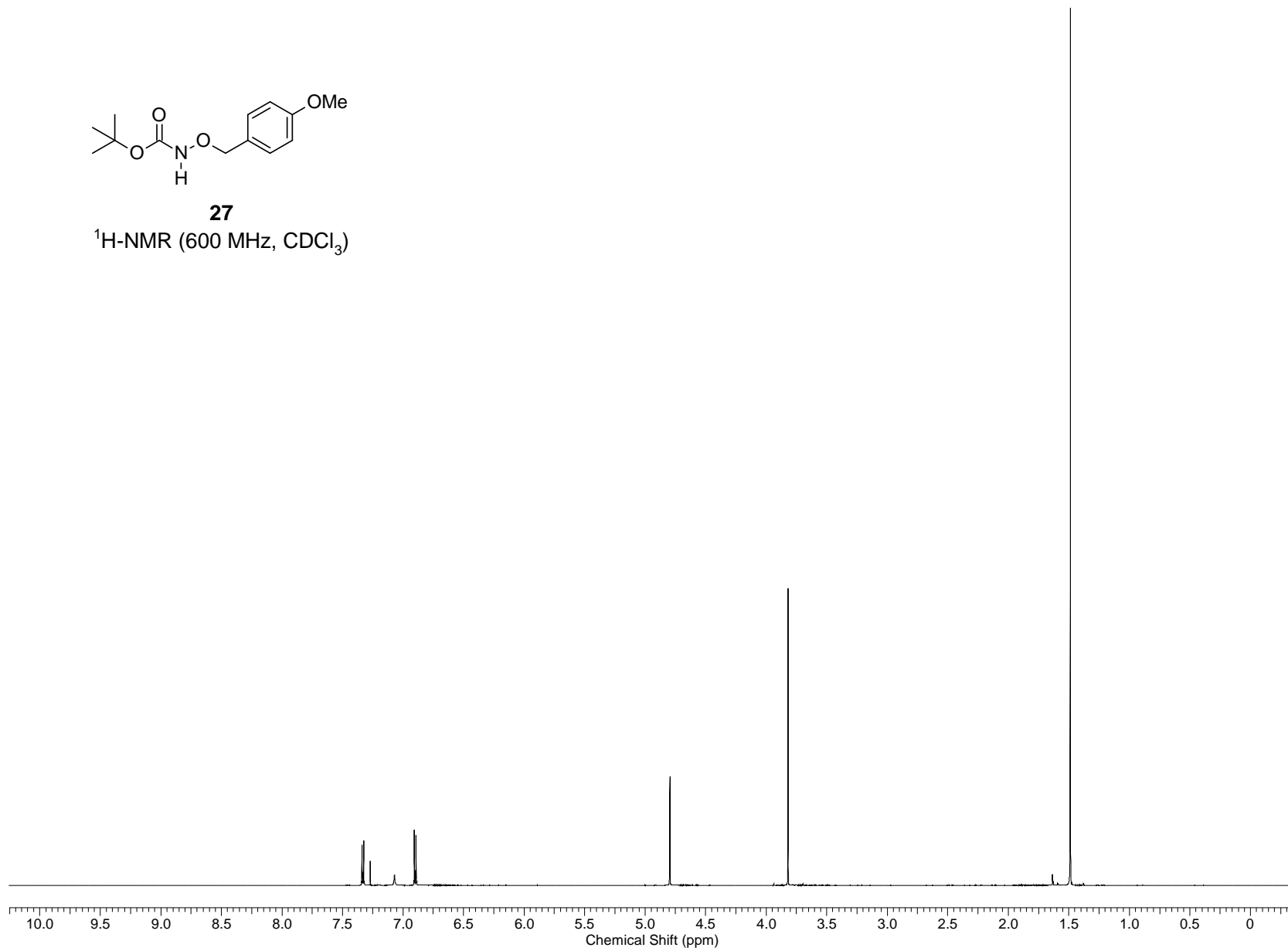


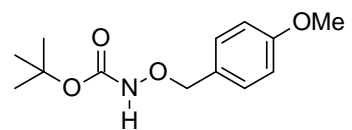




**27**

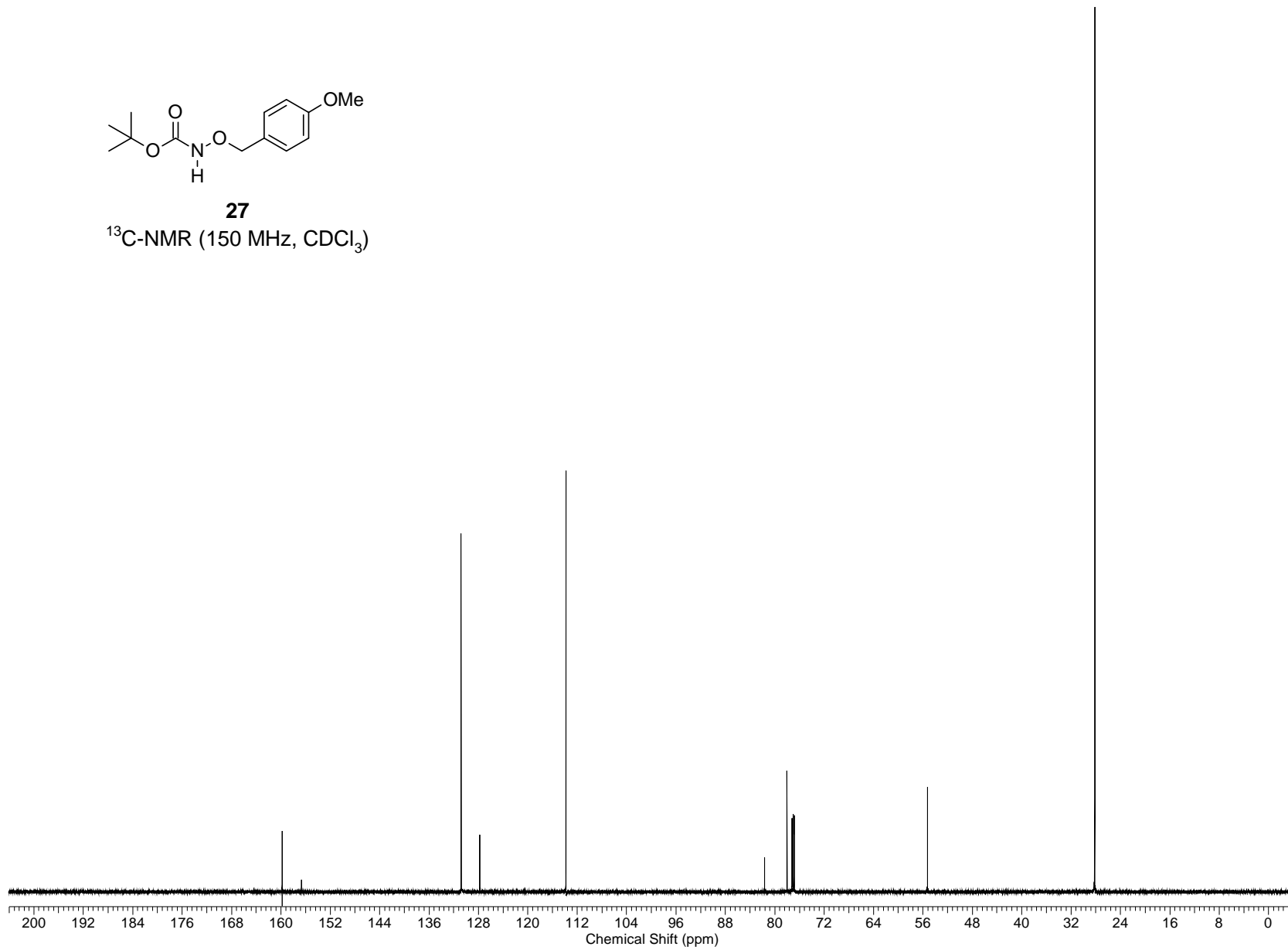
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

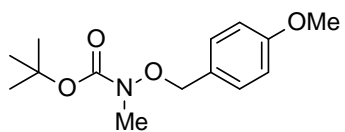




**27**

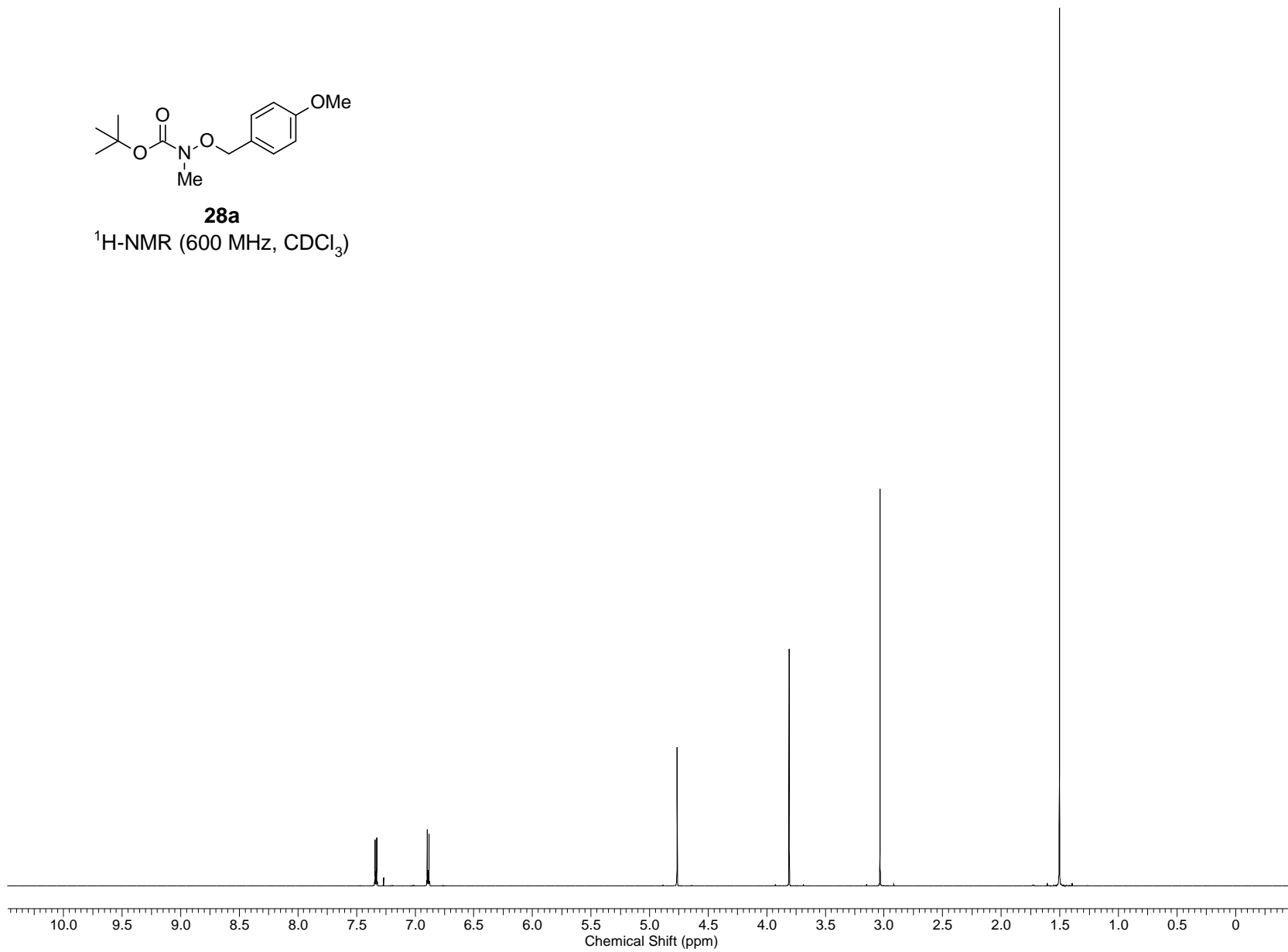
$^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )

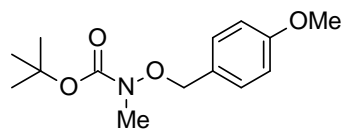




**28a**

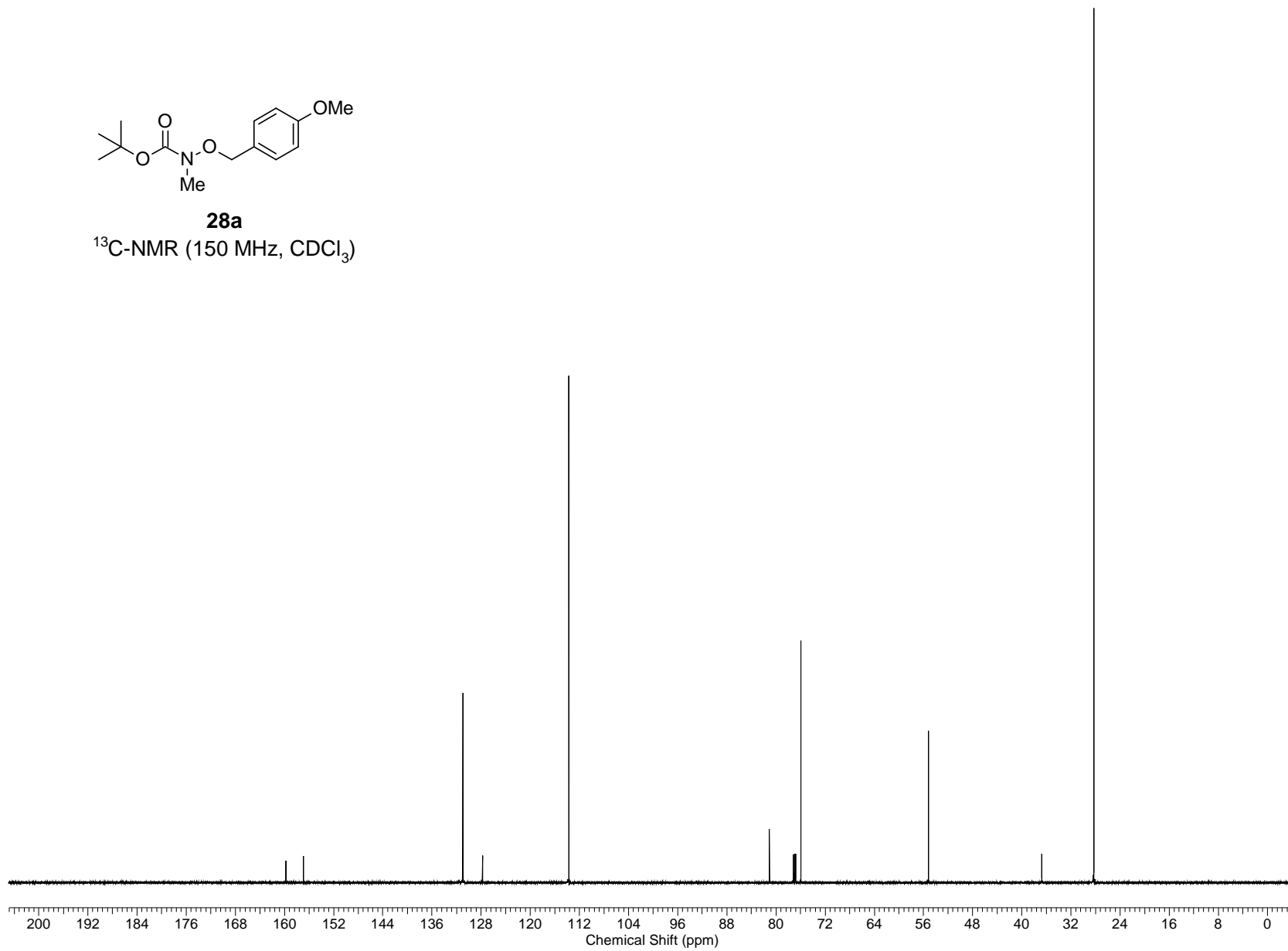
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

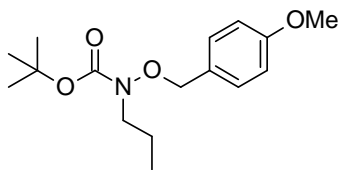




**28a**

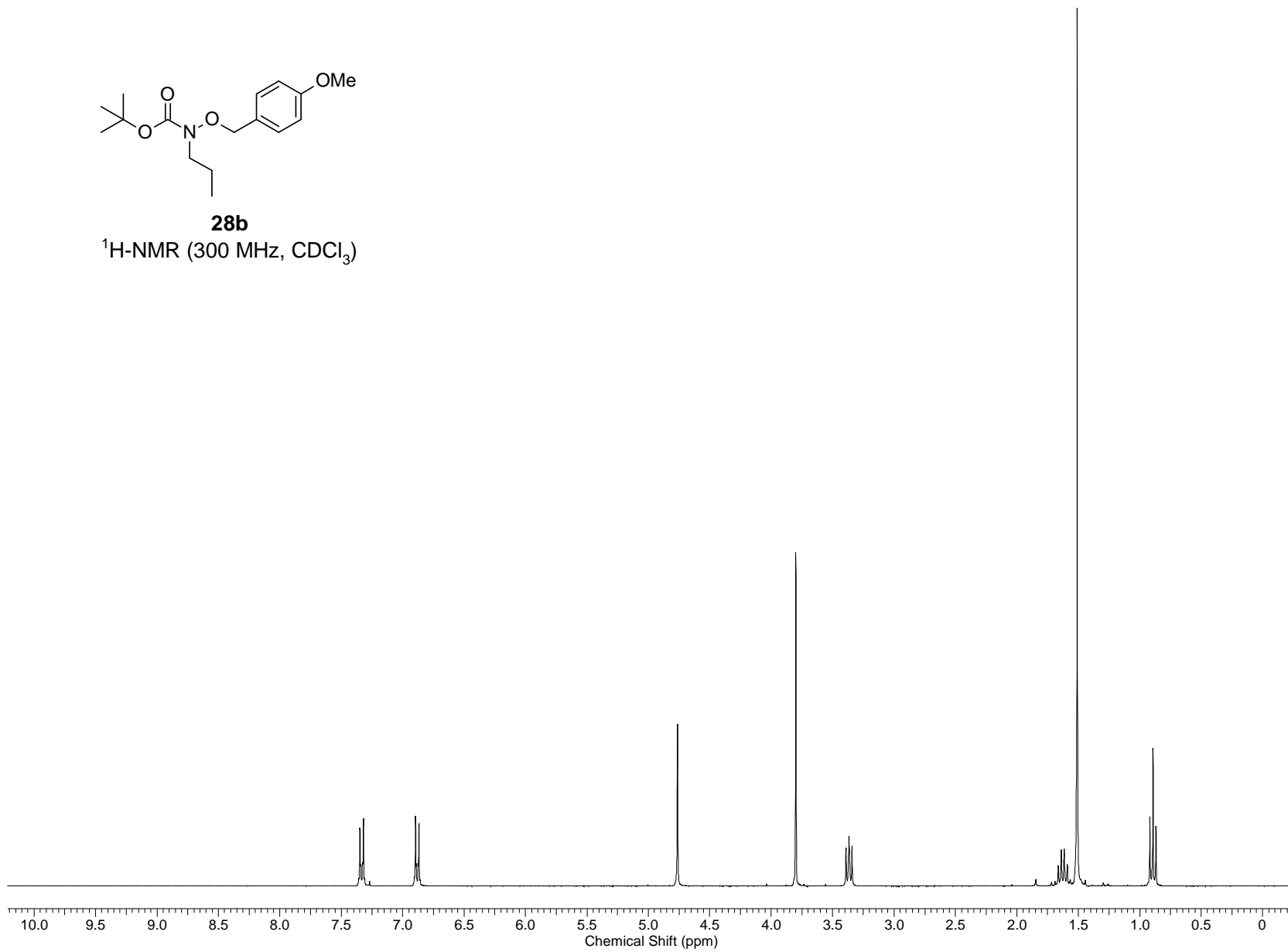
$^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )



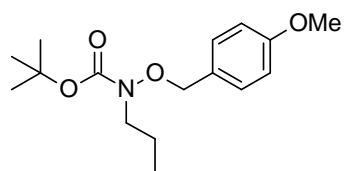


**28b**

<sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>)

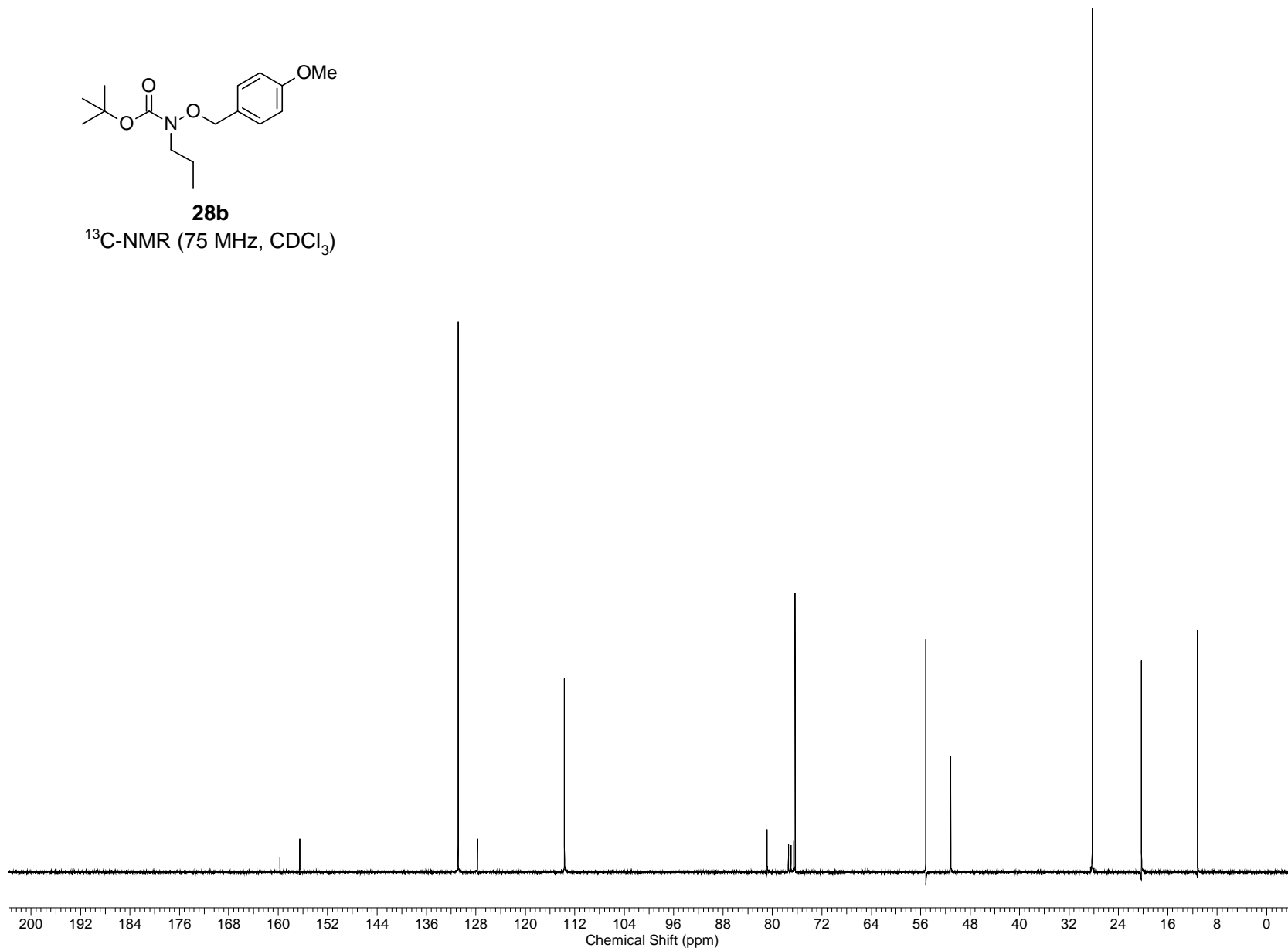


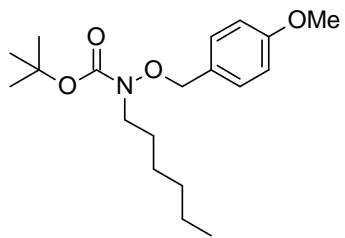




**28b**

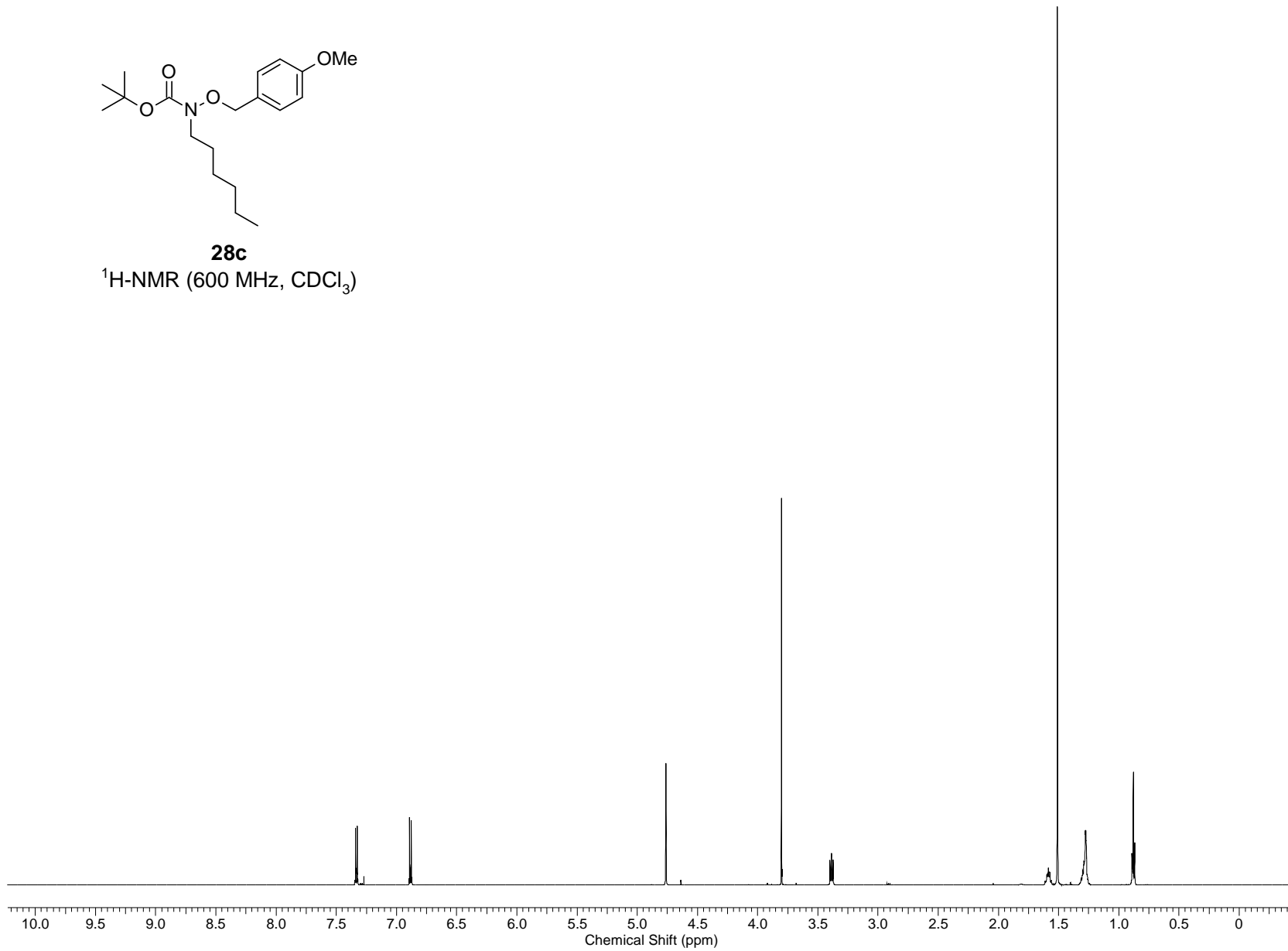
$^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )

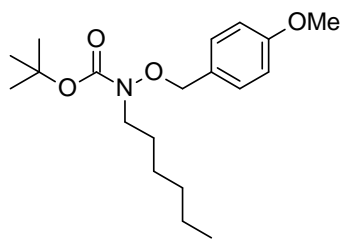




**28c**

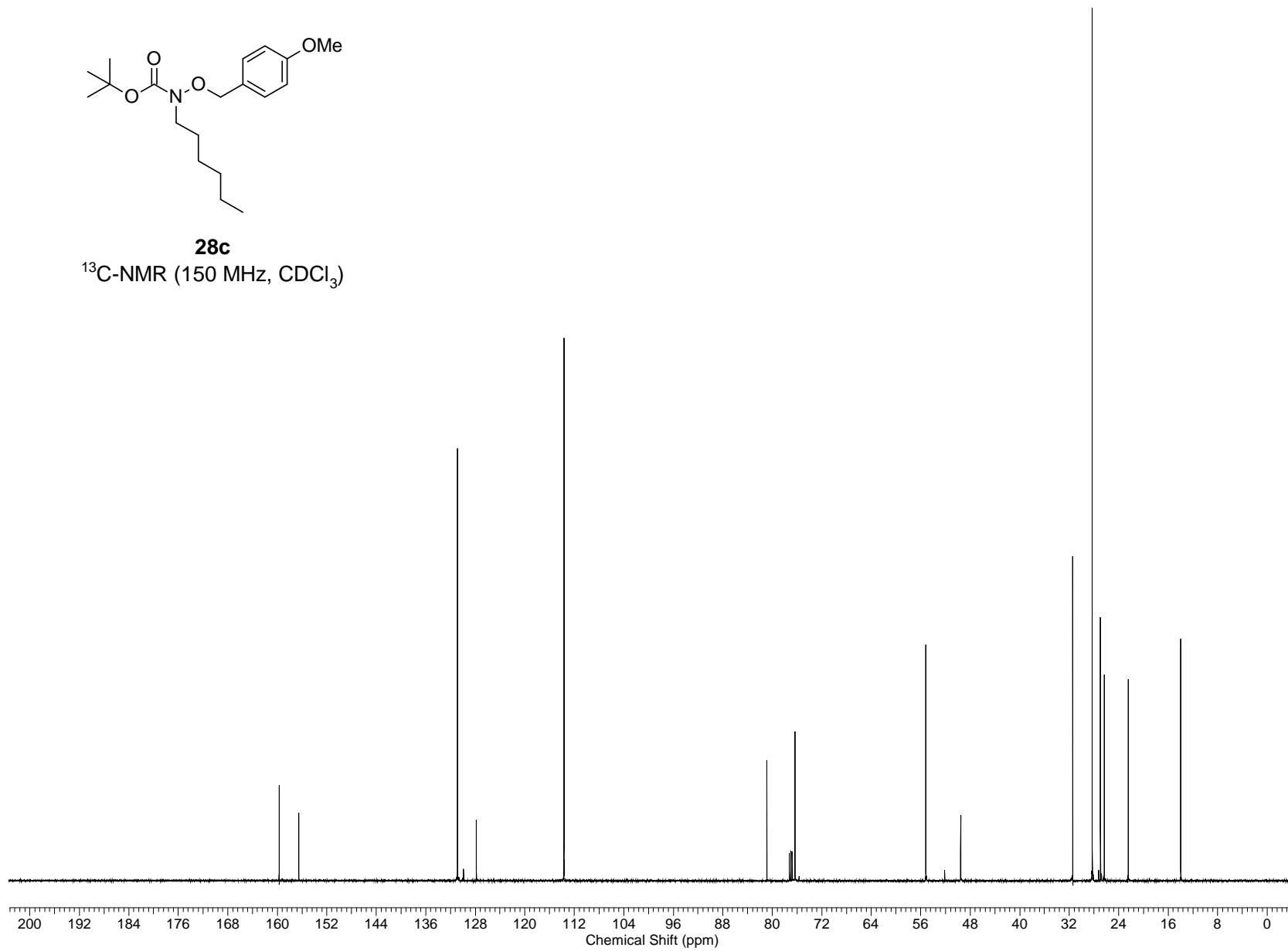
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

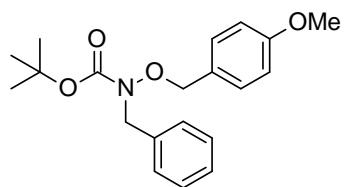




**28c**

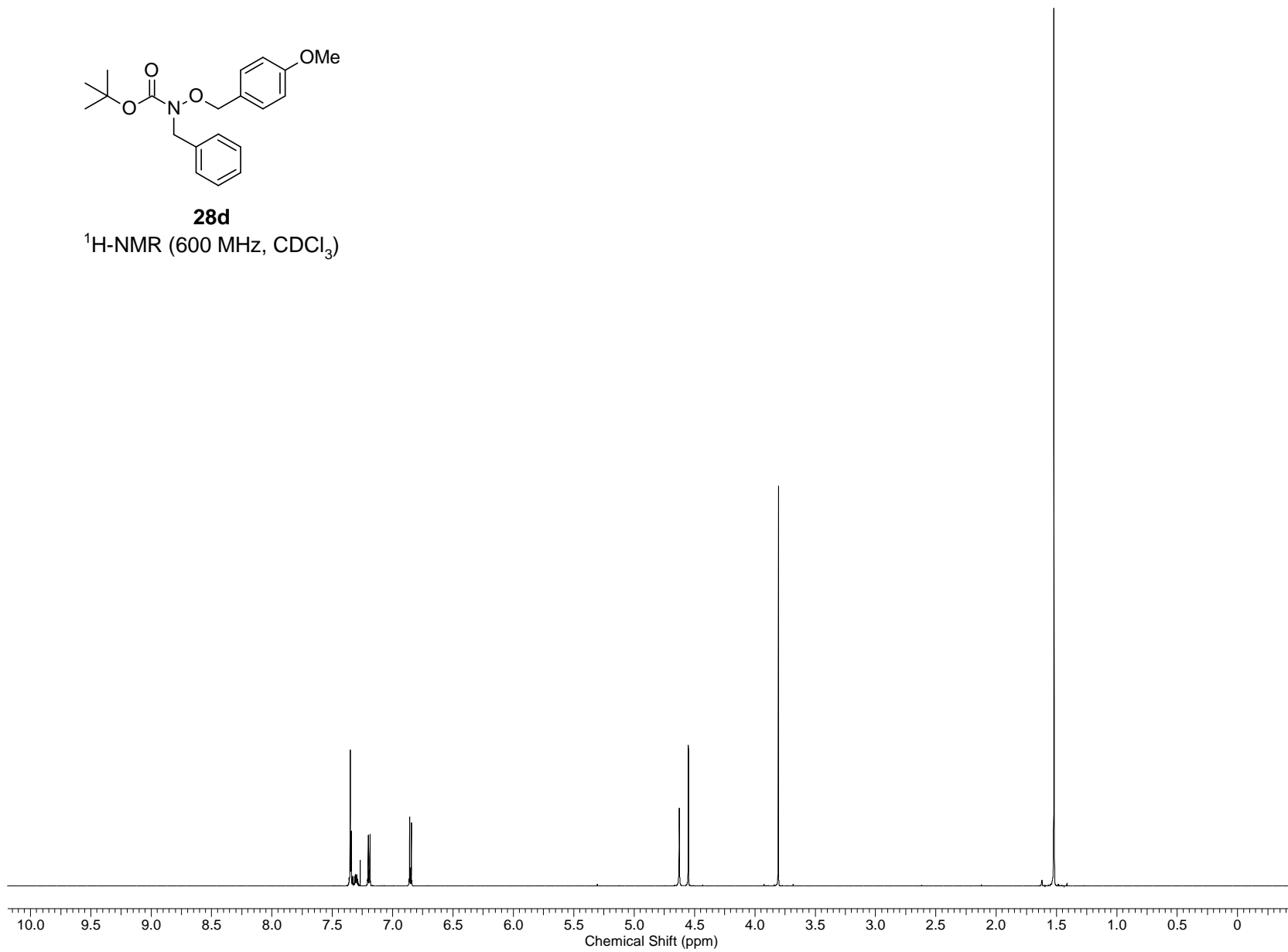
$^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )

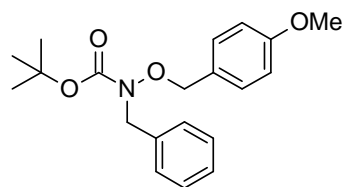




**28d**

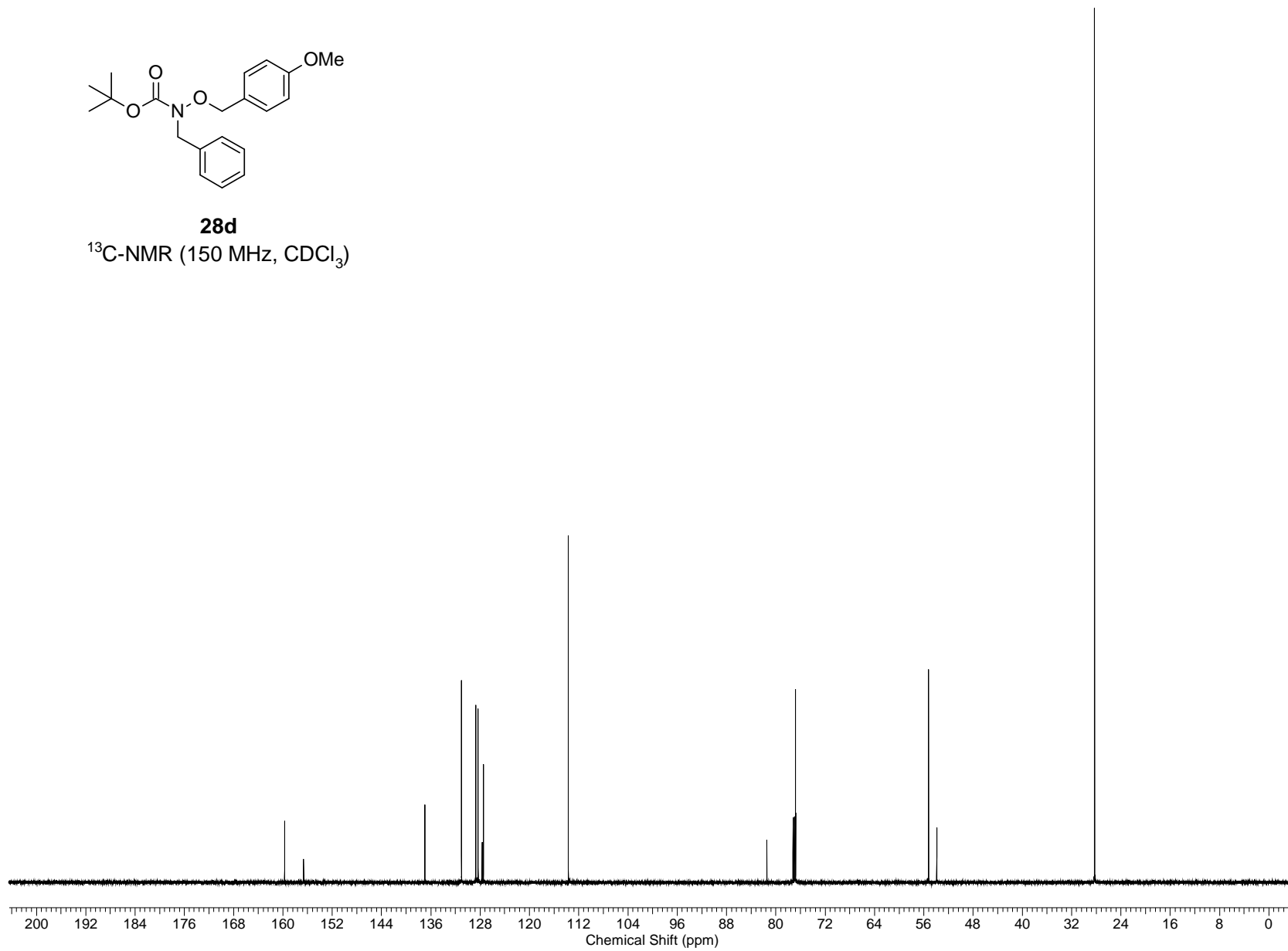
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

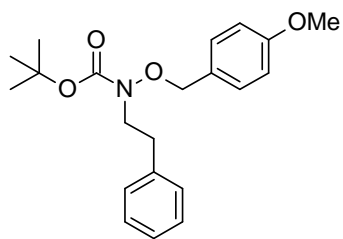




**28d**

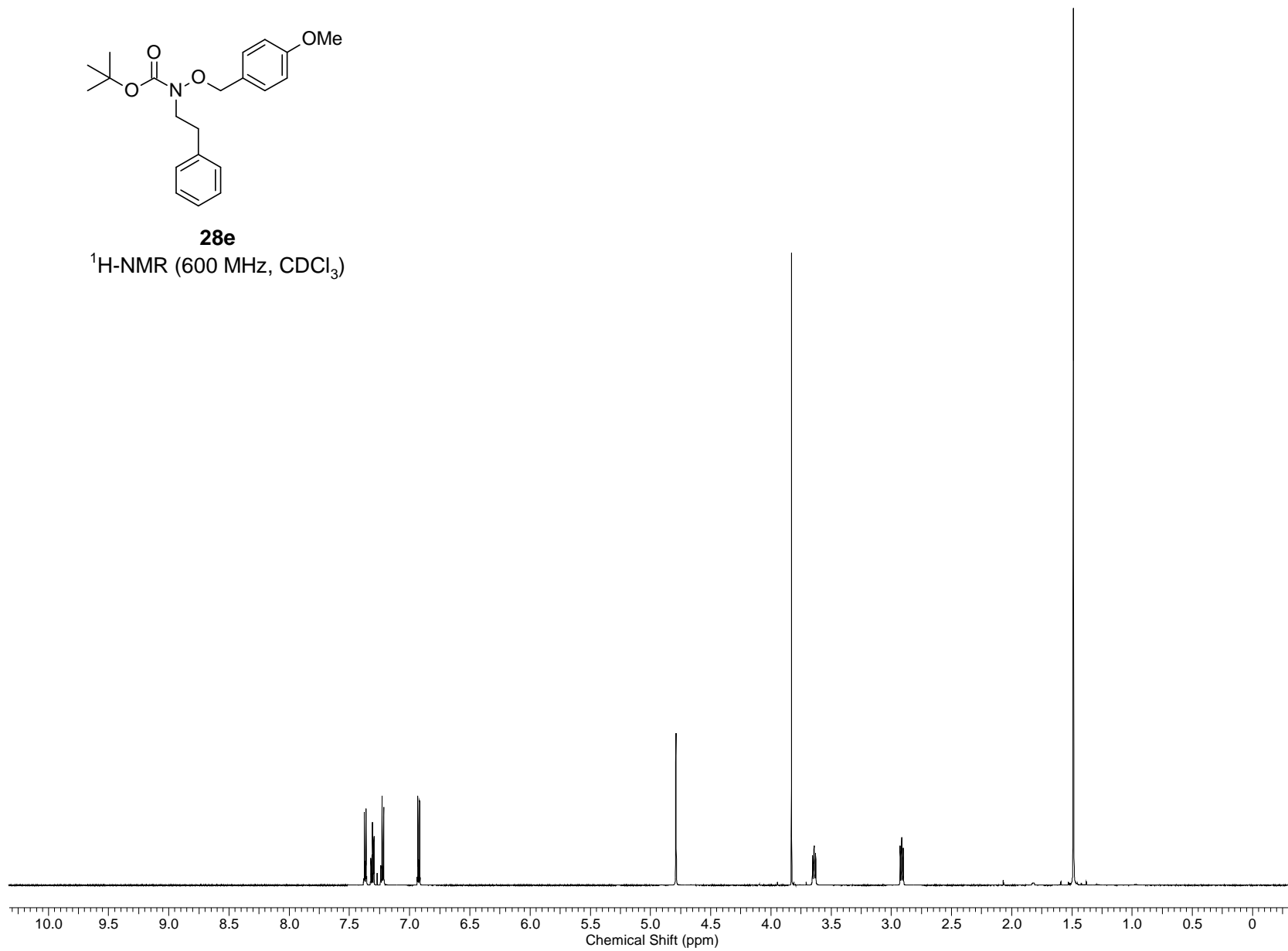
$^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )

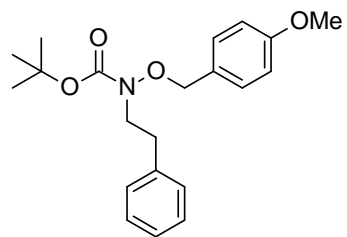




**28e**

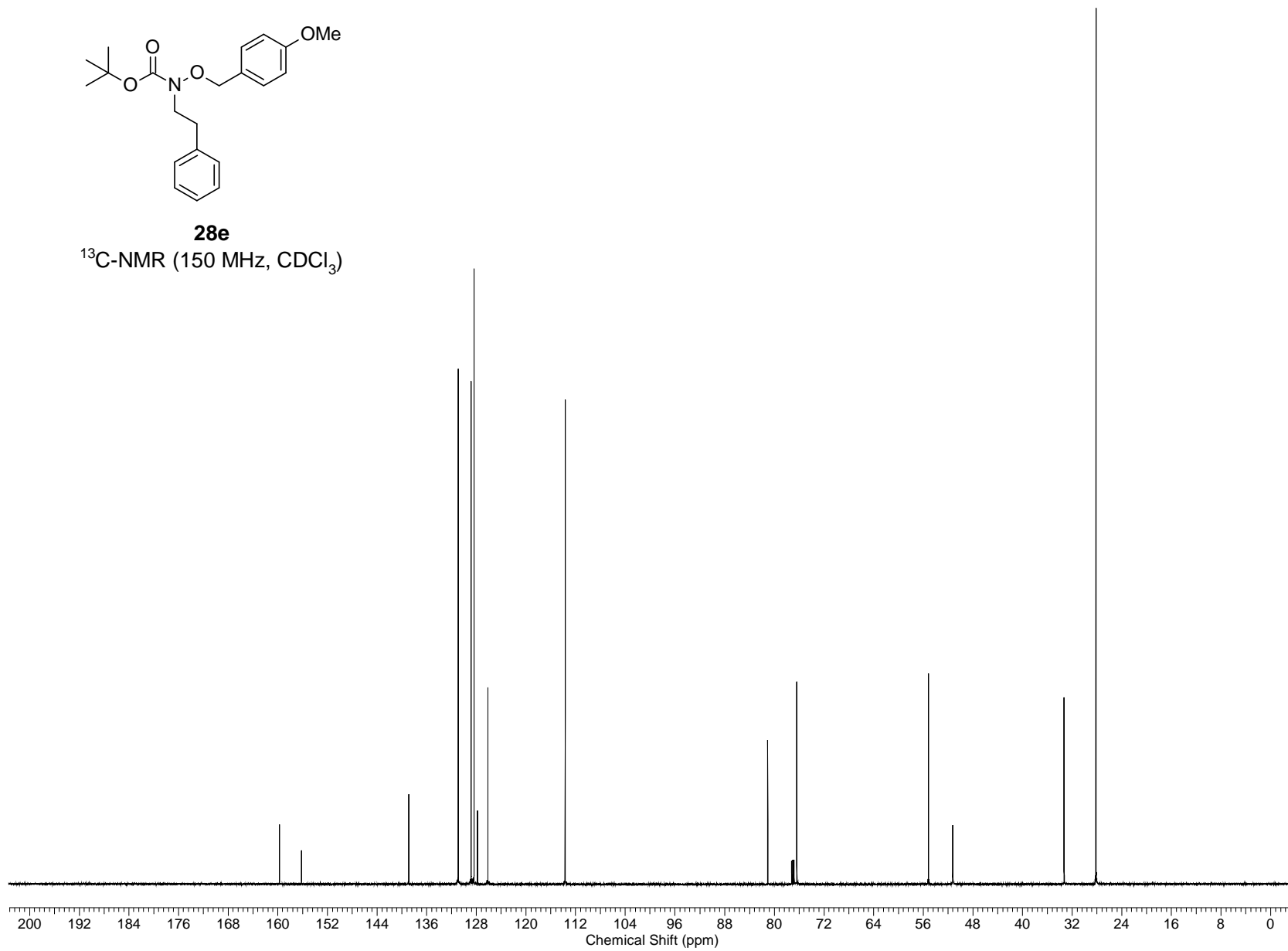
$^1\text{H-NMR}$  (600 MHz,  $\text{CDCl}_3$ )

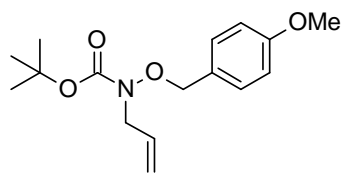




**28e**

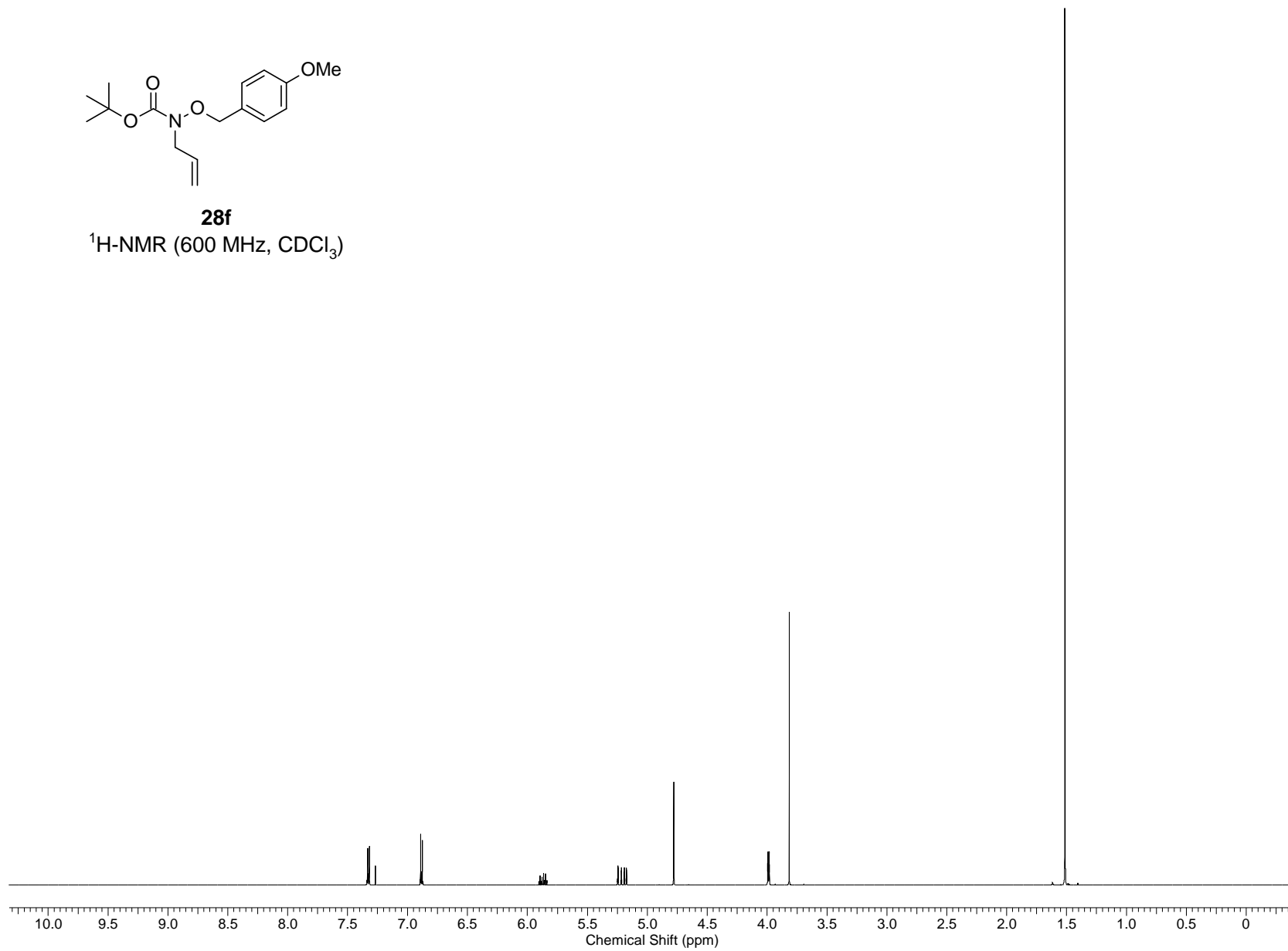
$^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )



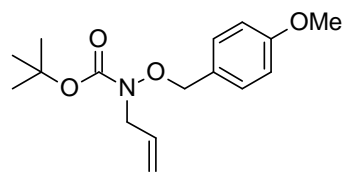


**28f**

<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

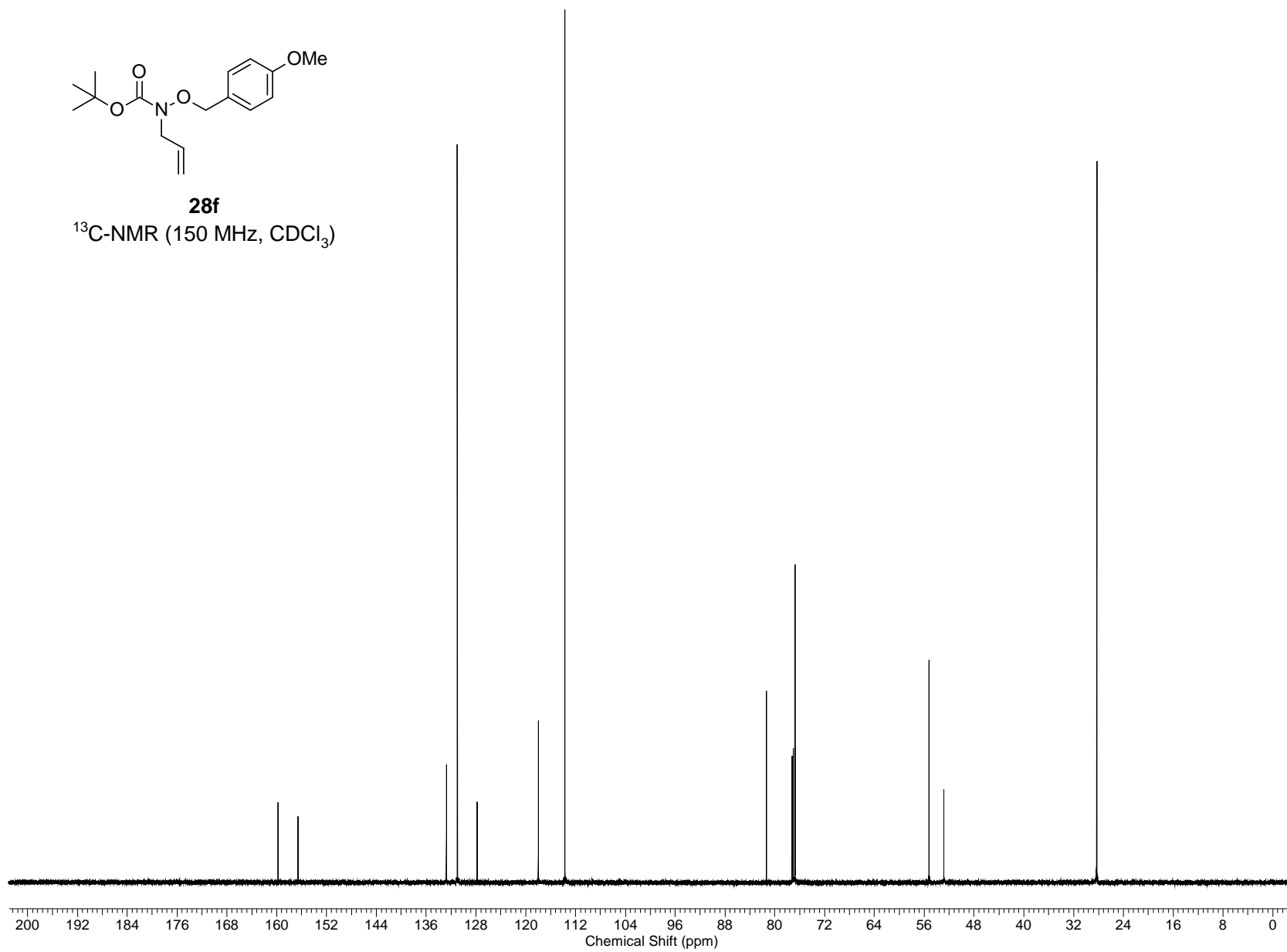


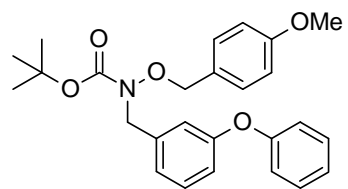




**28f**

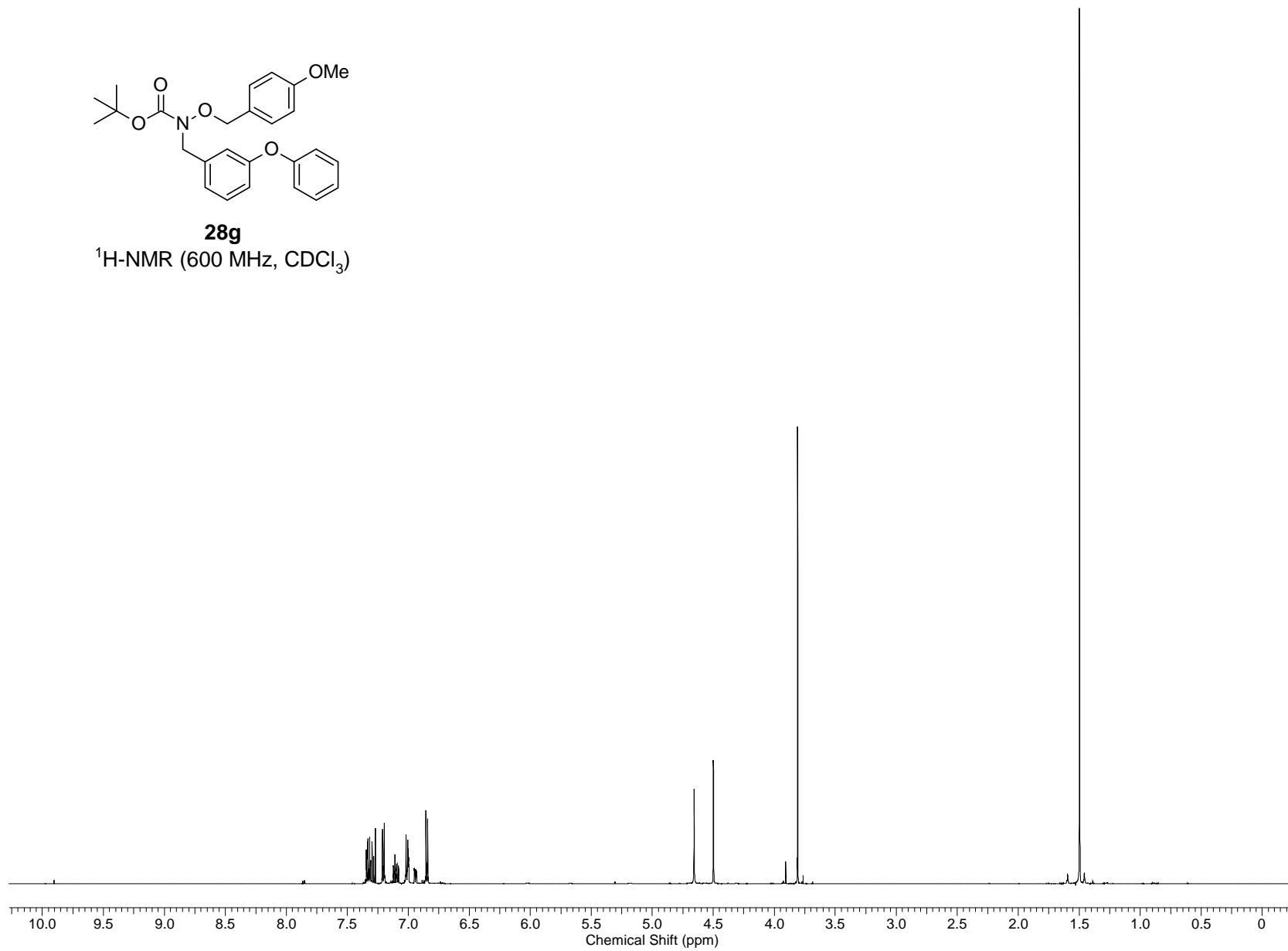
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

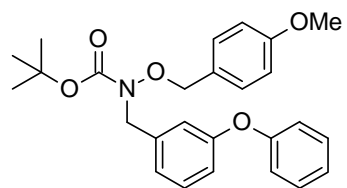




**28g**

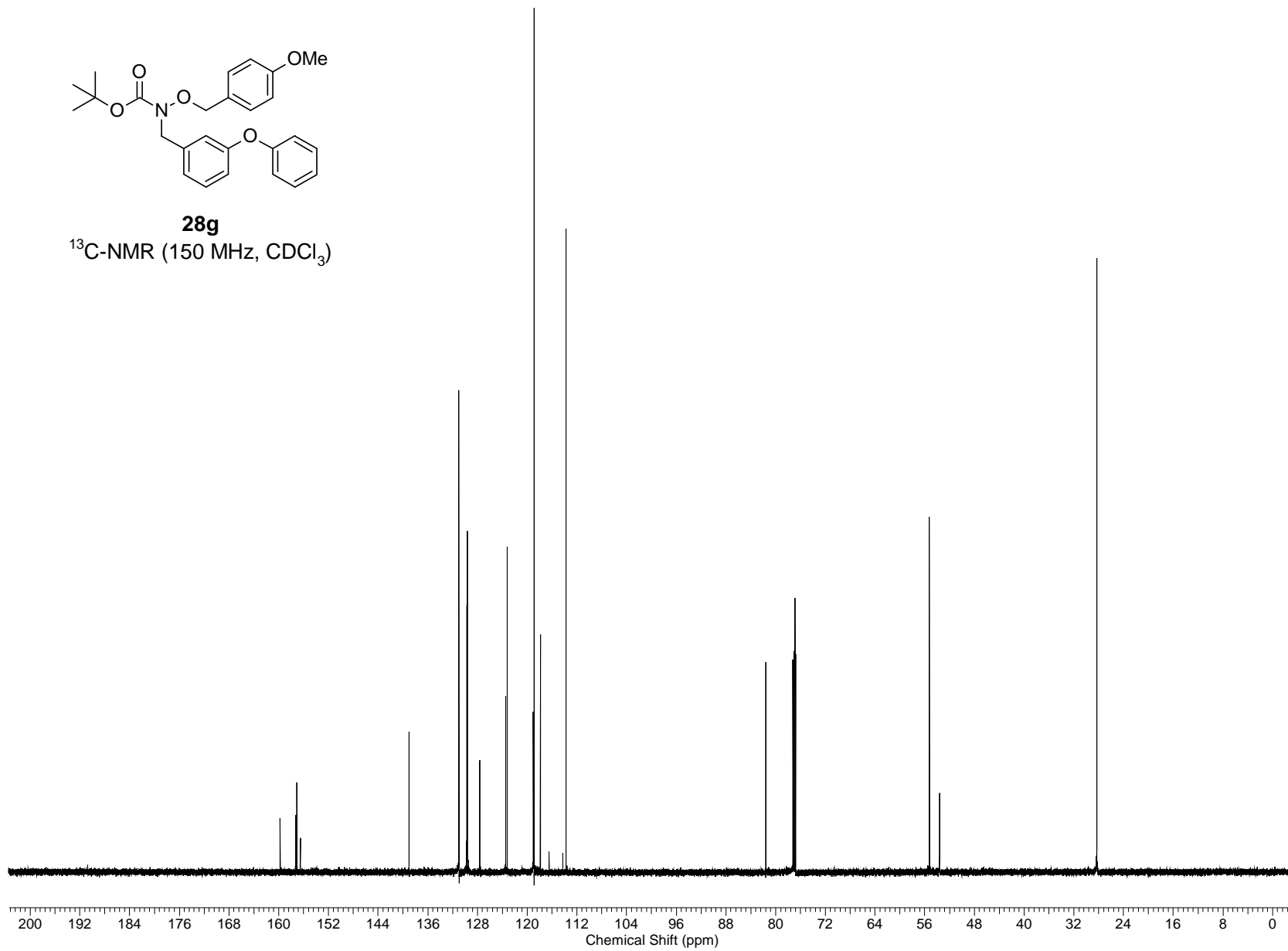
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

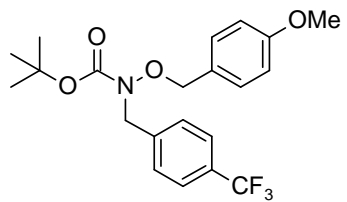




**28g**

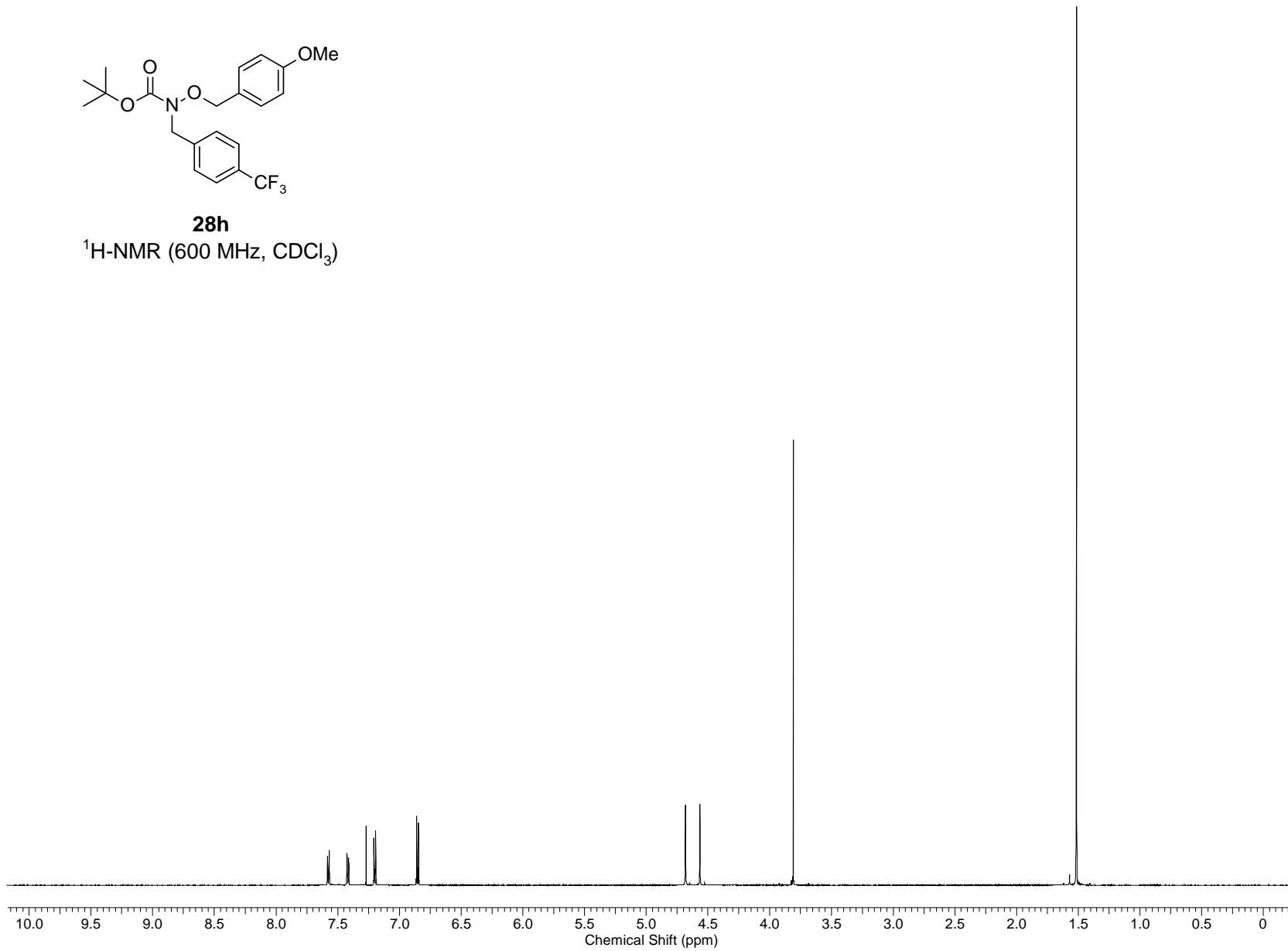
$^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )

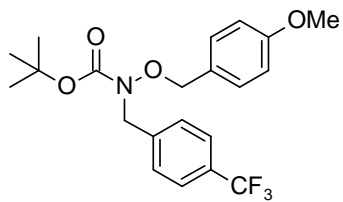




**28h**

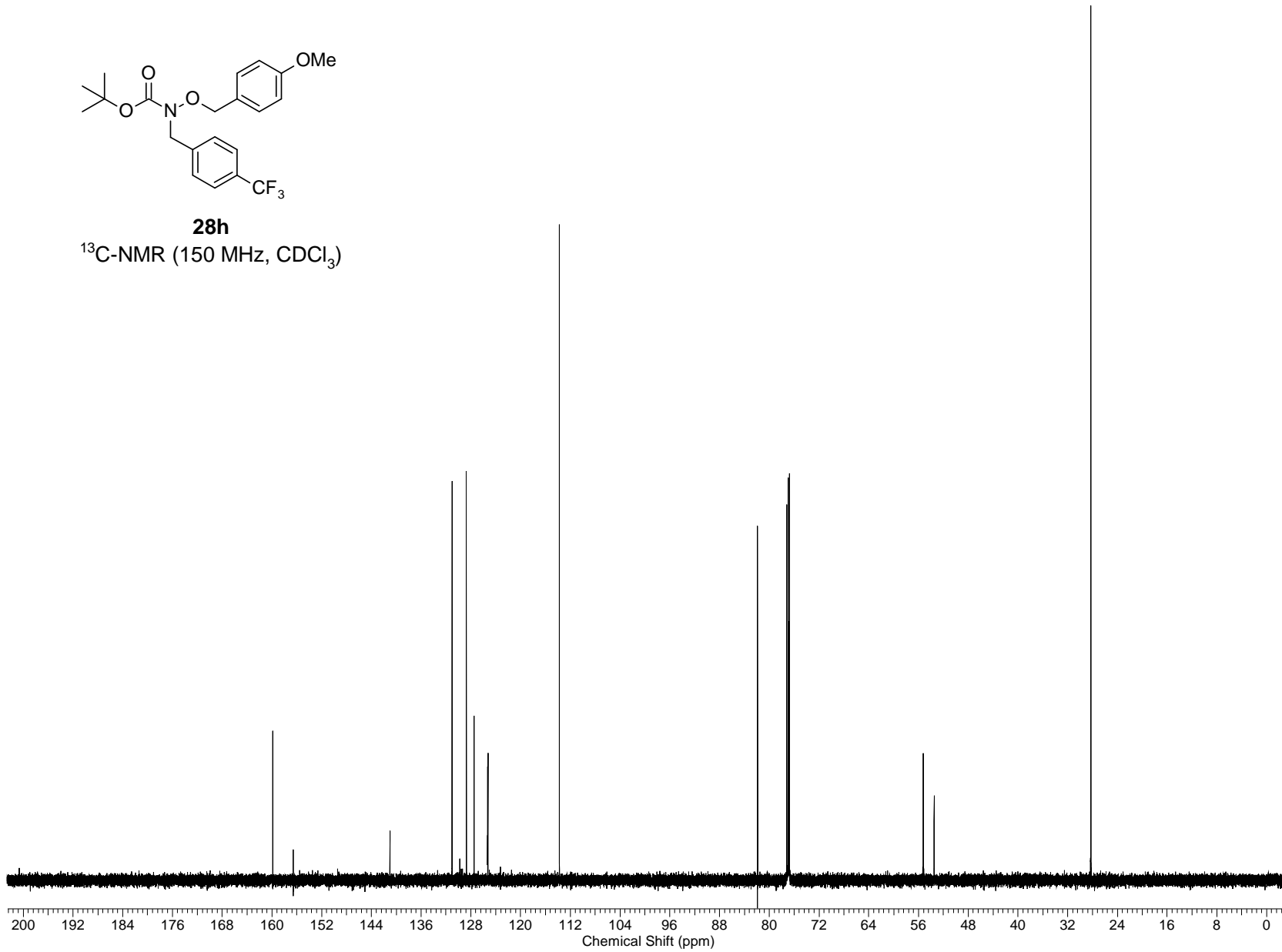
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

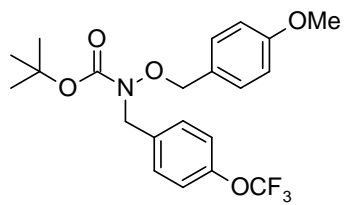




**28h**

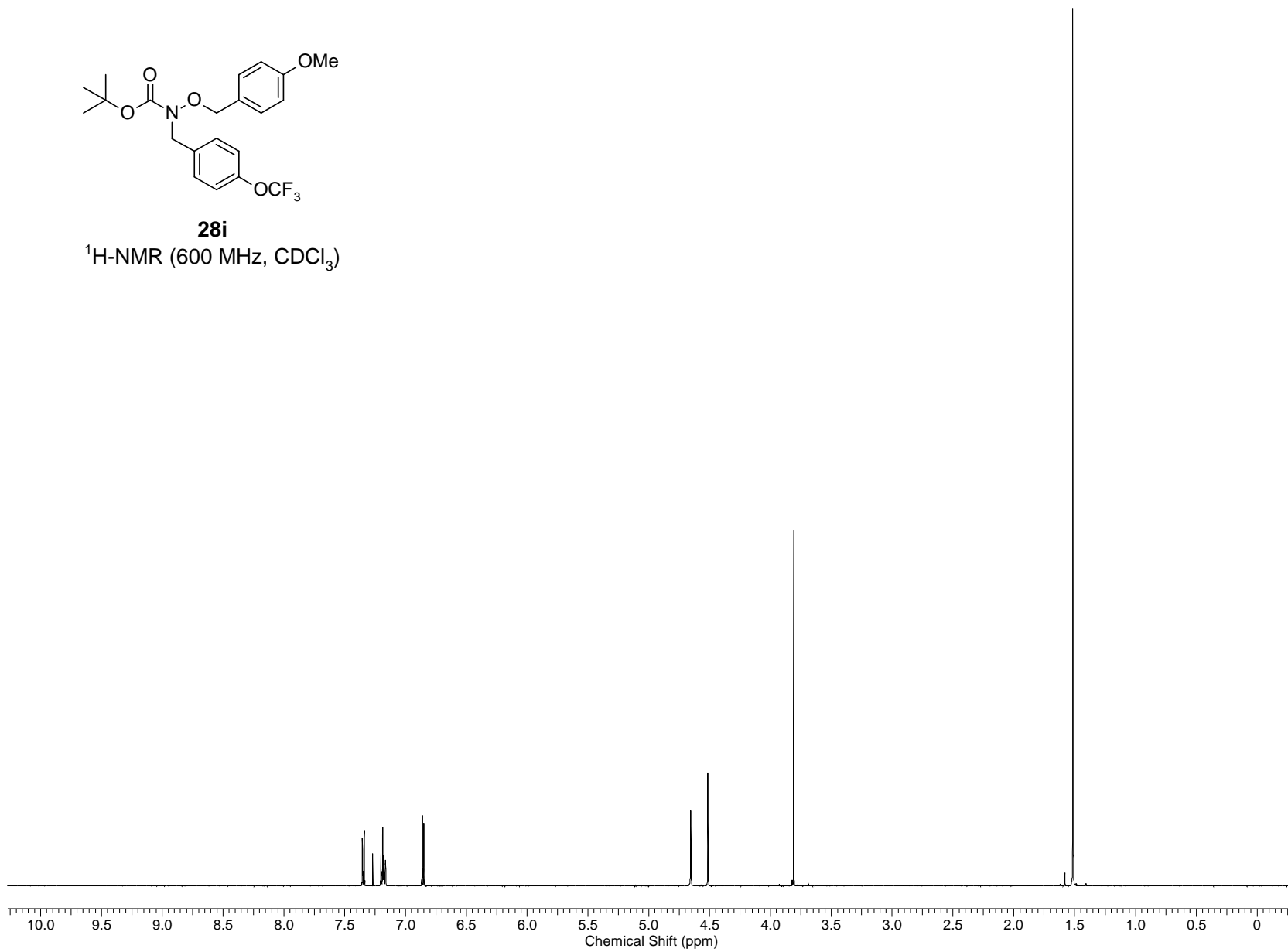
$^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )

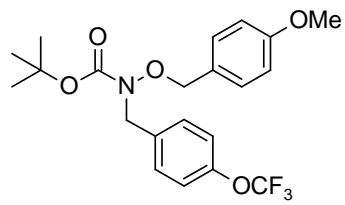




**28i**

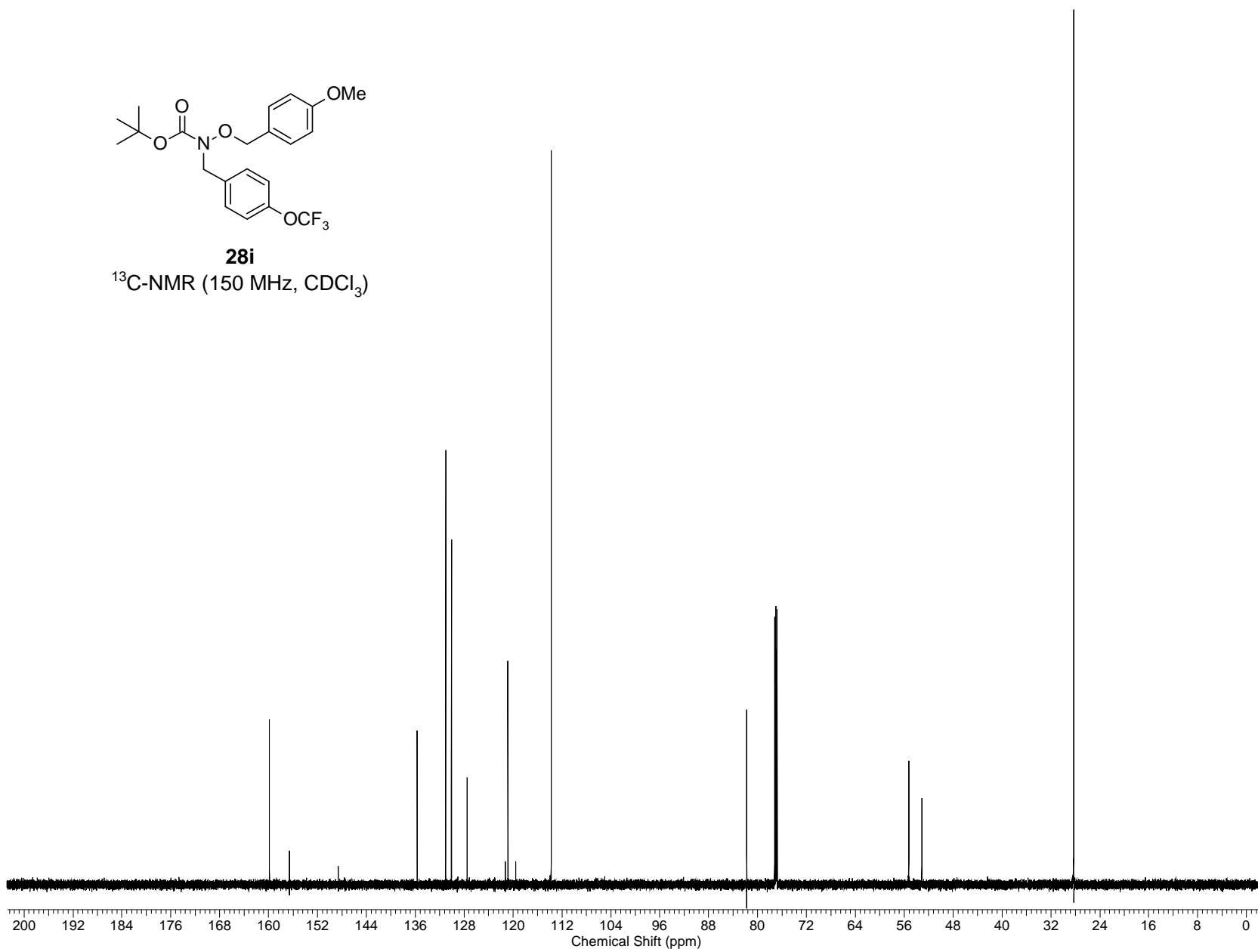
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

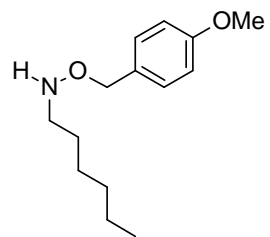




**28i**

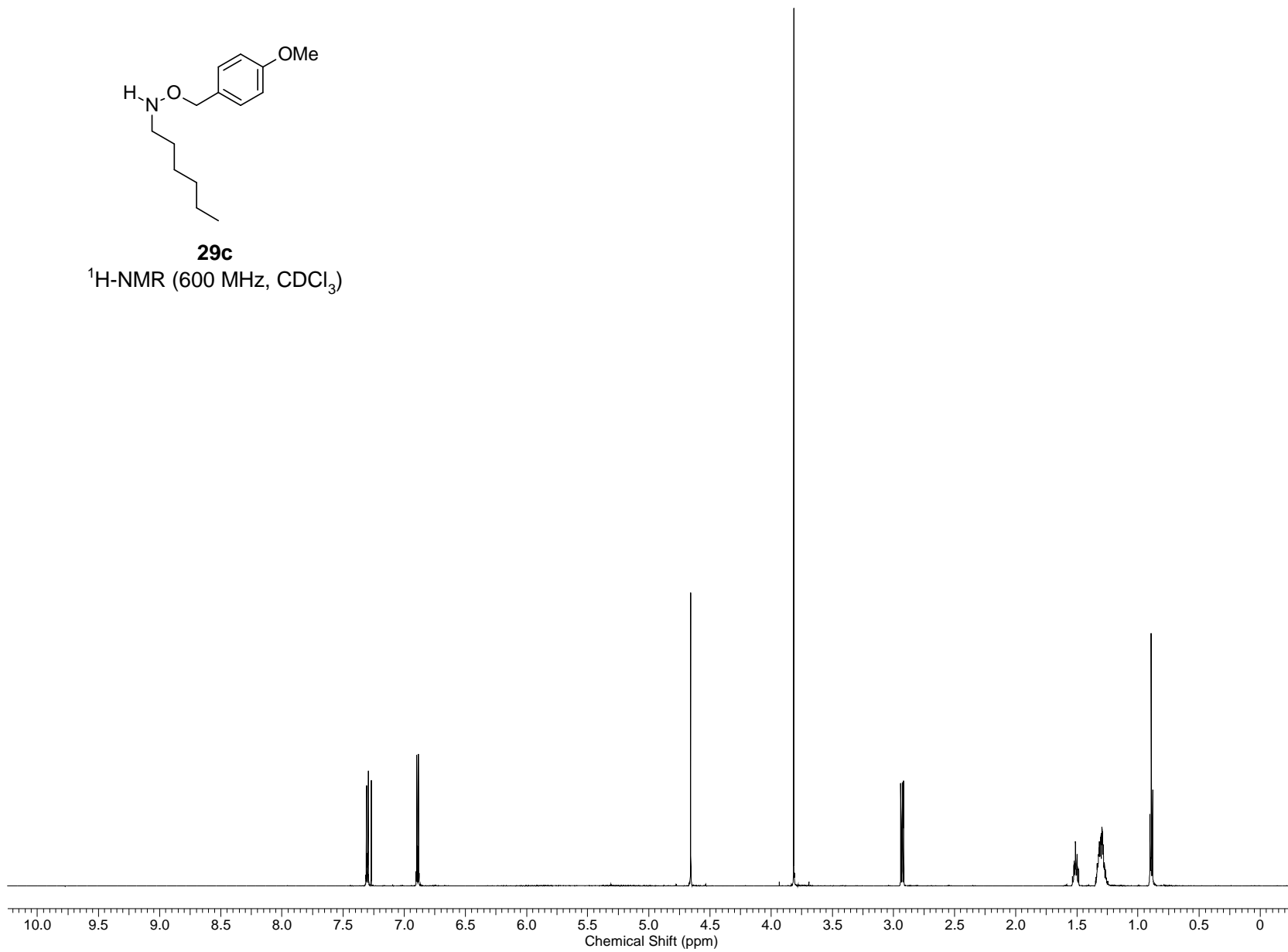
$^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )



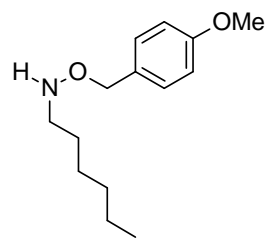


**29c**

<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

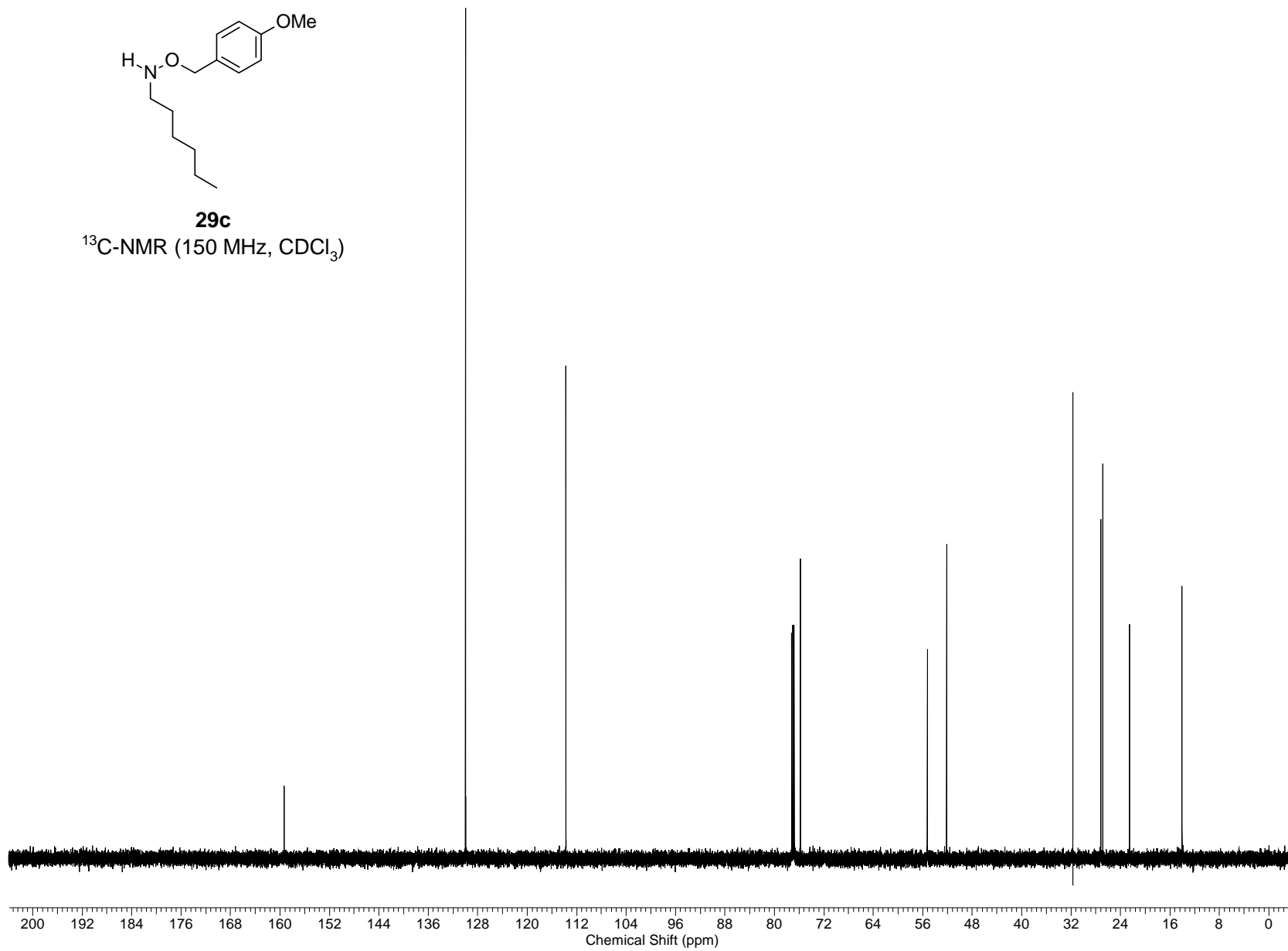


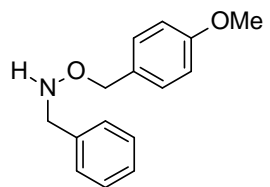




**29c**

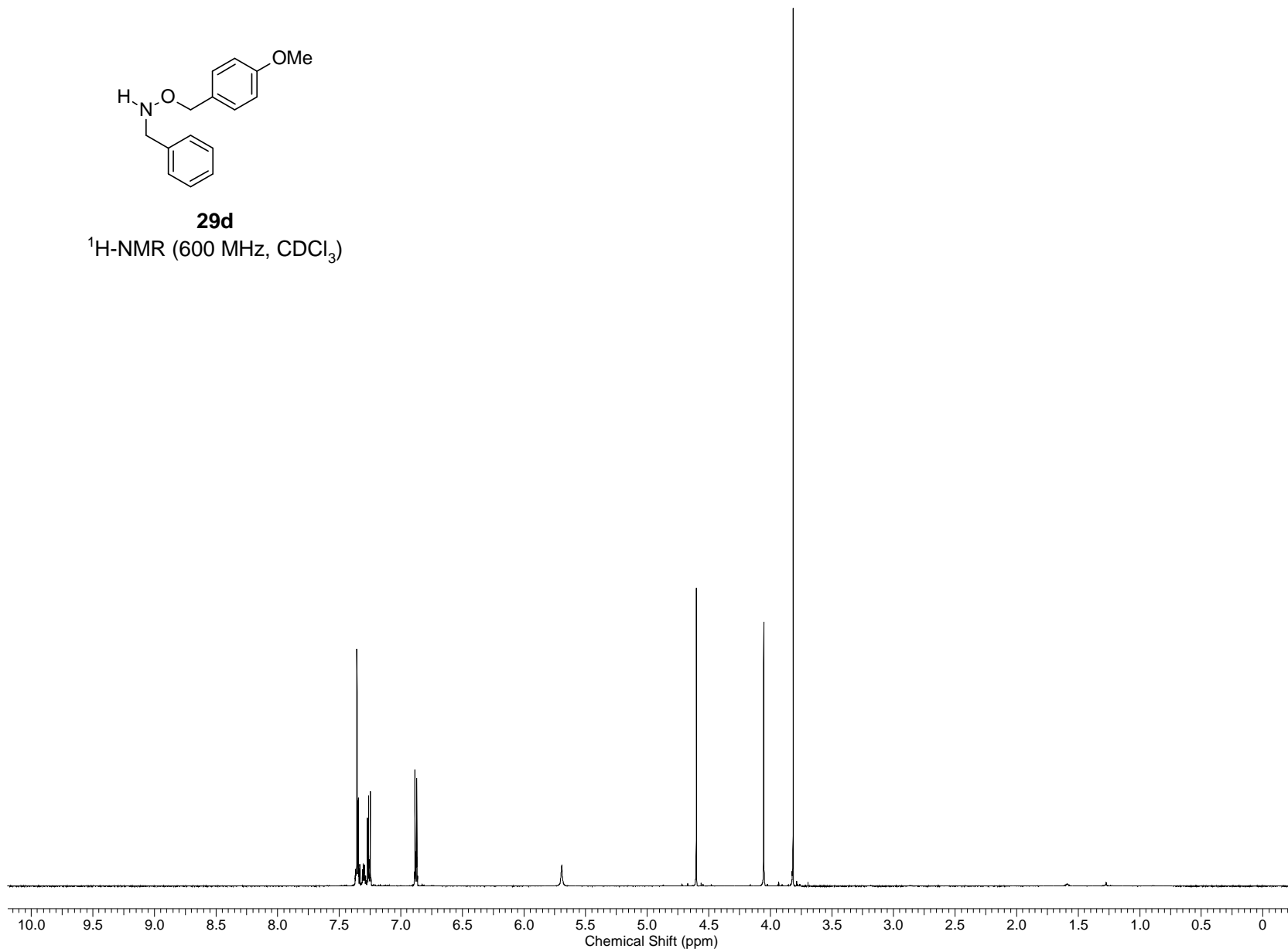
$^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )

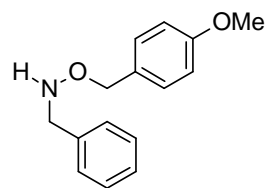




**29d**

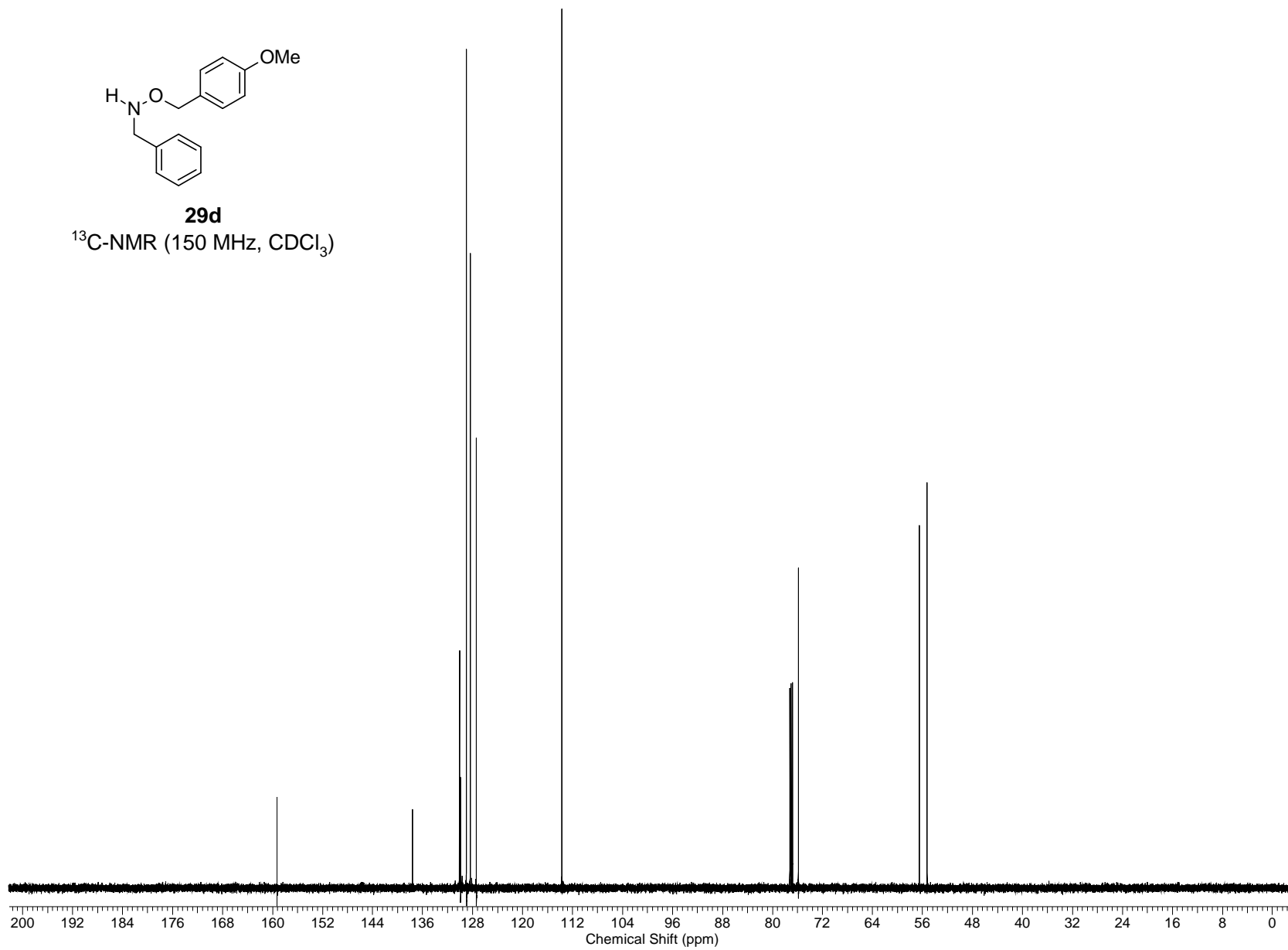
$^1\text{H-NMR}$  (600 MHz,  $\text{CDCl}_3$ )

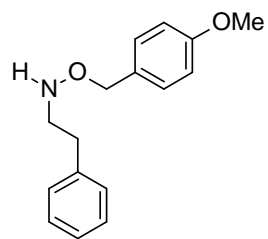




**29d**

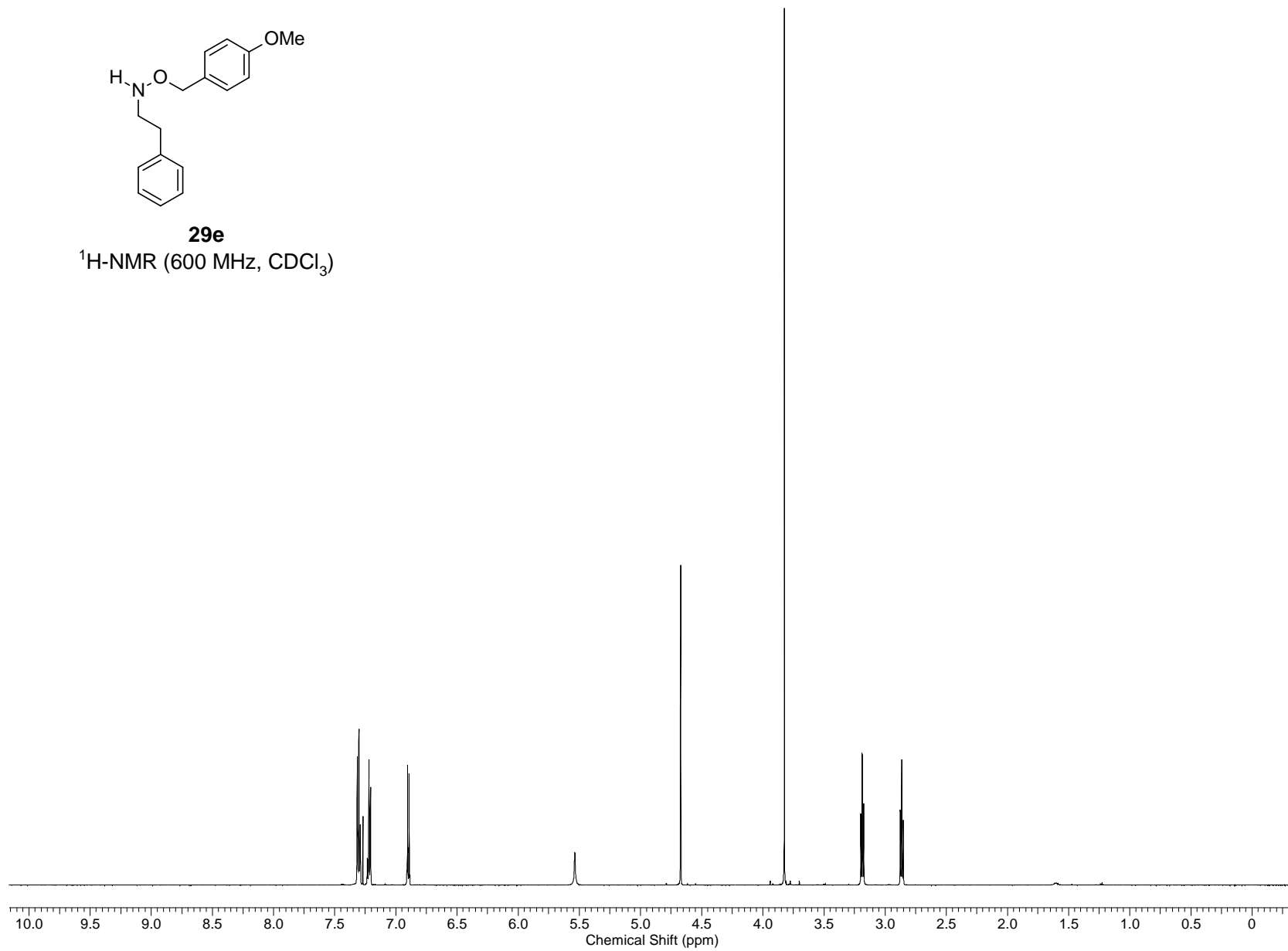
$^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )

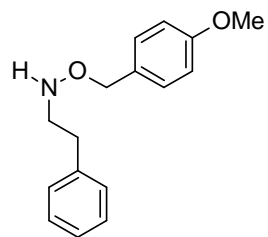




**29e**

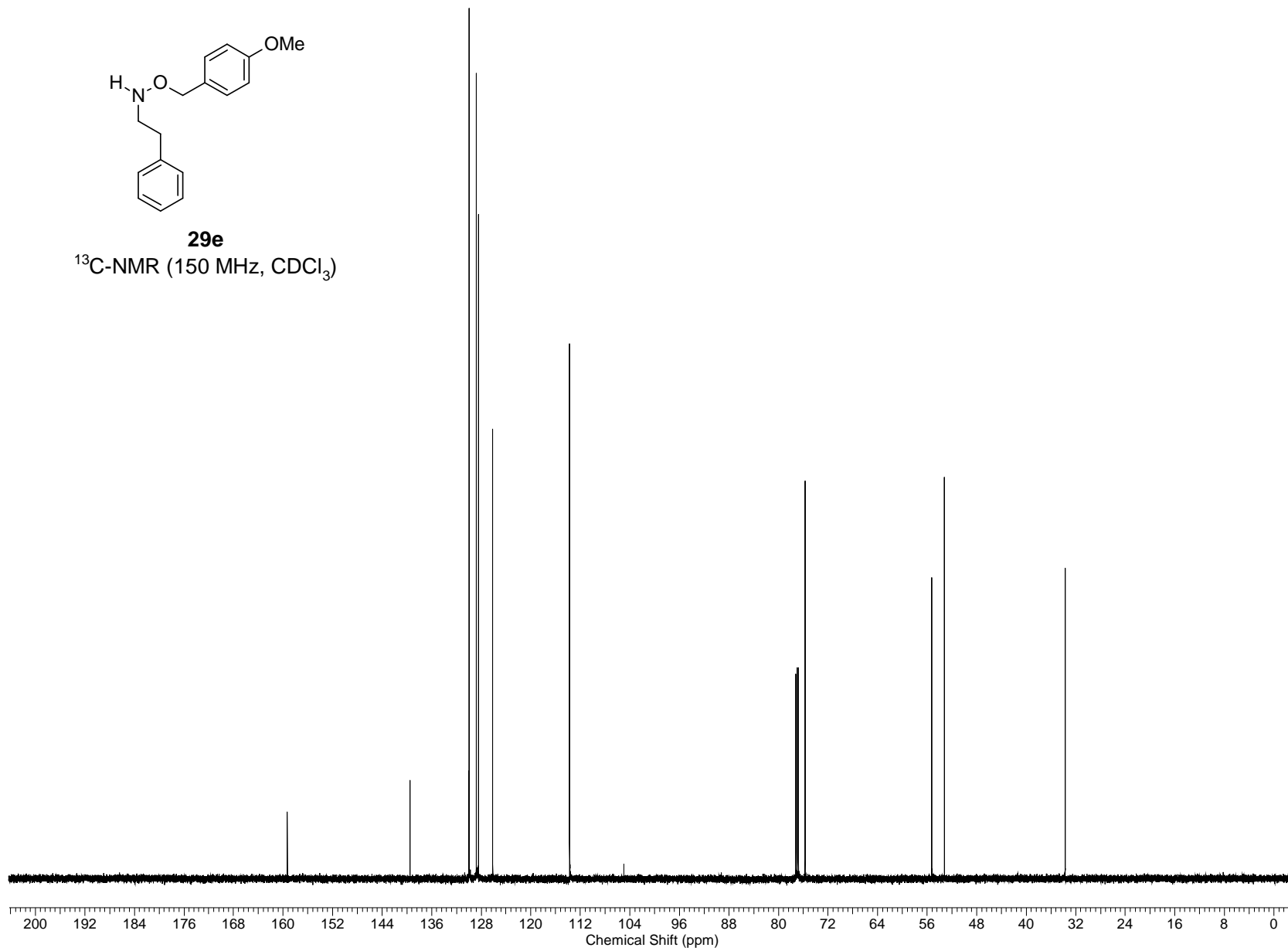
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

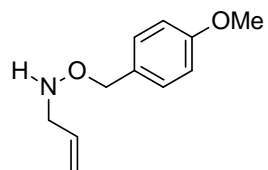




**29e**

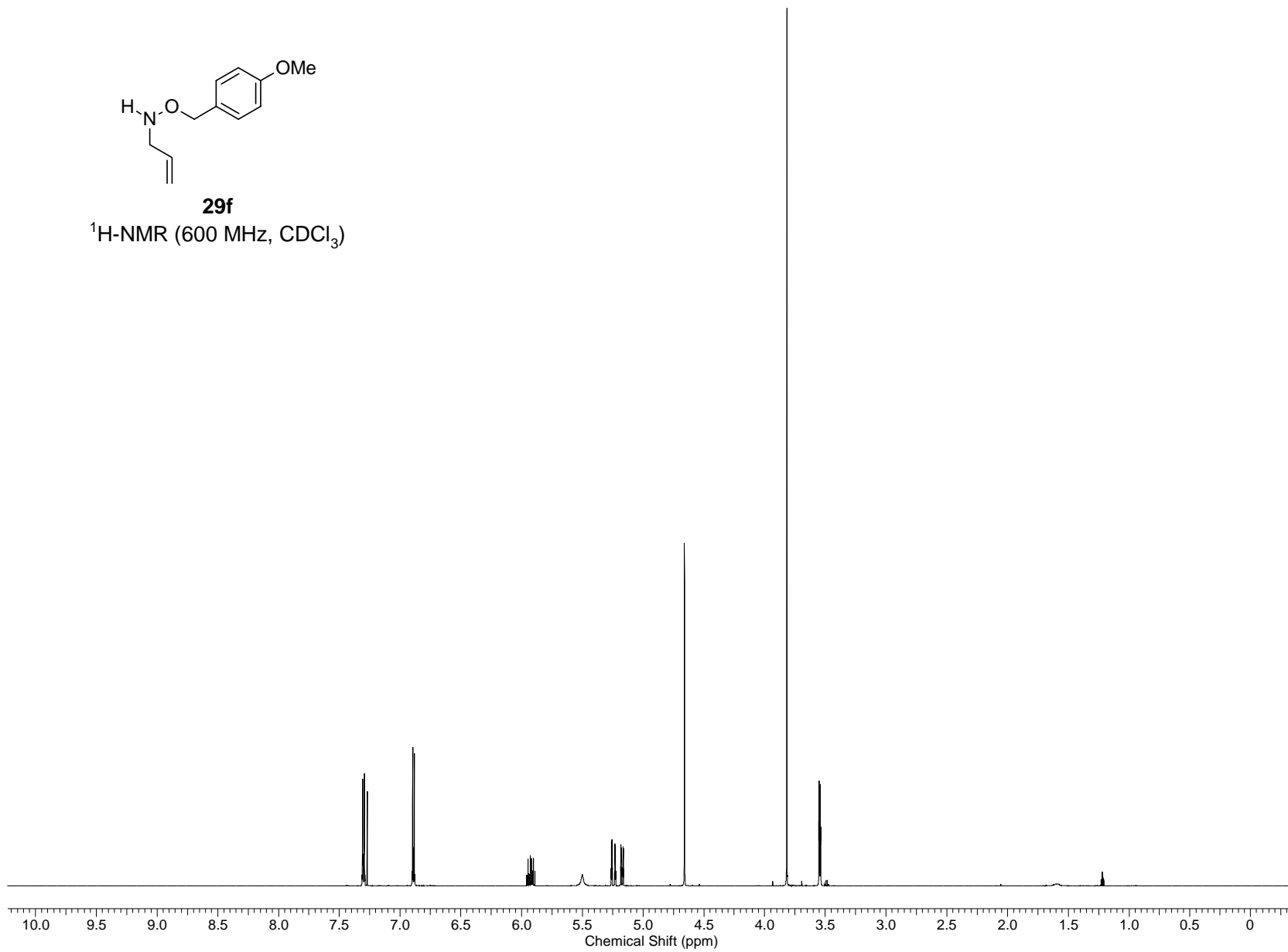
$^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )

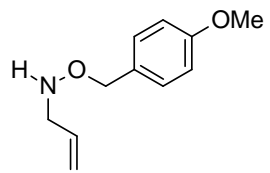




**29f**

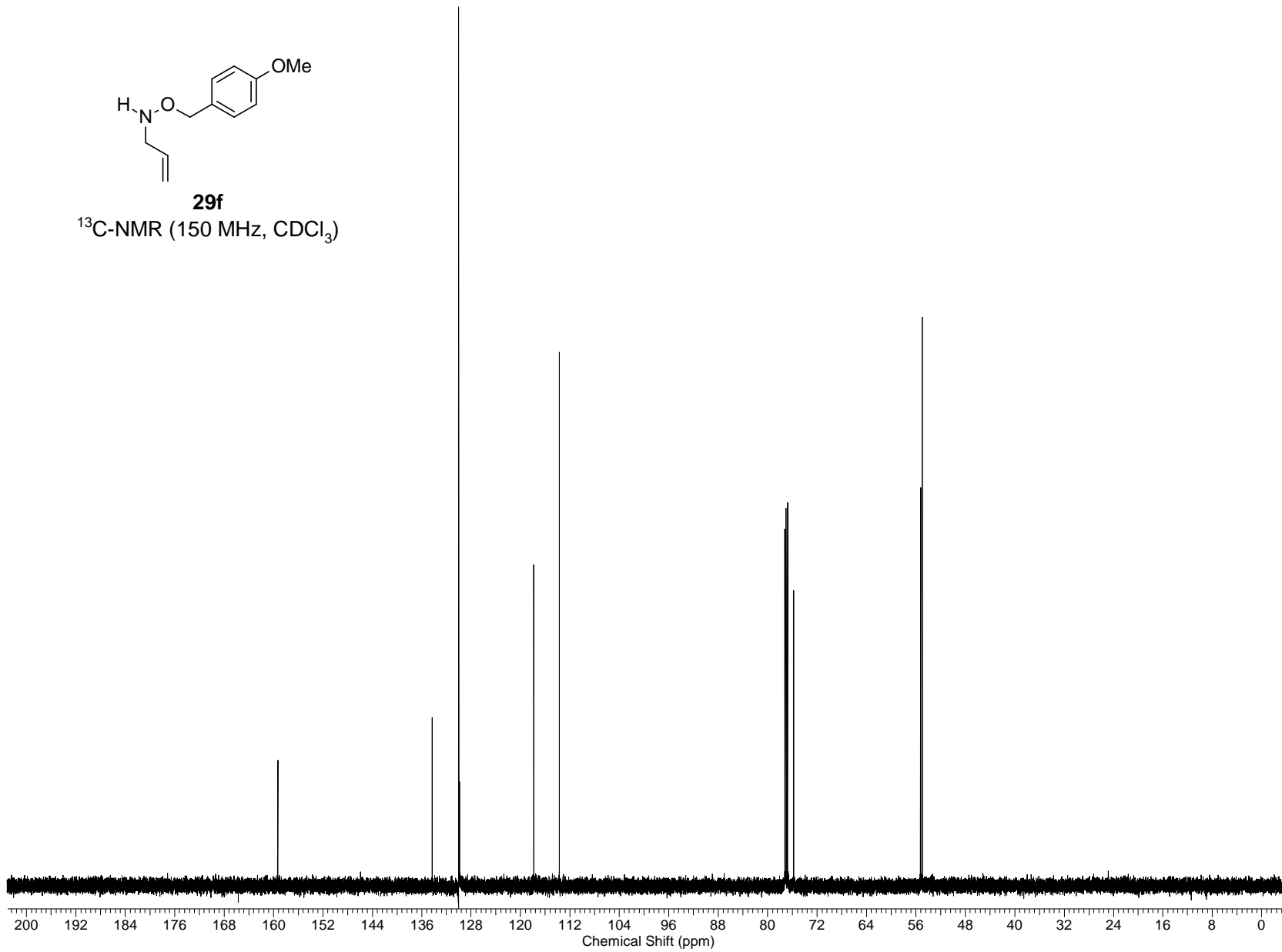
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

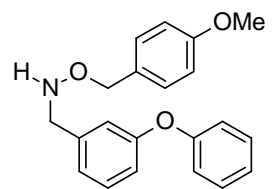




**29f**

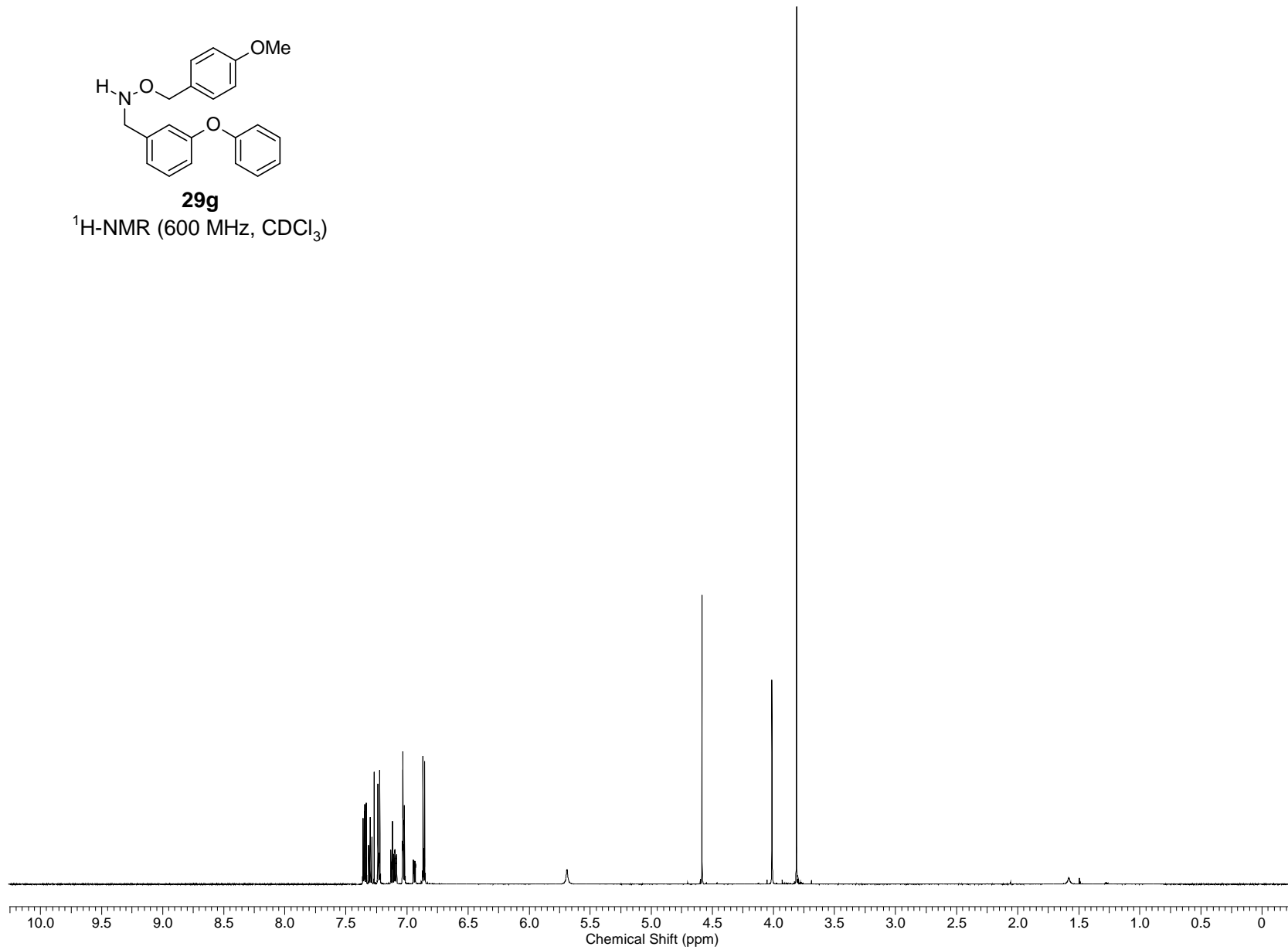
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)



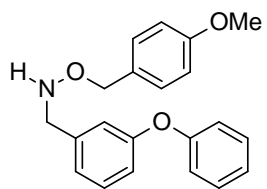


**29g**

<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

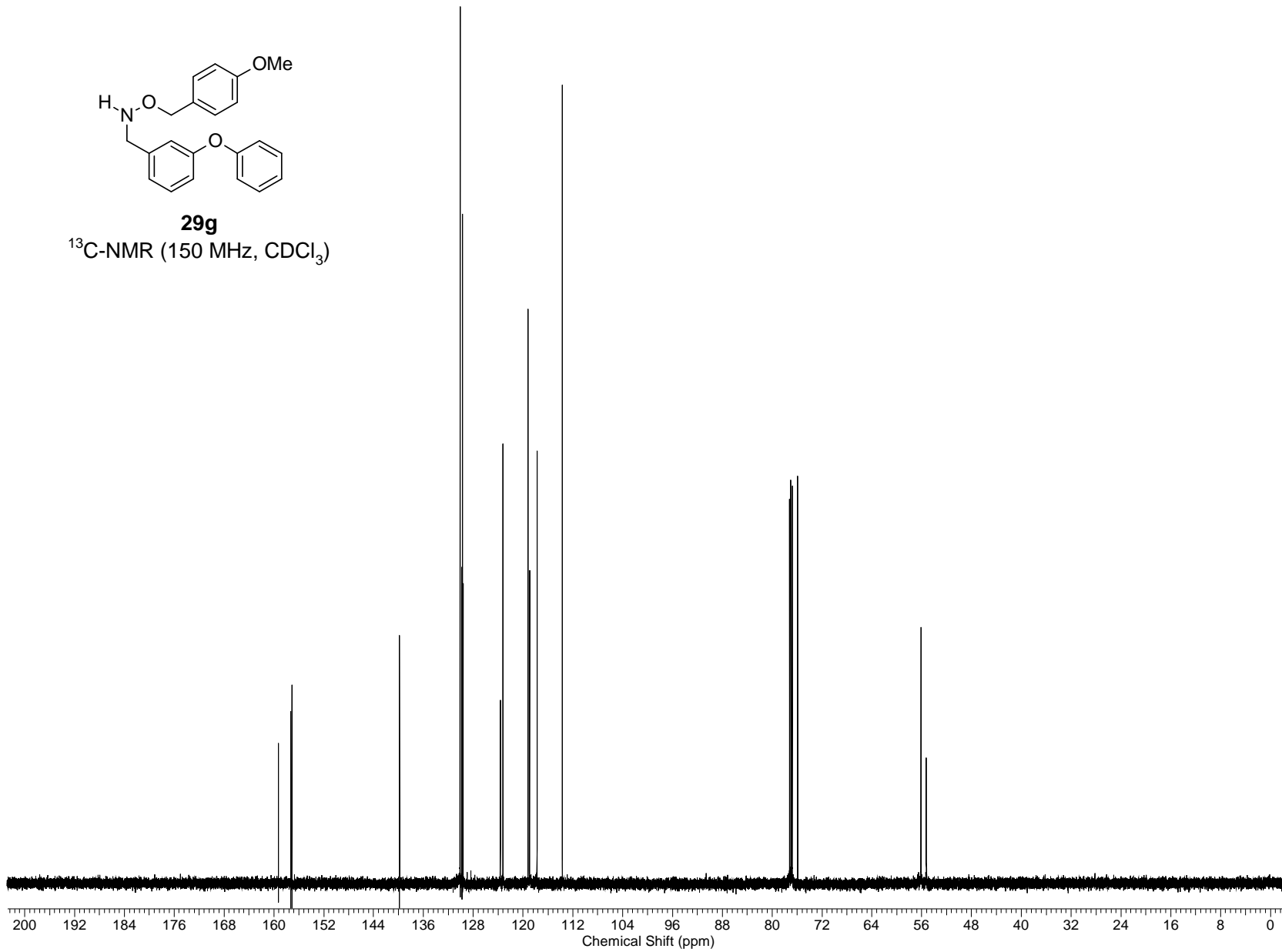


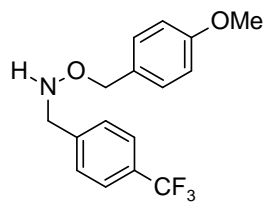




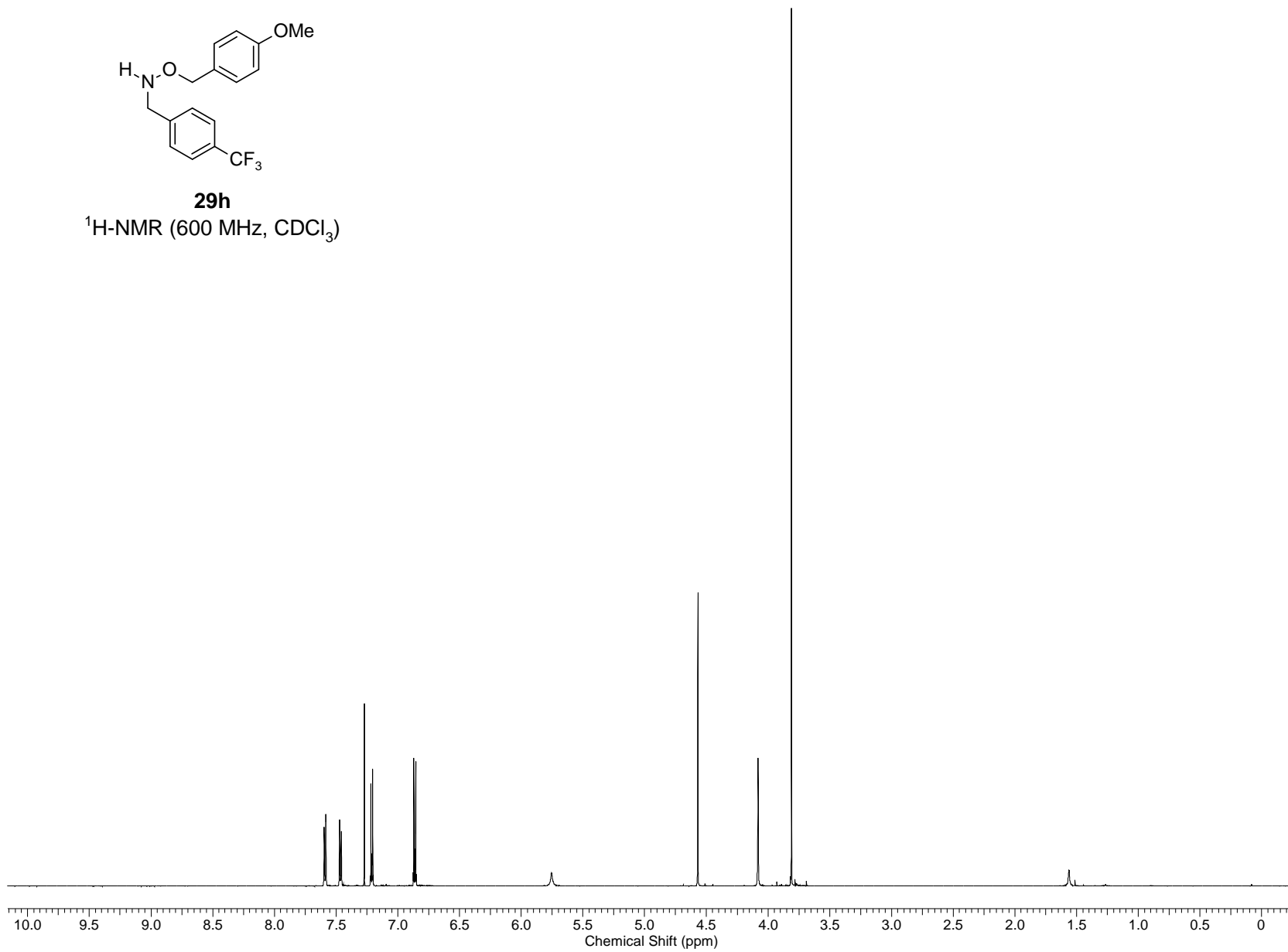
**29g**

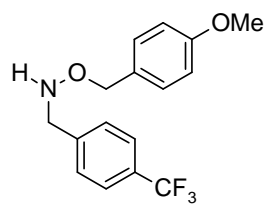
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)





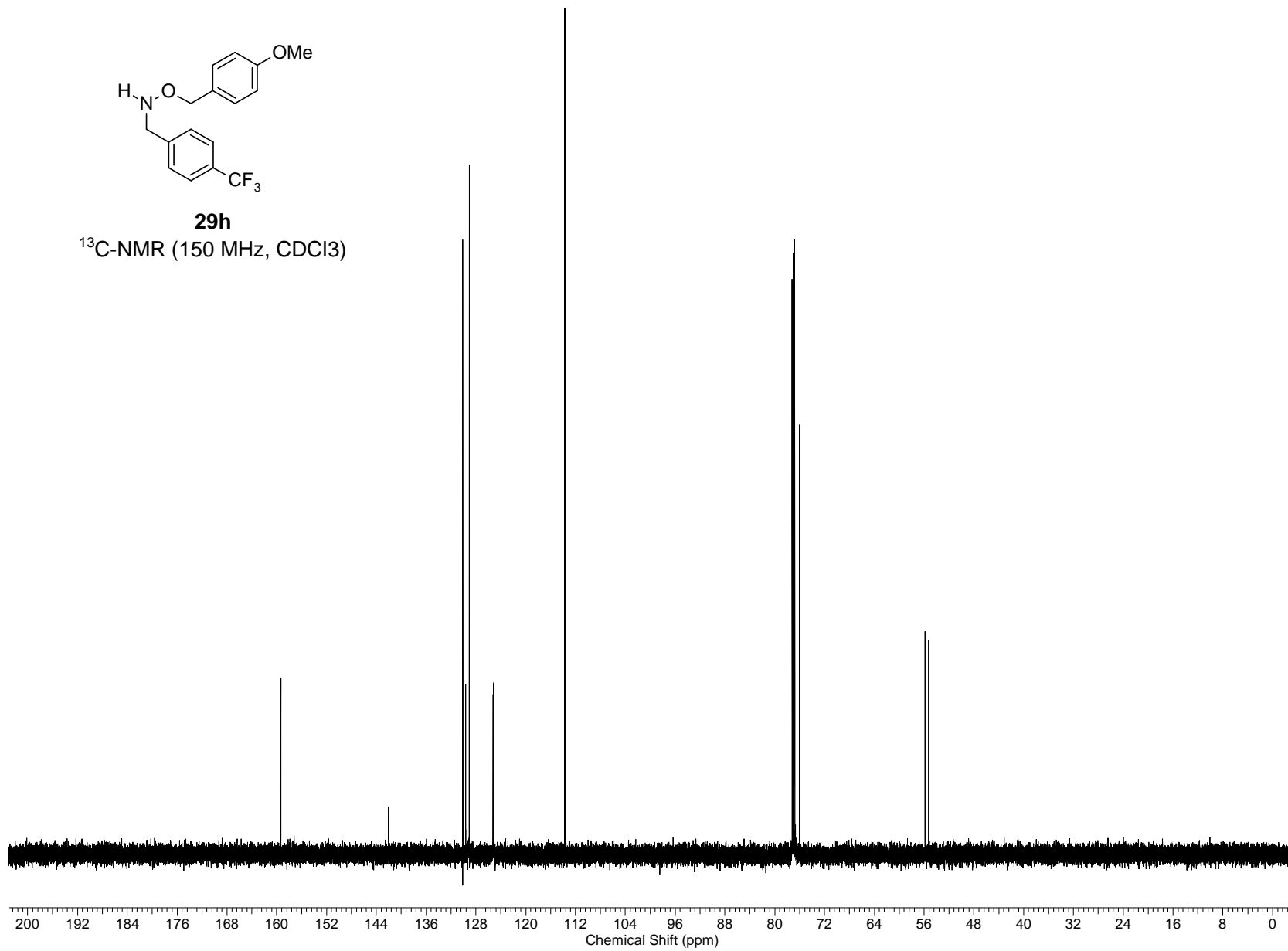
**29h**  
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

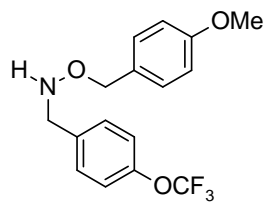




**29h**

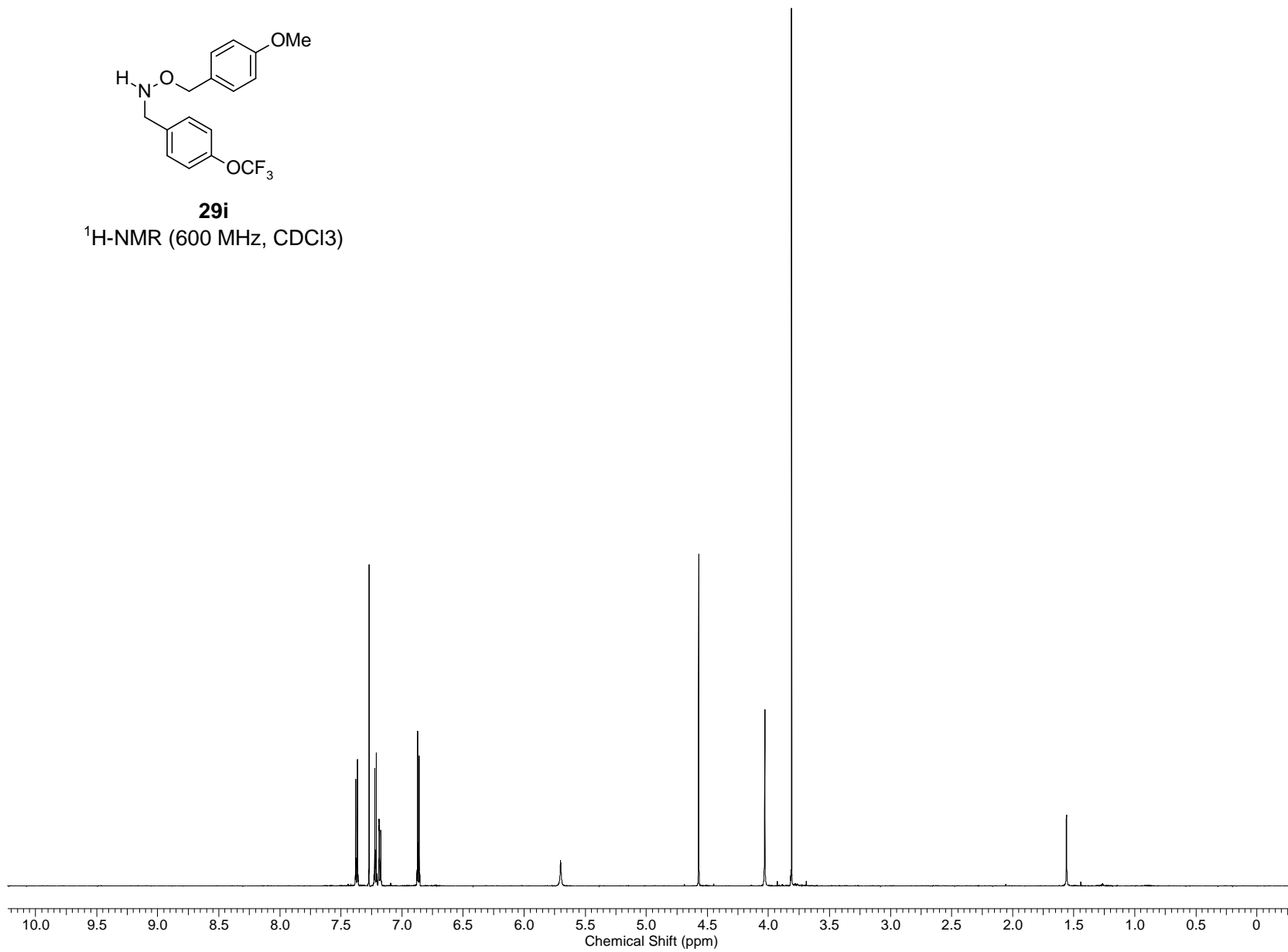
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

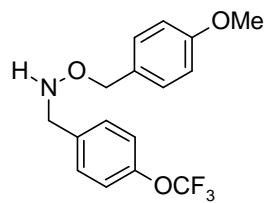




**29i**

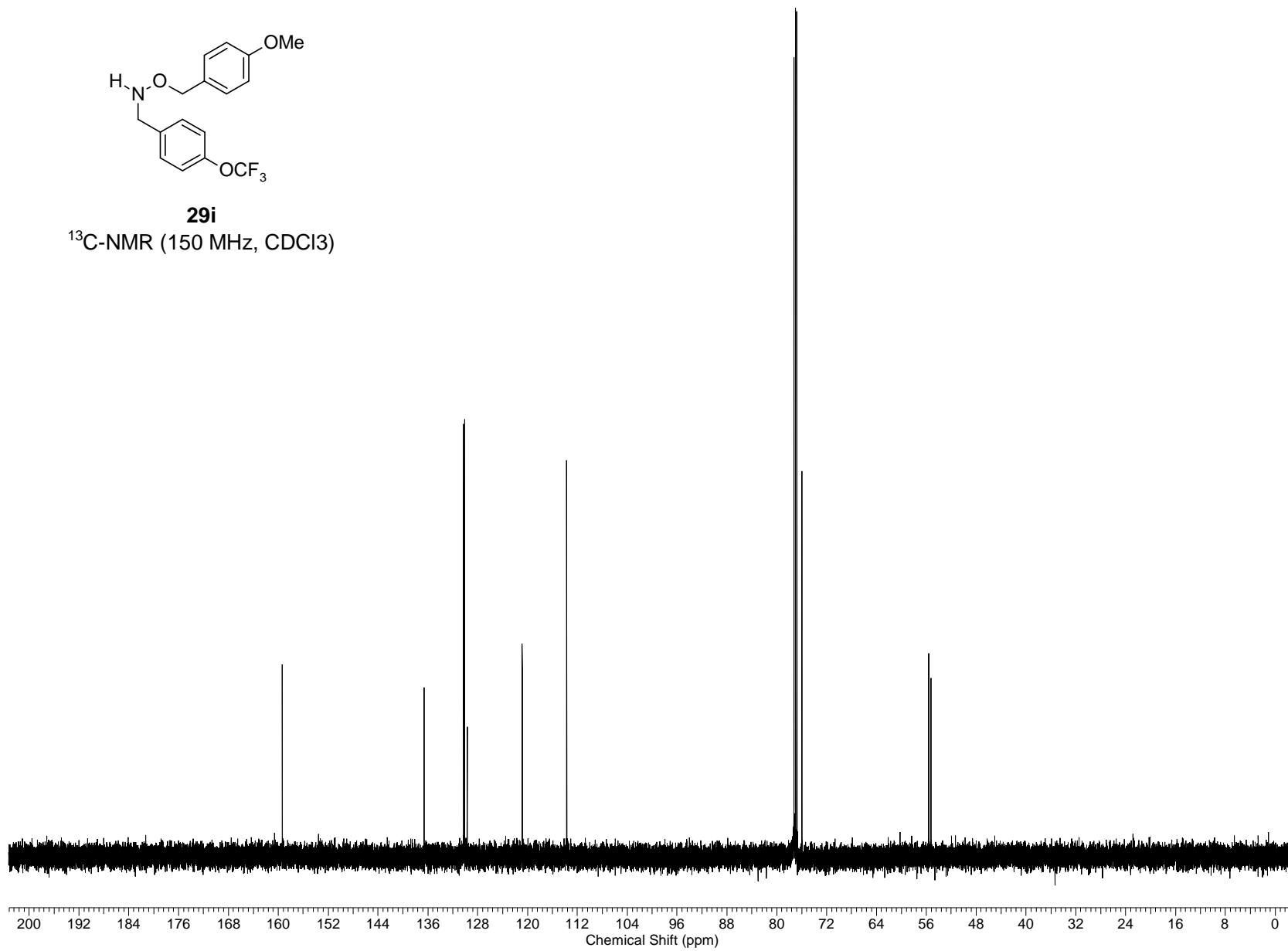
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

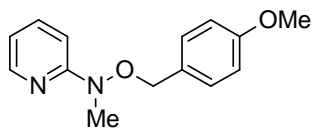




**29i**

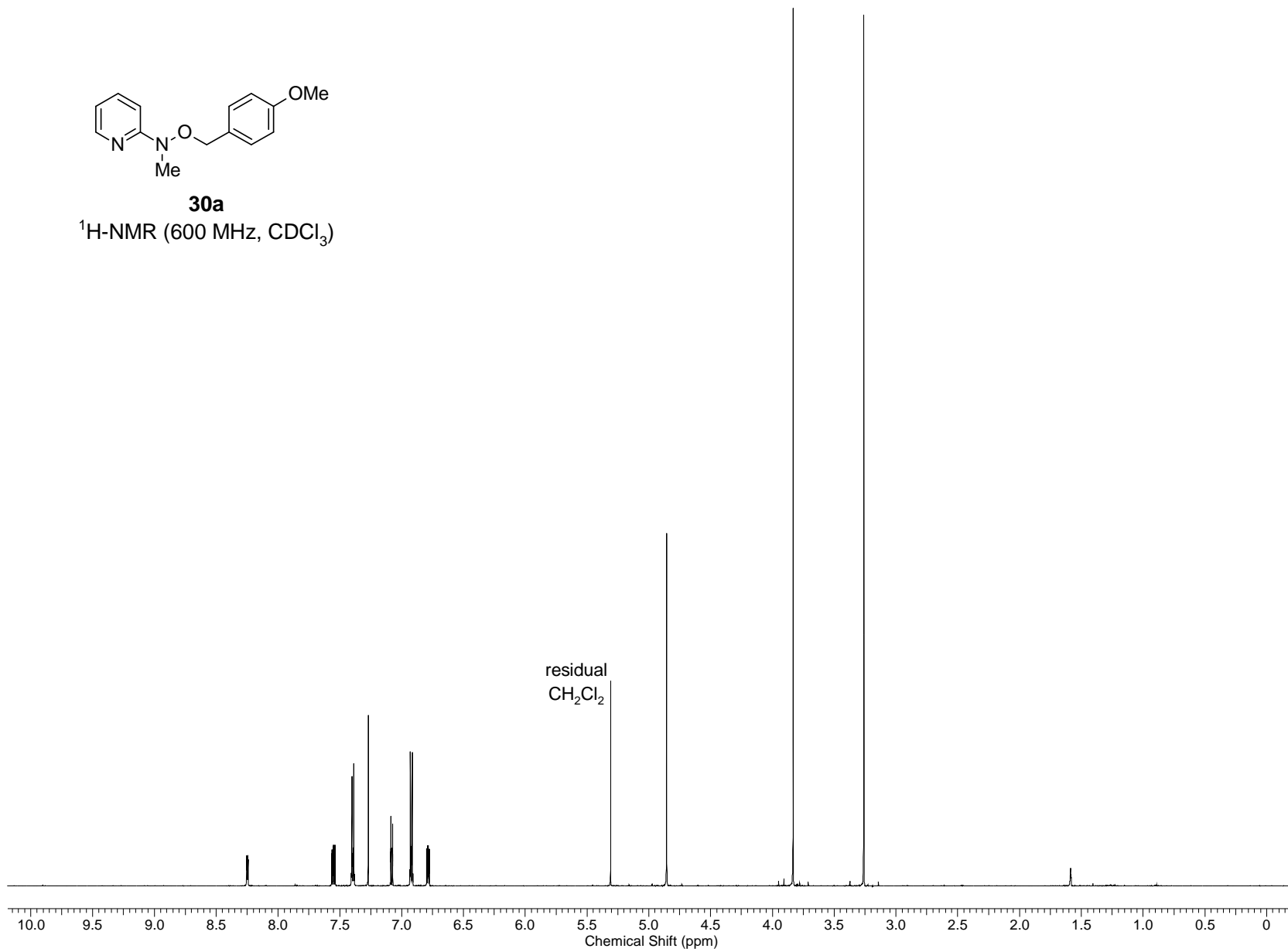
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

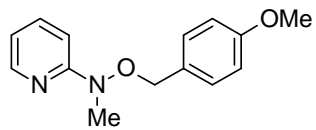




**30a**

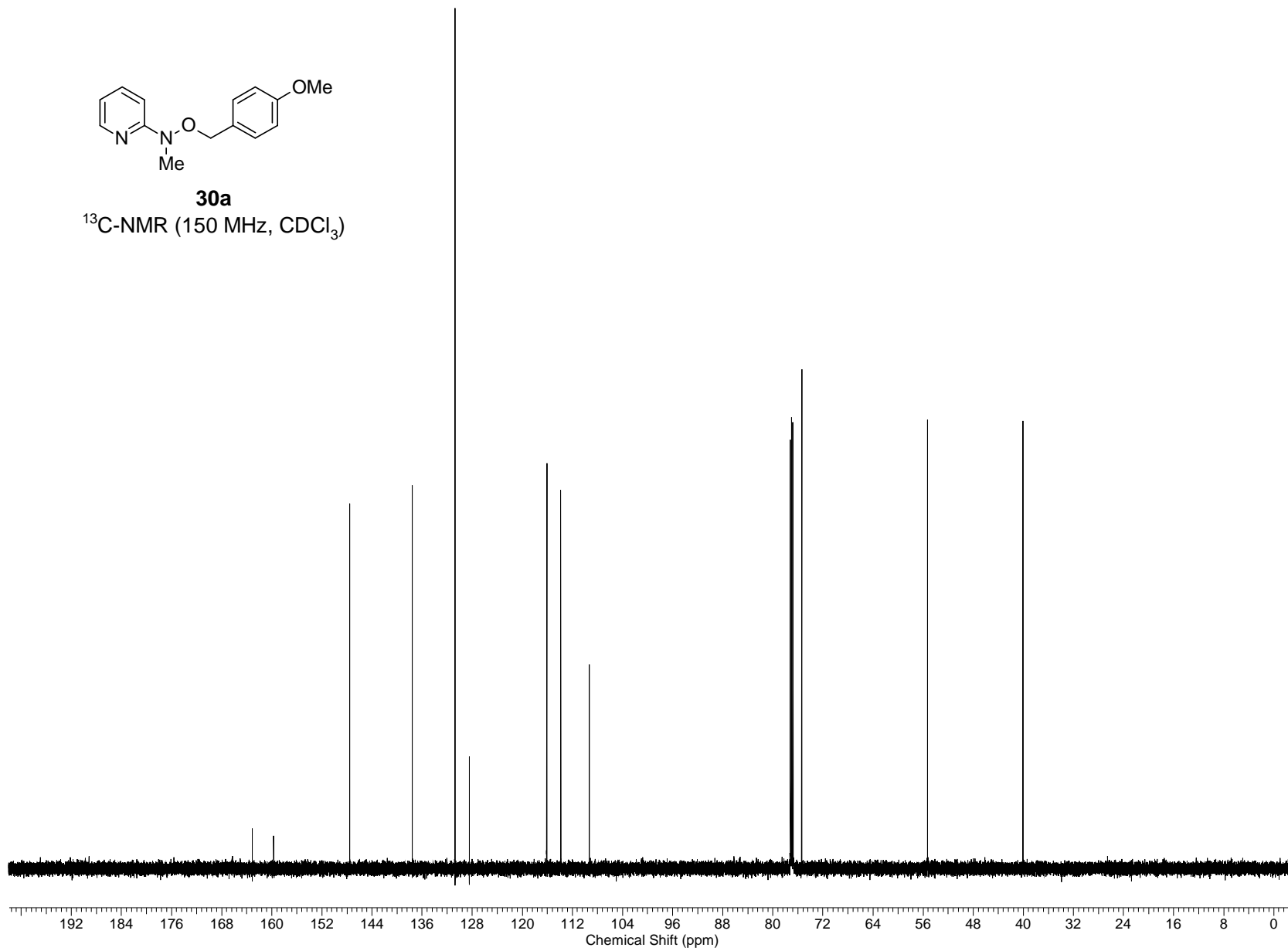
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

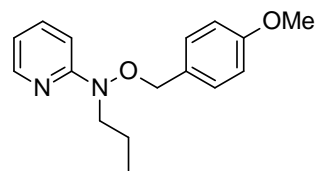




**30a**

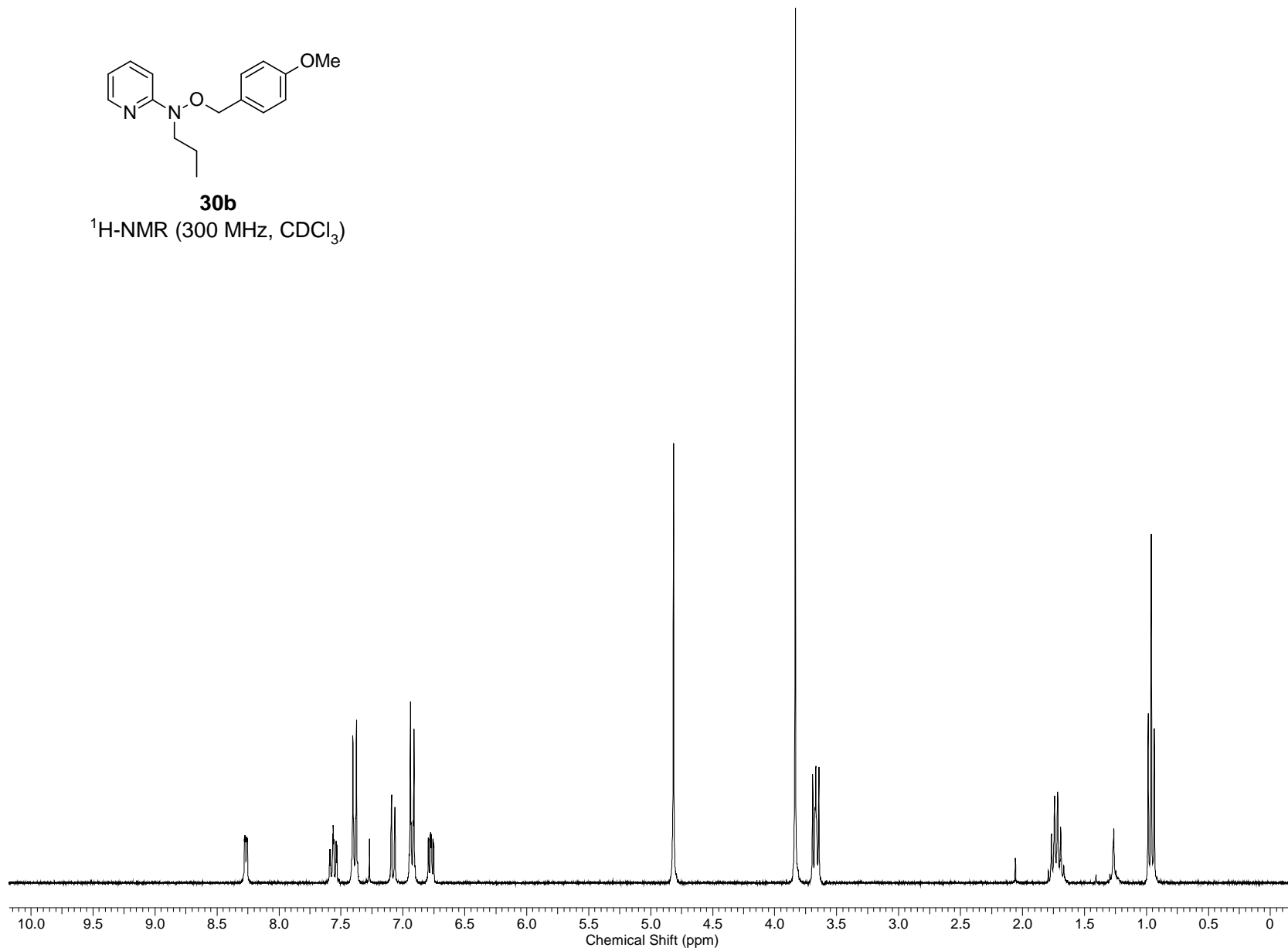
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)



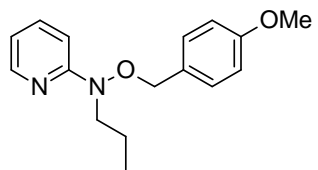


**30b**

<sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>)

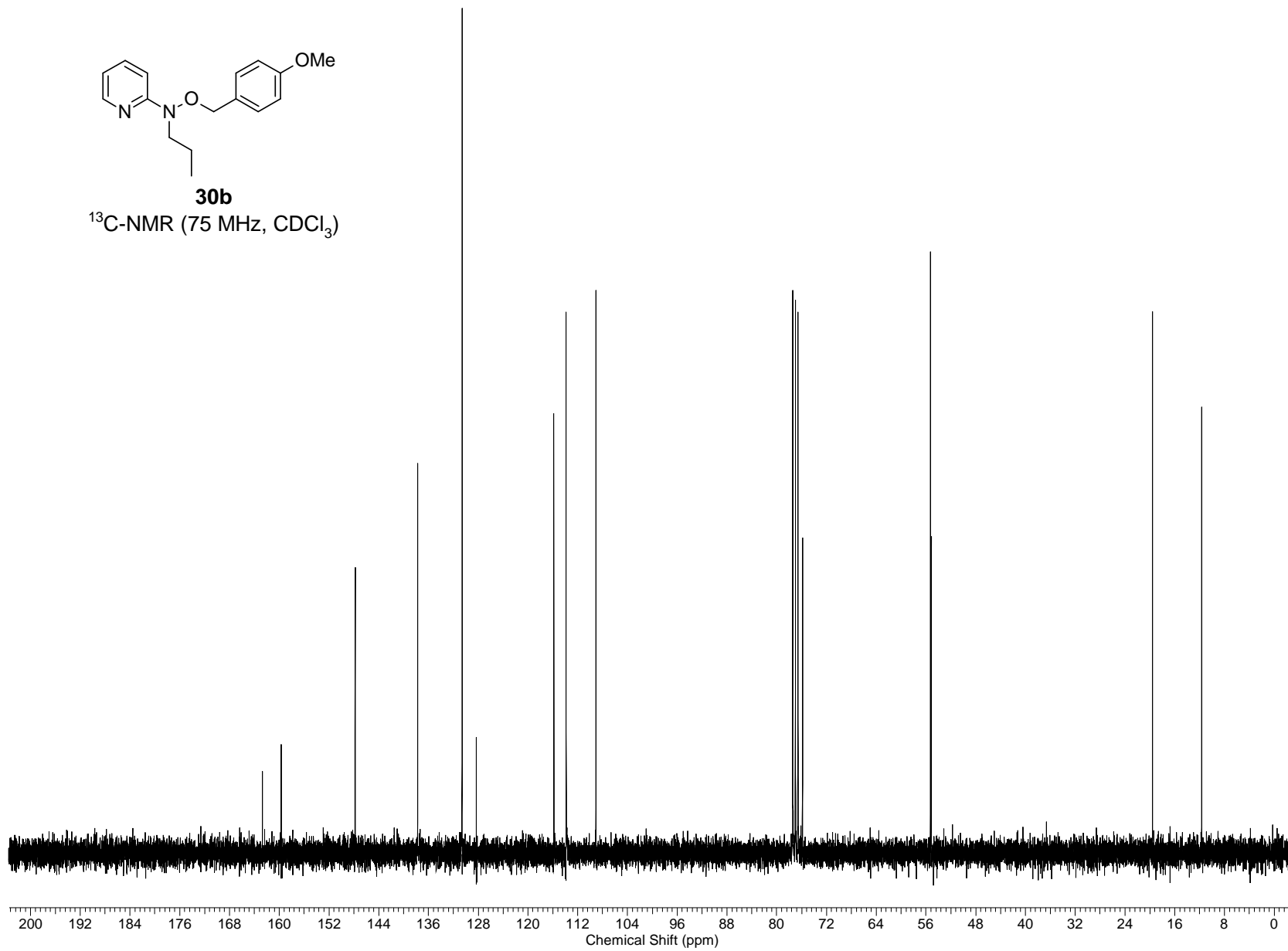


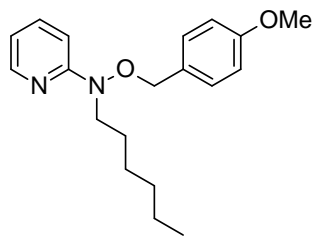




**30b**

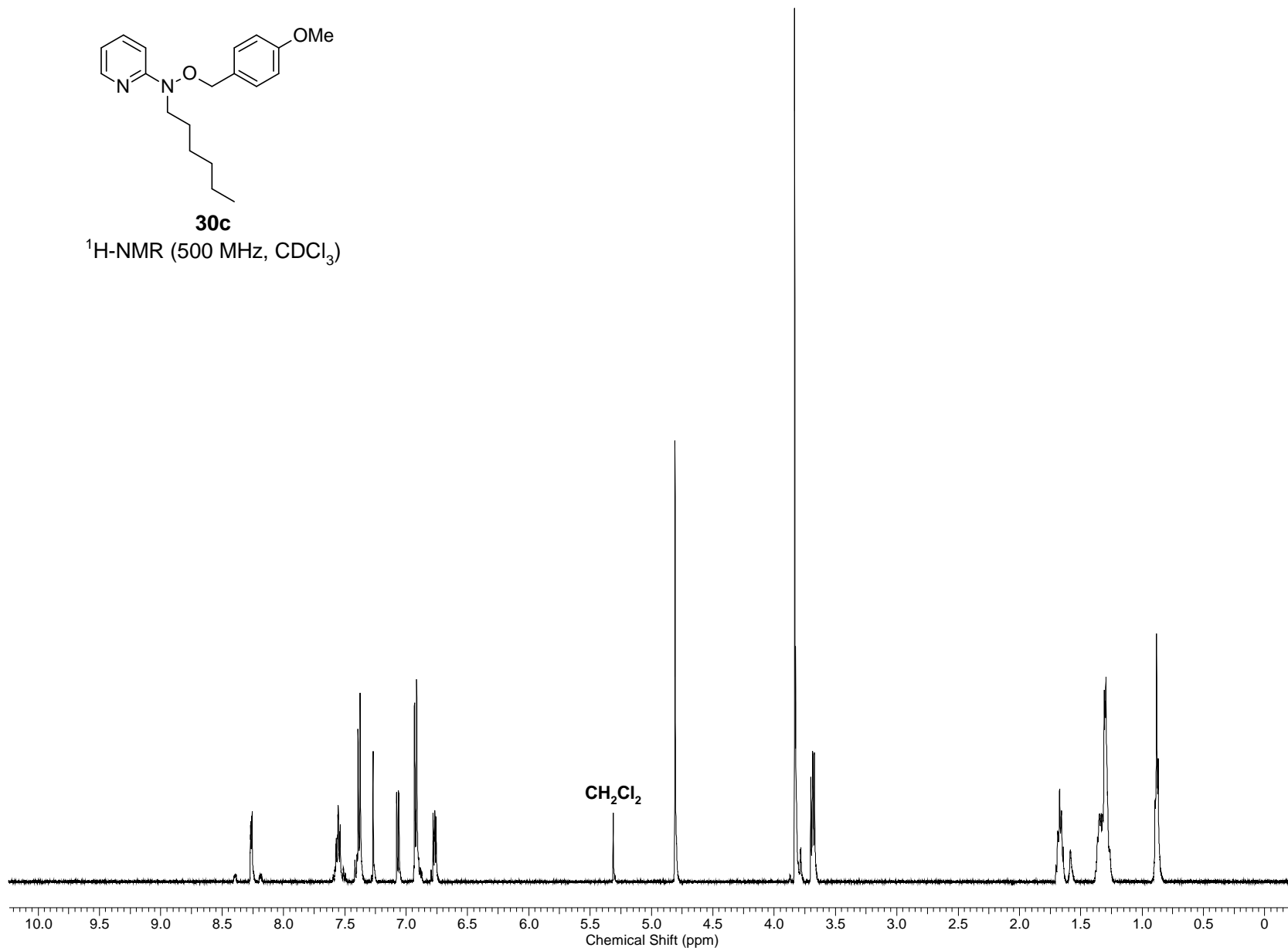
<sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>)

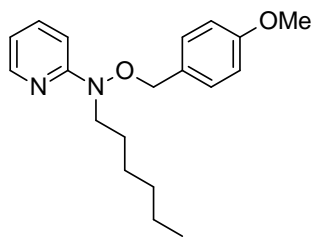




**30c**

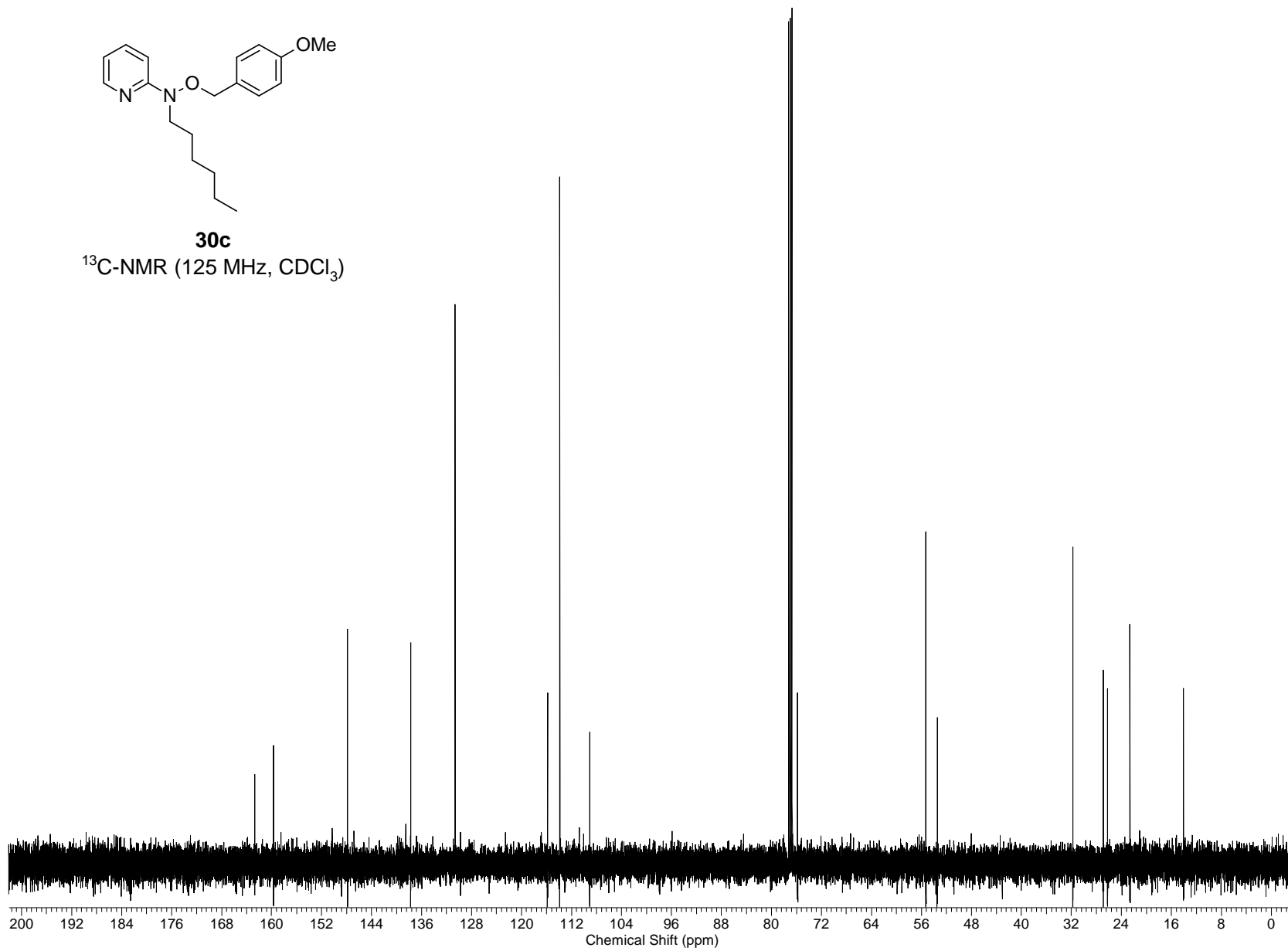
<sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)

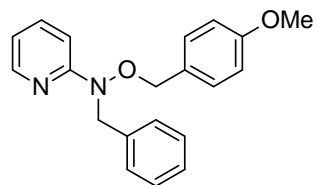




**30c**

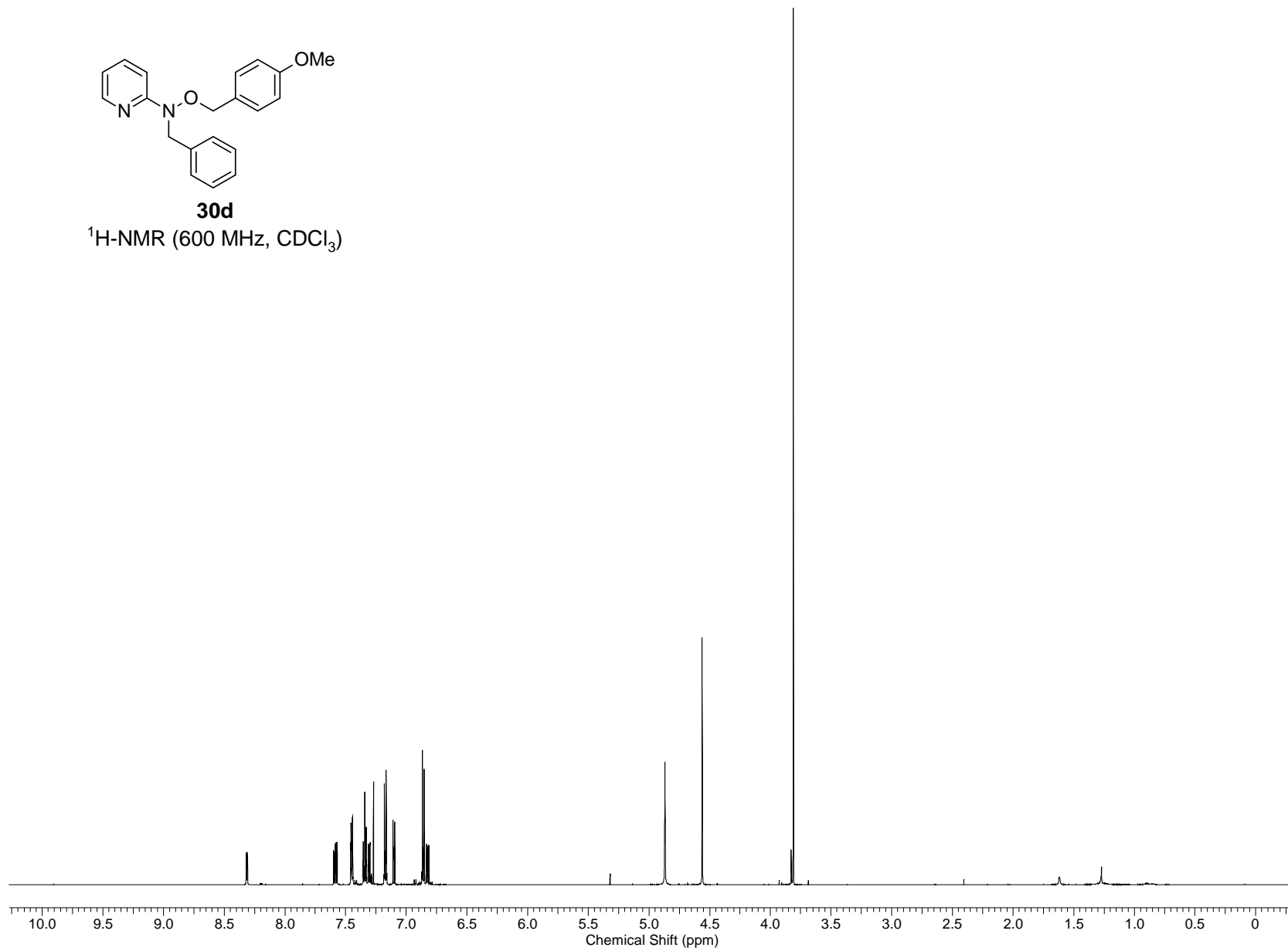
$^{13}\text{C}$ -NMR (125 MHz,  $\text{CDCl}_3$ )

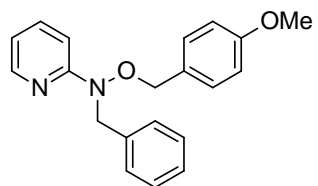




**30d**

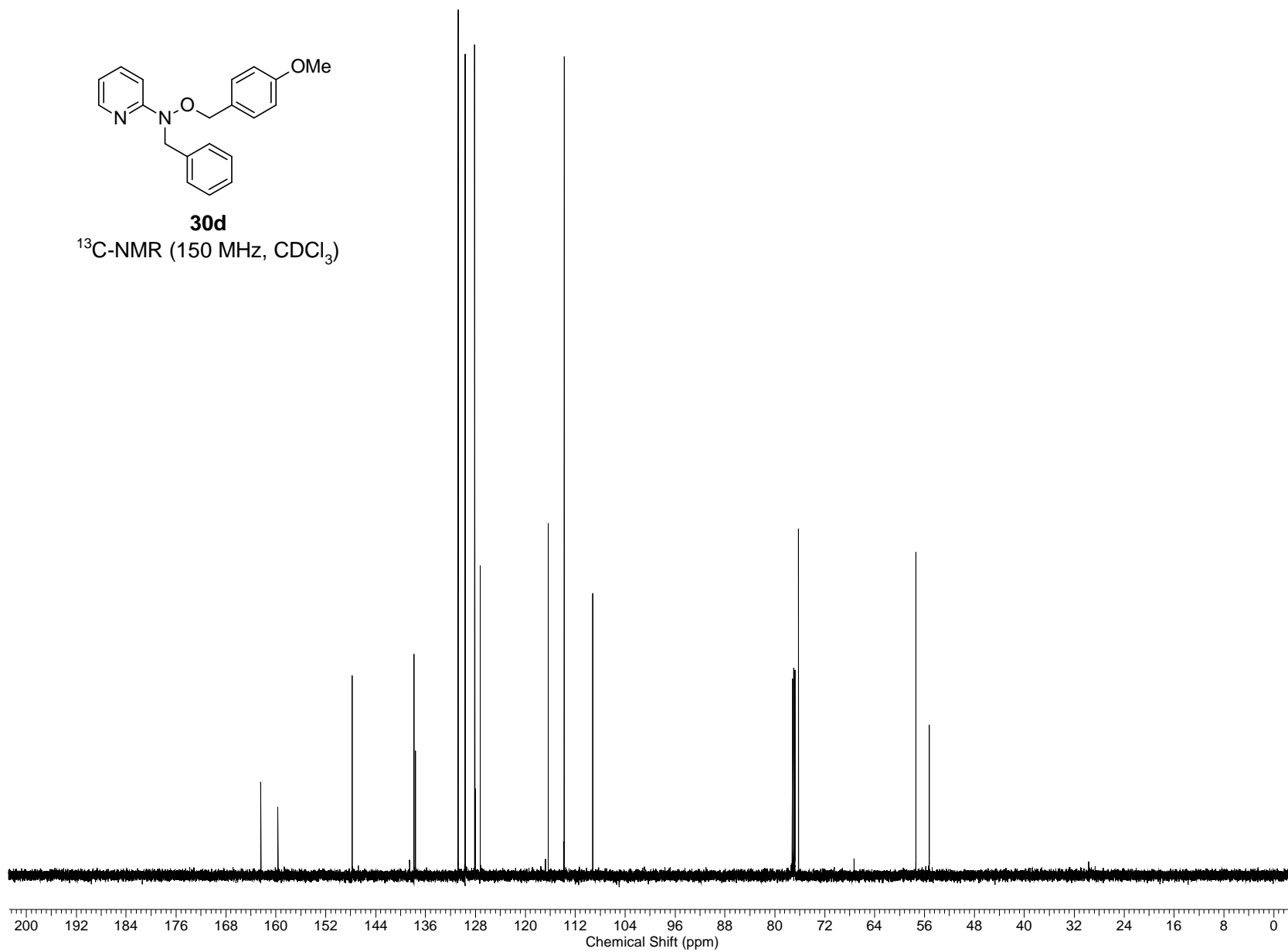
$^1\text{H-NMR}$  (600 MHz,  $\text{CDCl}_3$ )

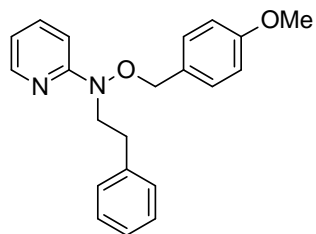




**30d**

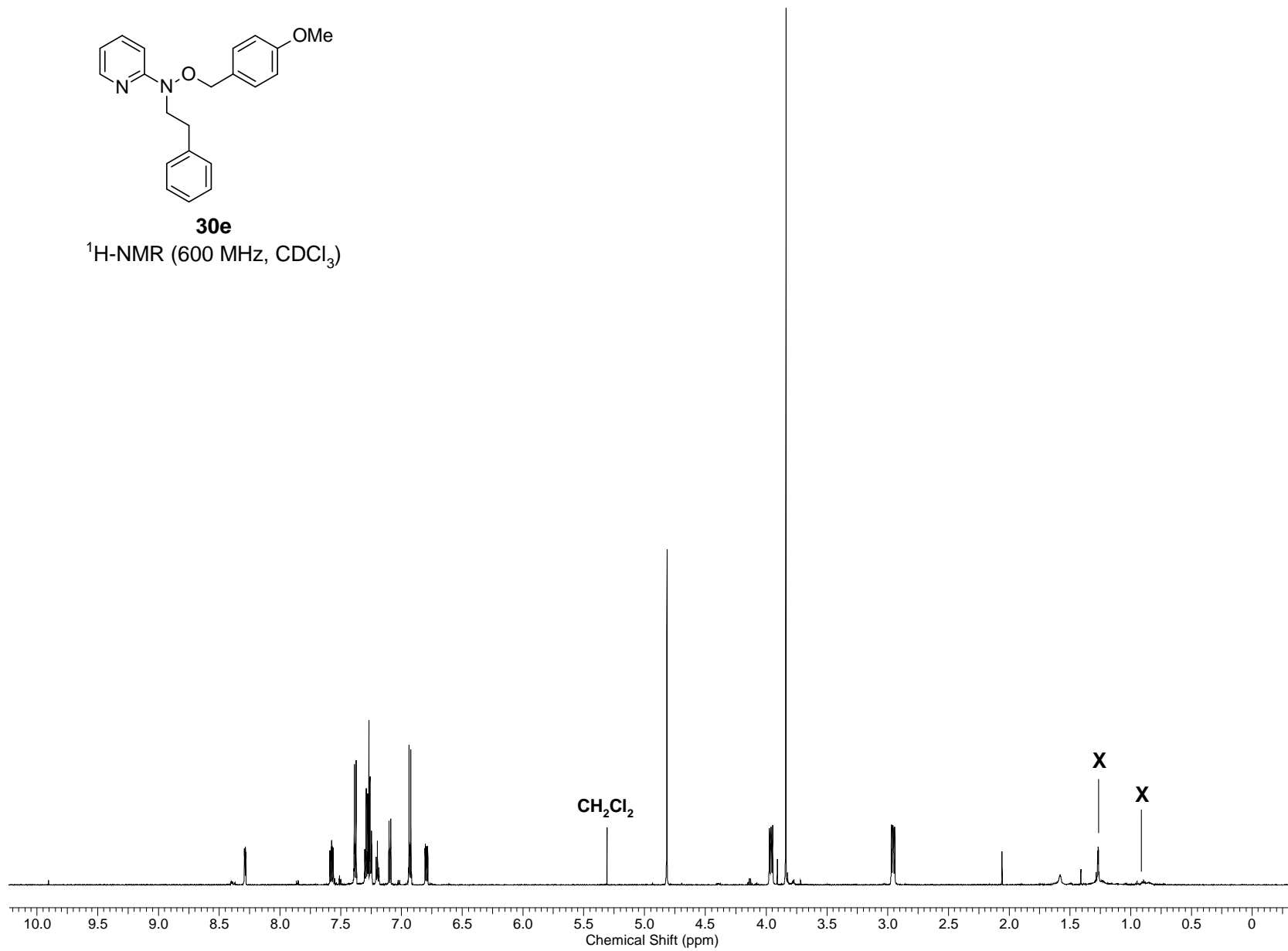
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

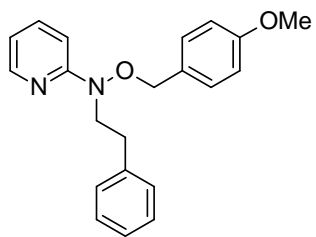




**30e**

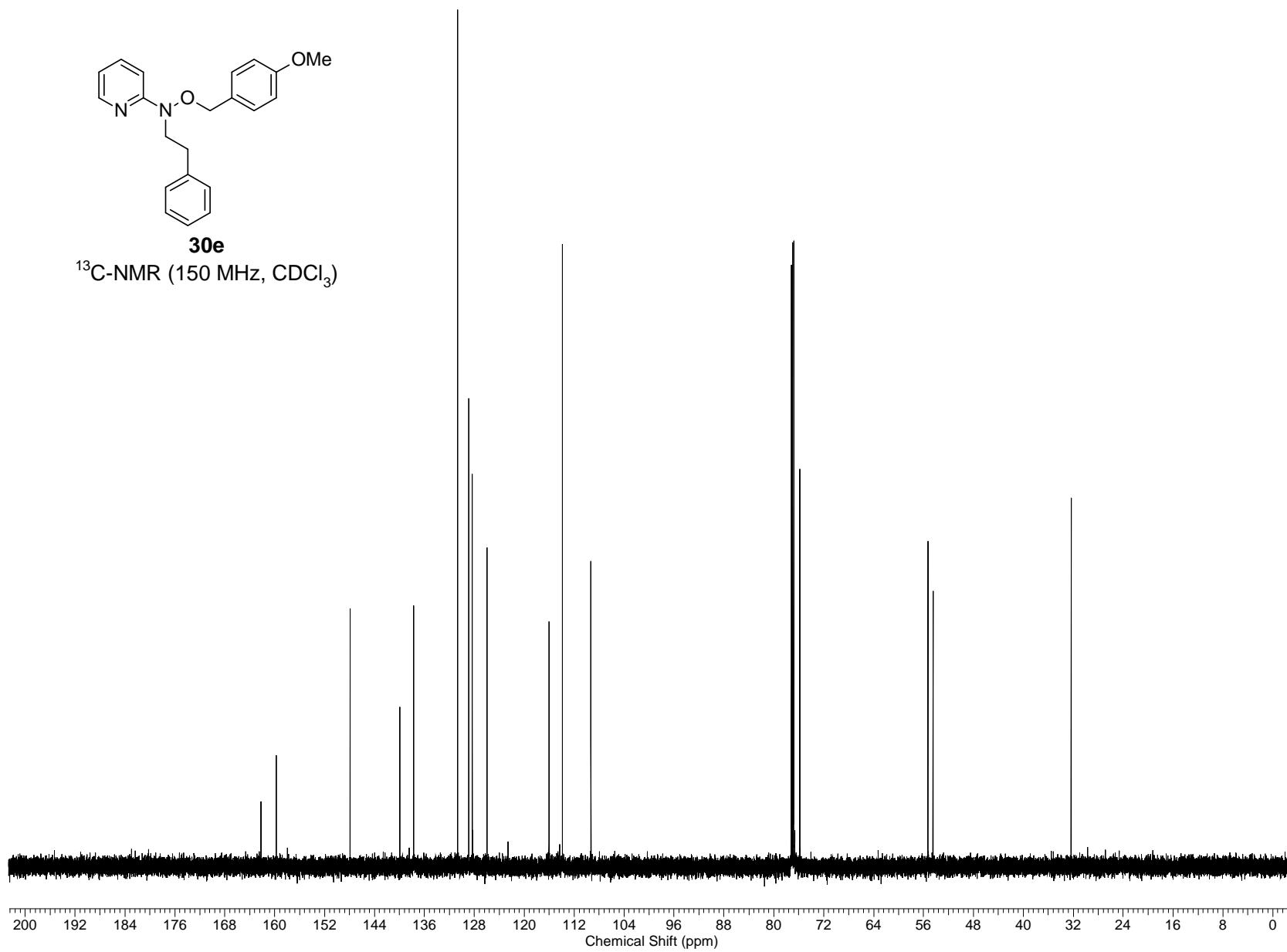
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

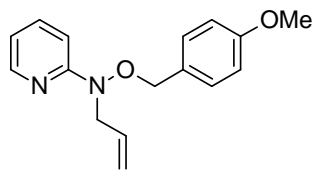




**30e**

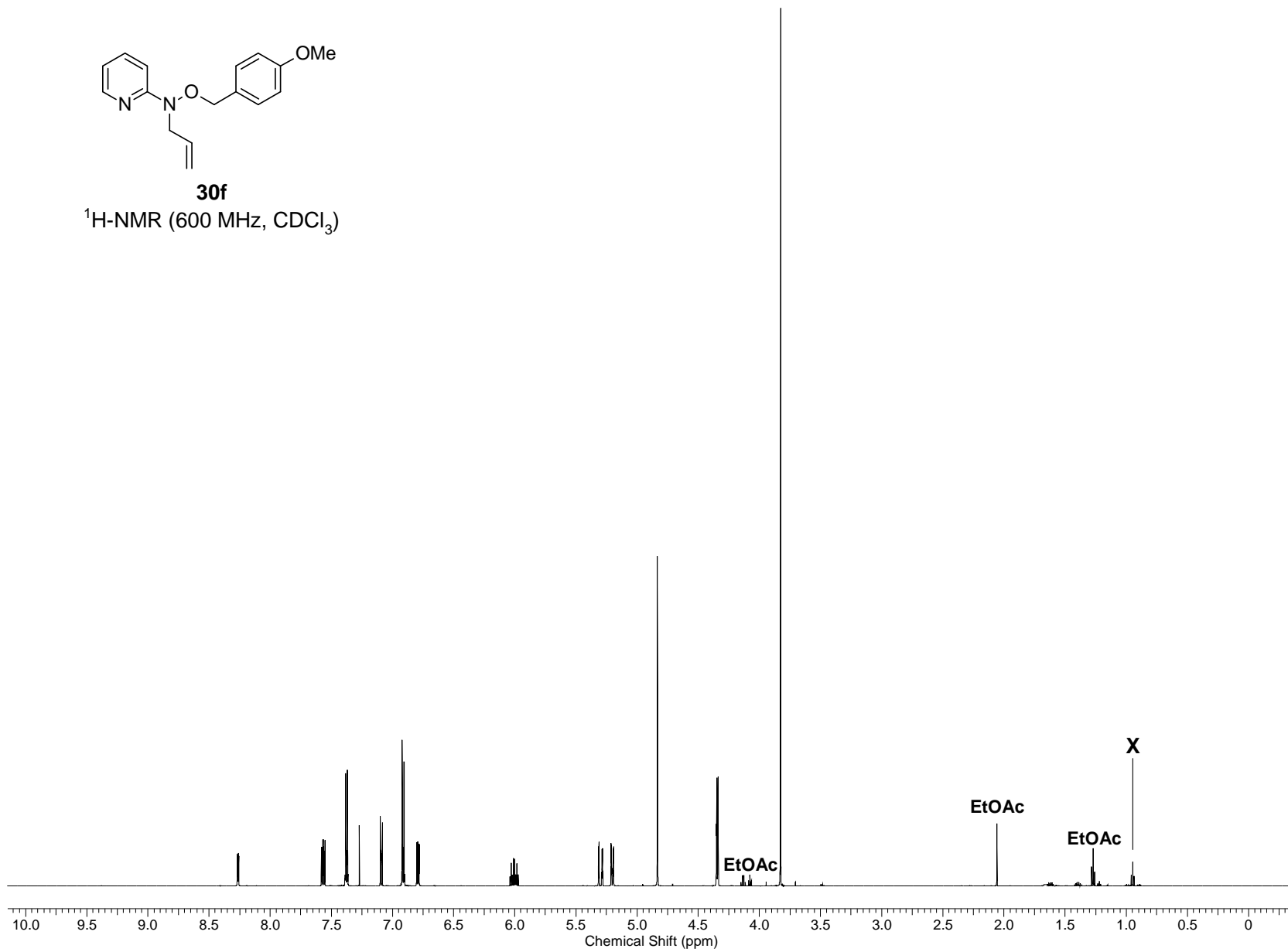
$^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )



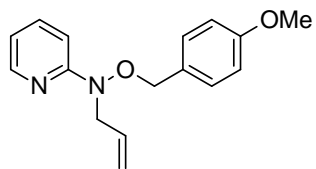


**30f**

<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

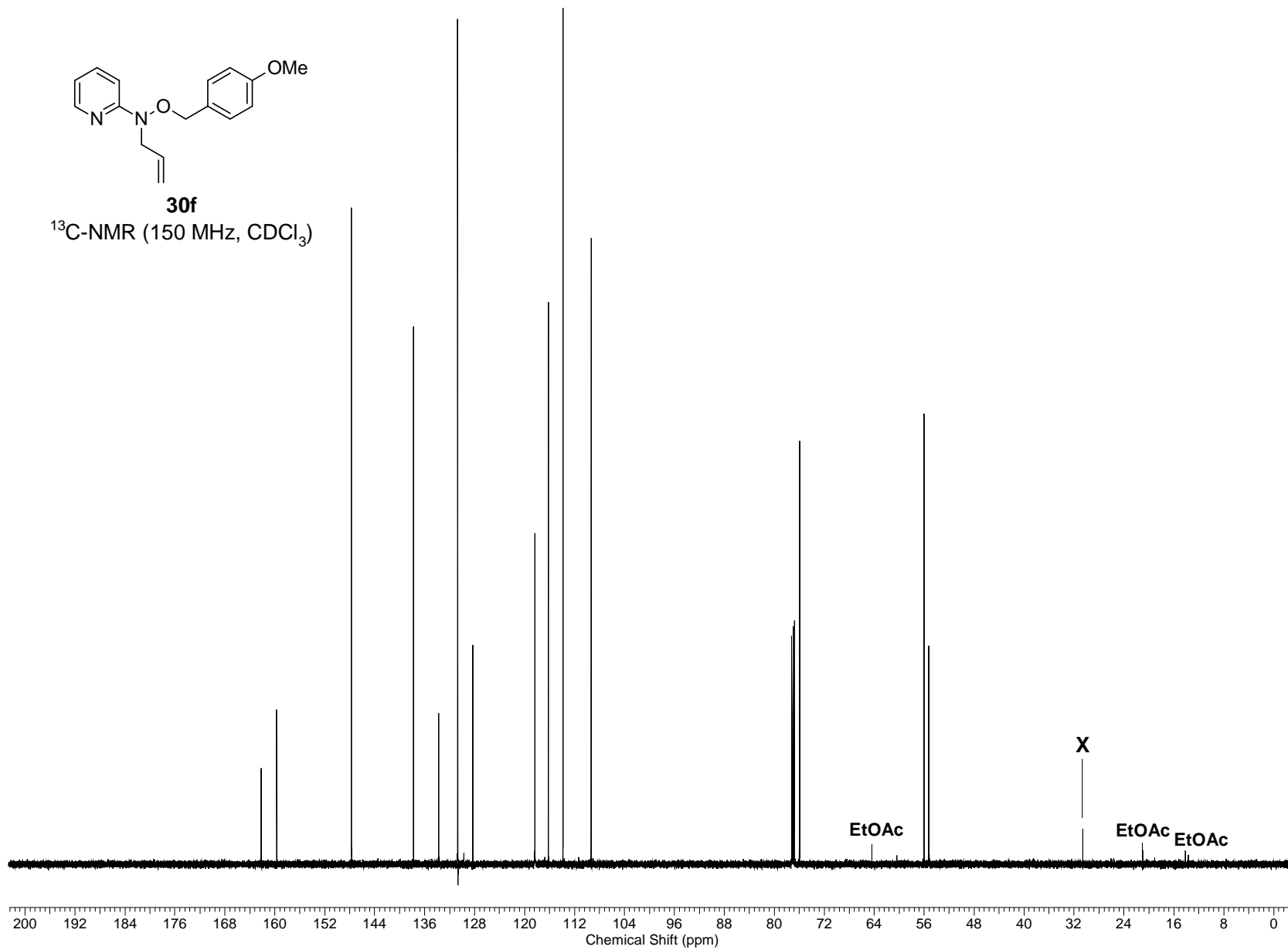


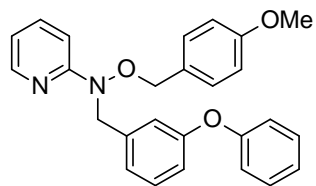




**30f**

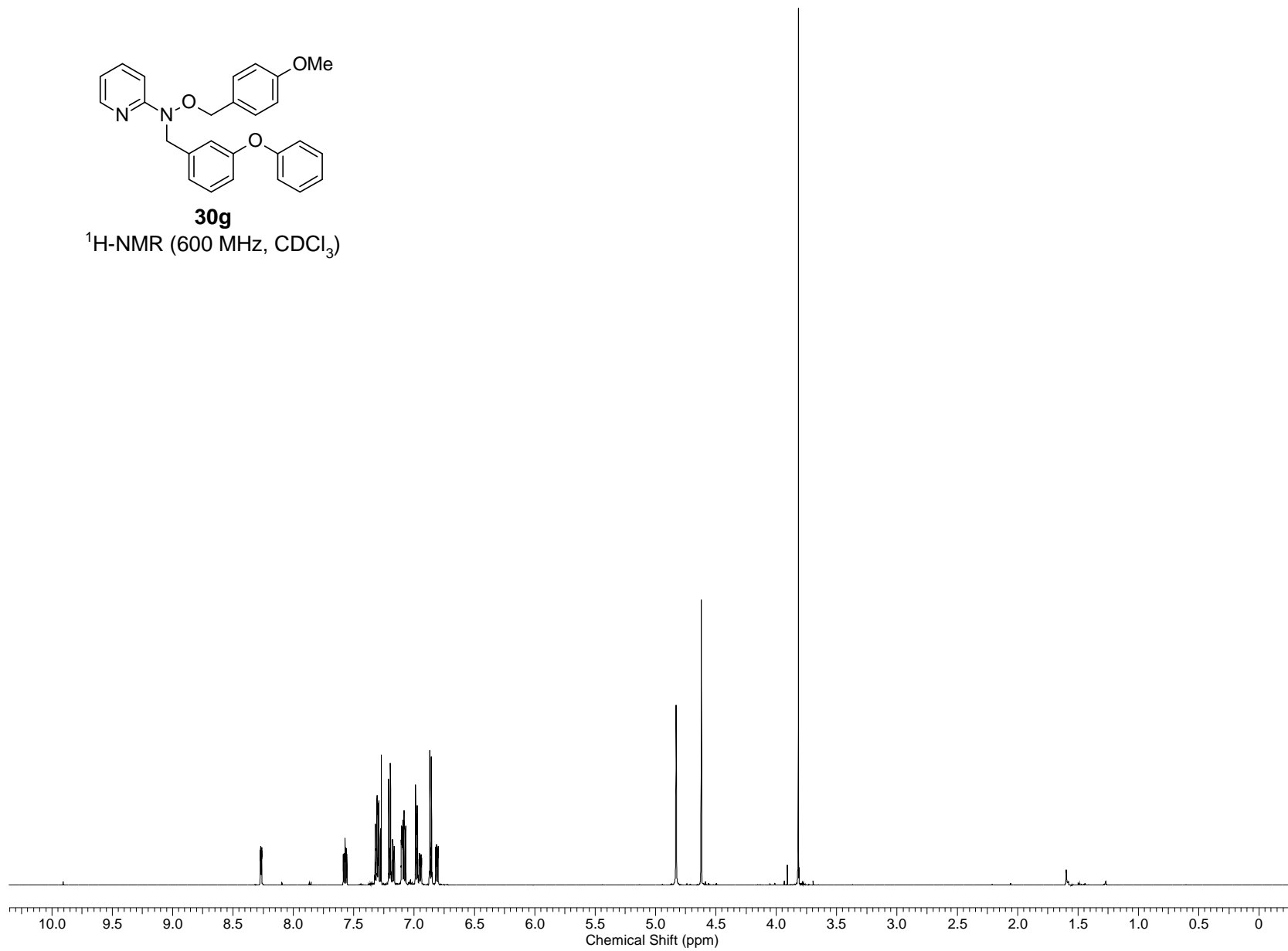
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

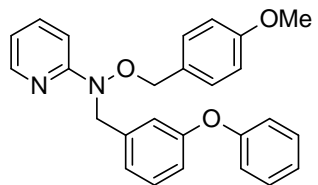




**30g**

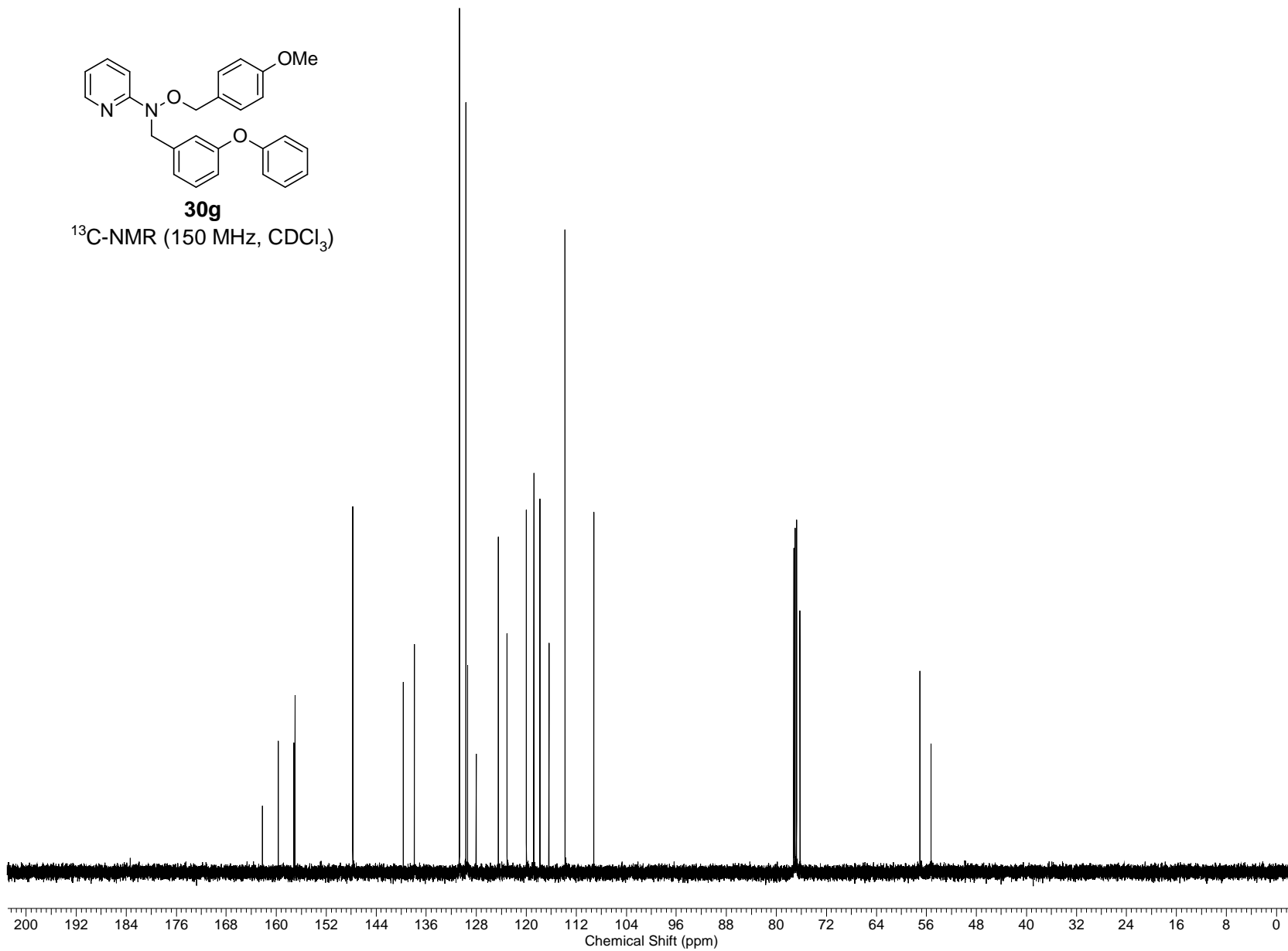
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

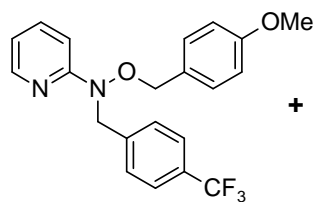




**30g**

<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

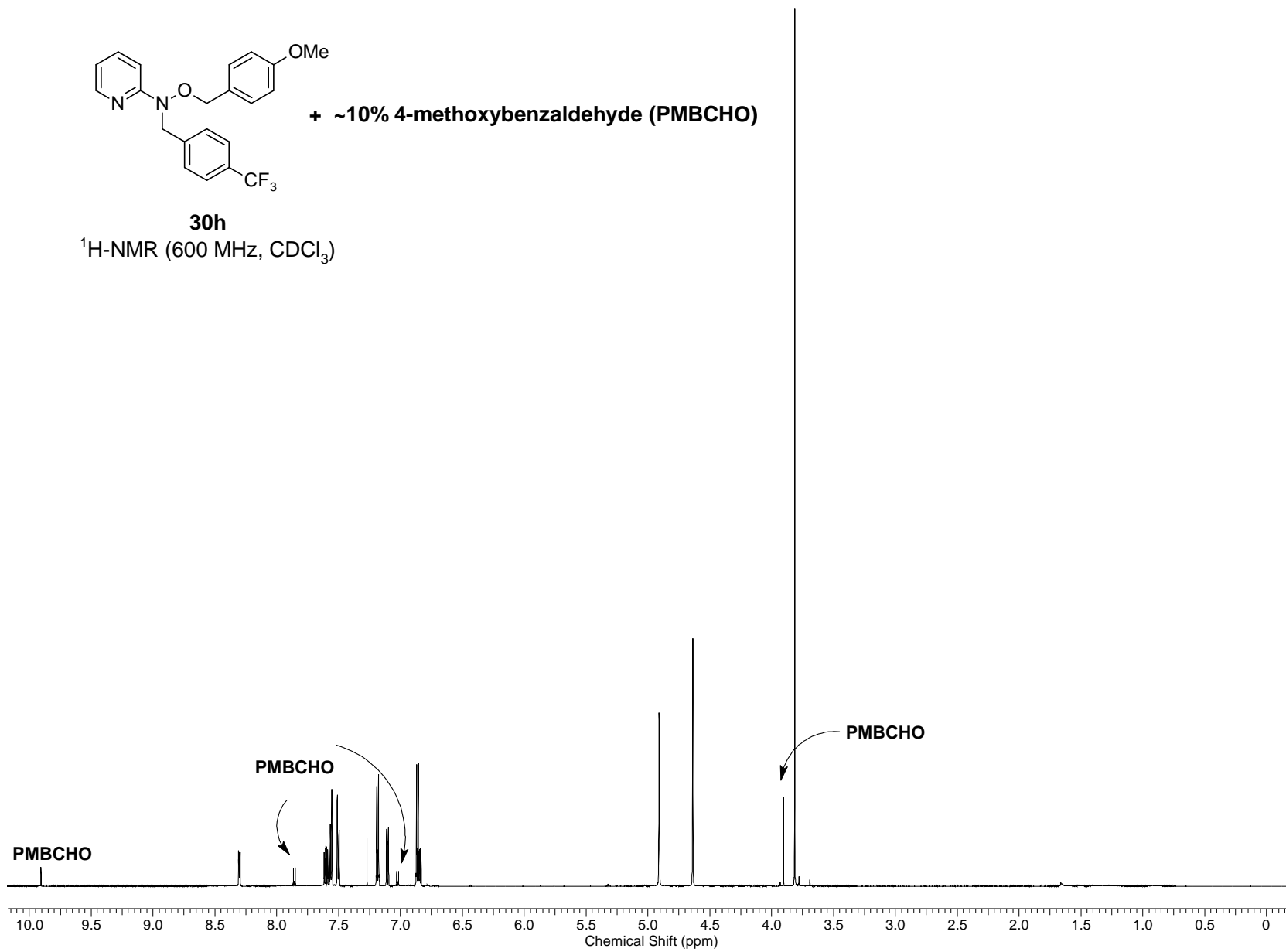


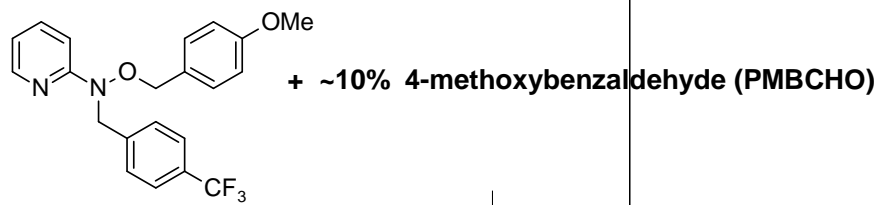


+ ~10% 4-methoxybenzaldehyde (PMBCHO)

**30h**

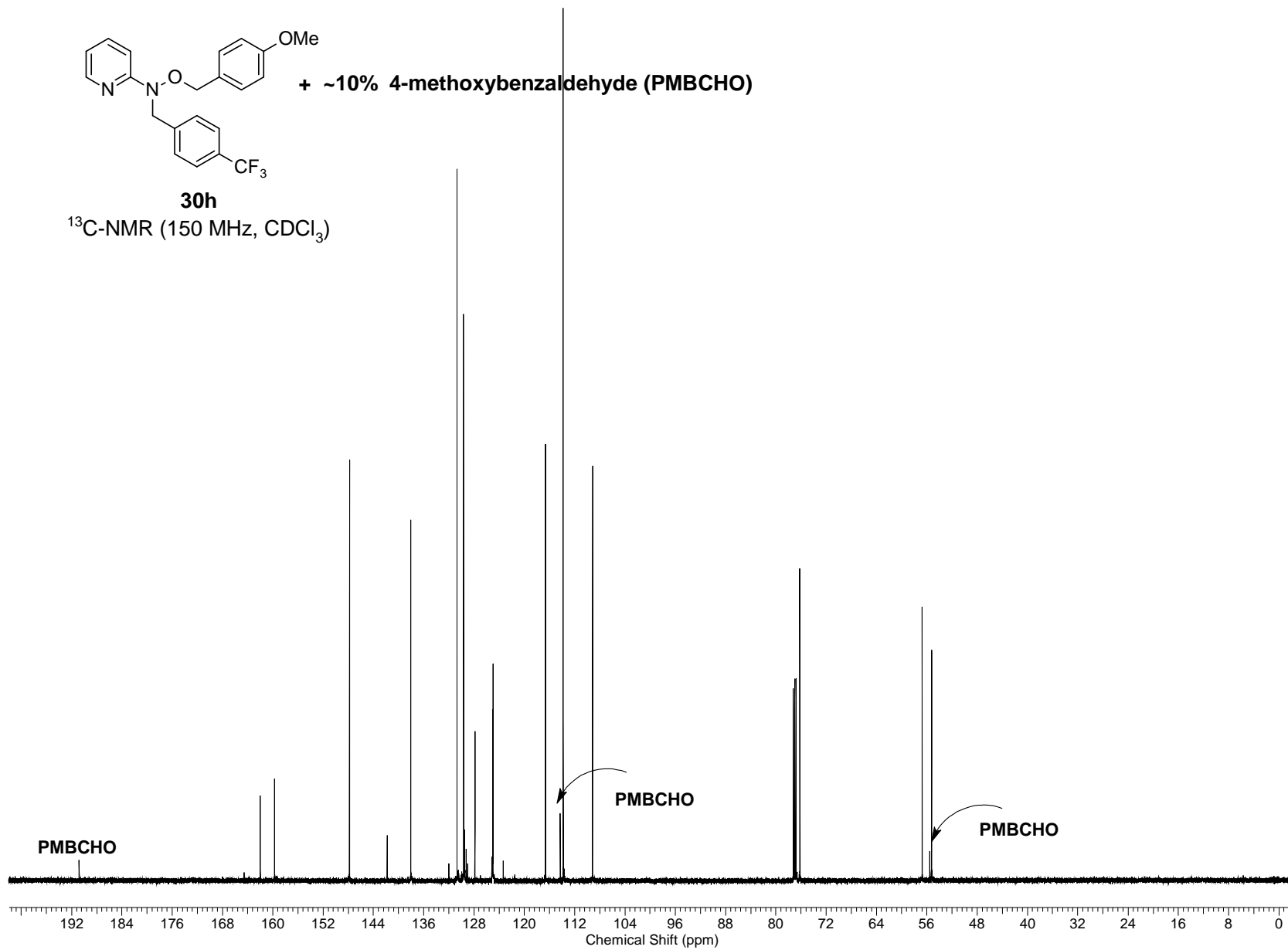
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

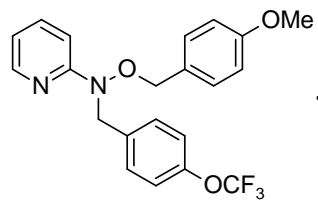




**30h**

<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

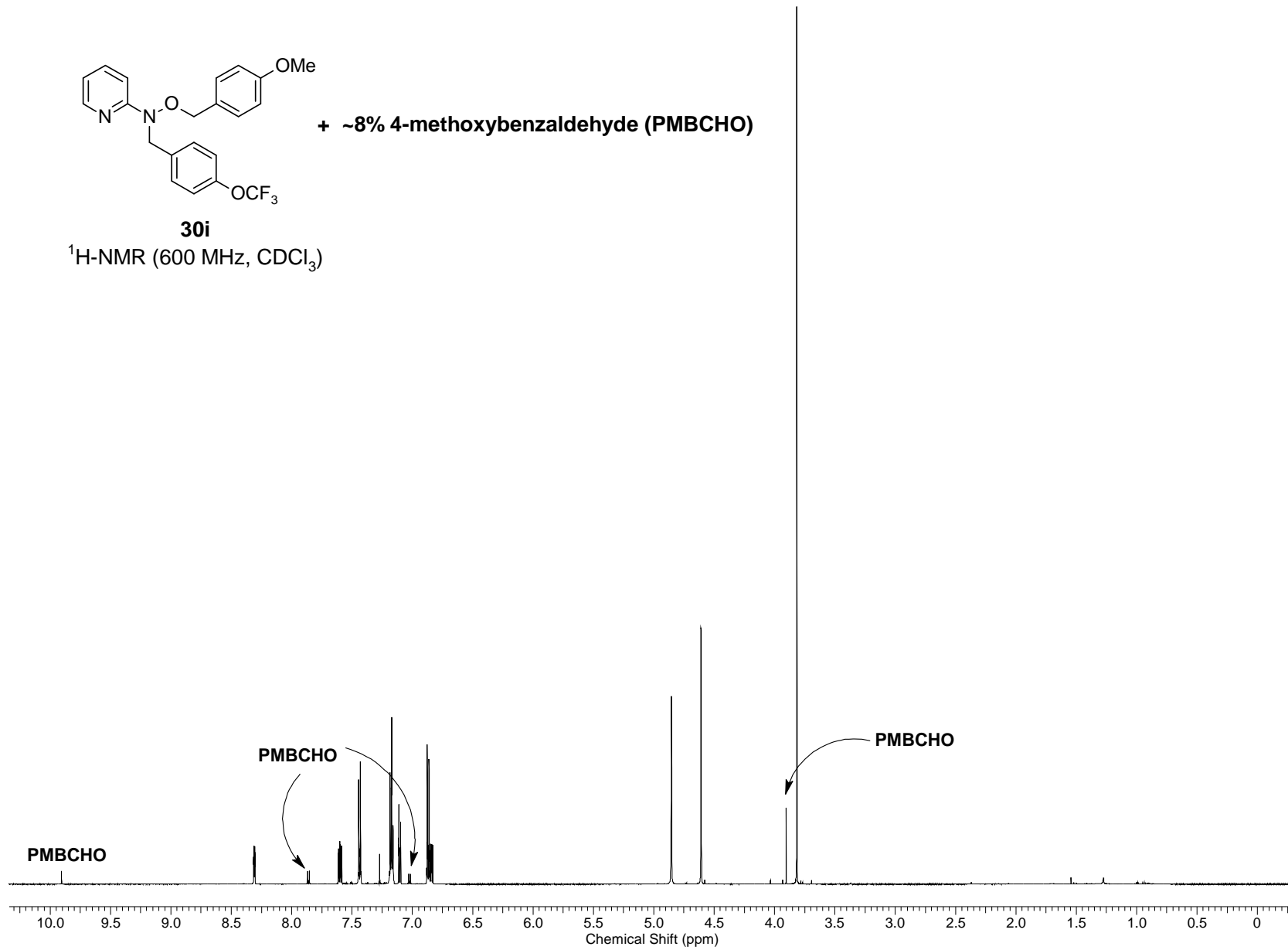


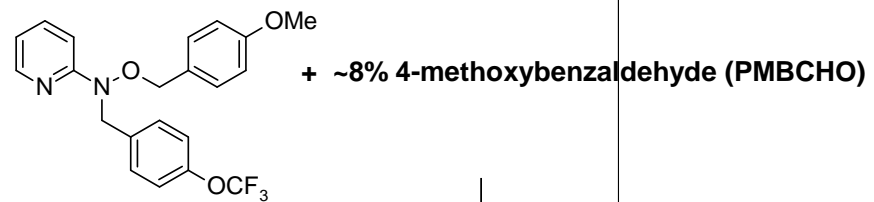


+ ~8% 4-methoxybenzaldehyde (PMBCHO)

**30i**

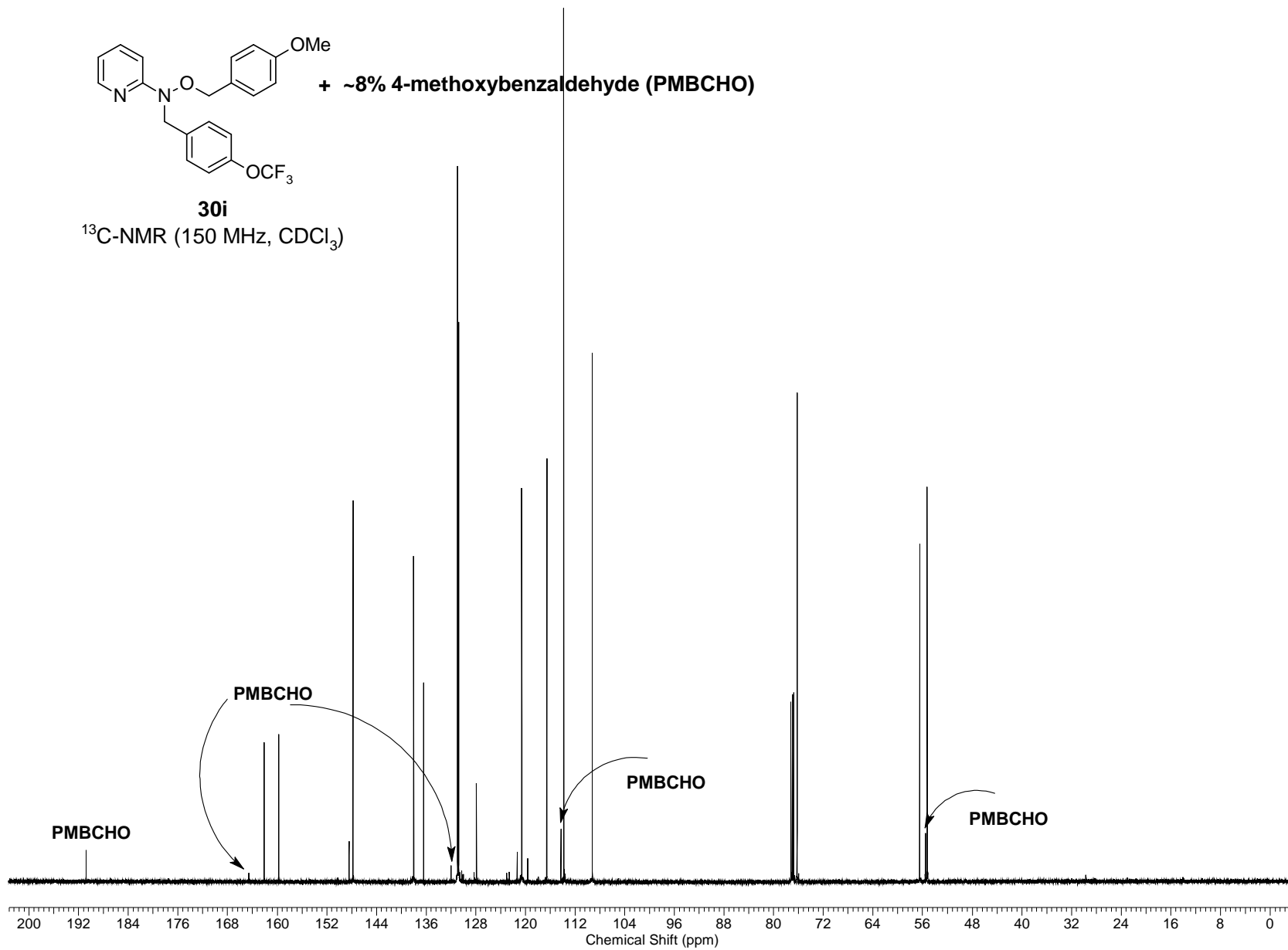
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

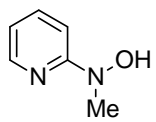




**30i**

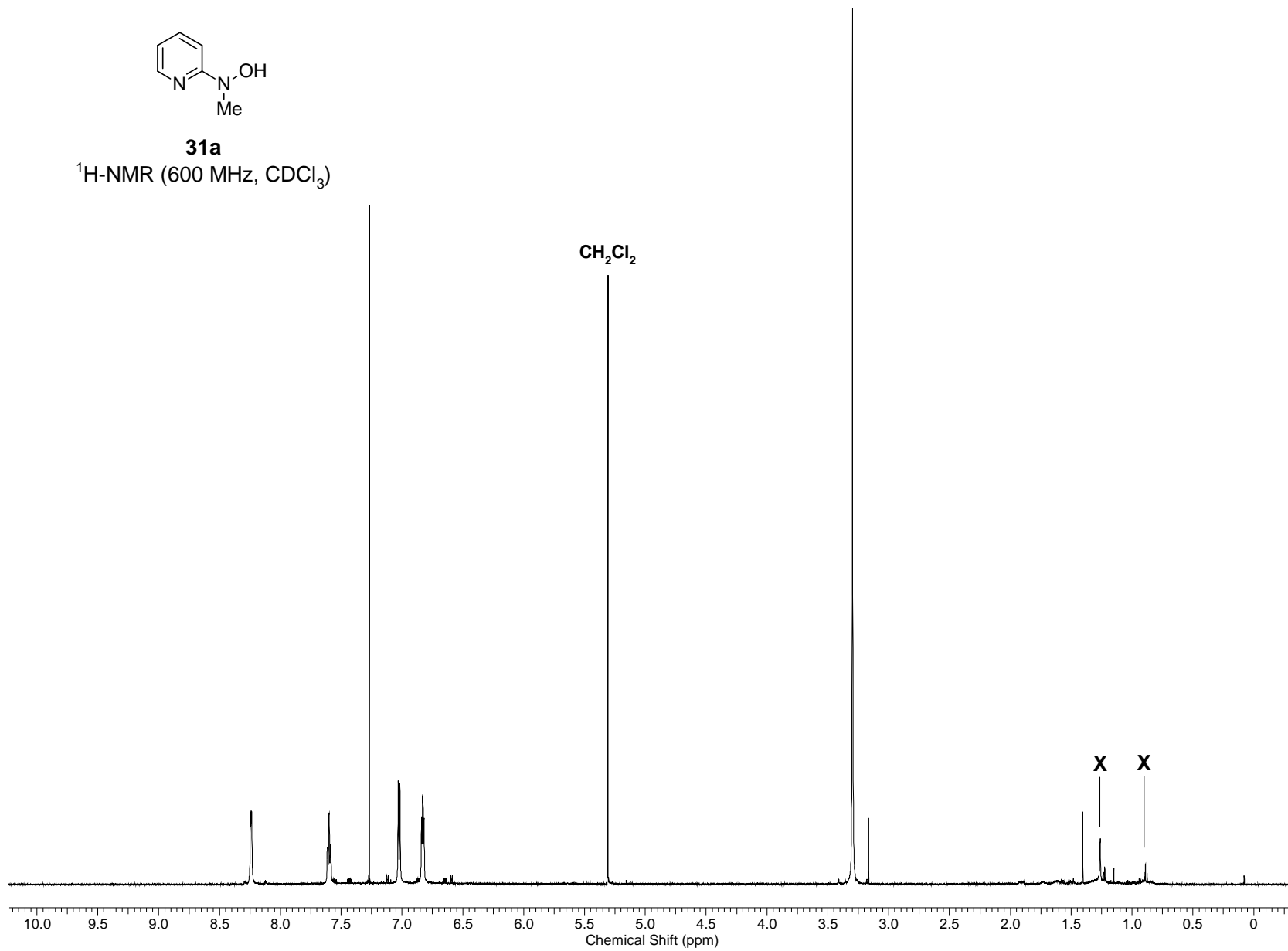
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)



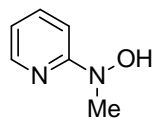


**31a**

<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

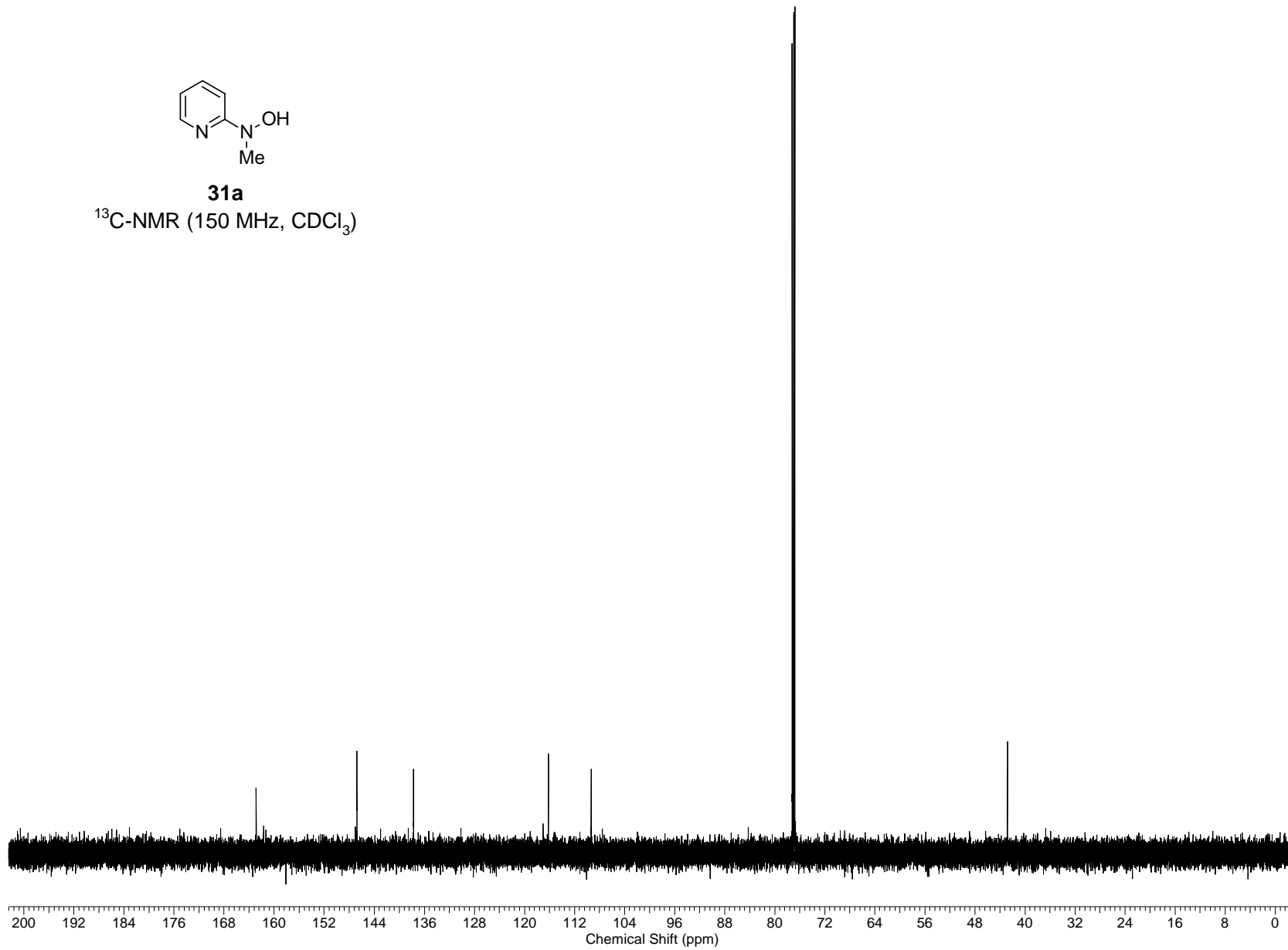


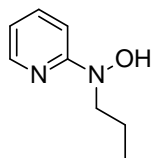




**31a**

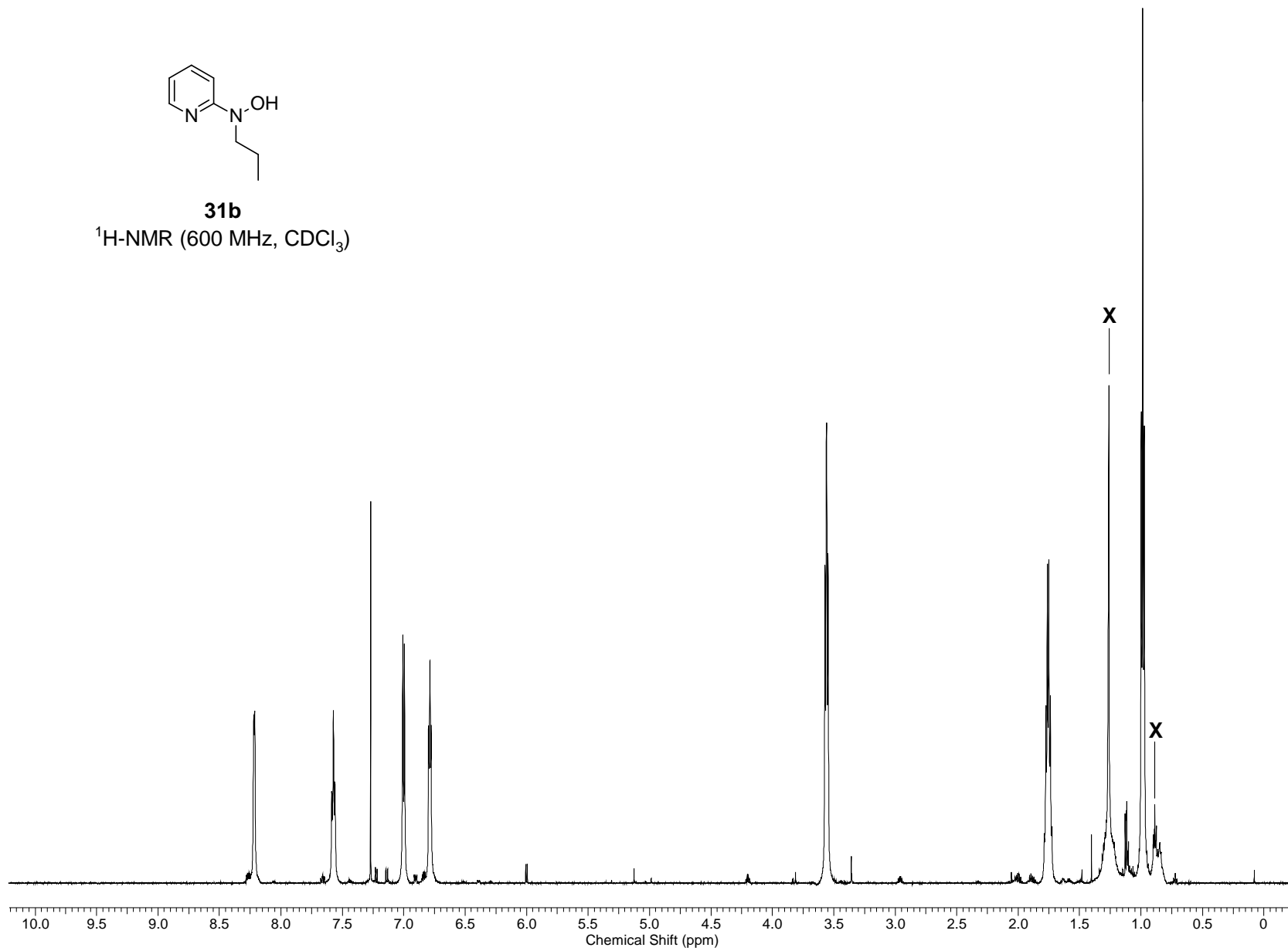
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

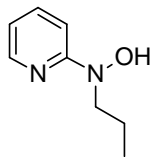




**31b**

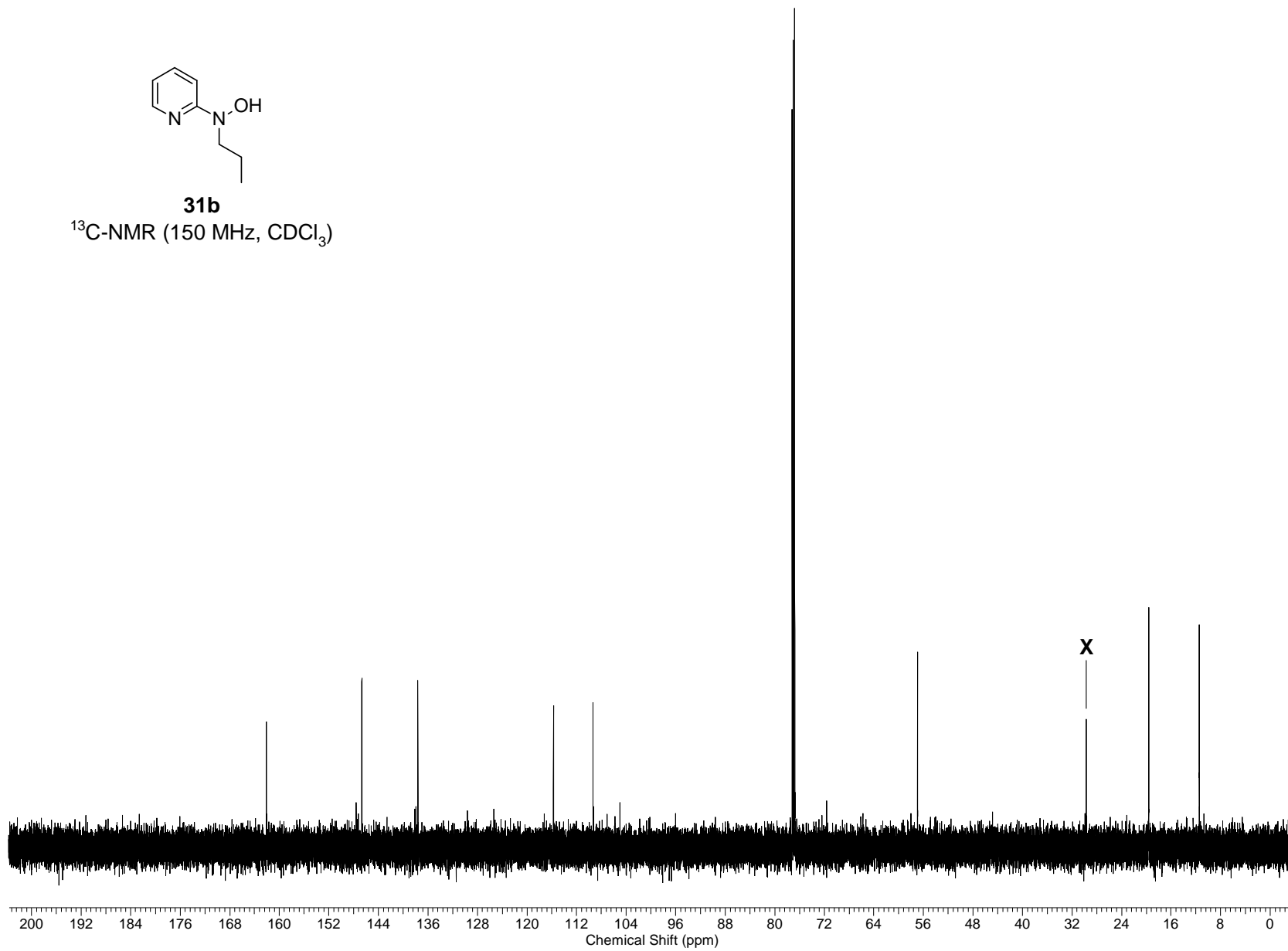
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

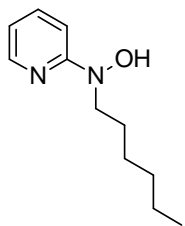




**31b**

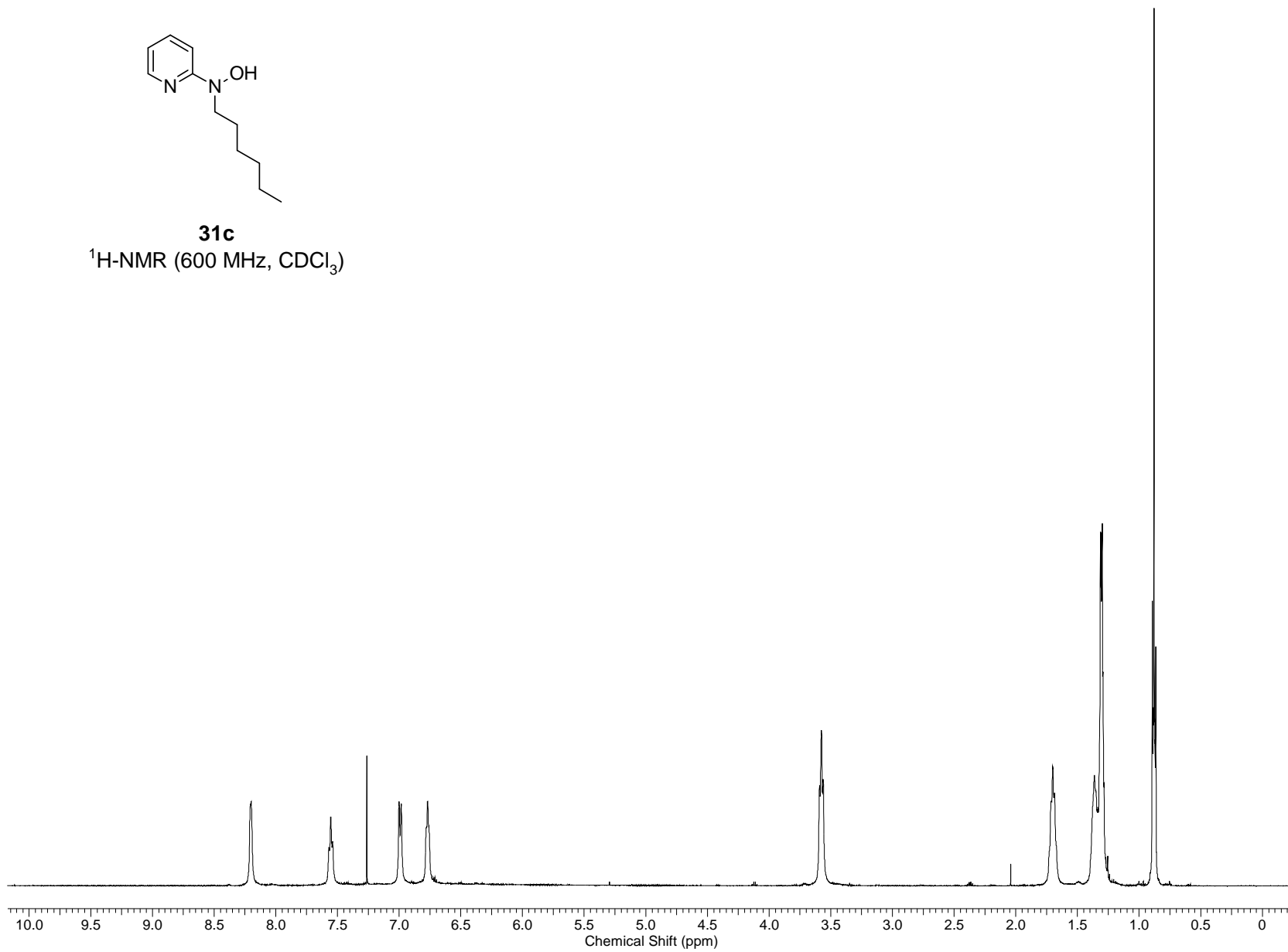
$^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )

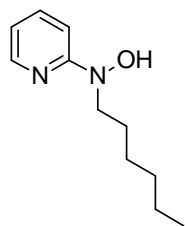




**31c**

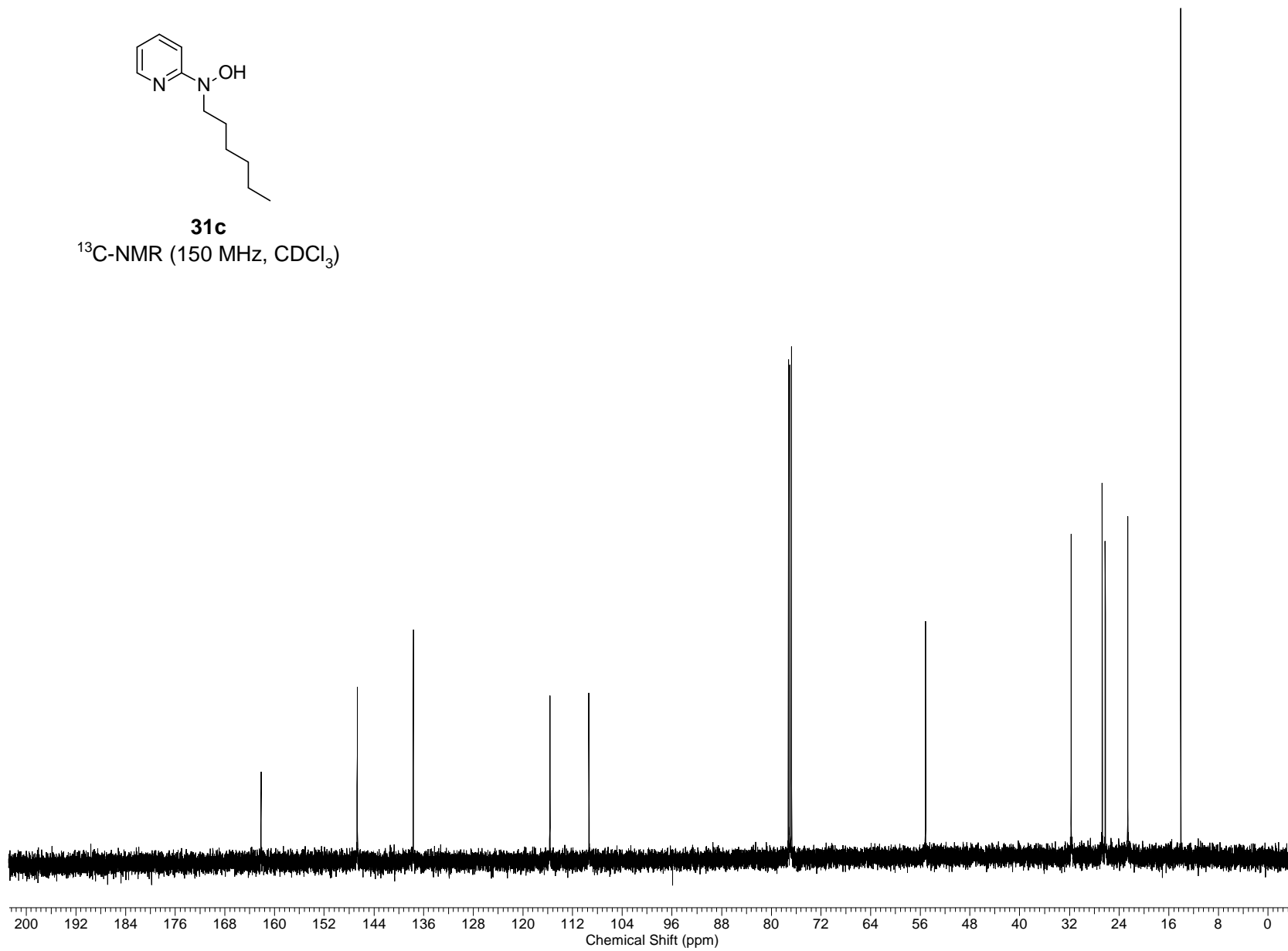
$^1\text{H-NMR}$  (600 MHz,  $\text{CDCl}_3$ )

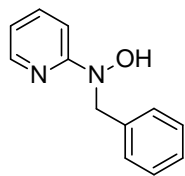




**31c**

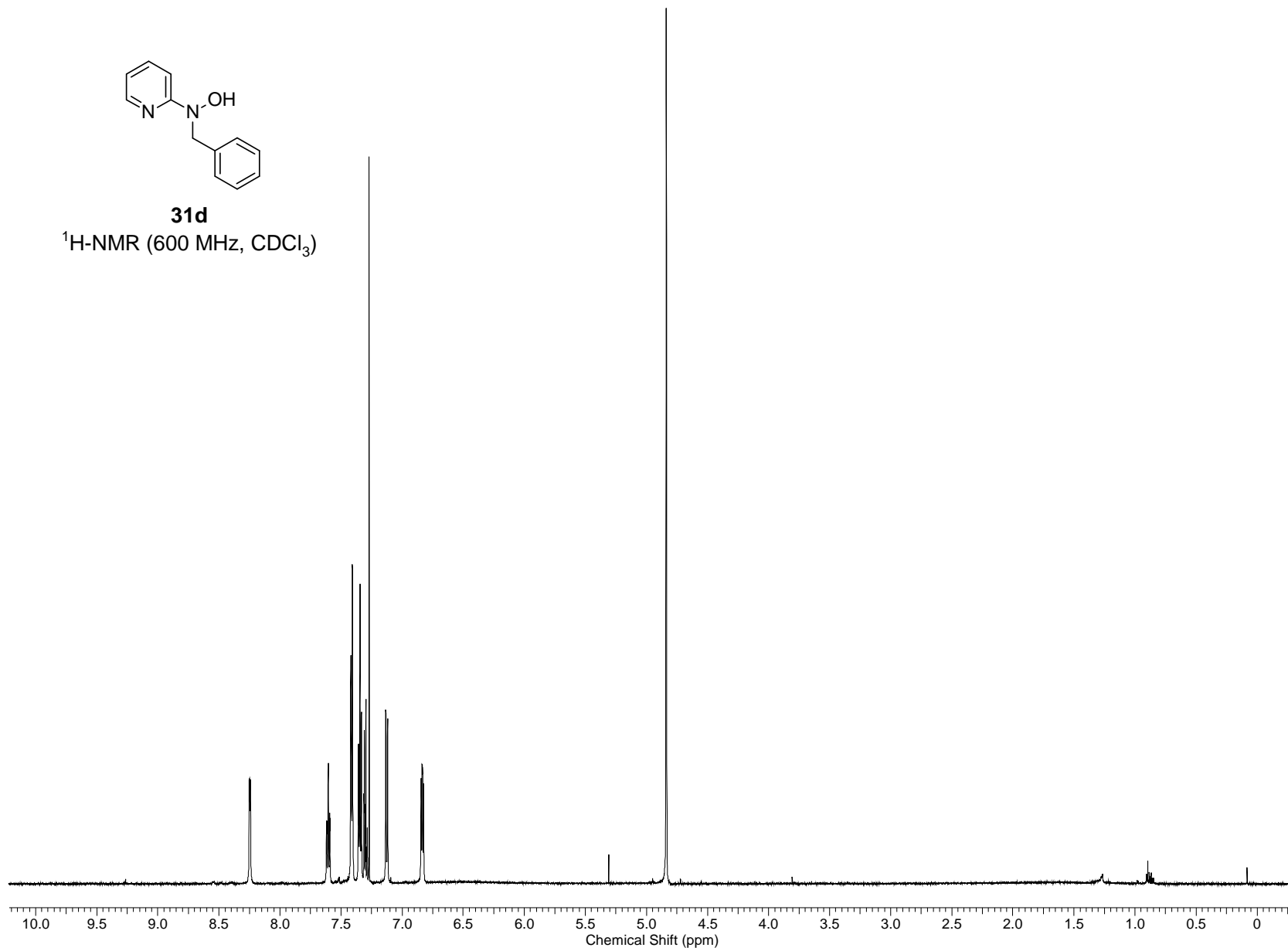
$^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )

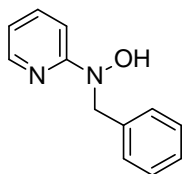




**31d**

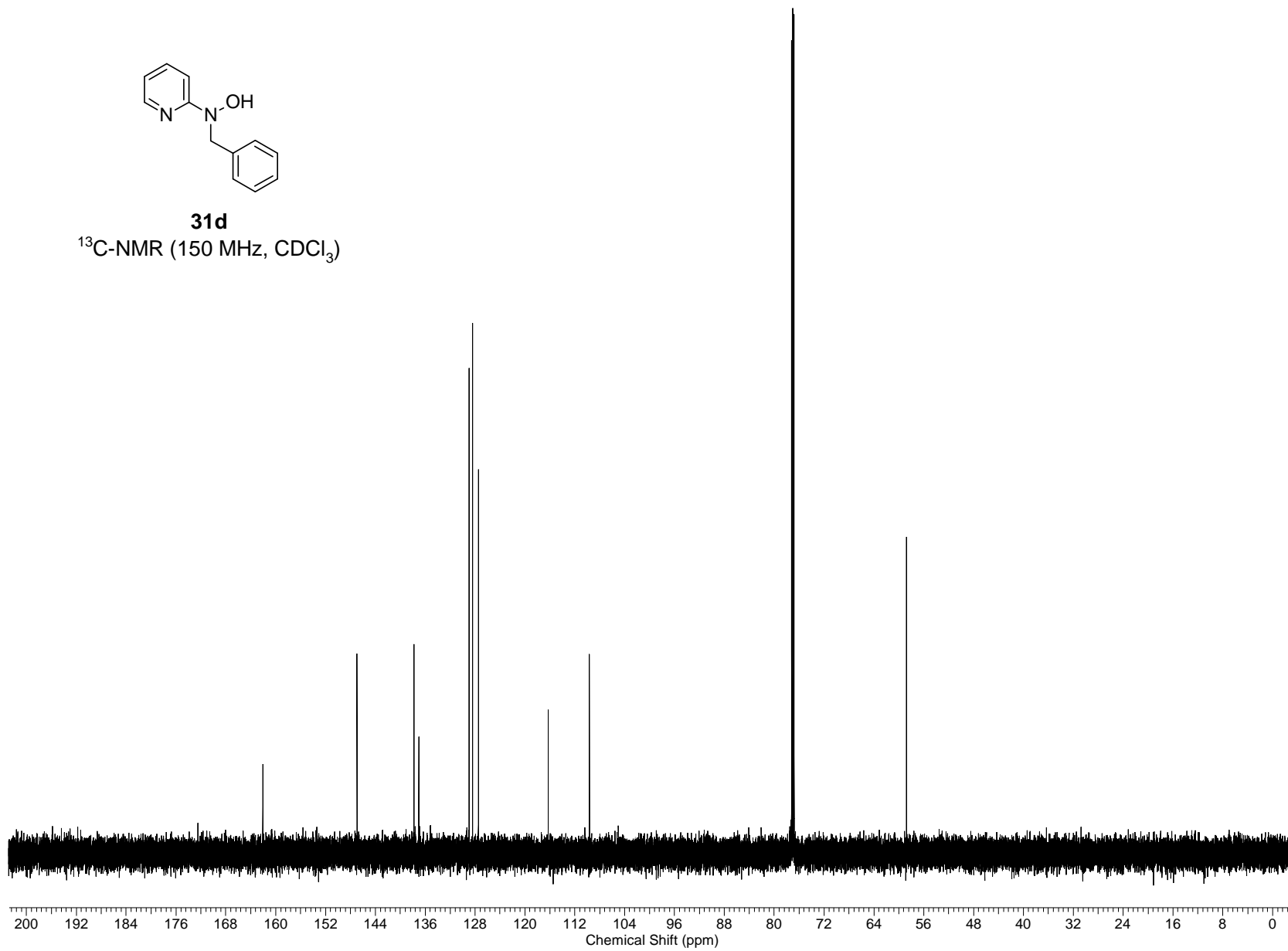
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

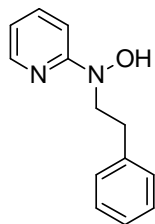




**31d**

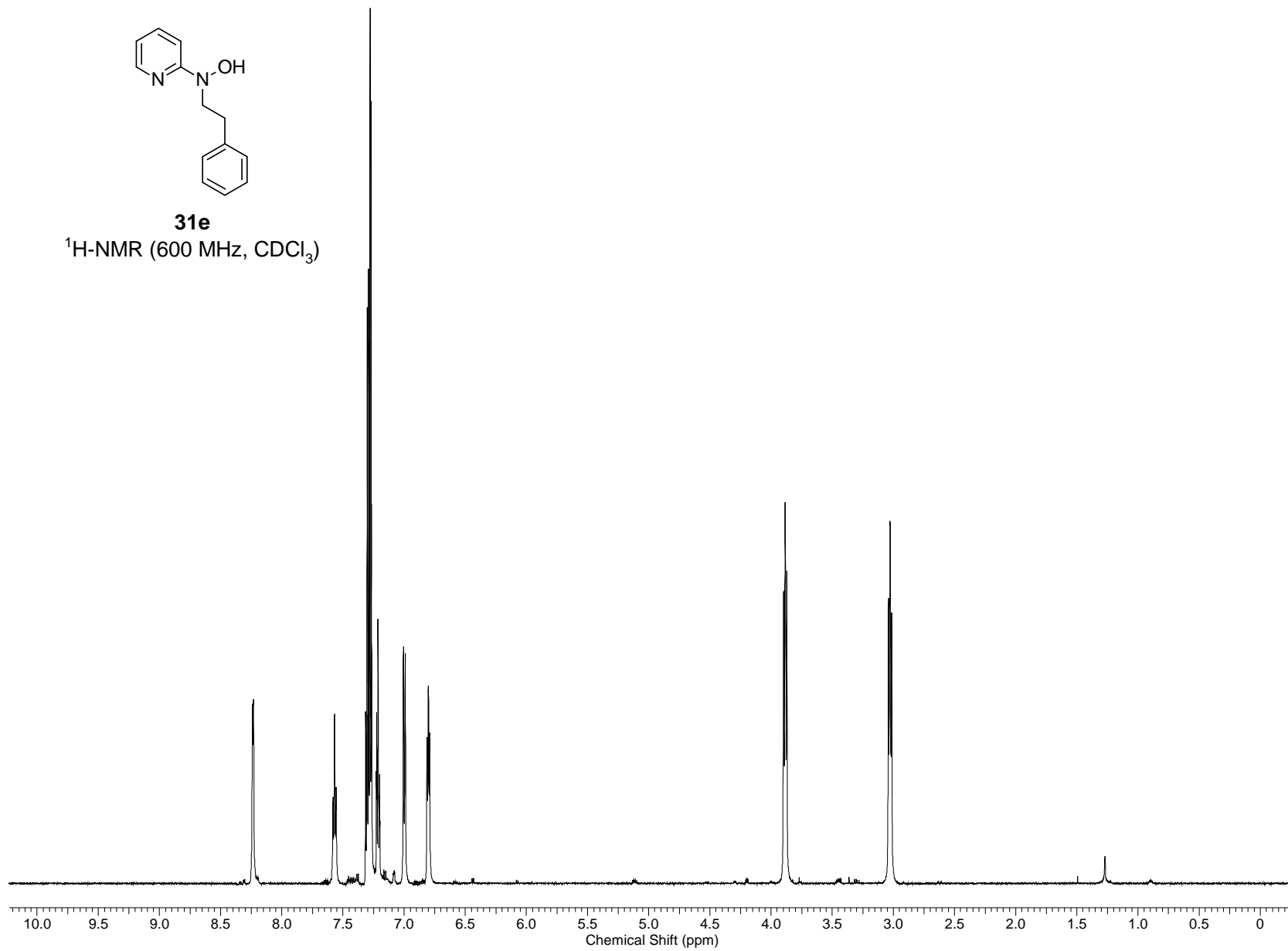
$^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )



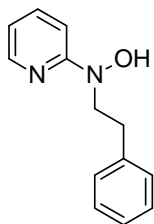


**31e**

<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

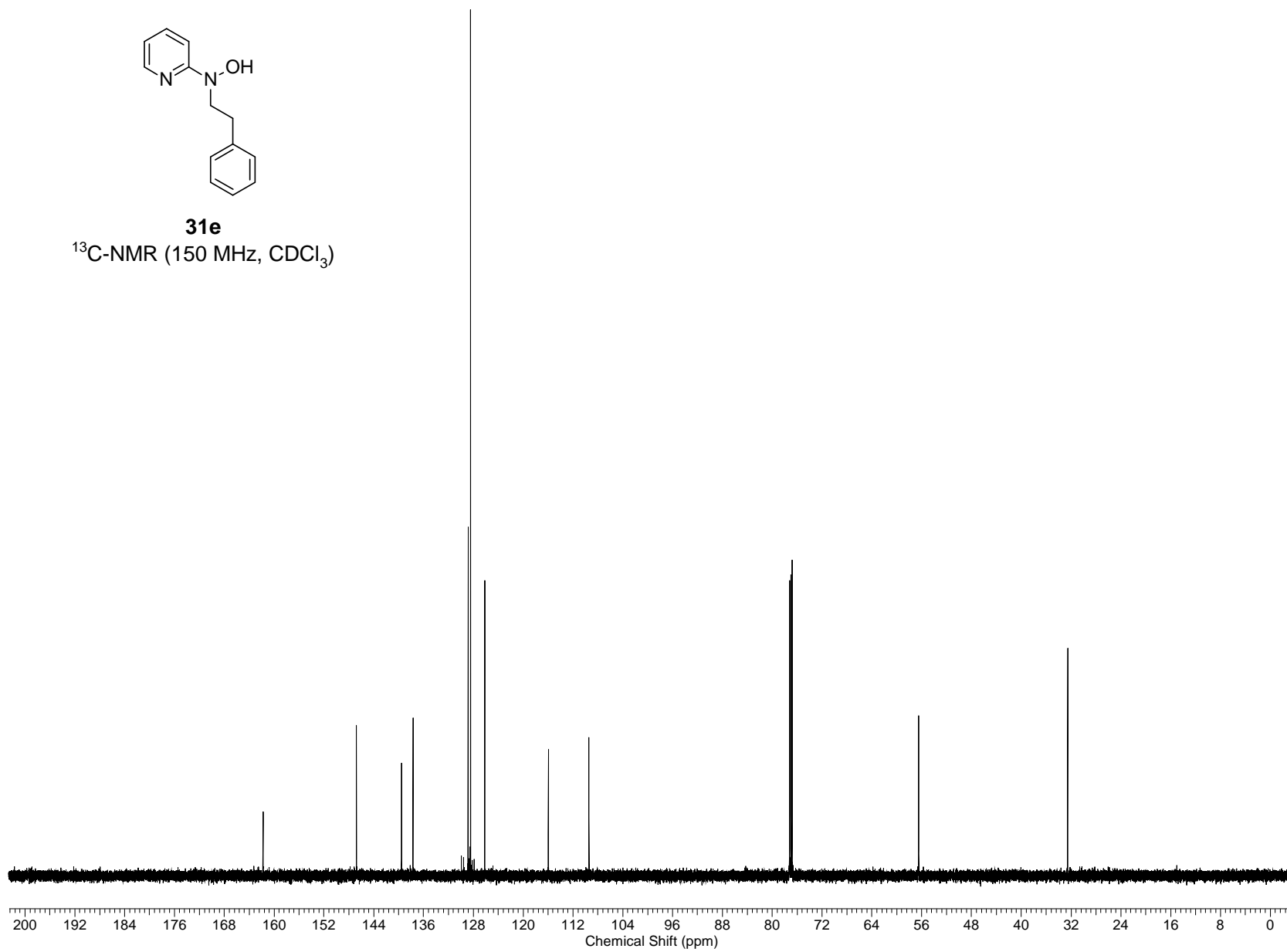


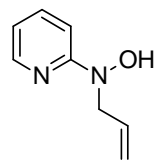




**31e**

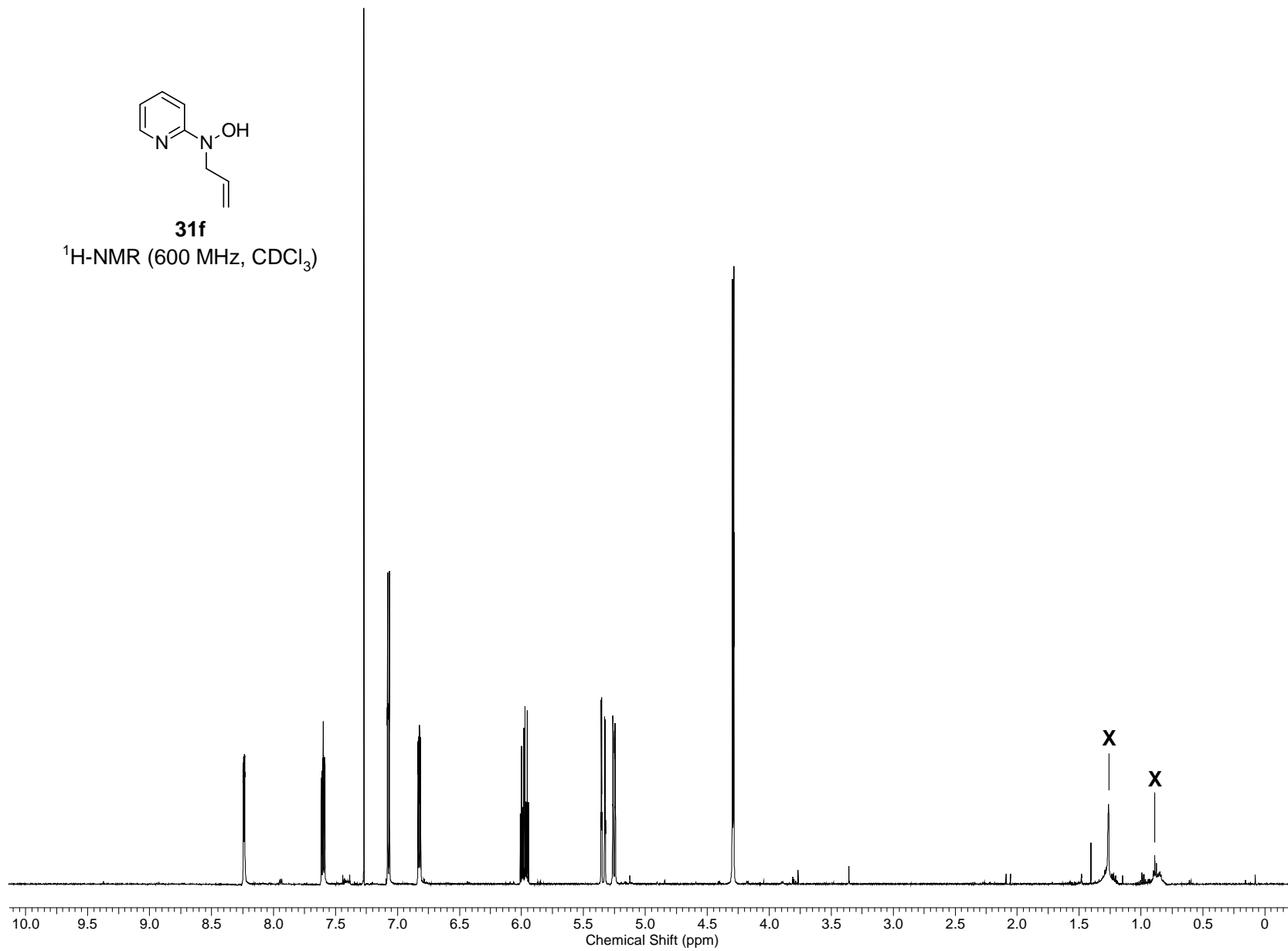
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

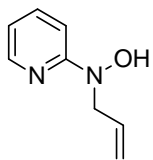




**31f**

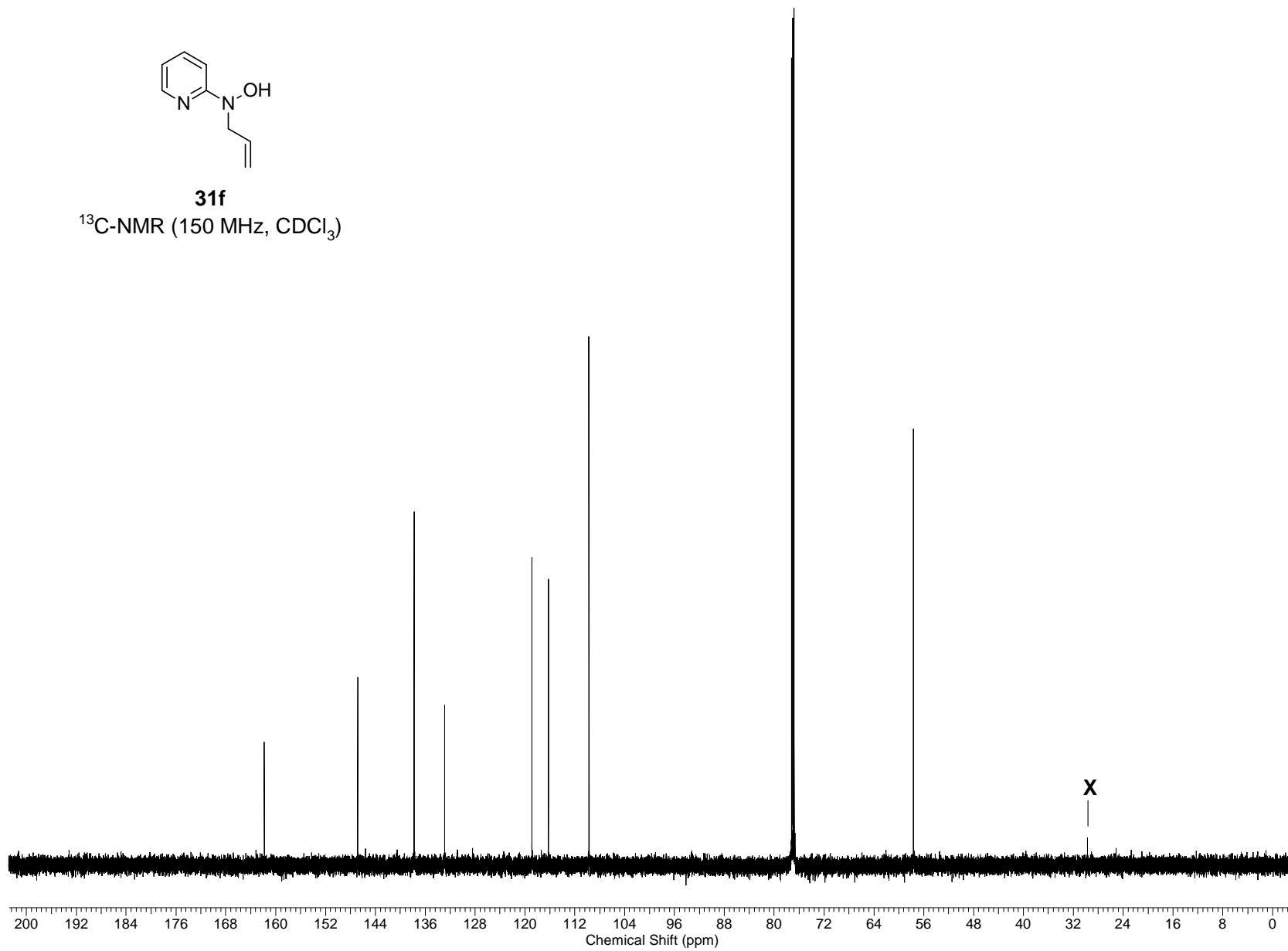
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

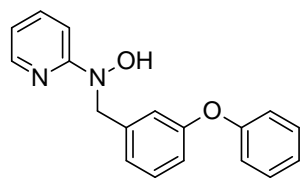




**31f**

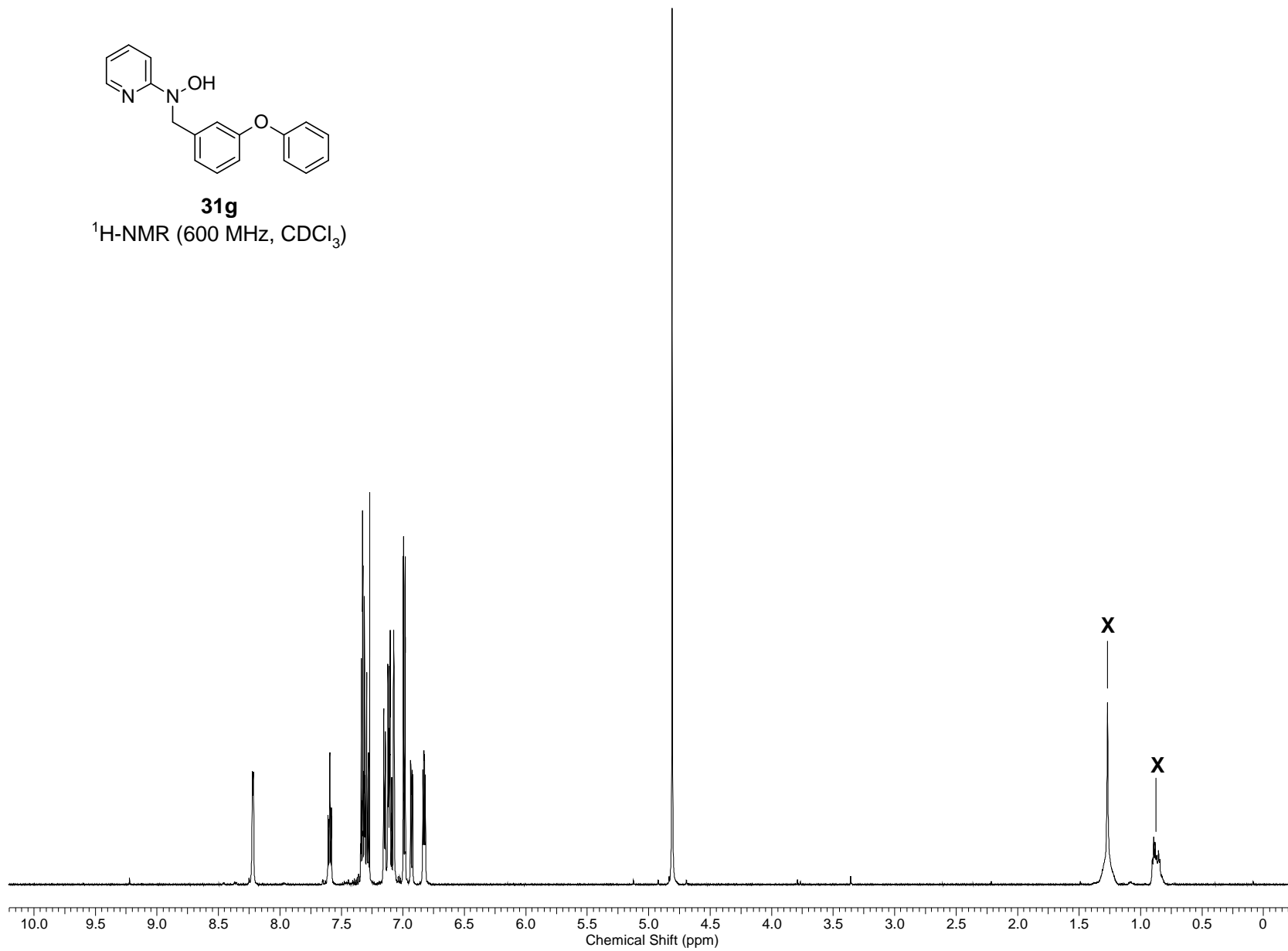
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

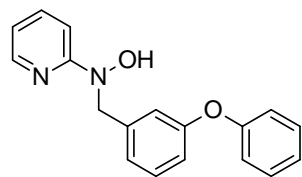




**31g**

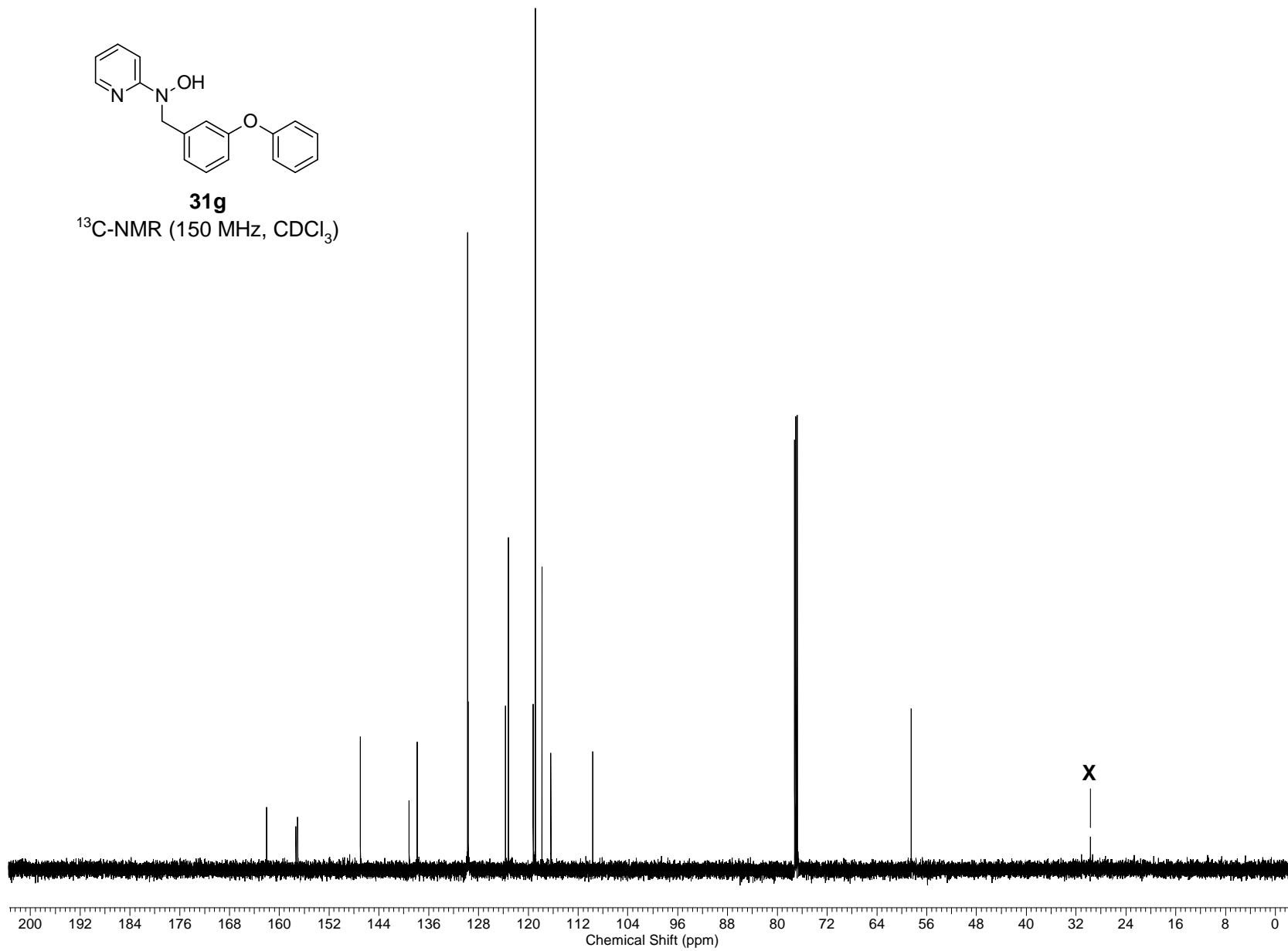
<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)



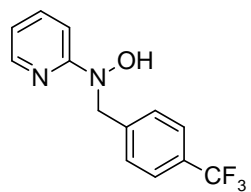


**31g**

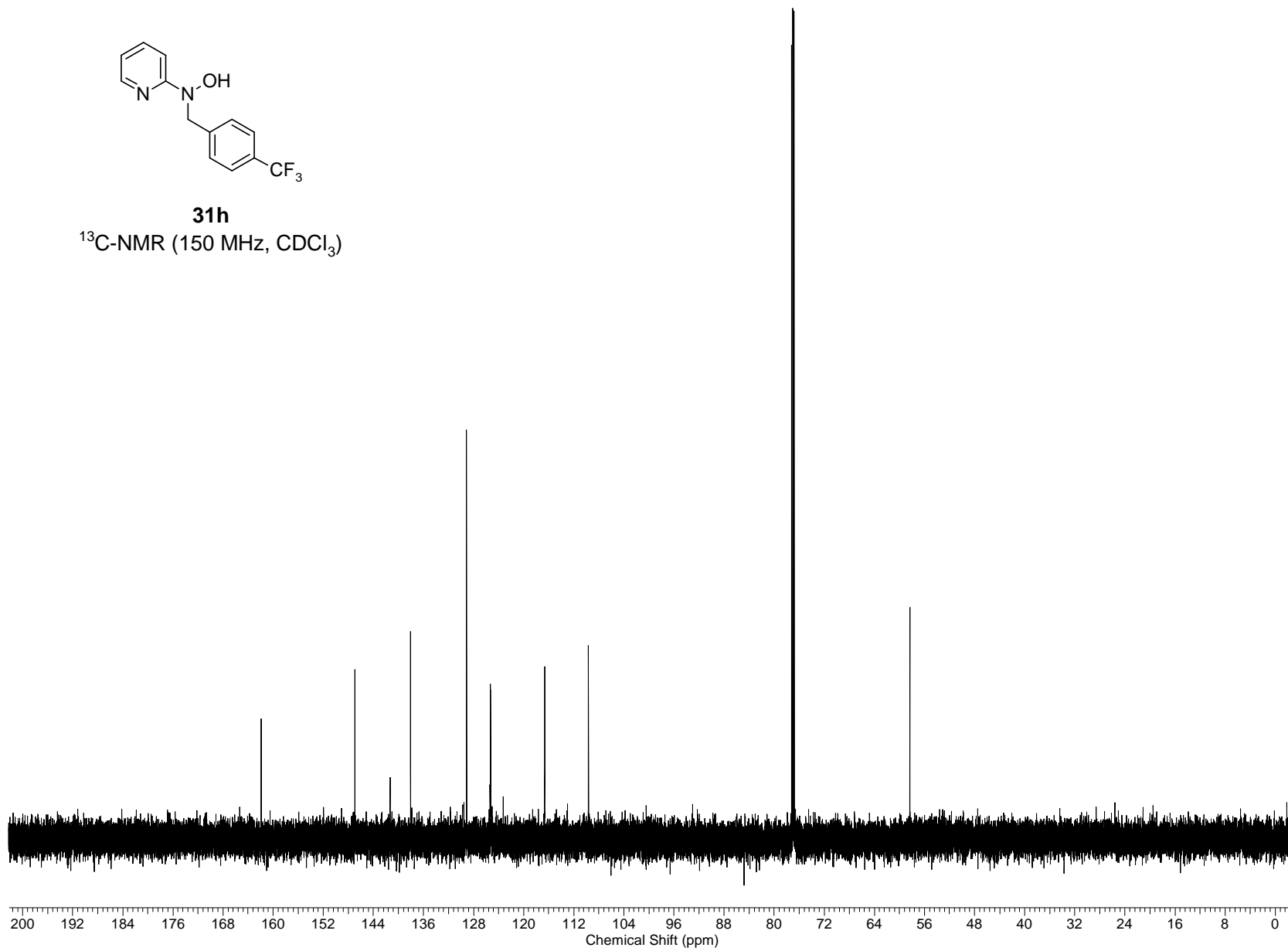
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)

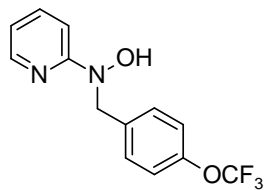






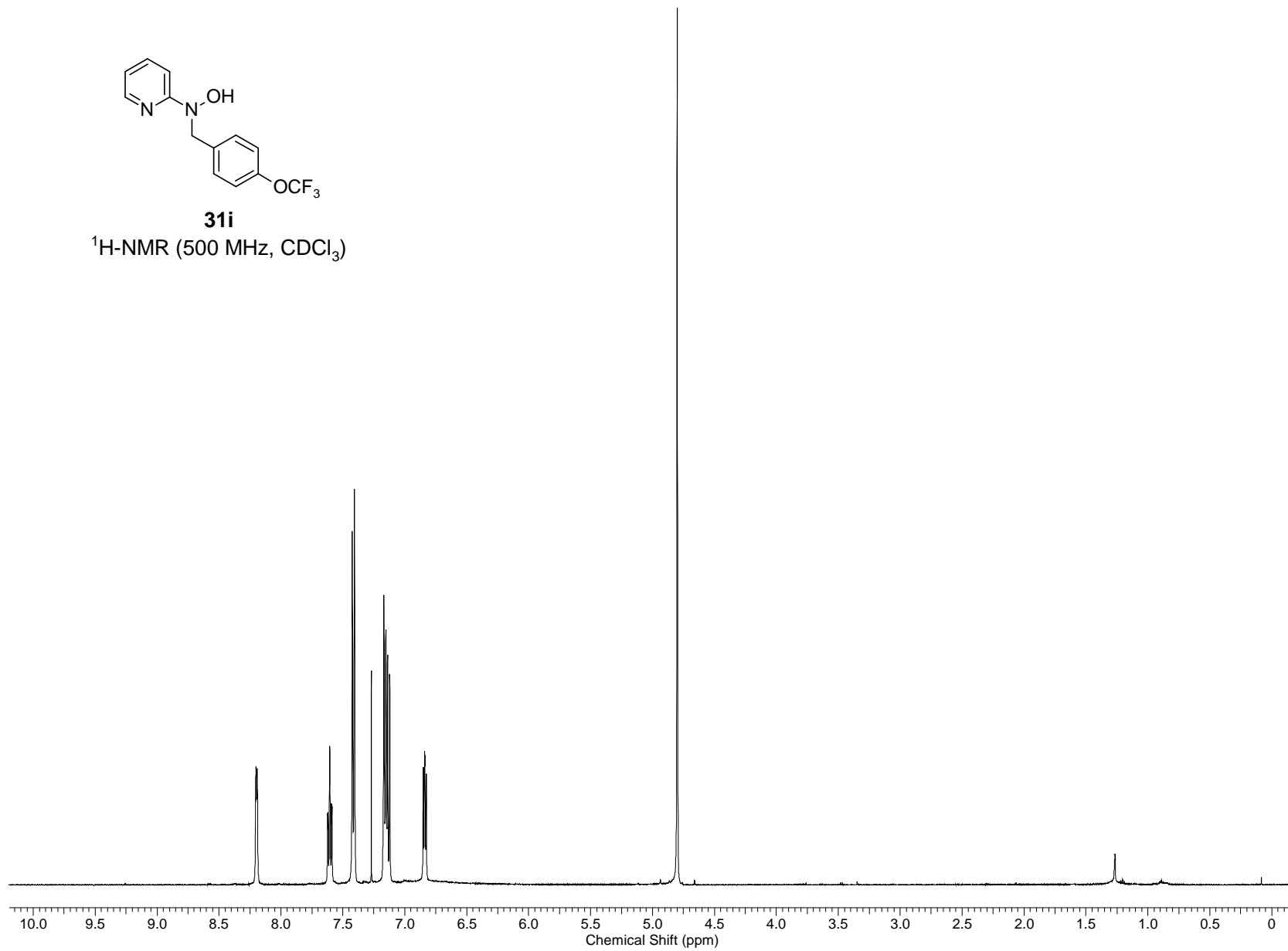
**31h**  
<sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)



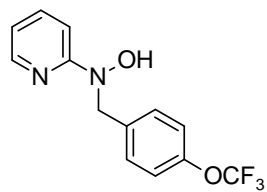


**31i**

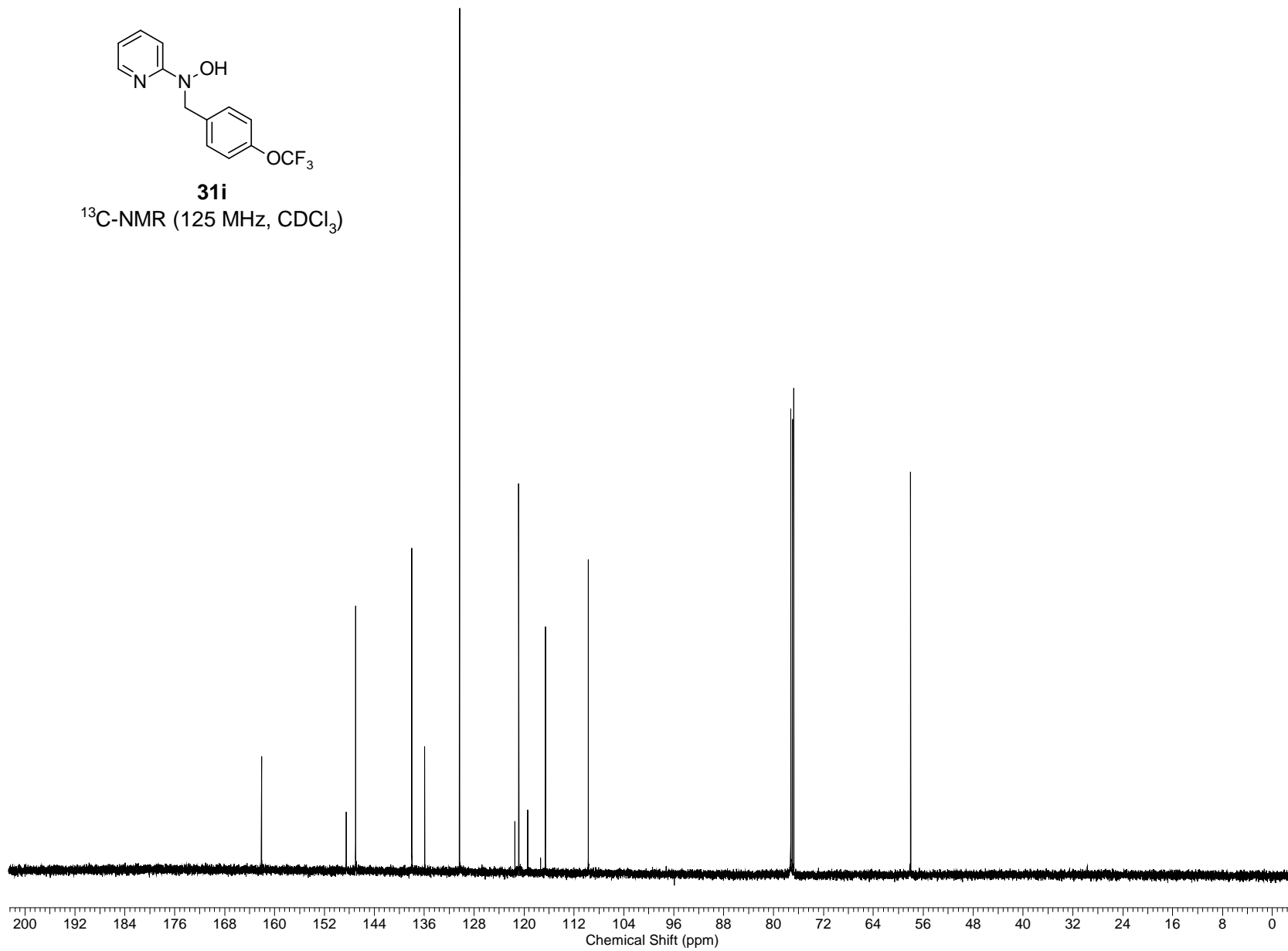
<sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)

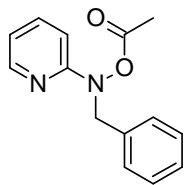






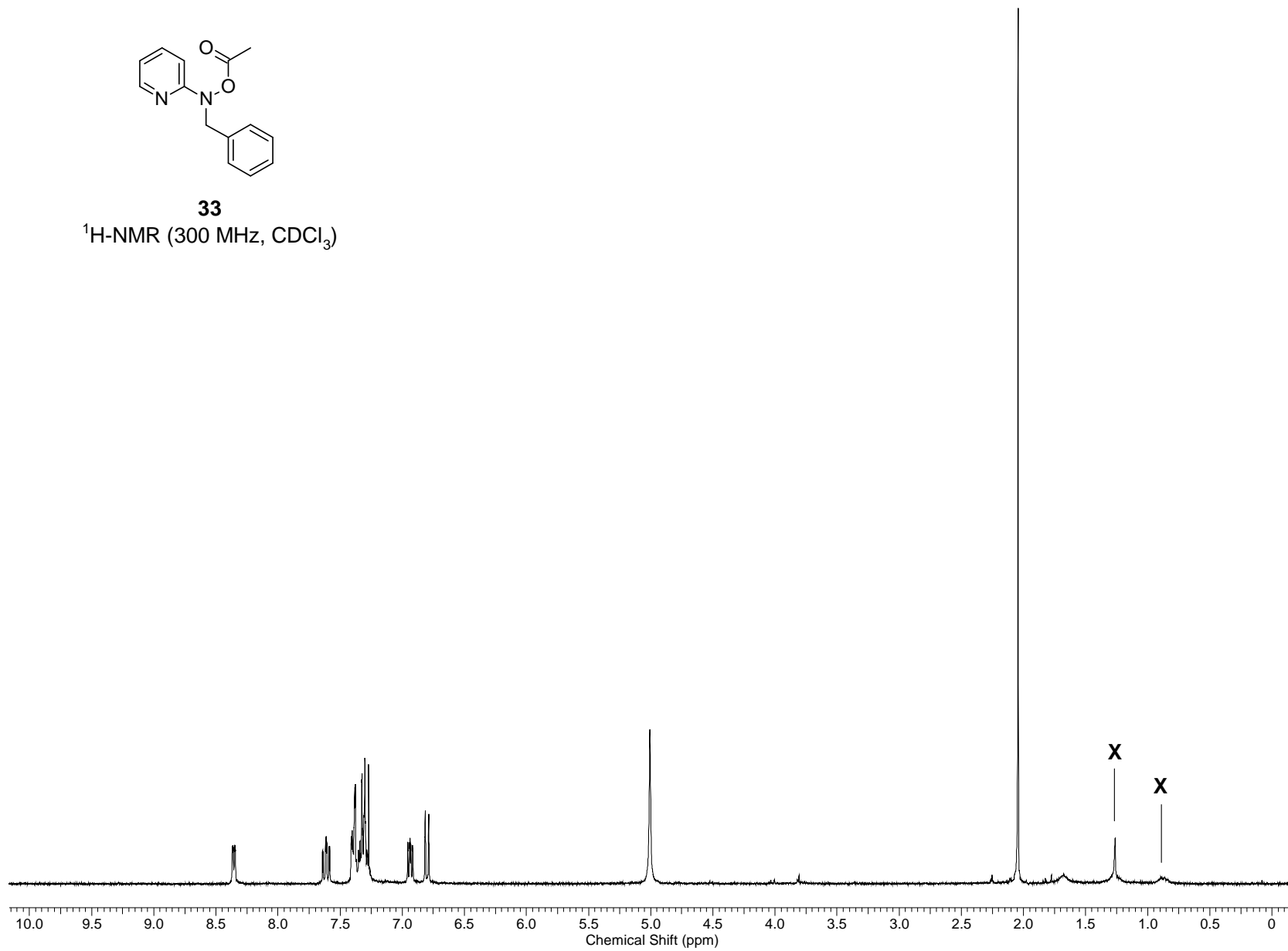
**31i**  
<sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>)

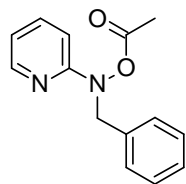




**33**

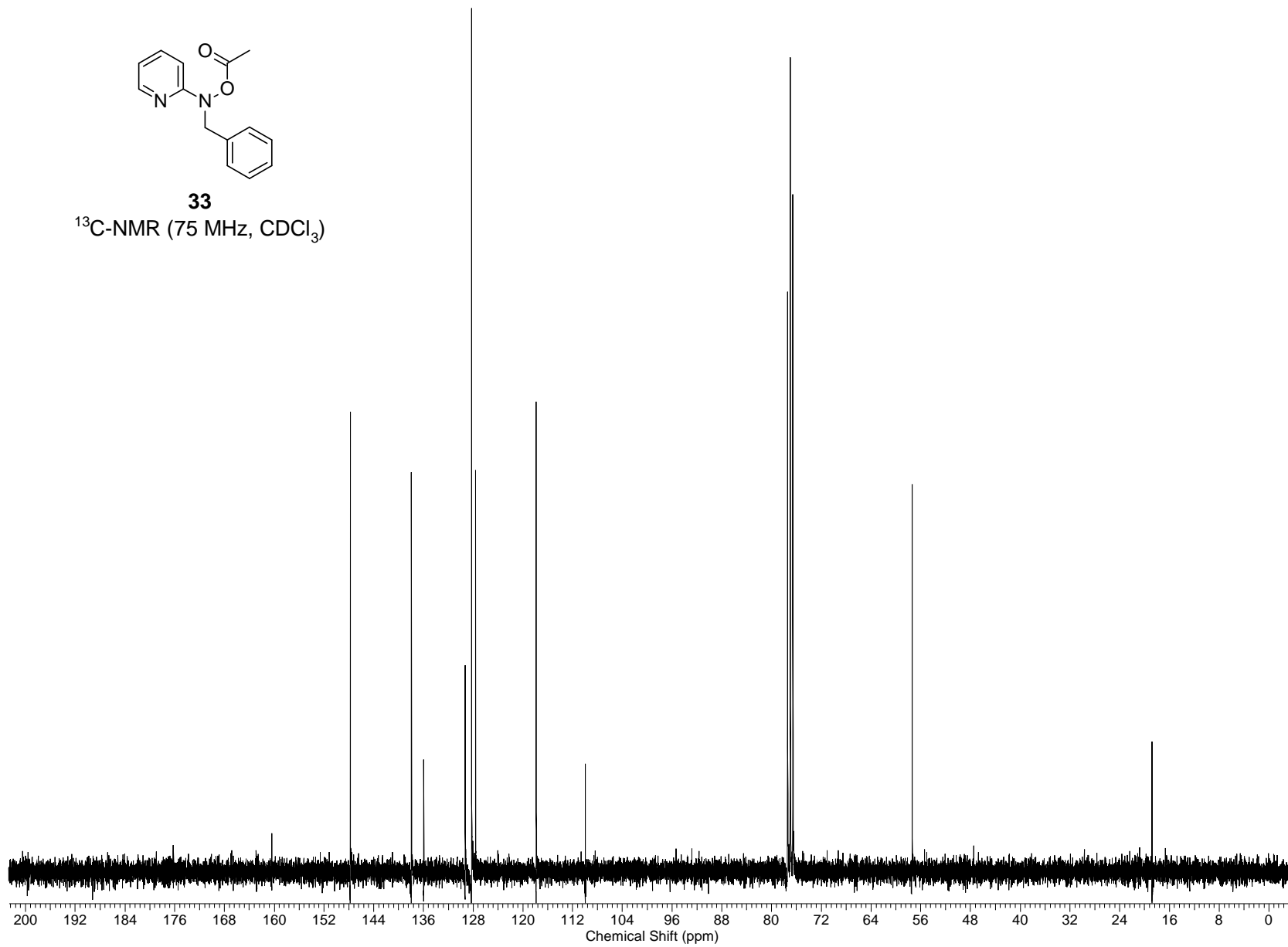
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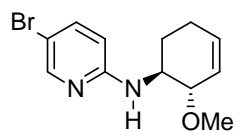




**33**

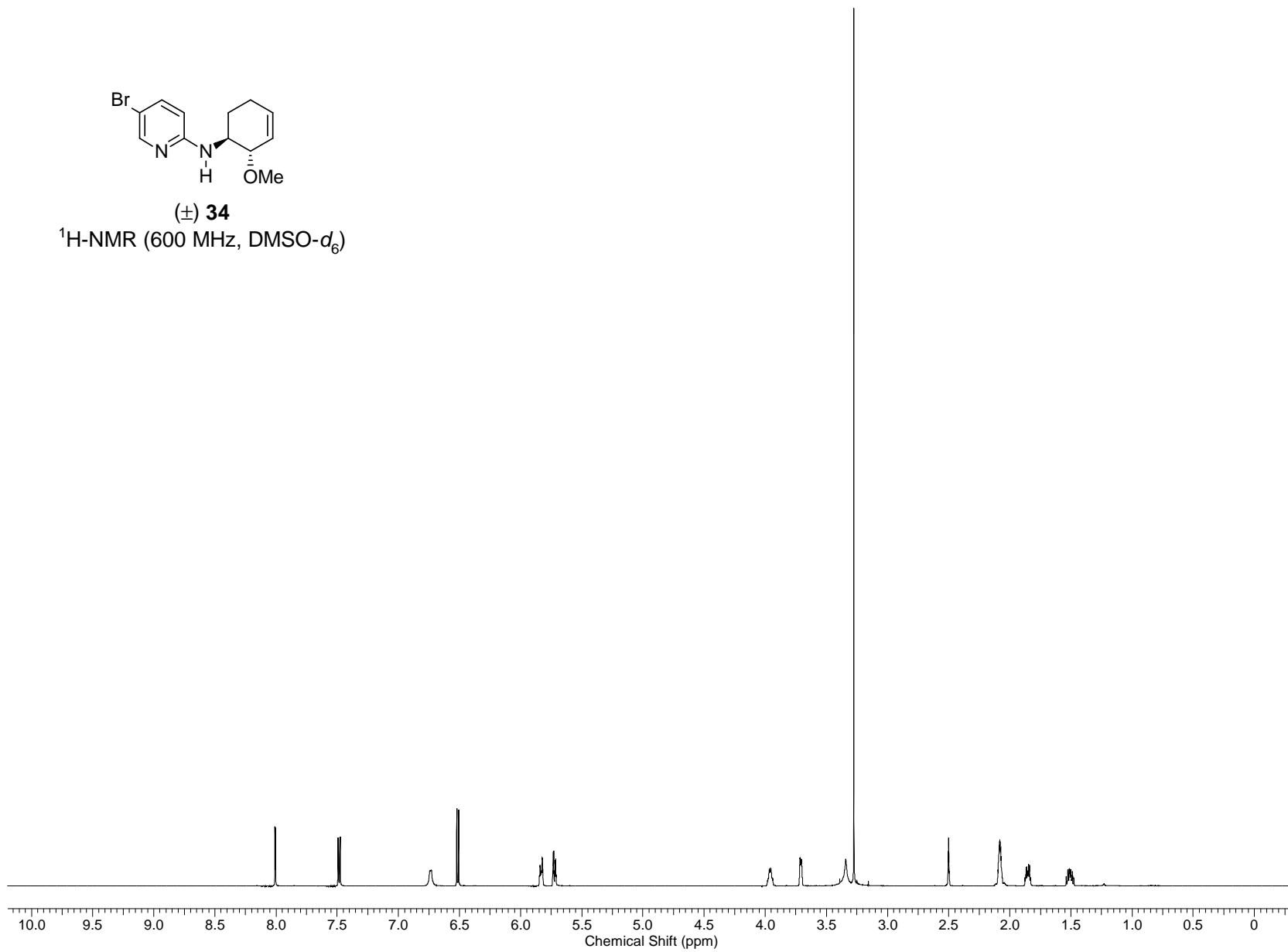
<sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>)

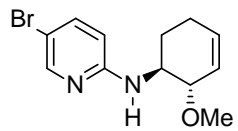




(±) **34**

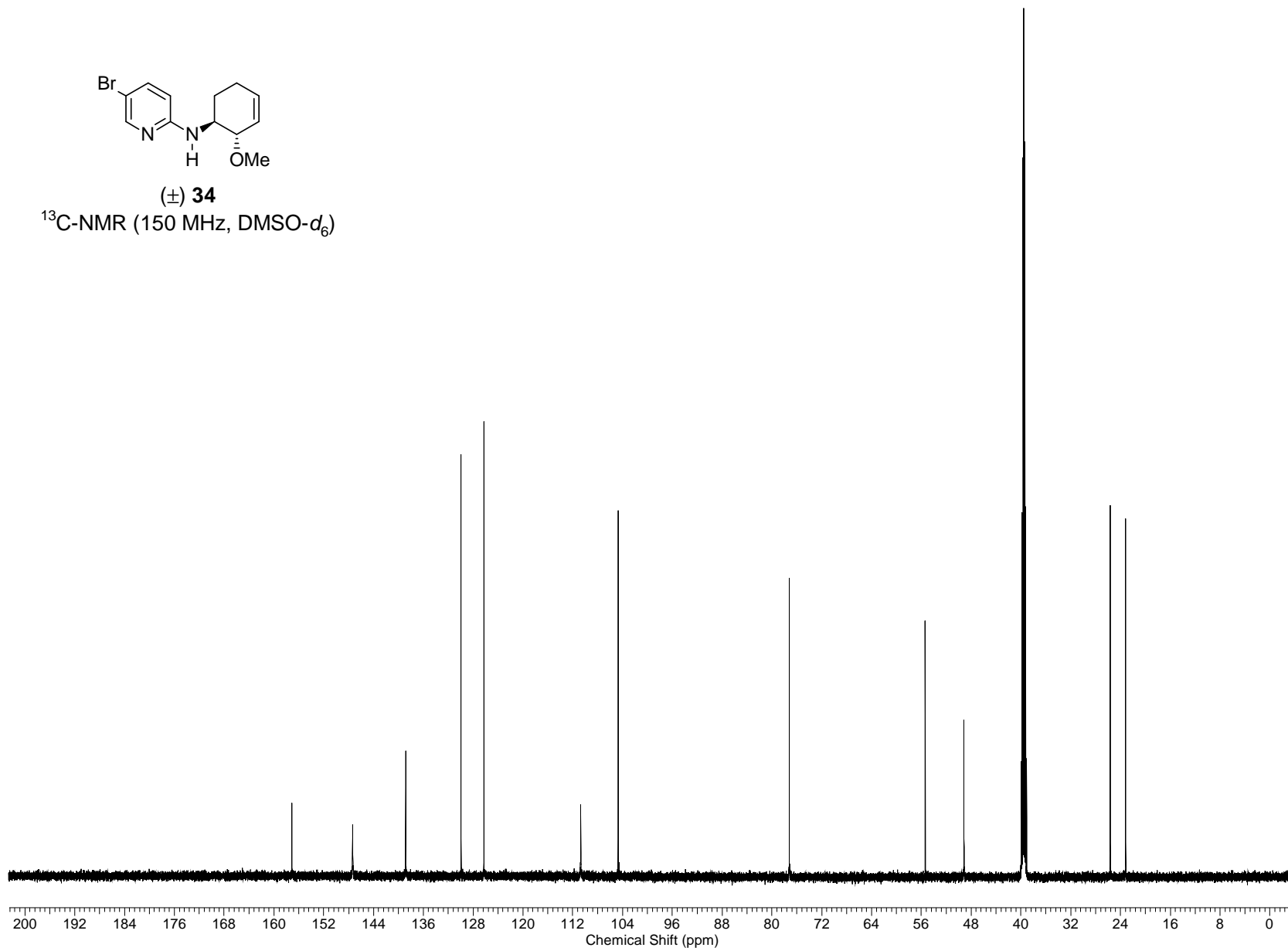
<sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)

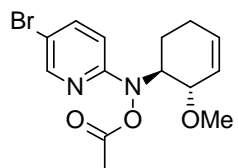




(±) **34**

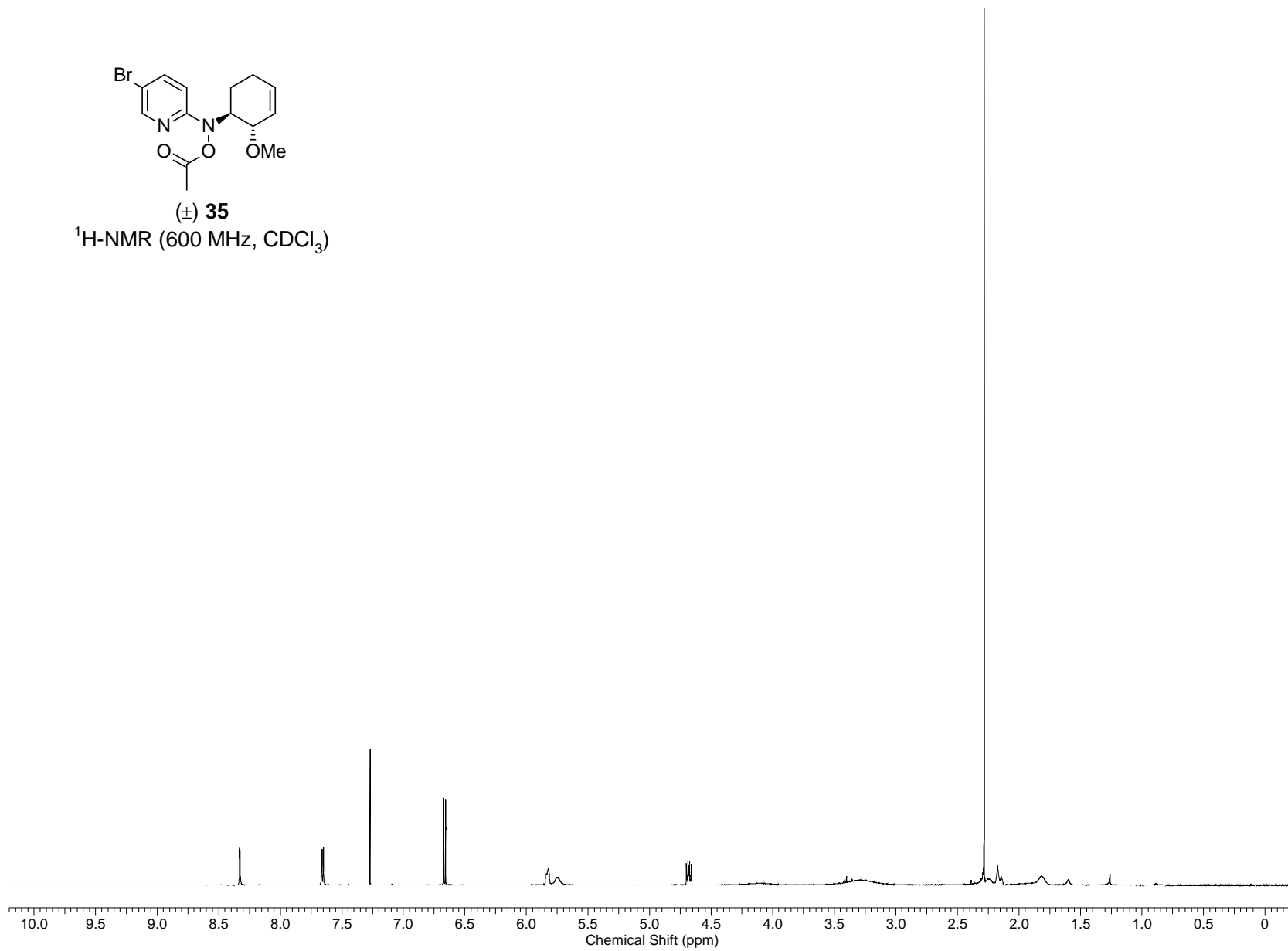
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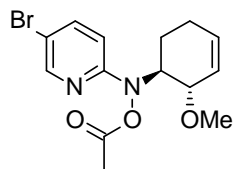




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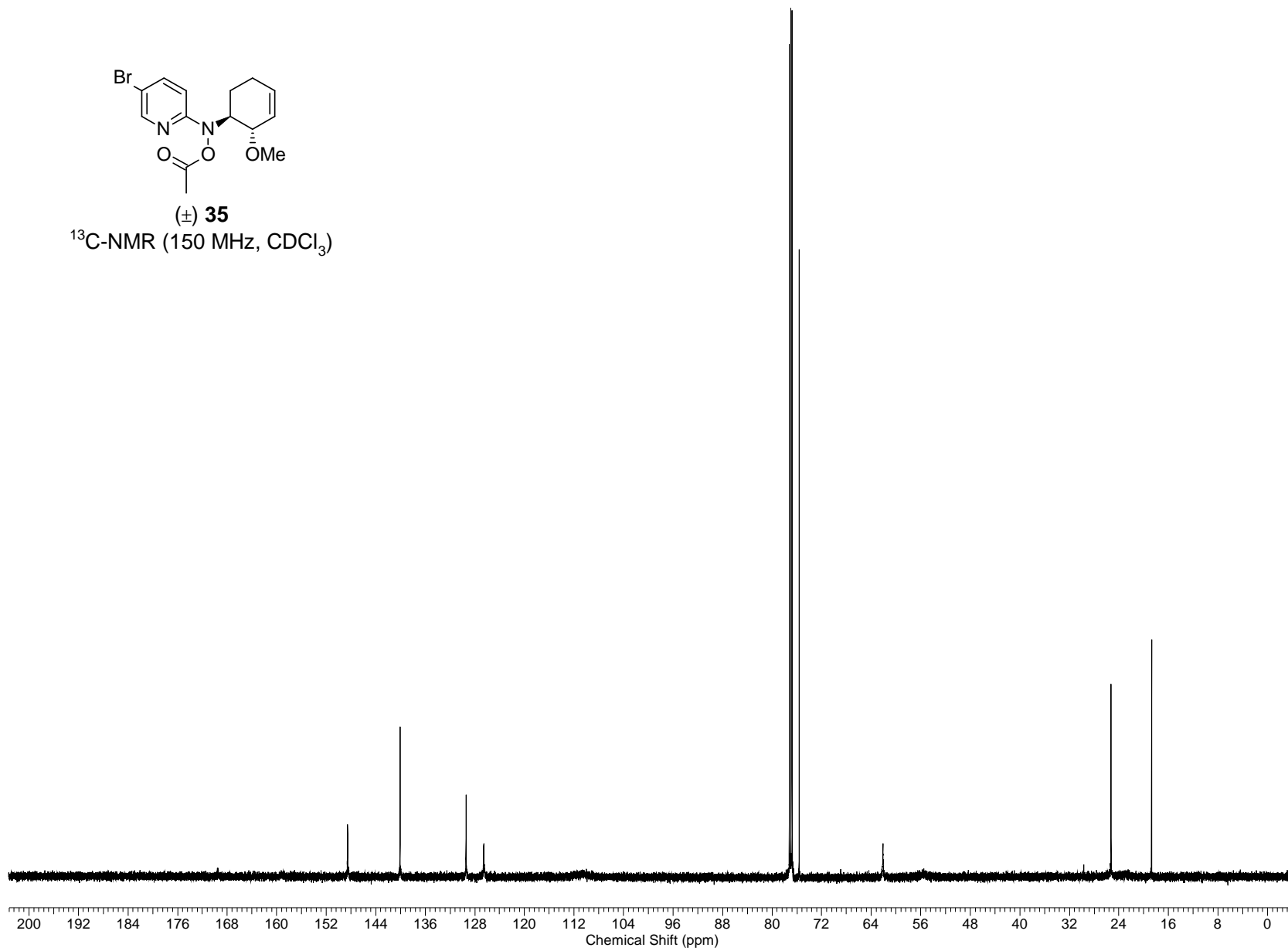
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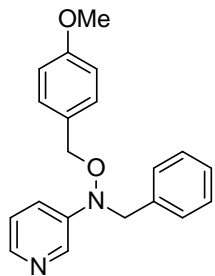




(±) **35**

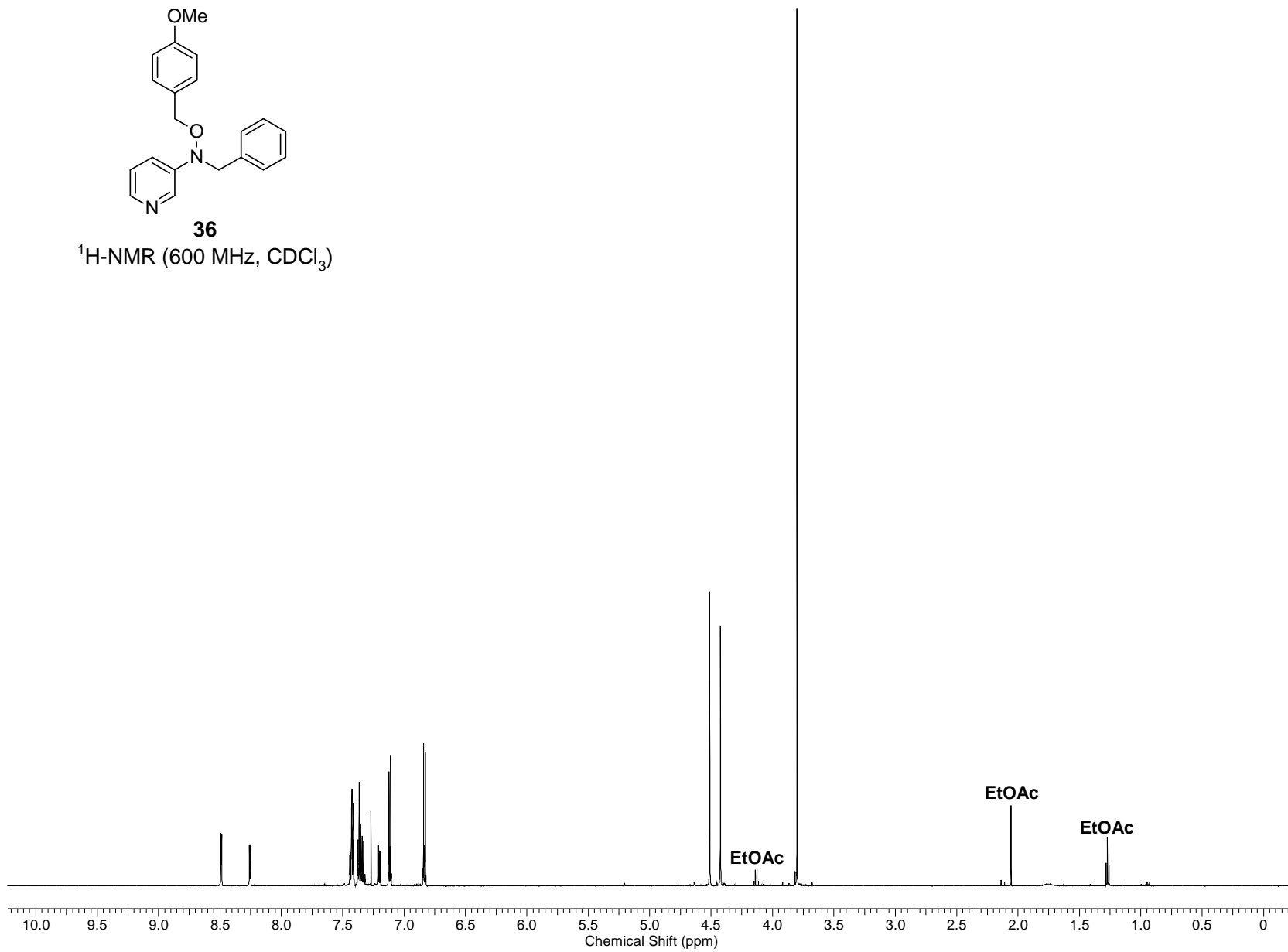
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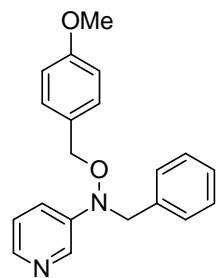


**36**

<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

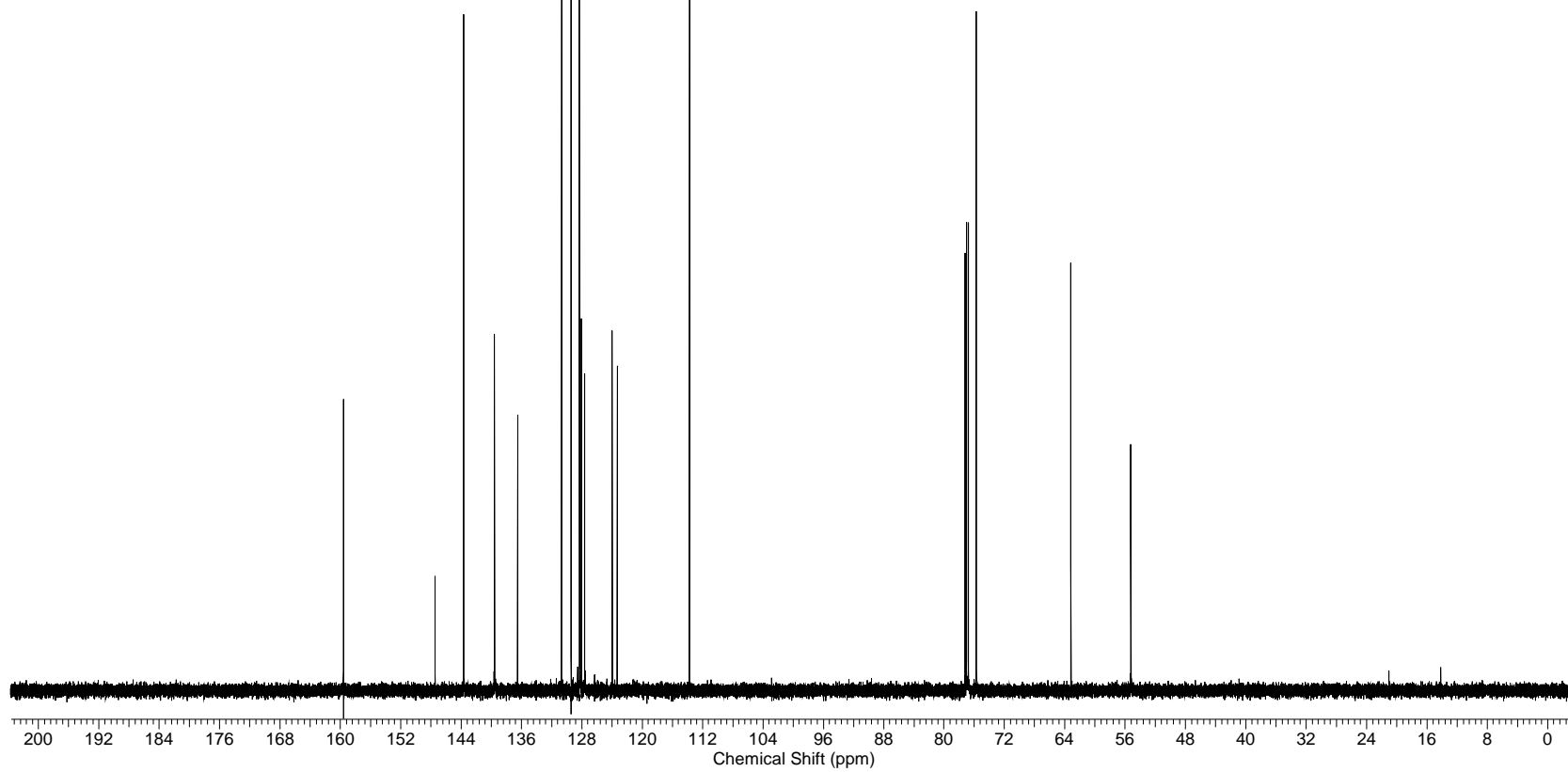


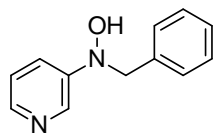




**36**

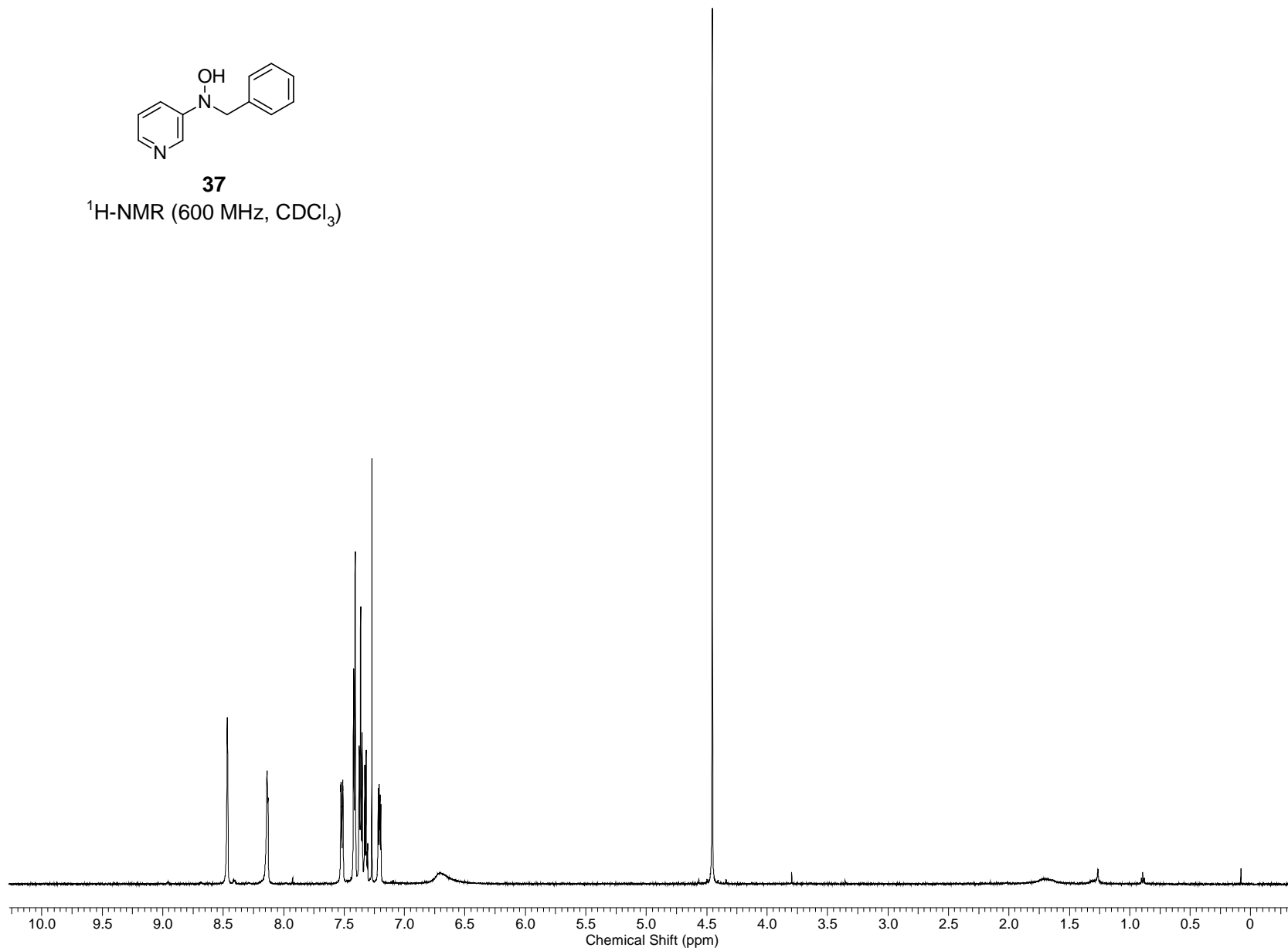
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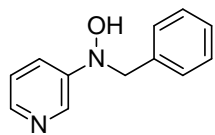




**37**

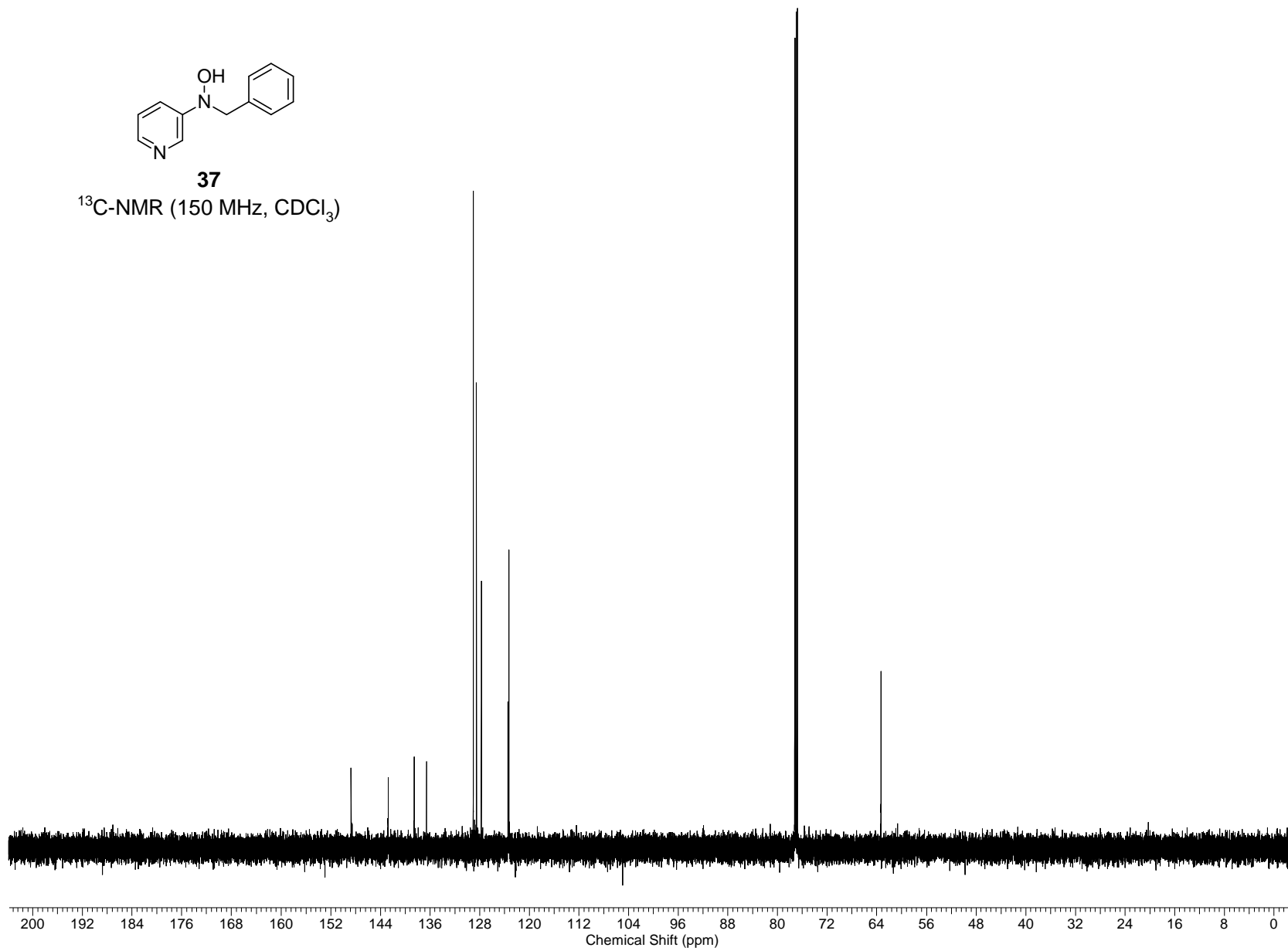
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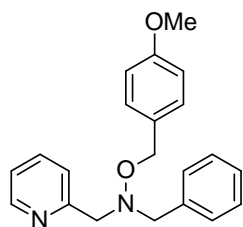




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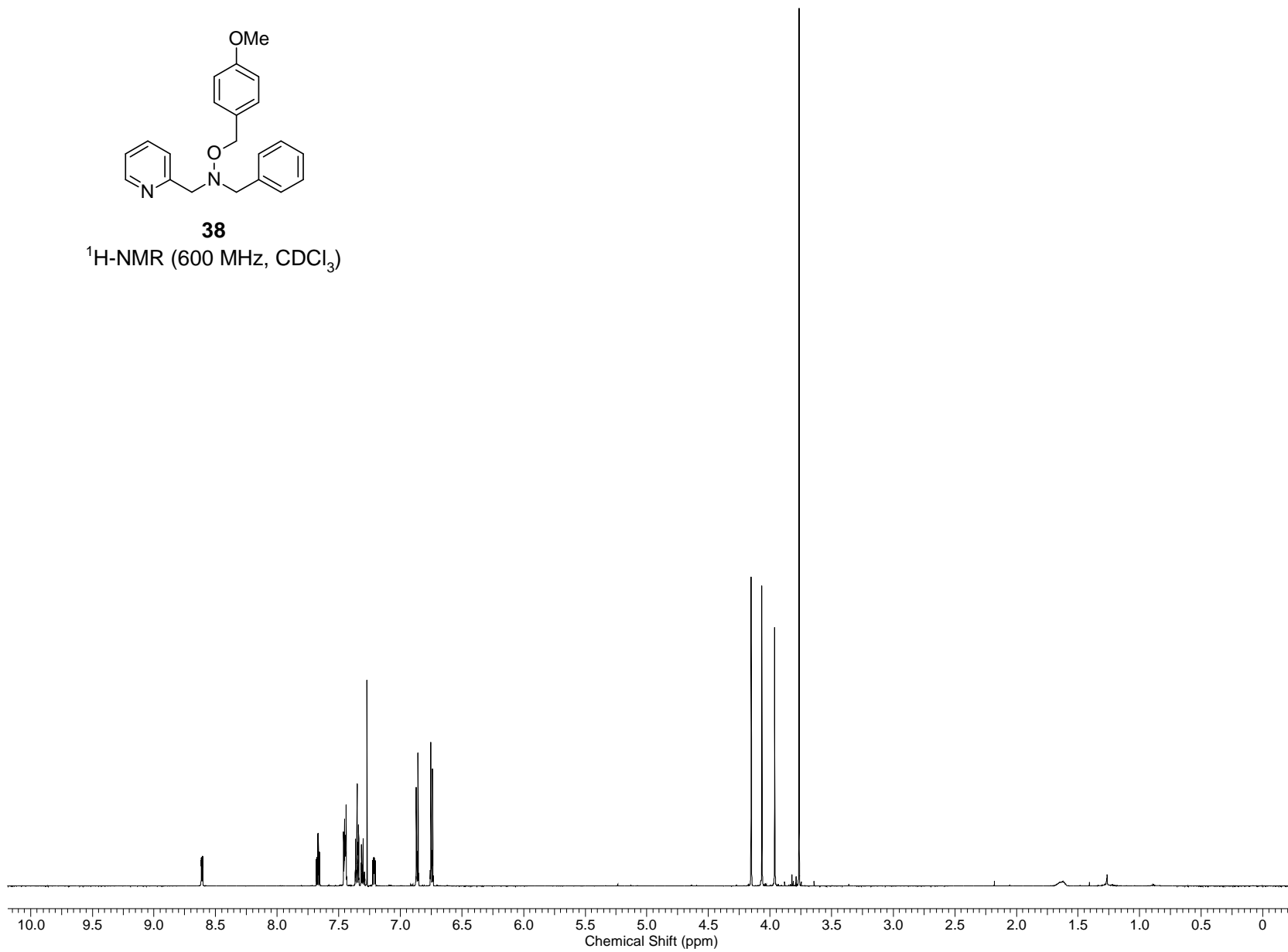
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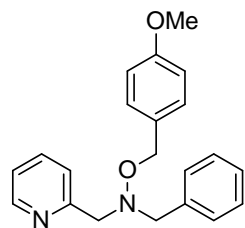




**38**

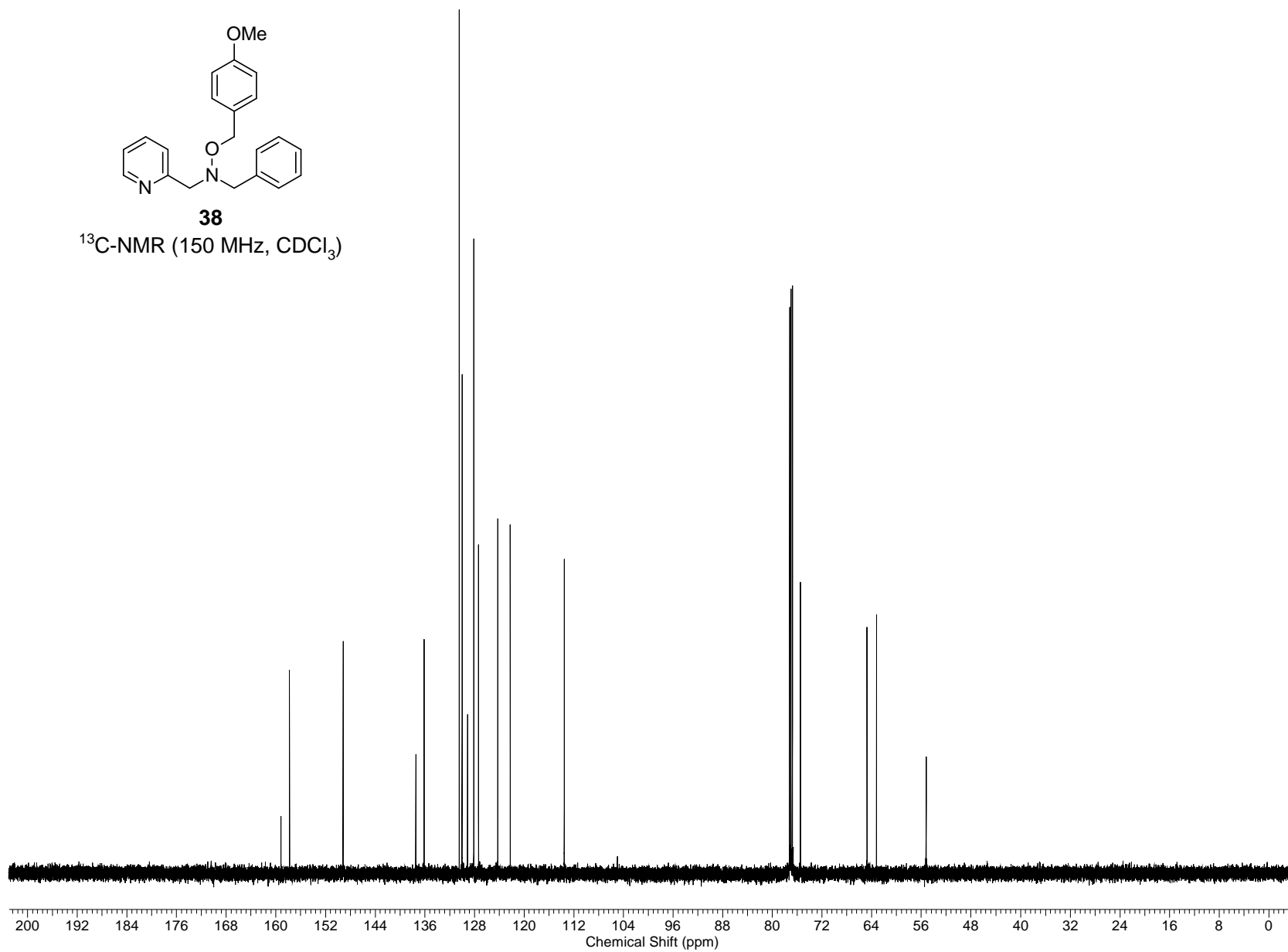
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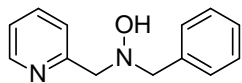




**38**

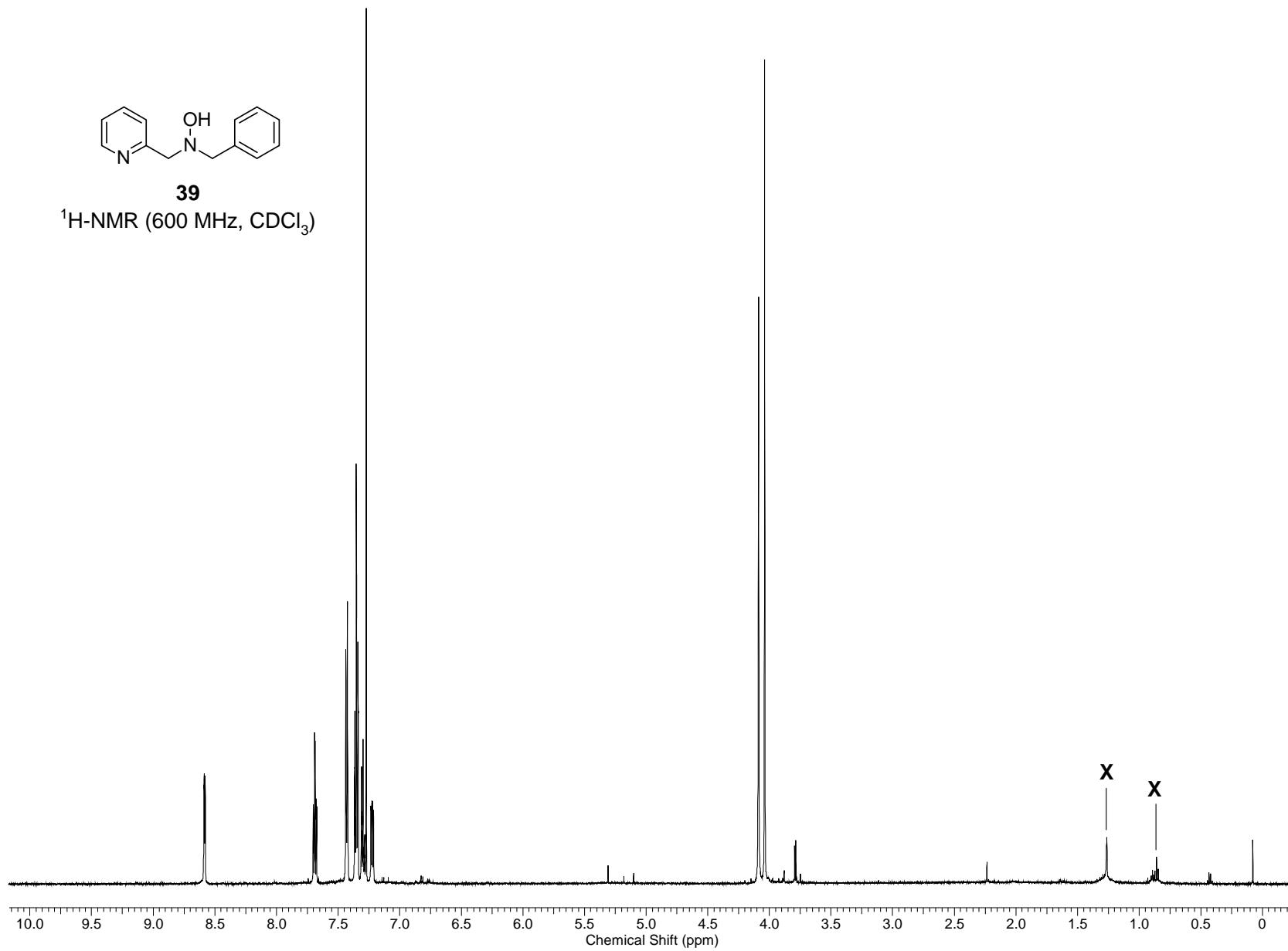
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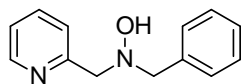




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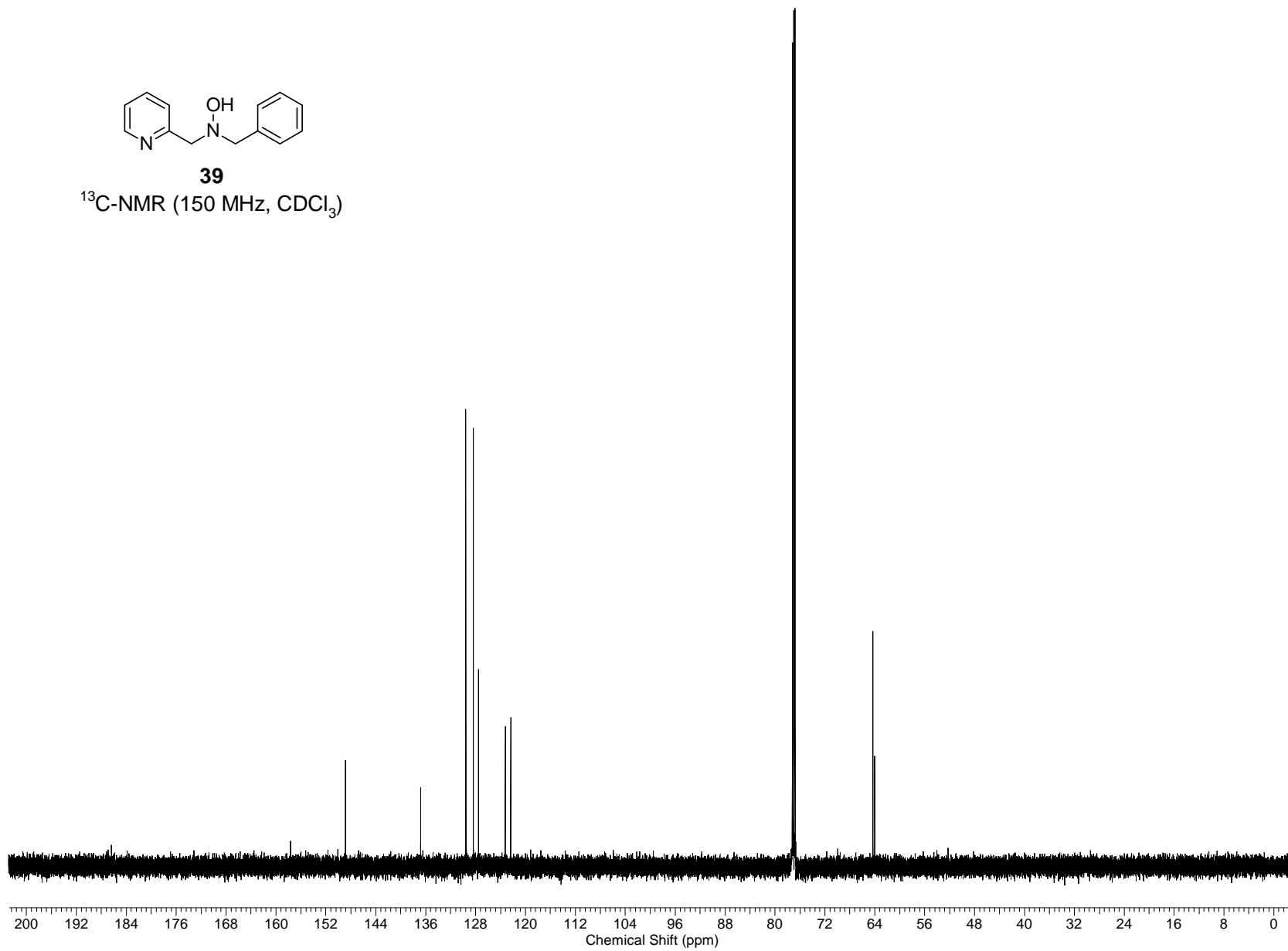
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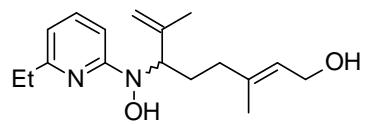




**39**

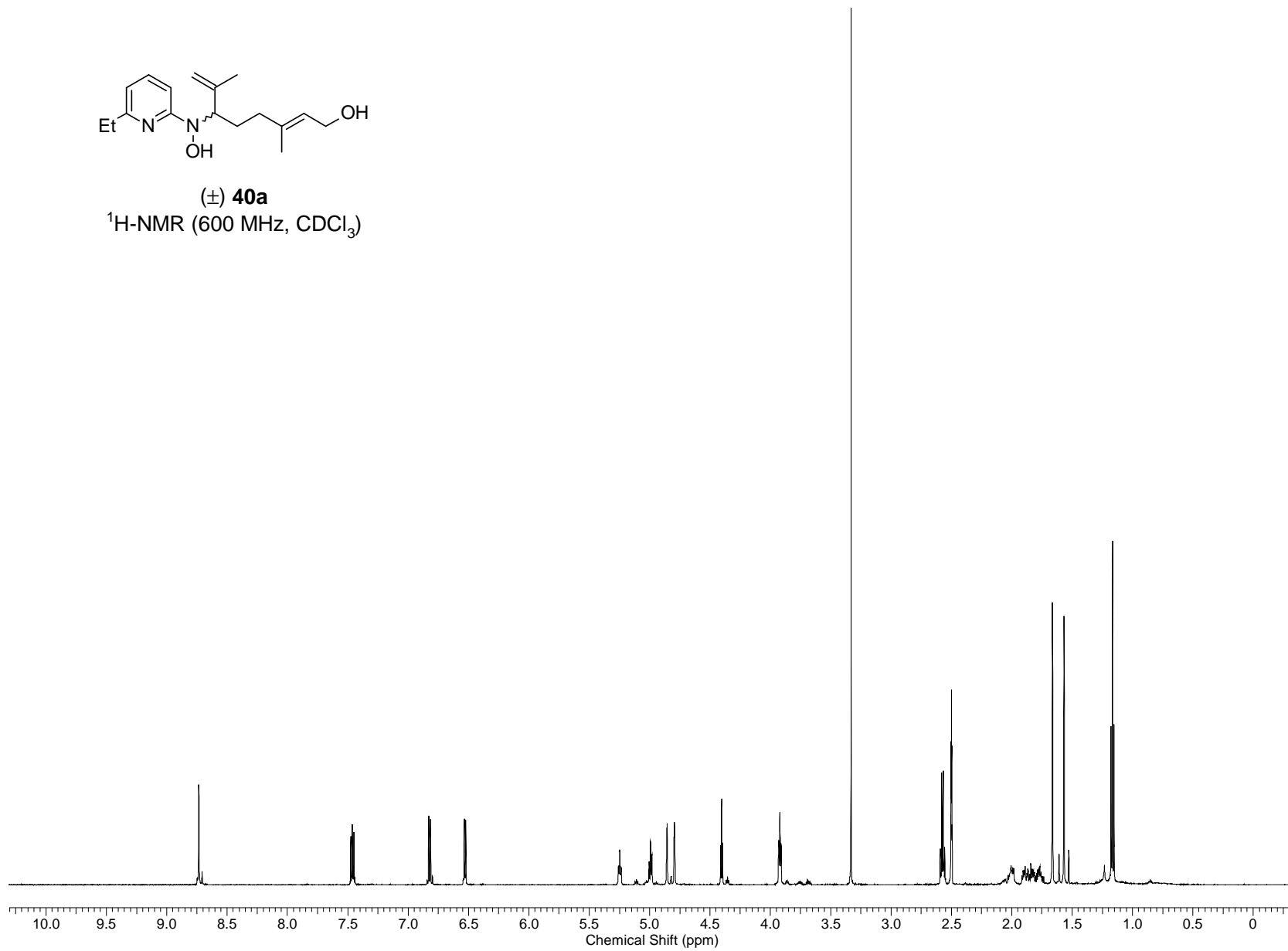
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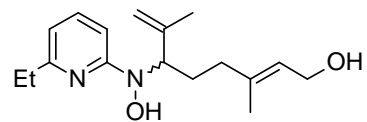


(±) **40a**

<sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)

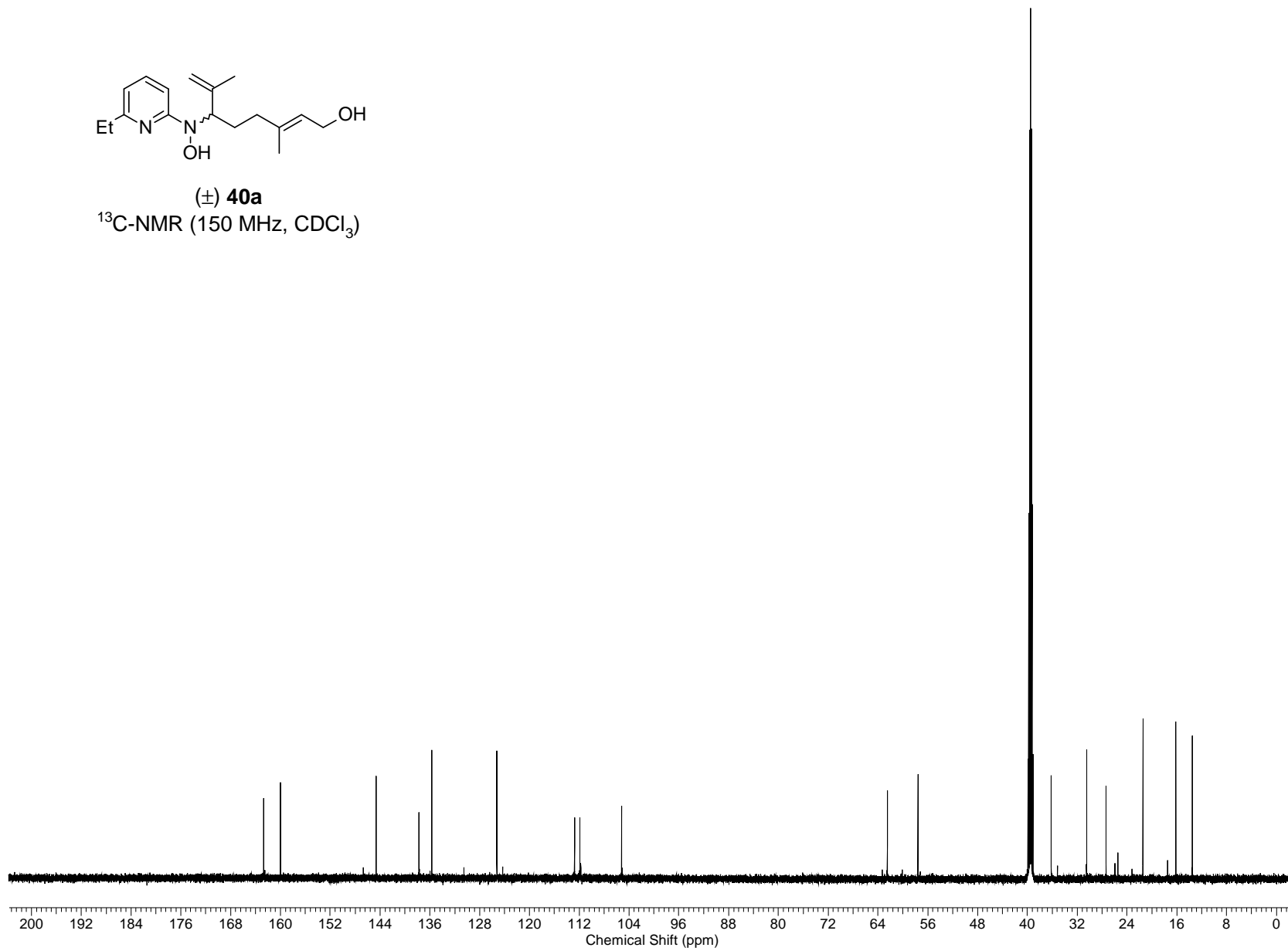




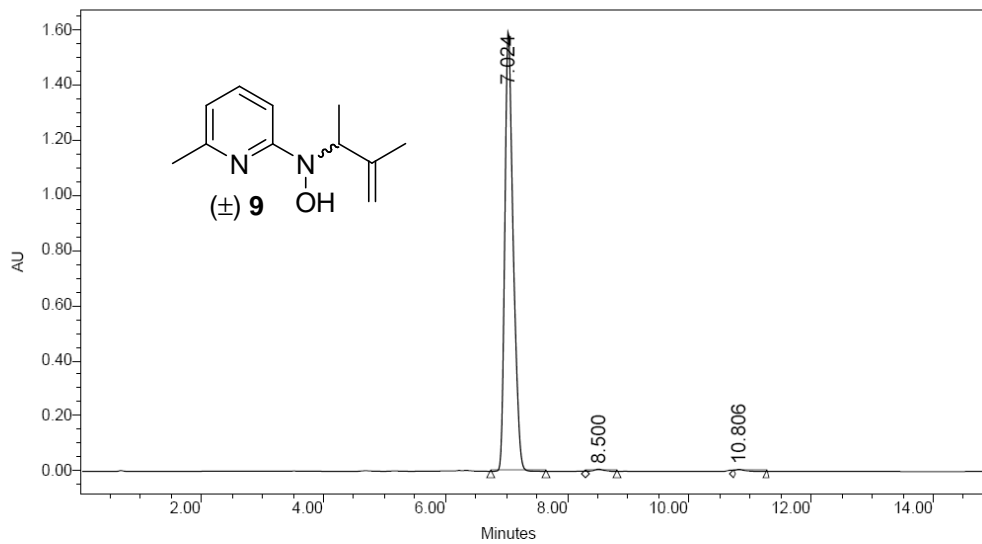
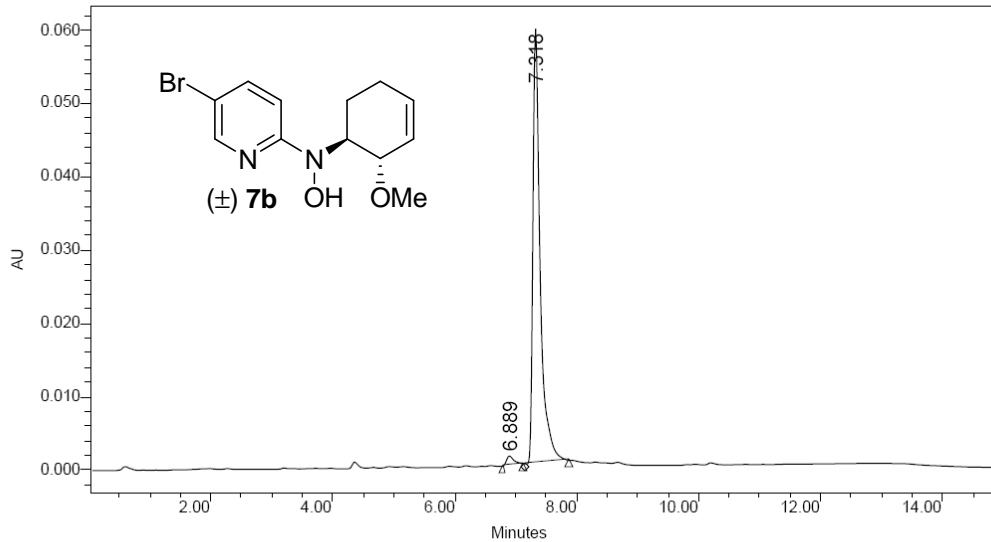
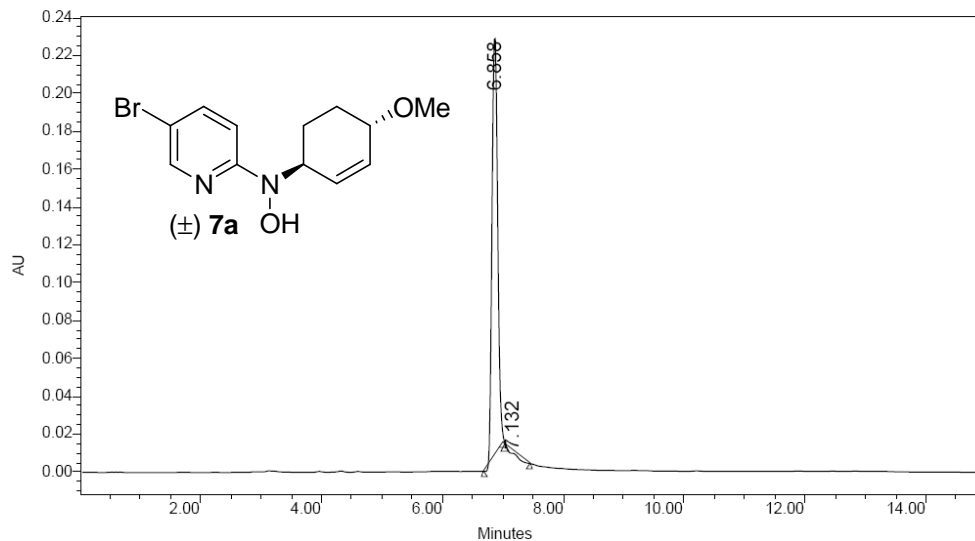


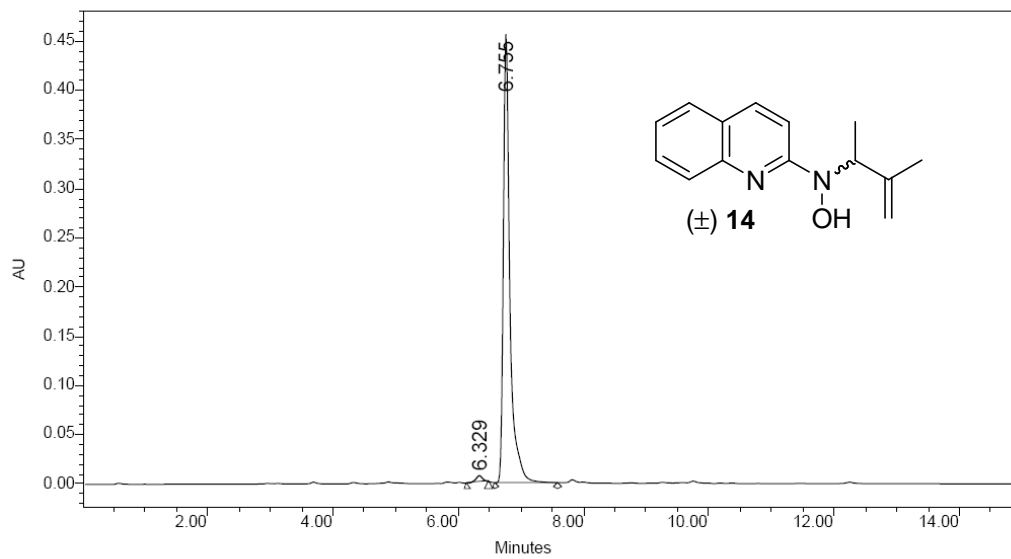
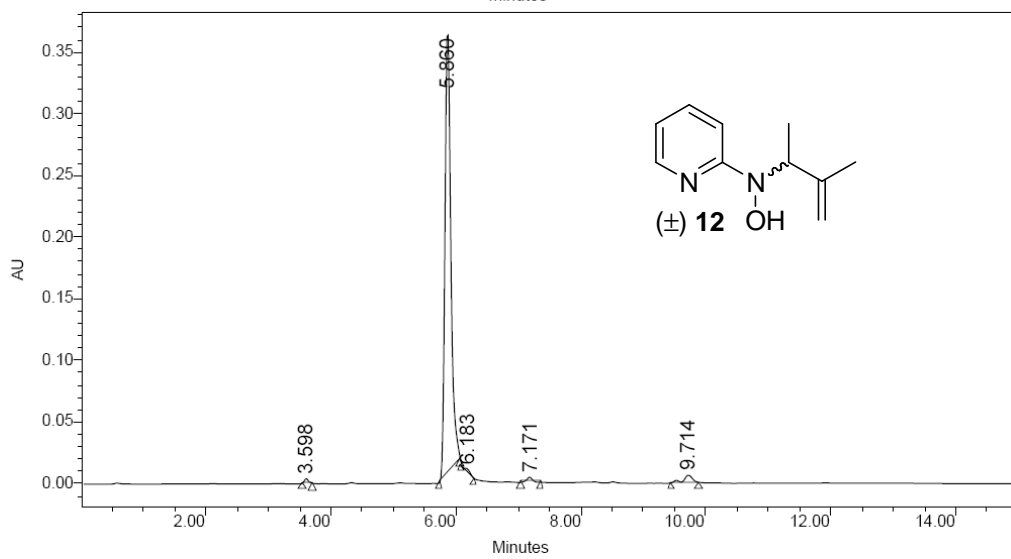
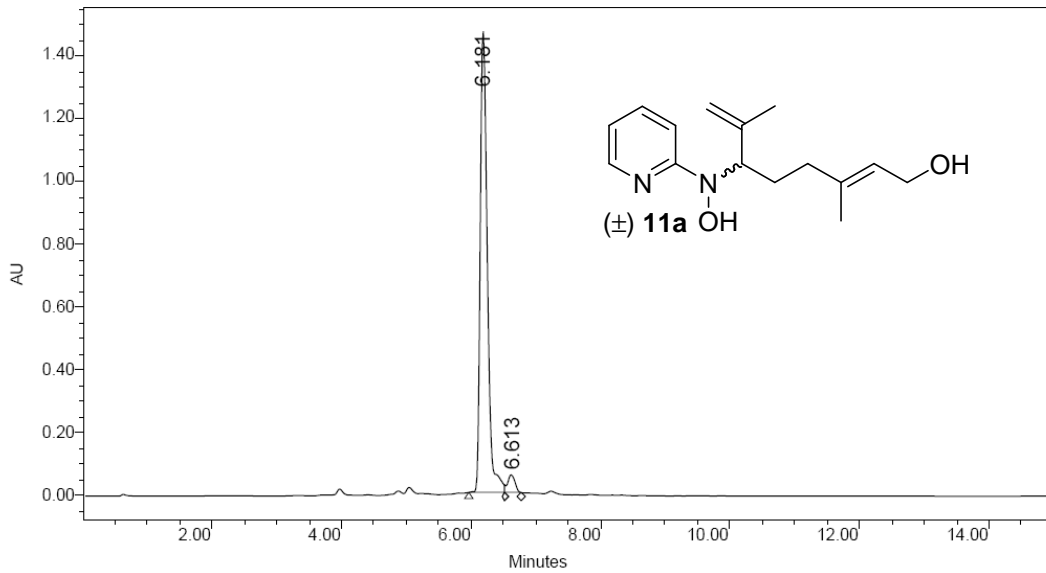
(±) **40a**

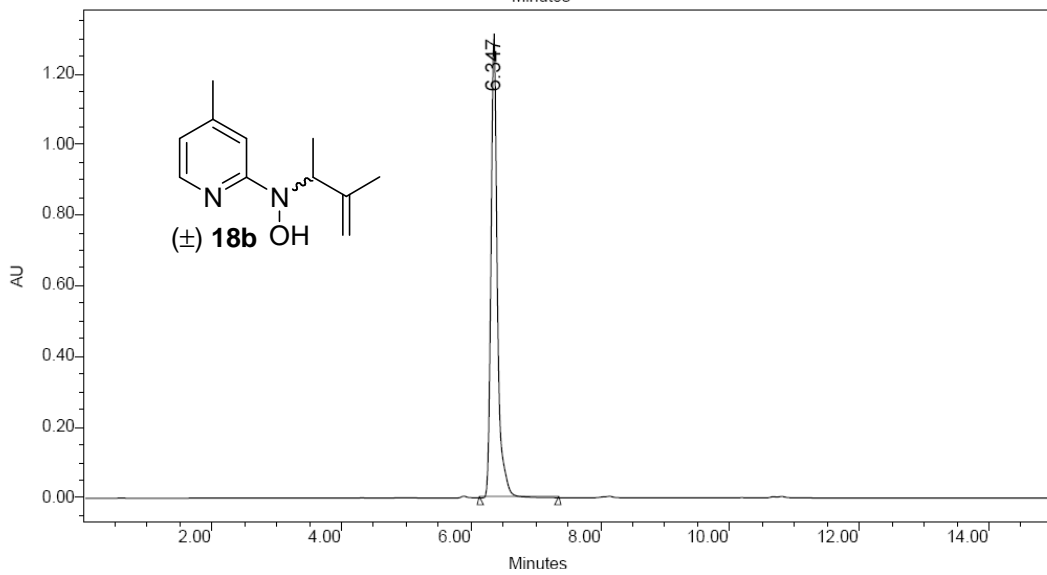
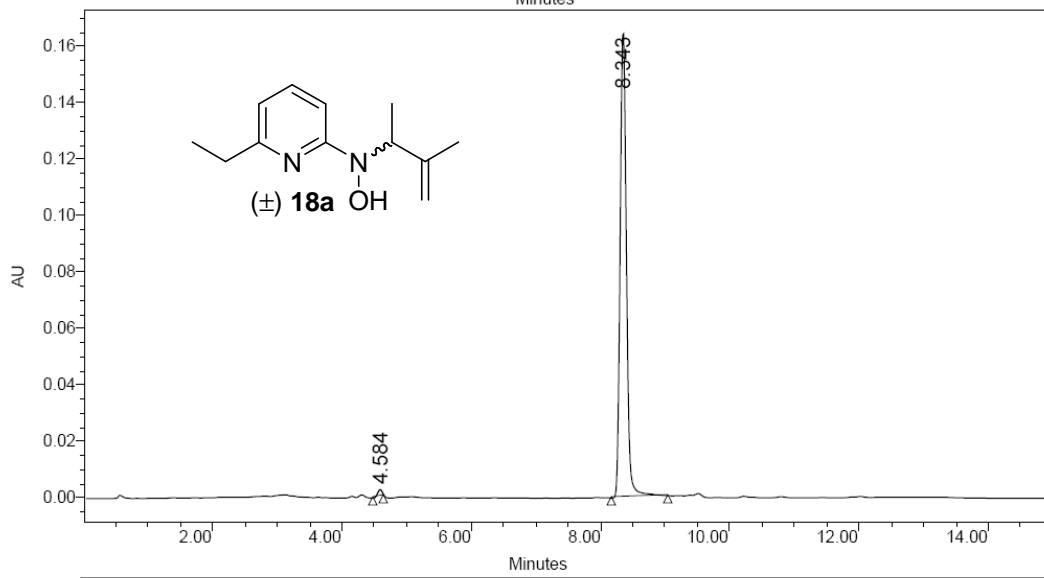
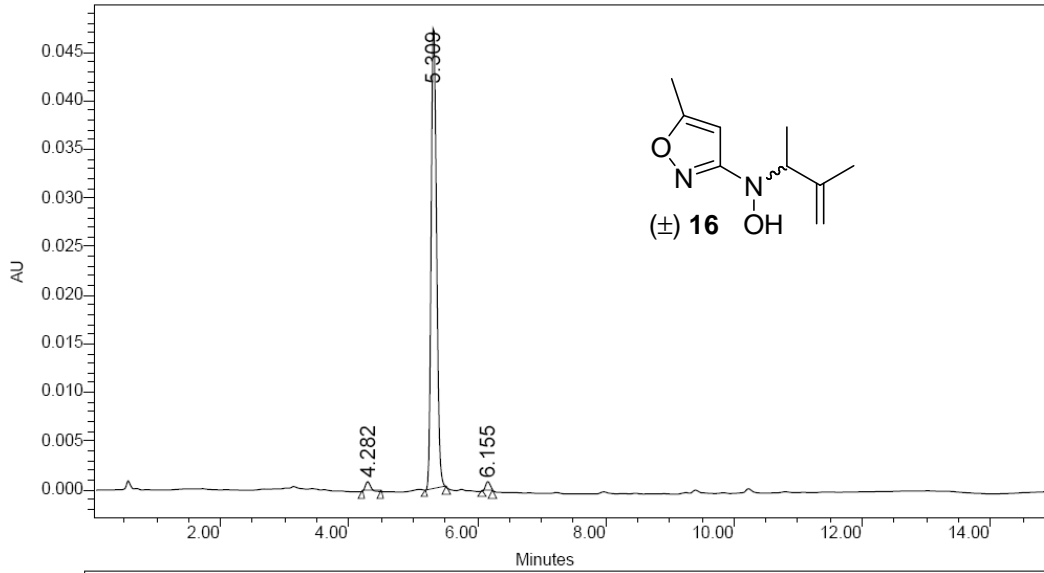
$^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )

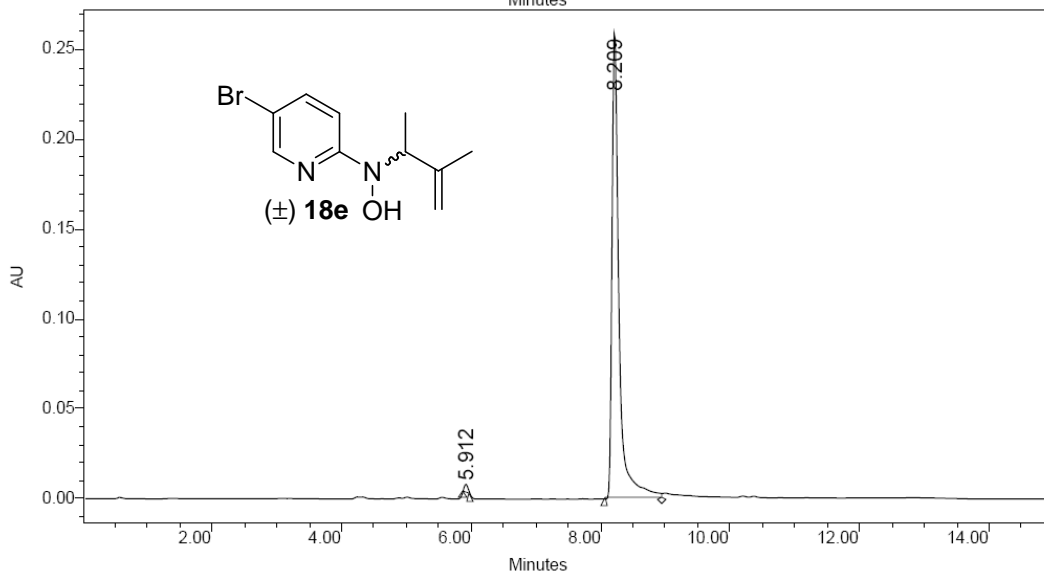
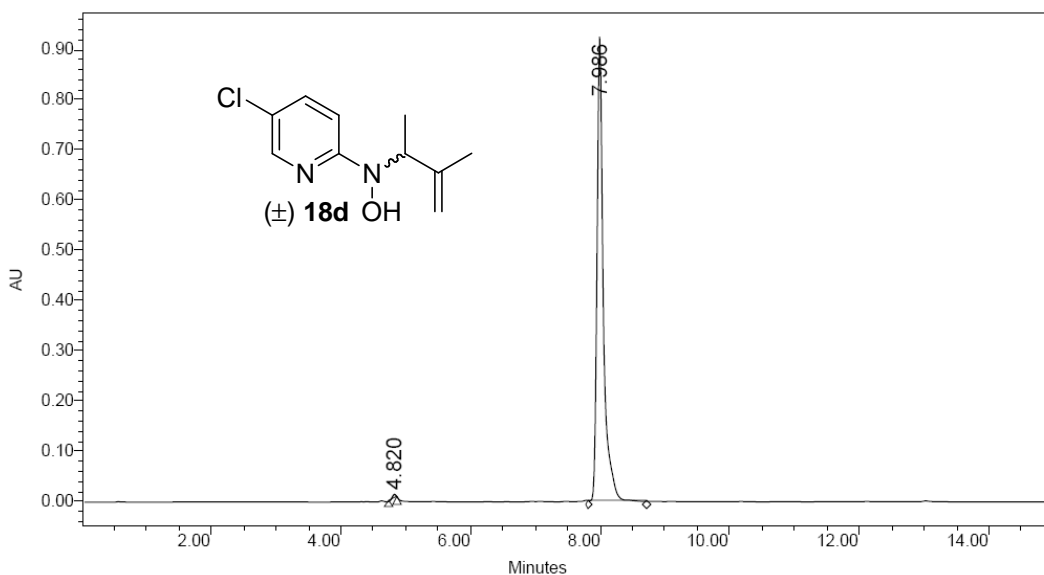
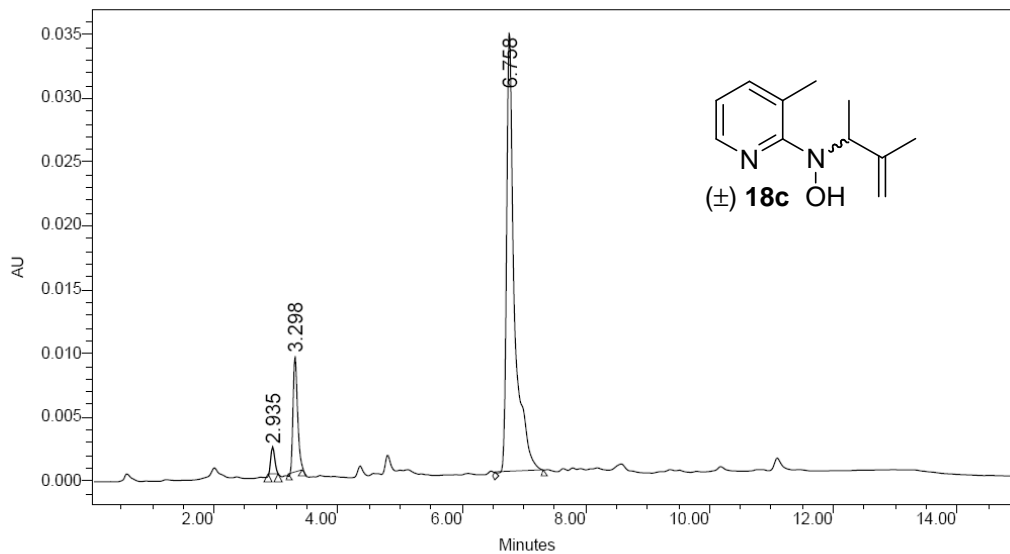


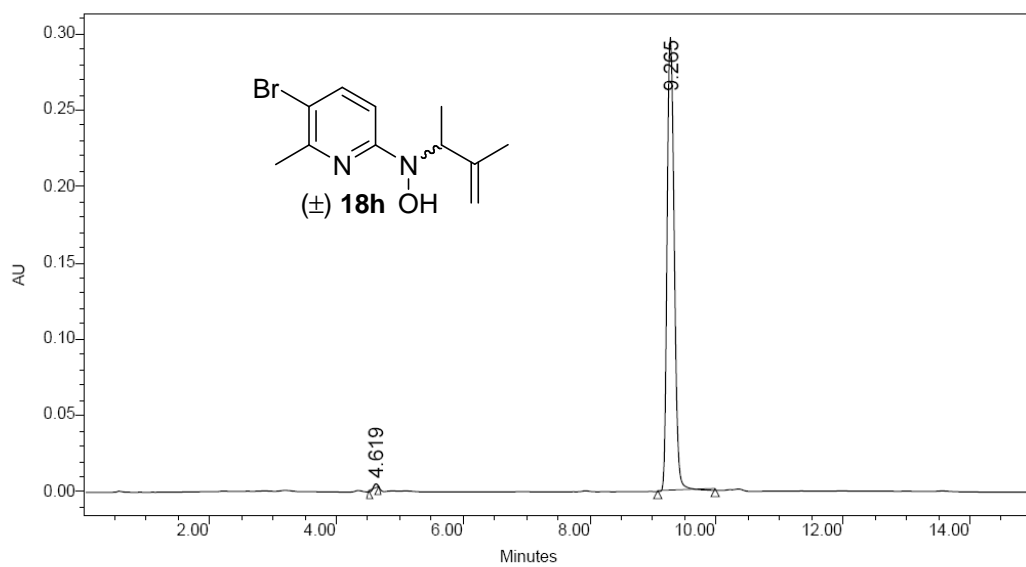
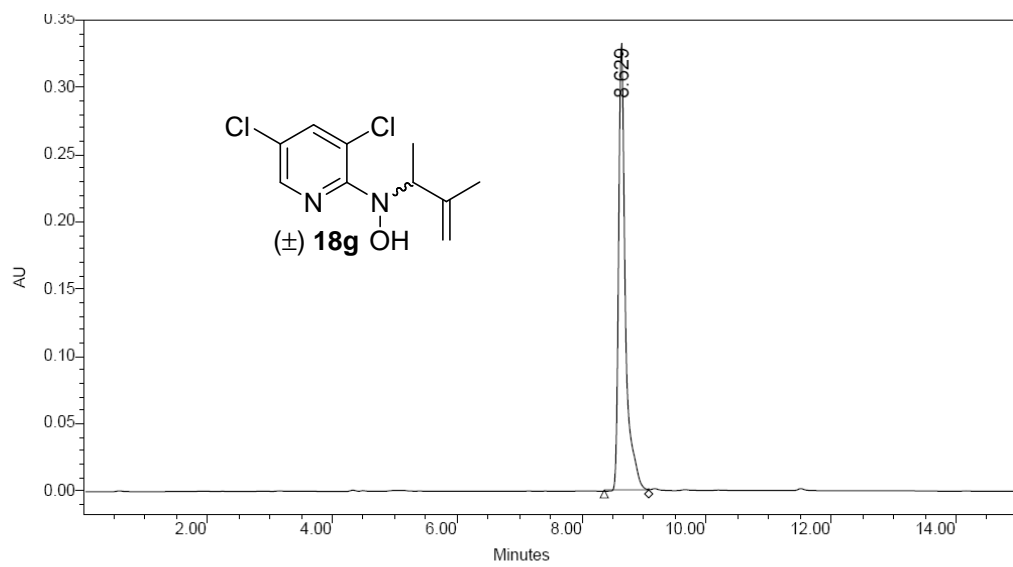
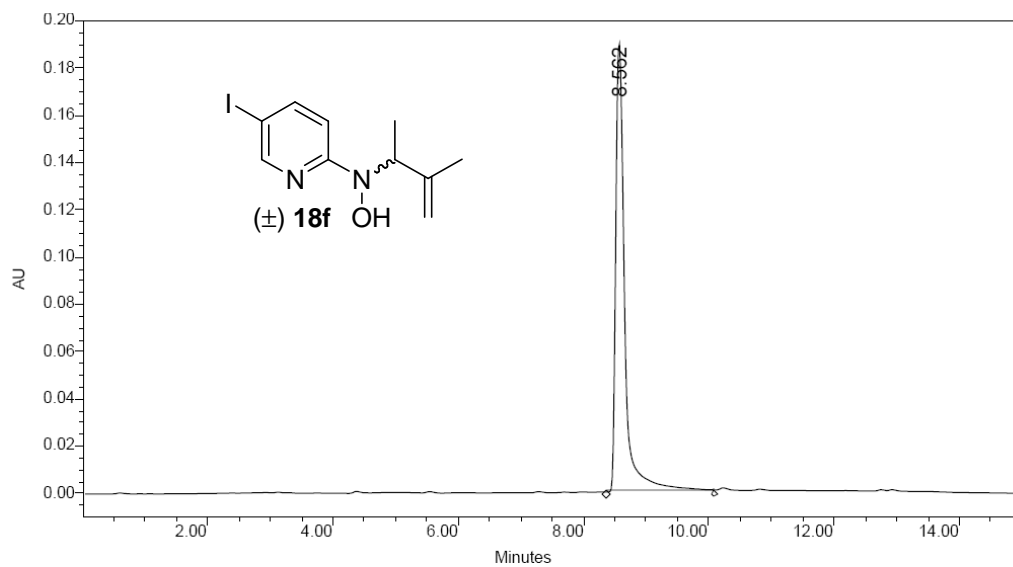
## VI. Copies of HPLC Chromatograms

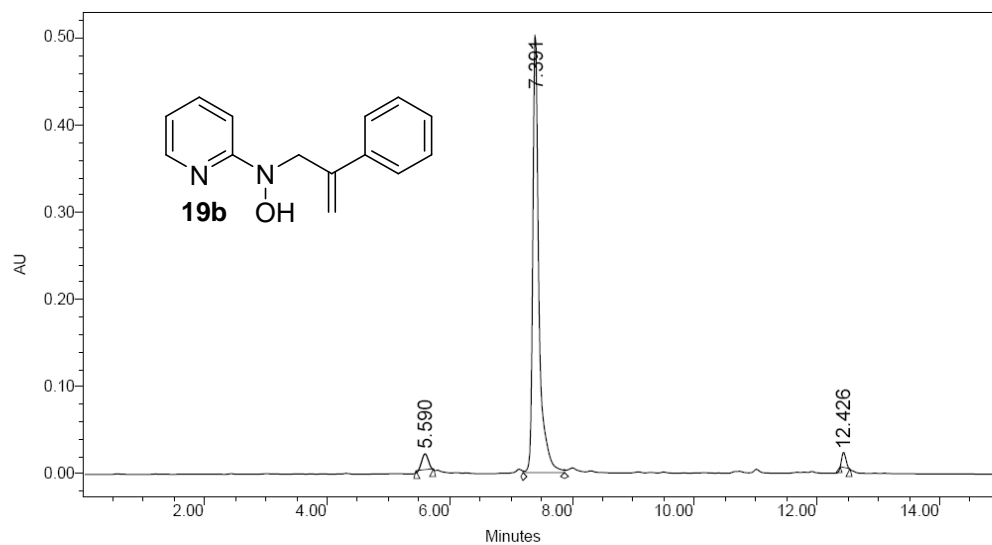
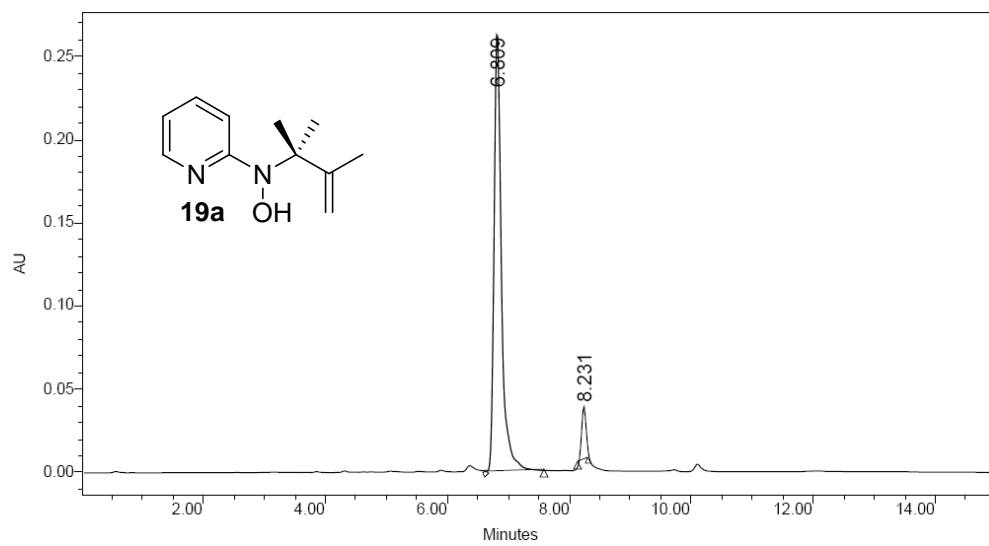
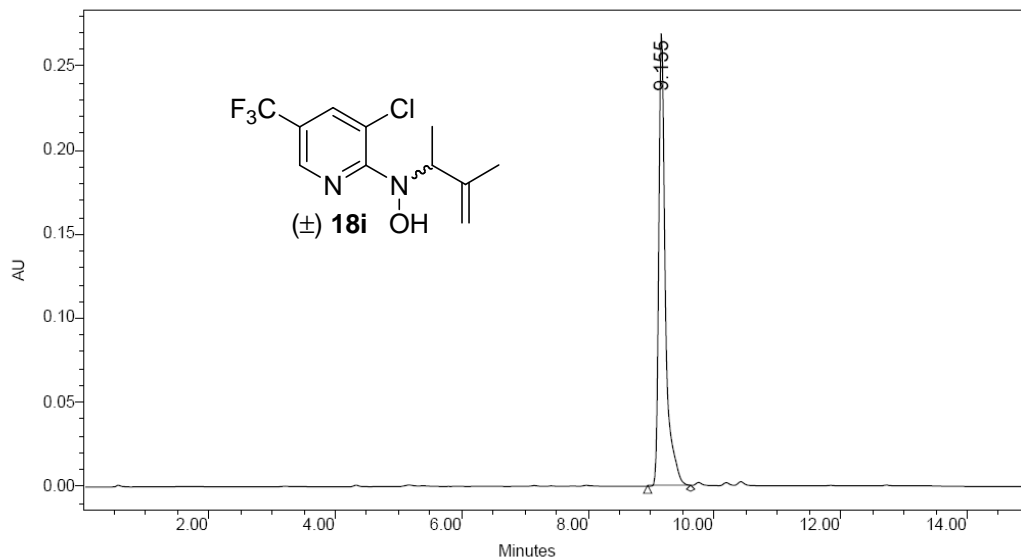


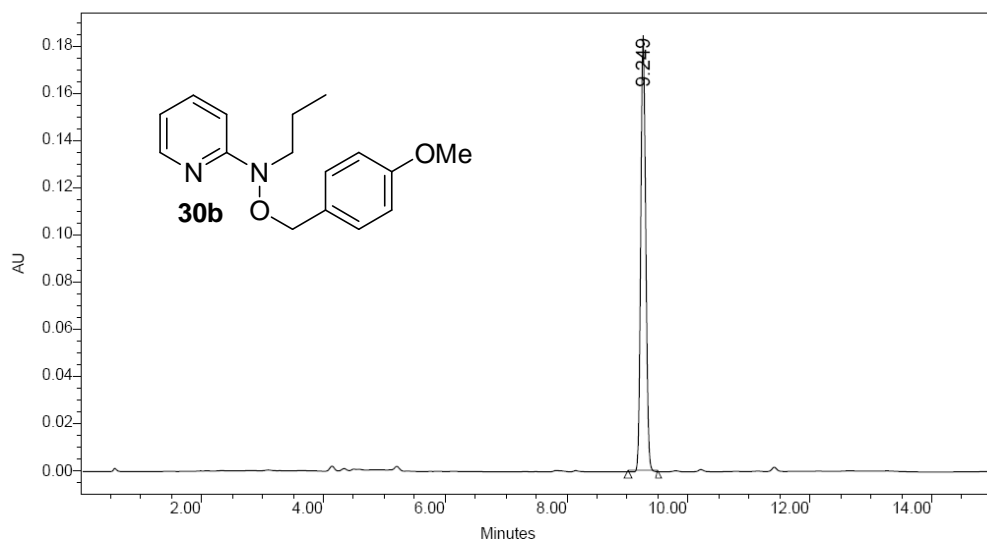
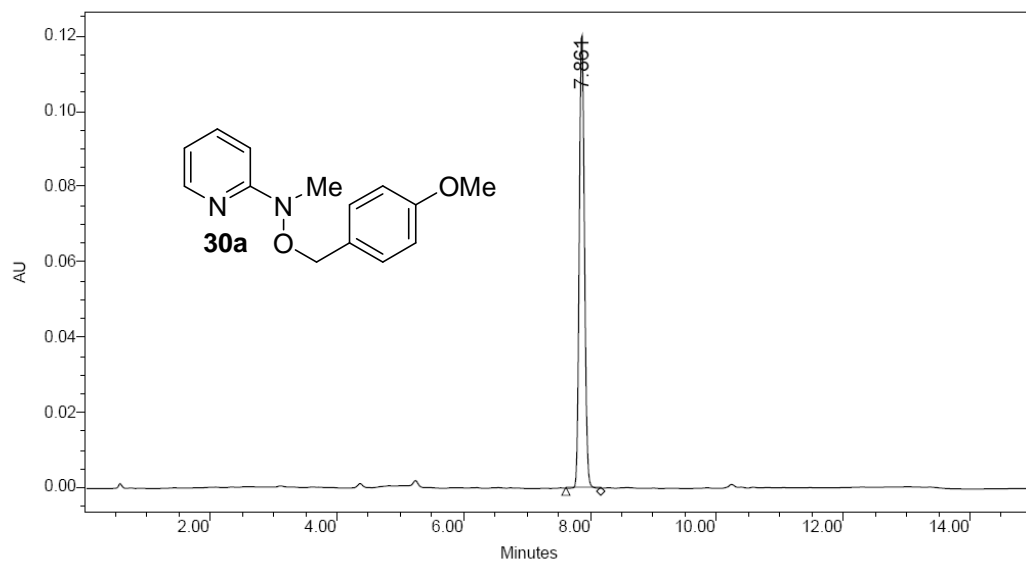
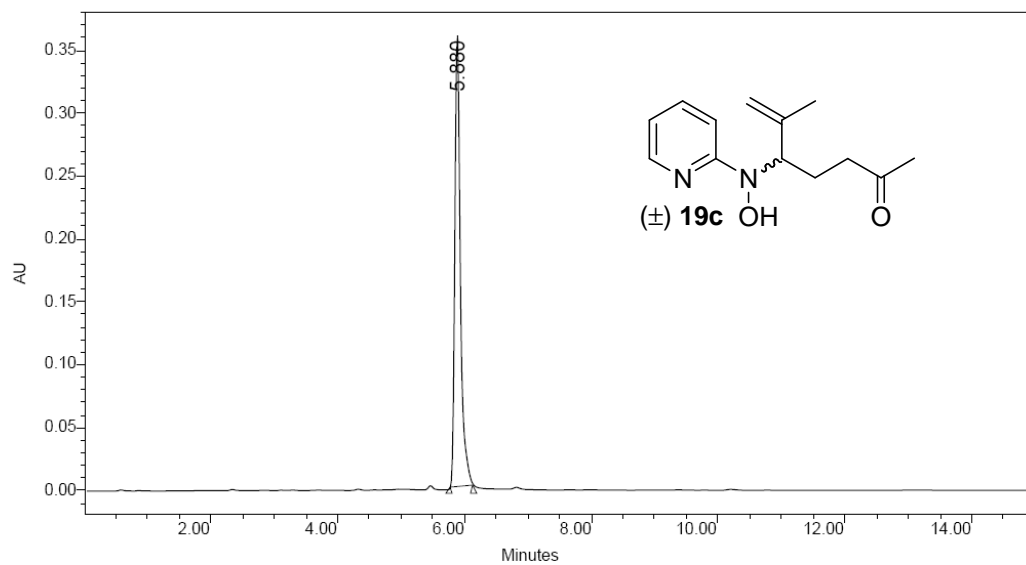




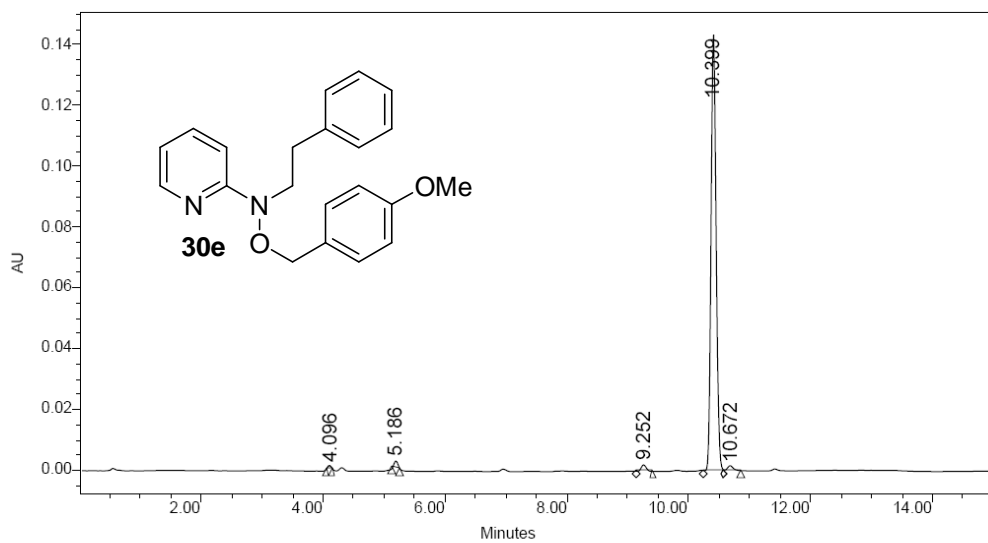
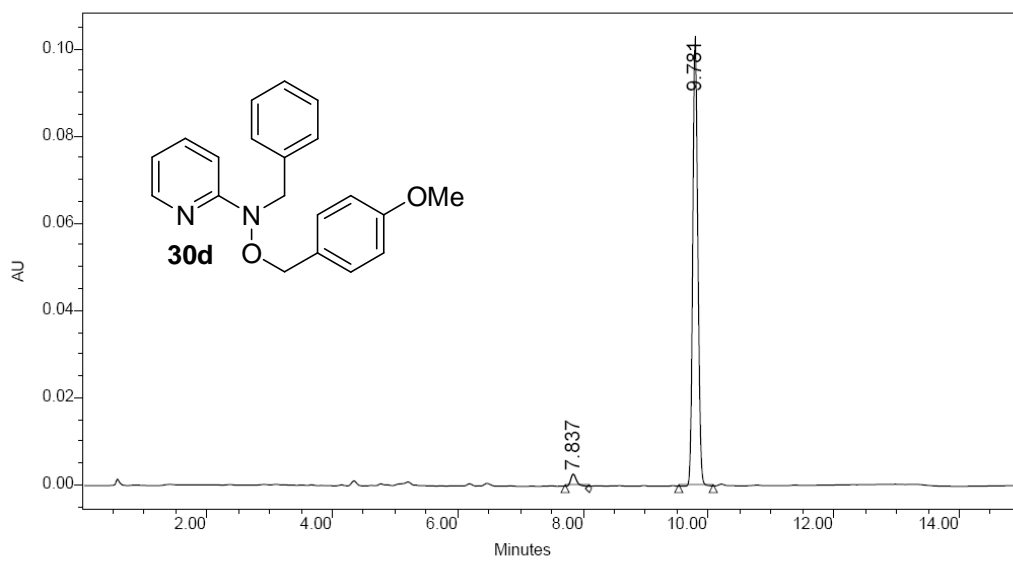
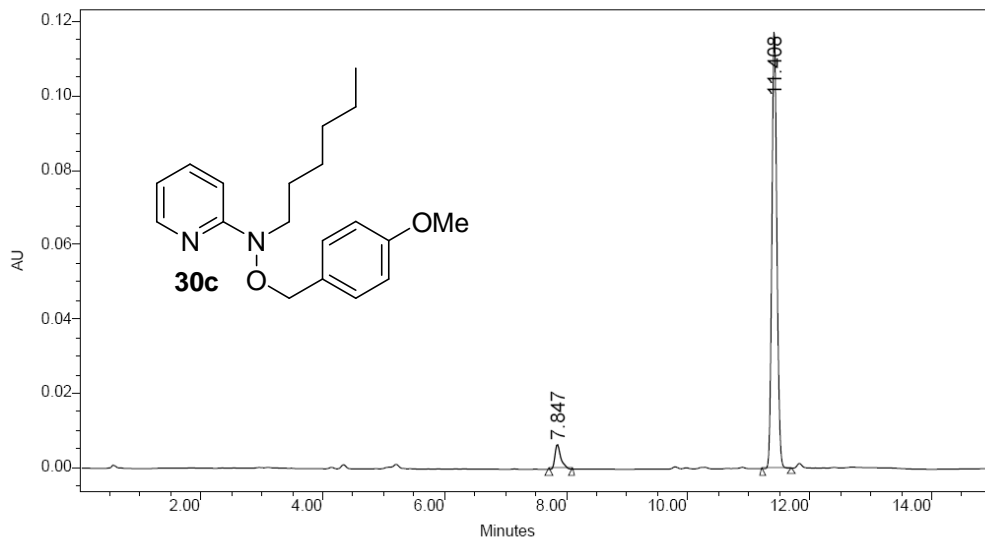


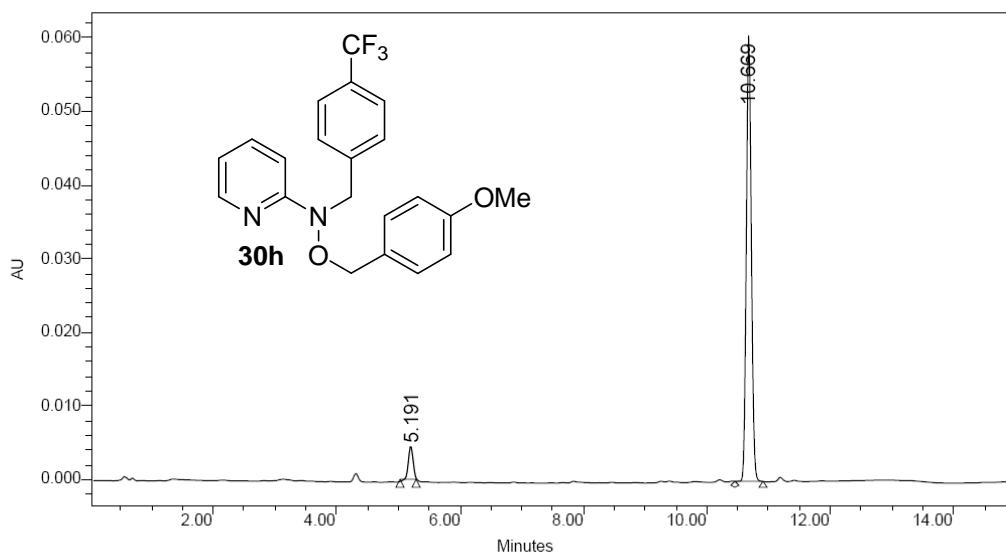
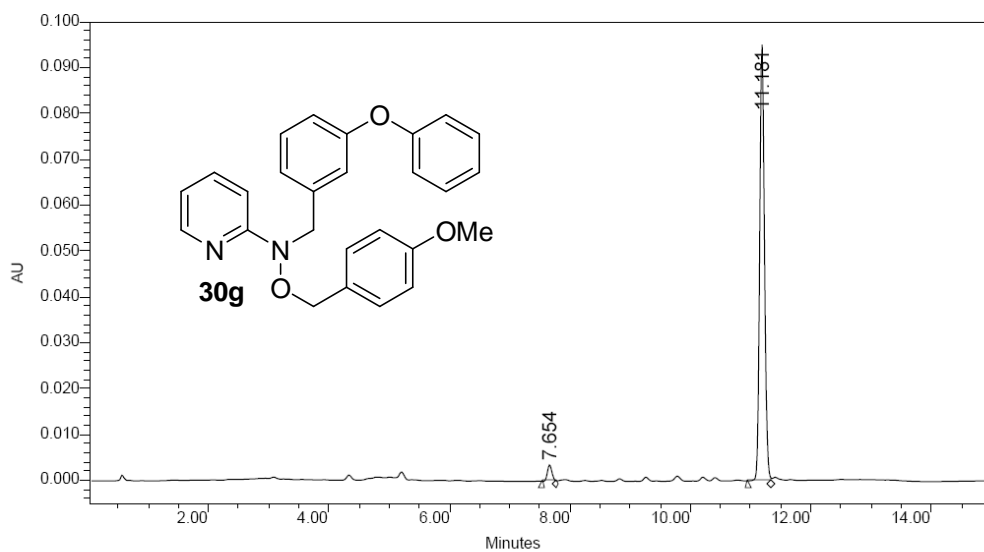
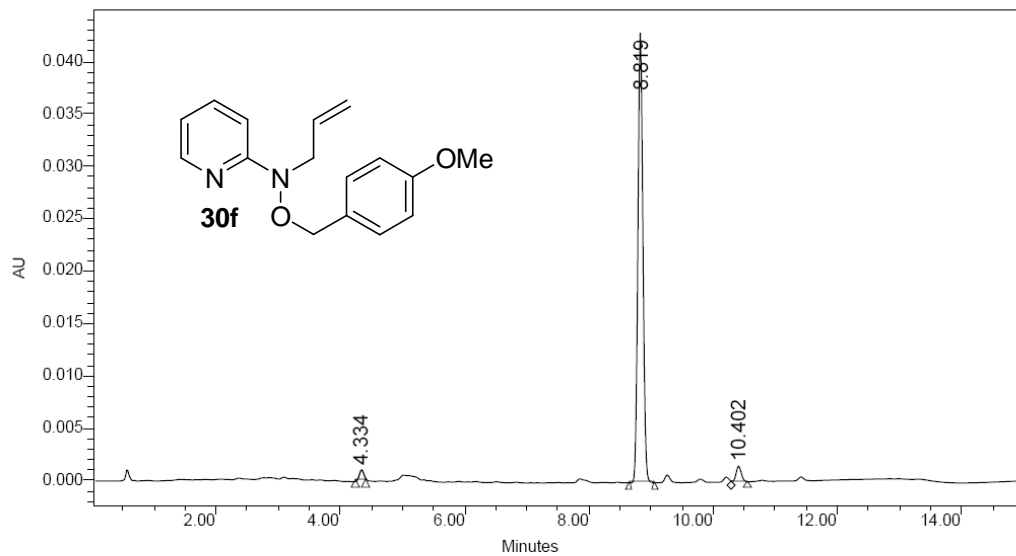


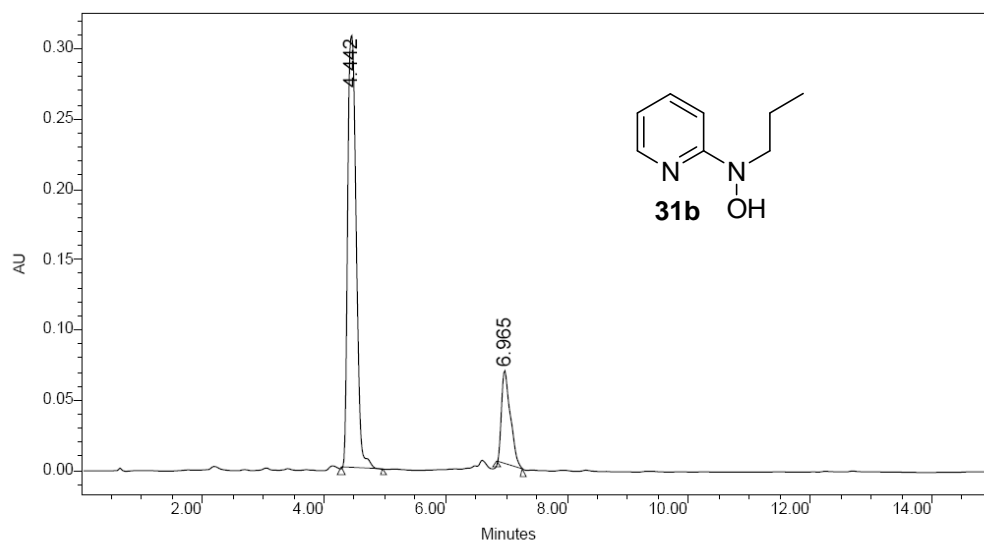
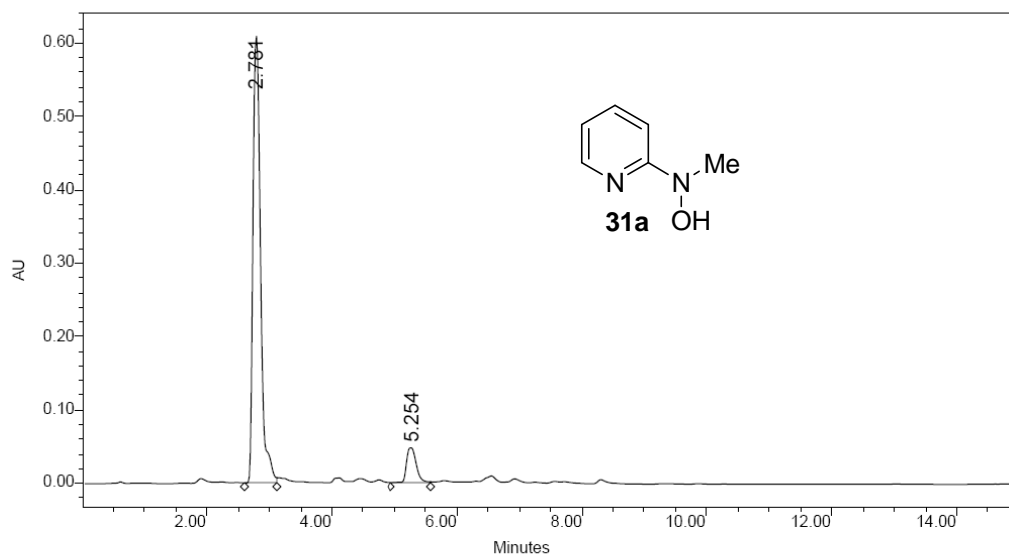
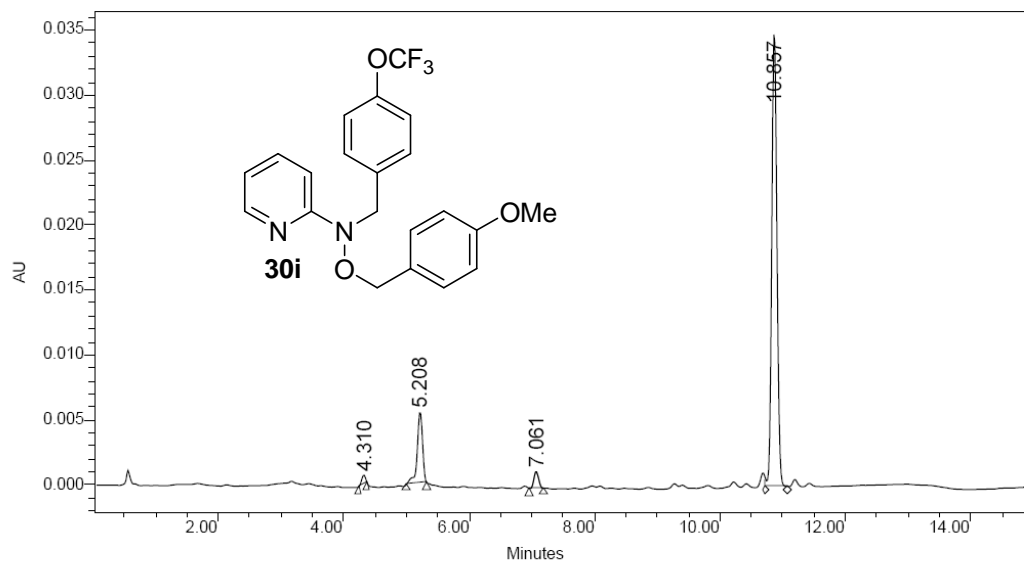


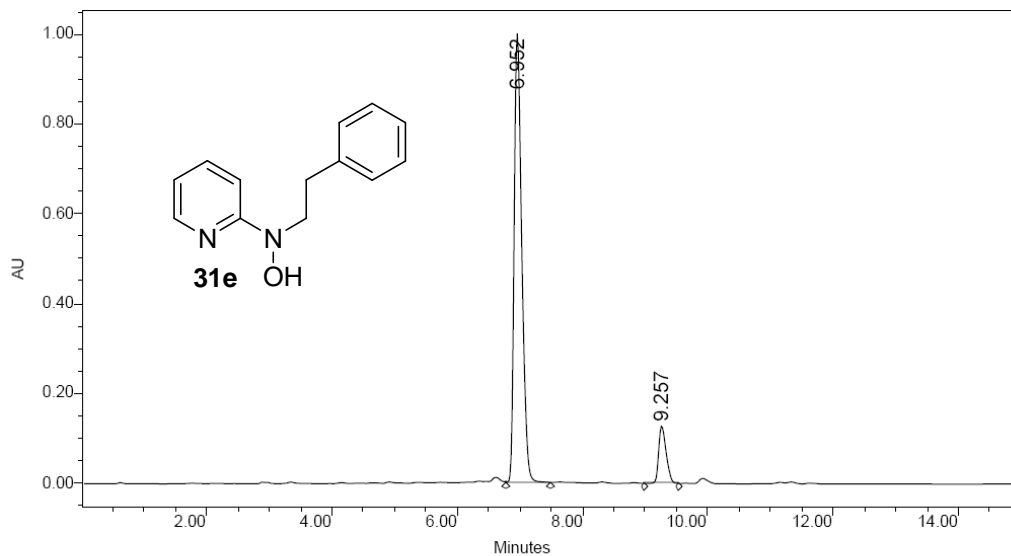
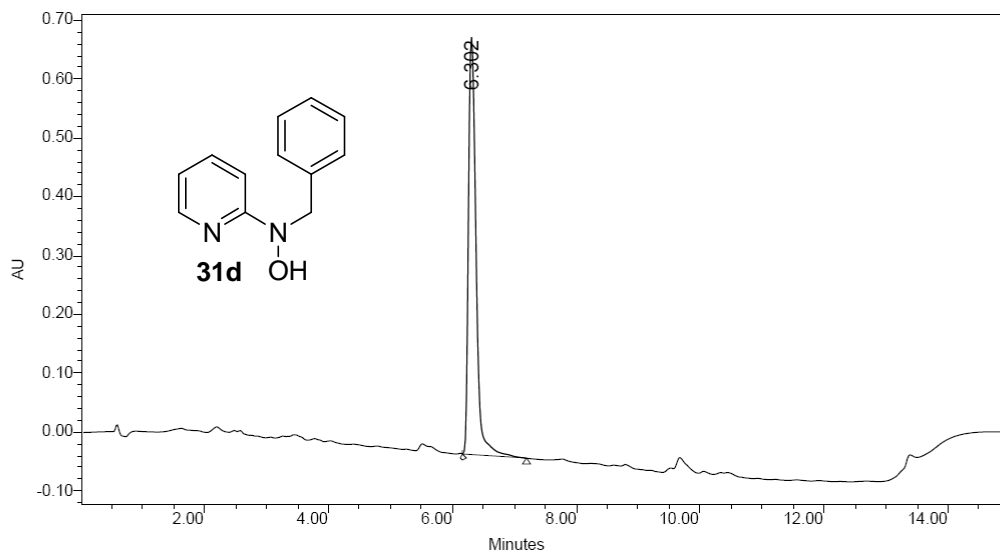
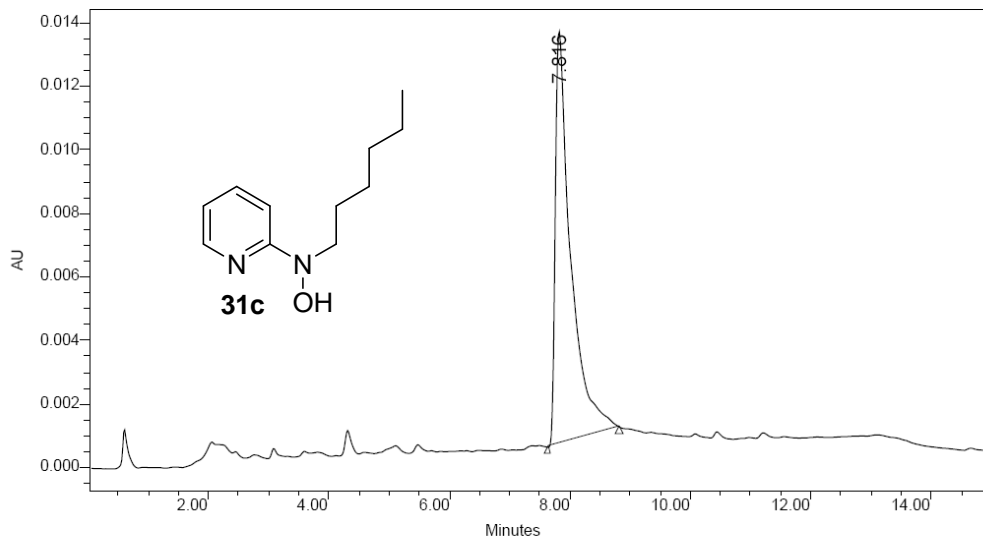


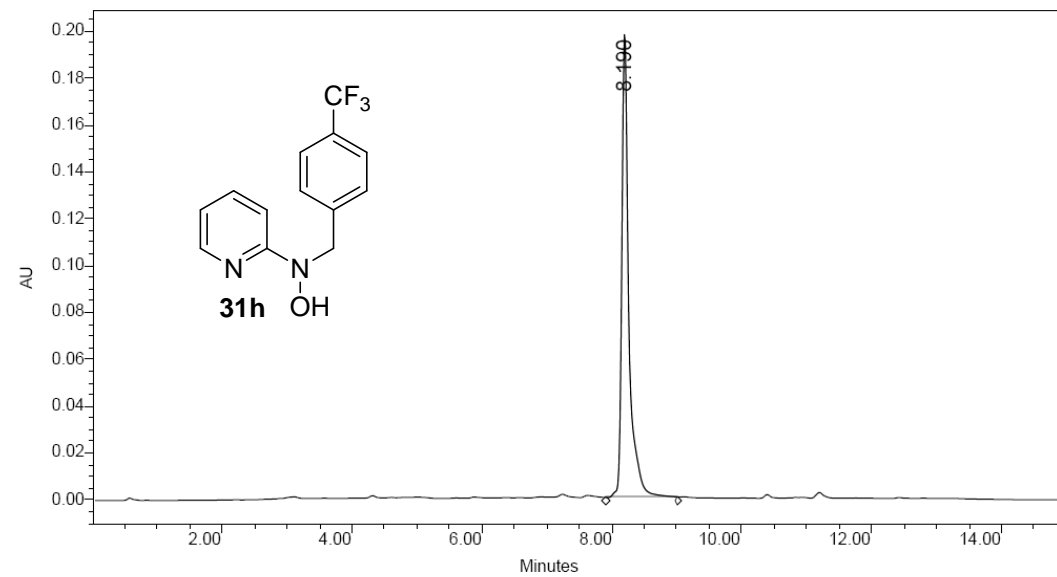
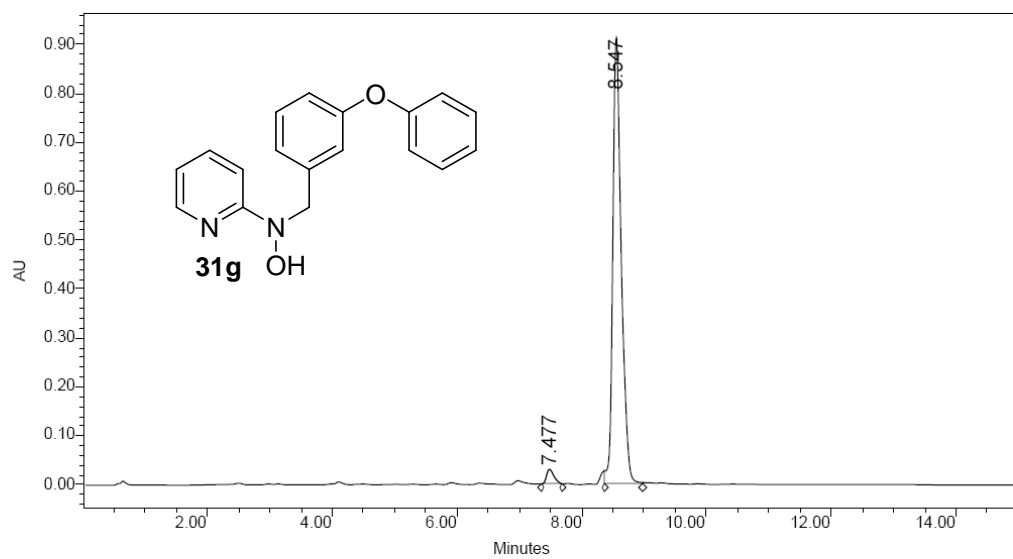
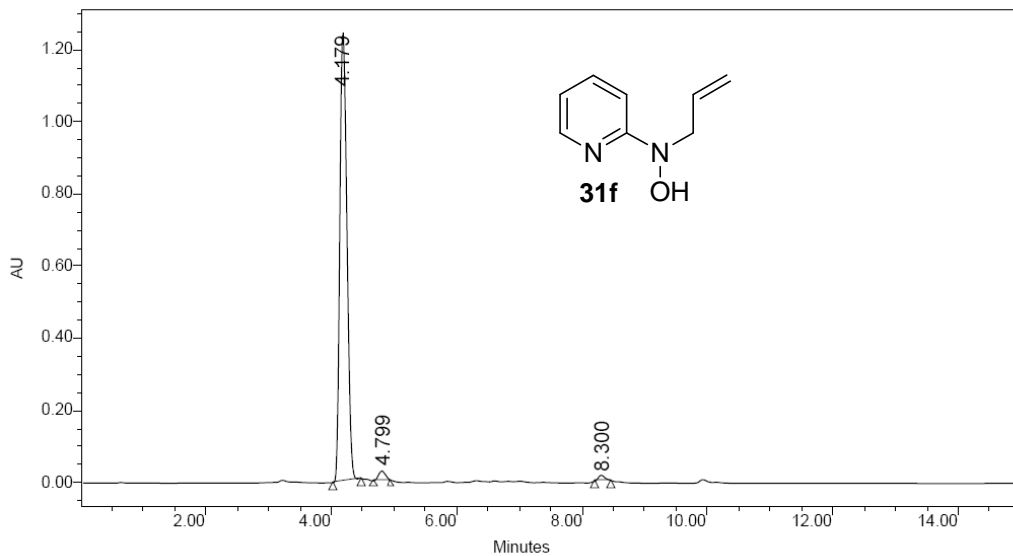


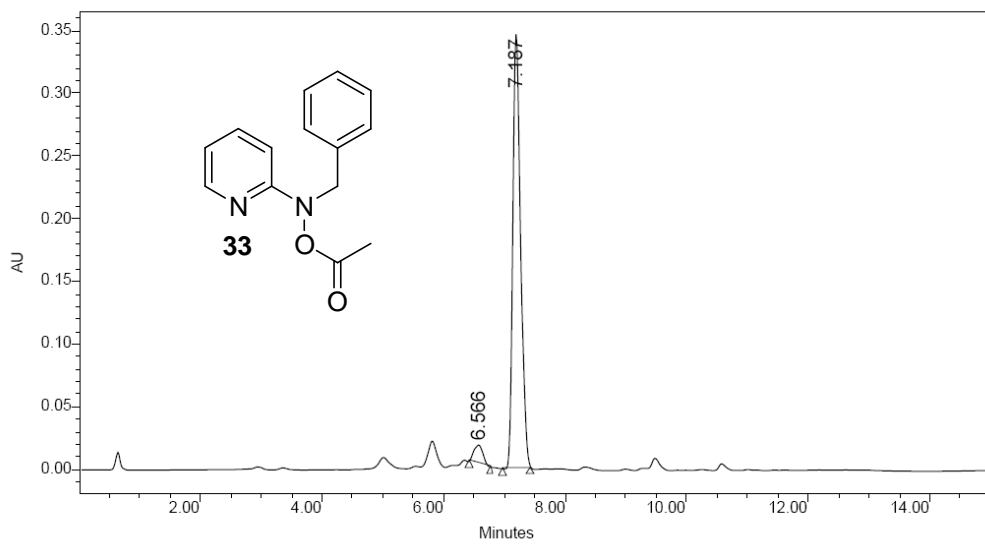
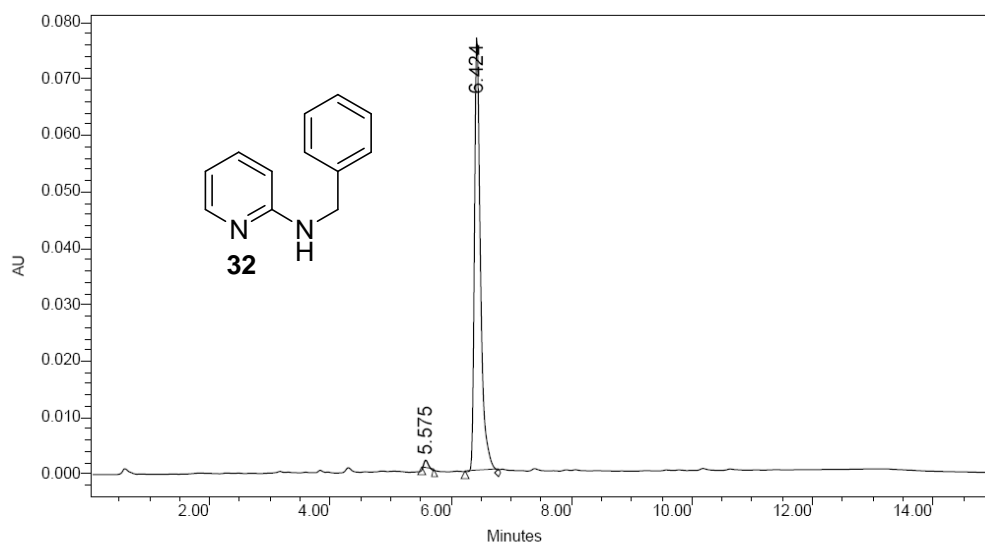
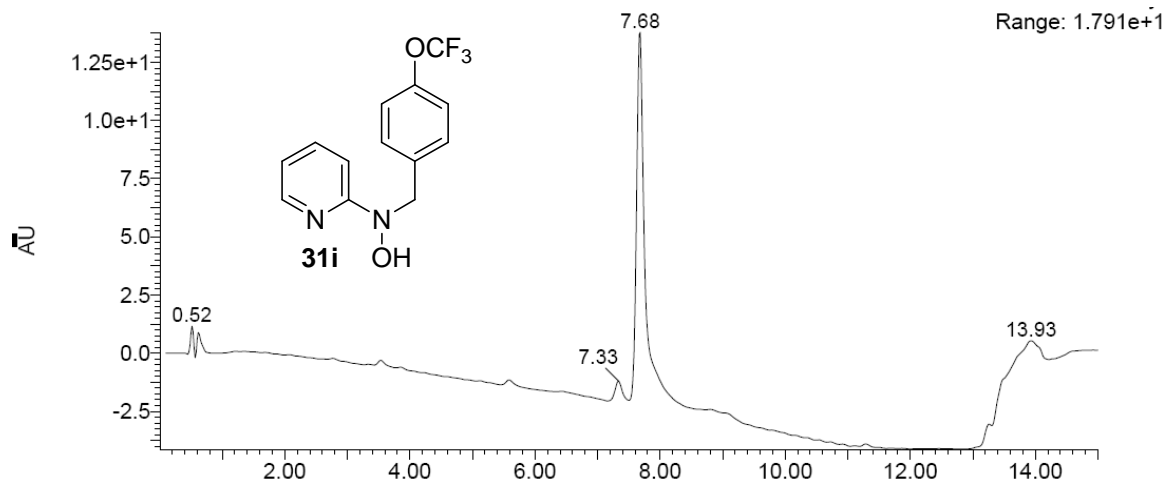


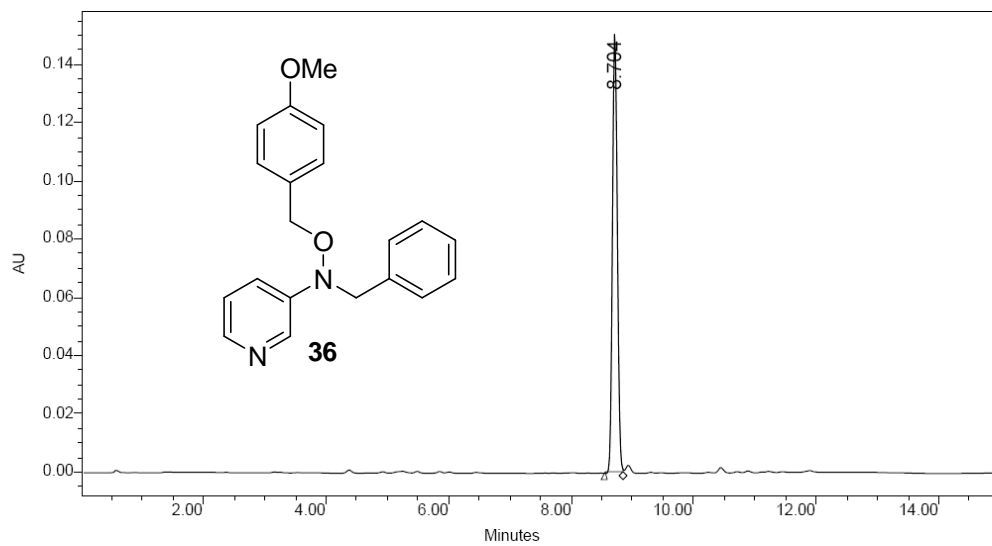
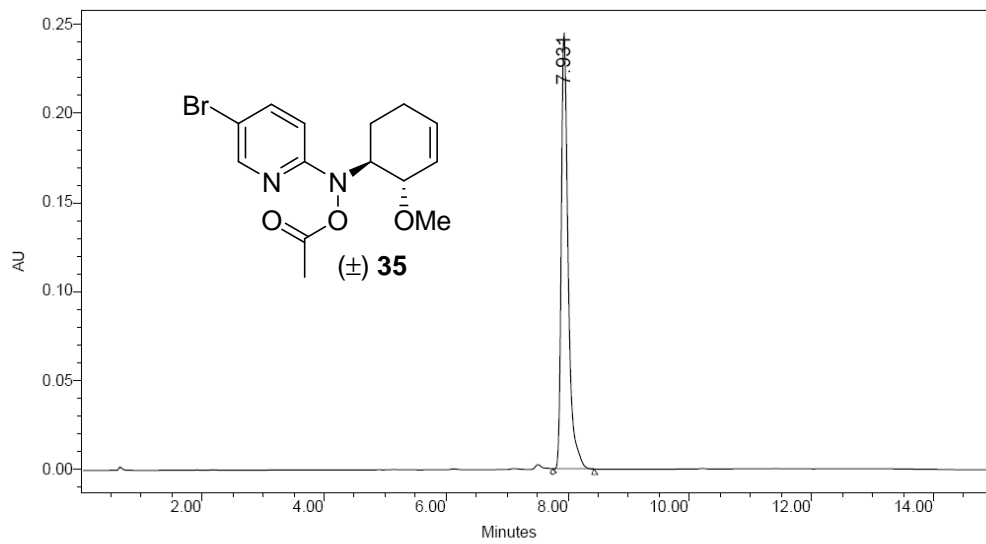
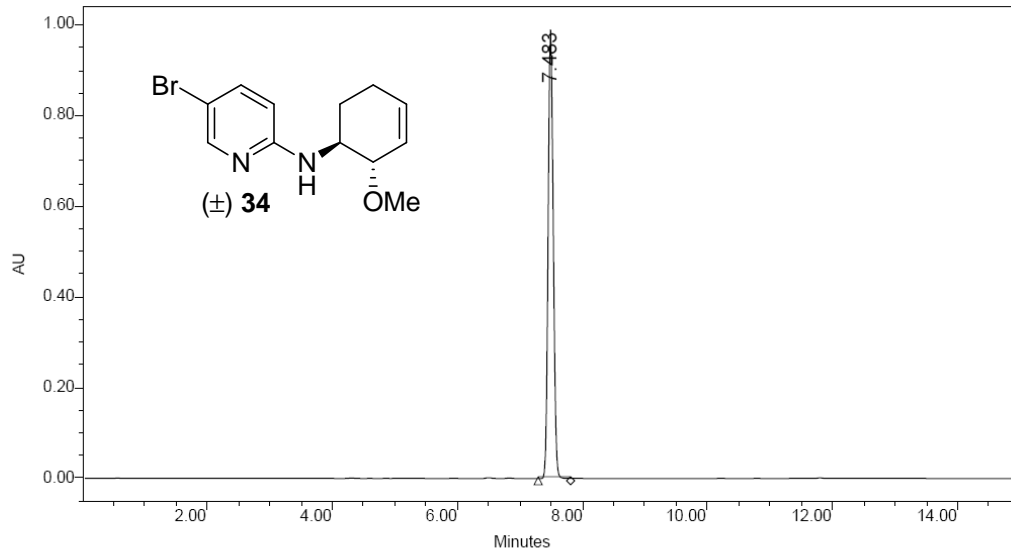


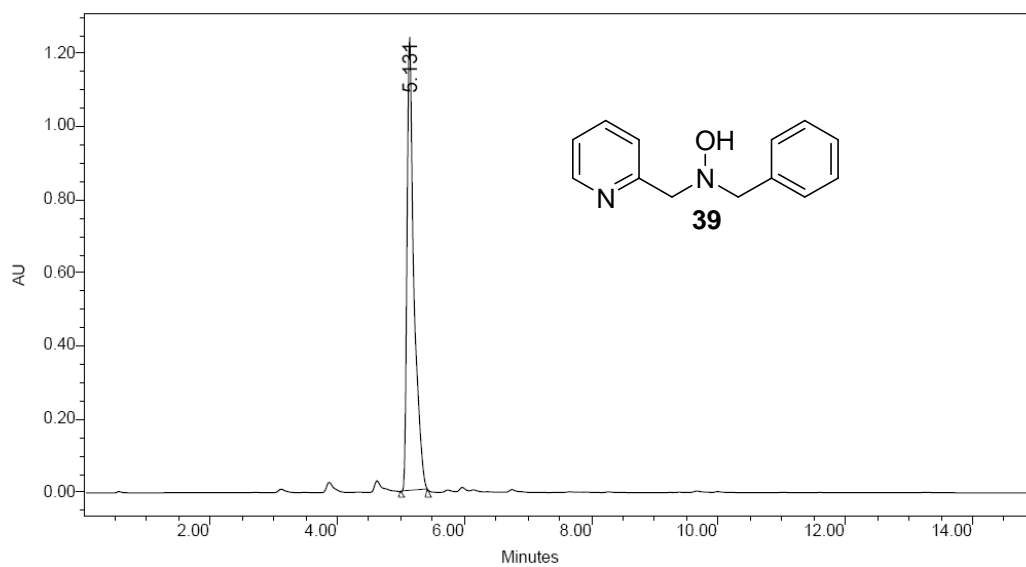
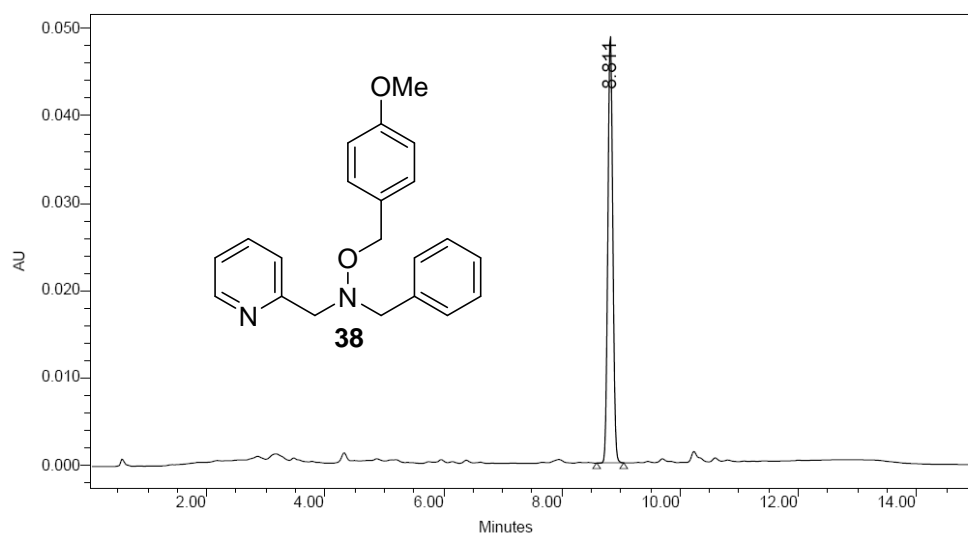
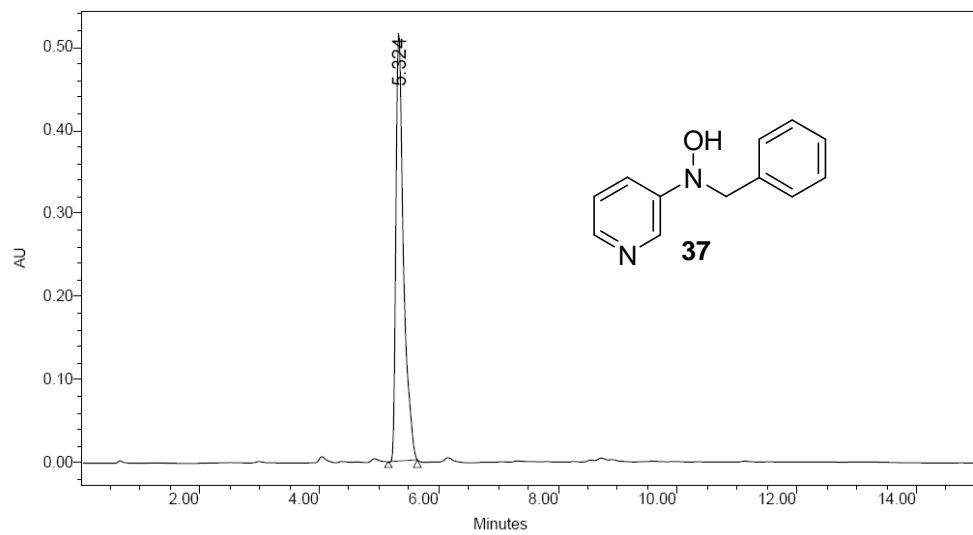




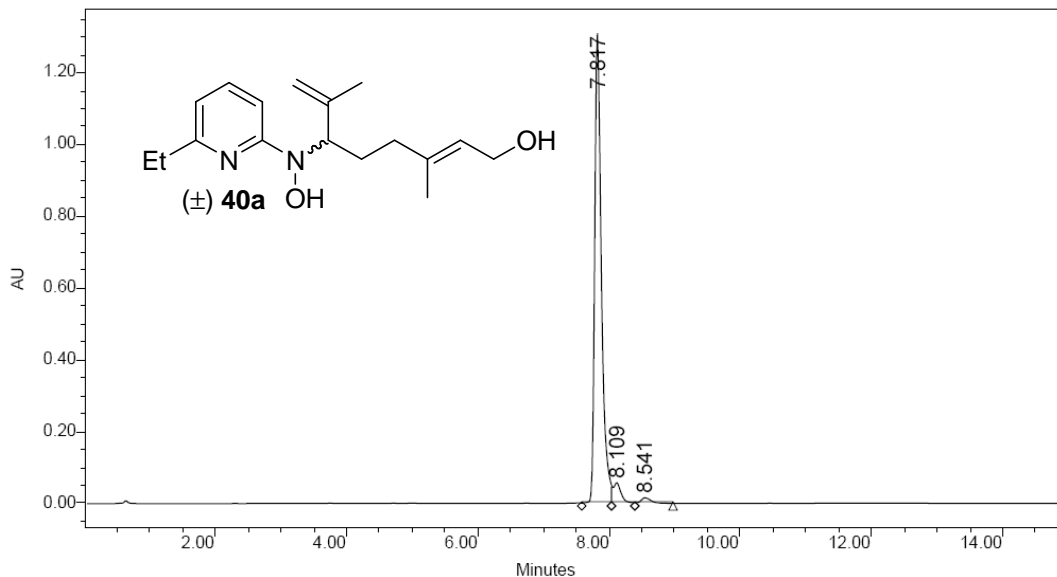












## VII. X-ray Diffraction Data for Compound 40a

### Discussion

Compound **40a** crystallizes as colorless block-like crystals from a dichloromethane/hexanes solution. There are two molecules of the compound in the unit cell of the primitive, centrosymmetric space group P-1.

The structure of compound **40a** is as expected (see **Figures 4, S1, and S2**). Due to the center of symmetry present in the lattice, the compound is a racemate. The C12-C13 bond distance clearly shows the presence of a vinyl group (C12-C13 = 1.323(2) Å). The hydroxyl, O2, forms an H-bond to the pyridyl nitrogen of a neighboring molecule related by inversion symmetry, which in turn also donates an H-bond from the its hydroxyl O2' back to the pyridine nitrogen N2. These H-bonded dimers are then linked in chains along the crystallographic a-axis by hydrogen bonds from the hydroxyl O1, to the hydroxyl oxygen O2'' of a neighboring H-bonded dimer (see **Table S10** for details of Hydrogen-bonds).

The bond distances and angles within the molecule are as expected. Data were collected at the slightly elevated (with respect to routine experiments) temperature of 200 K. Below this temperature the crystals were observed to undergo a phase transition resulting in the crystal shattering.

### Crystal Summary

Crystal data for C<sub>17</sub>H<sub>26</sub>N<sub>2</sub>O<sub>2</sub>; M<sub>r</sub> = 290.40; Triclinic; space group P-1; *a* = 8.7695(3) Å; *b* = 9.2936(3) Å; *c* = 11.3422(4) Å; α = 102.466(2)°; β = 108.736(2)°; γ = 94.007(2)°; V = 845.03(5)

$\text{\AA}^3$ ;  $Z = 2$ ;  $T = 200(2)$  K;  $\lambda(\text{Mo-K}\alpha) = 0.71073$   $\text{\AA}$ ;  $\mu(\text{Mo-K}\alpha) = 0.075$   $\text{mm}^{-1}$ ;  $d_{\text{calc}} = 1.141$   $\text{g}\cdot\text{cm}^{-3}$ ; 11449 reflections collected; 3470 unique ( $R_{\text{int}} = 0.0208$ ); giving  $R_1 = 0.0435$ ,  $wR_2 = 0.1137$  for 2827 data with  $[I > 2\sigma(I)]$  and  $R_1 = 0.0536$ ,  $wR_2 = 0.1217$  for all 3470 data. Residual electron density ( $e^- \cdot \text{\AA}^{-3}$ ) max/min: 0.215/-0.211.

An arbitrary sphere of data were collected on a colorless block-like crystal, having approximate dimensions of  $0.65 \times 0.21 \times 0.17$  mm, on a Bruker APEX-II diffractometer using a combination of  $\omega$ - and  $\phi$ -scans of  $0.5^\circ$ . Data were corrected for absorption and polarization effects and analyzed for space group determination. The structure was solved by direct methods and expanded routinely. The model was refined by full-matrix least-squares analysis of  $F^2$  against all reflections. All non-hydrogen atoms were refined with anisotropic thermal displacement parameters. Unless otherwise noted, hydrogen atoms were included in calculated positions. Thermal parameters for the hydrogens were tied to the isotropic thermal parameter of the atom to which they are bonded ( $1.5 \times$  for methyl,  $1.2 \times$  for all others).

**Table S4.** Crystal data and structure refinement for compound **40a**.

Identification code	nd1105	
Empirical formula	$\text{C}_{17}\text{H}_{26}\text{N}_2\text{O}_2$	
Formula weight	290.40	
Temperature	200(2) K	
Wavelength	0.71073 $\text{\AA}$	
Crystal system	Triclinic	
Space group	P-1	
Unit cell dimensions	$a = 8.7695(3)$ $\text{\AA}$	$\alpha = 102.466(2)^\circ$
	$b = 9.2936(3)$ $\text{\AA}$	$\beta = 108.736(2)^\circ$
	$c = 11.3422(4)$ $\text{\AA}$	$\gamma = 94.007(2)^\circ$
Volume	845.03(5) $\text{\AA}^3$	
Z	2	
Density (calculated)	1.141 $\text{g}\cdot\text{cm}^{-3}$	
Absorption coefficient ( $\mu$ )	0.075 $\text{mm}^{-1}$	
F(000)	316	
Crystal size	$0.65 \times 0.21 \times 0.17$ $\text{mm}^3$	
$\theta$ range for data collection	1.96 to $26.59^\circ$	
Index ranges	$-10 \leq h \leq 10$ , $-11 \leq k \leq 11$ , $-14 \leq l \leq 14$	
Reflections collected	11449	
Independent reflections	3470 [ $R_{\text{int}} = 0.0208$ ]	
Completeness to $\theta = 26.59^\circ$	98.2 %	
Absorption correction	numerical	
Max. and min. transmission	1.0000 and 0.9331	
Refinement method	Full-matrix least-squares on $F^2$	
Data / restraints / parameters	3470 / 0 / 195	

Goodness-of-fit on $F^2$	1.049
Final R indices [ $I > 2\sigma(I)$ ]	$R_1 = 0.0435$ , $wR_2 = 0.1137$
R indices (all data)	$R_1 = 0.0536$ , $wR_2 = 0.1217$
Largest diff. peak and hole	0.215 and -0.211 $e^- \cdot \text{\AA}^{-3}$

**Table S5.** Atomic coordinates and equivalent isotropic displacement parameters ( $\text{\AA}^2$ ) for compound **40a**.  $U(\text{eq})$  is defined as one third of the trace of the orthogonalized  $U_{ij}$  tensor.

	x	y	z	$U(\text{eq})$
O(1)	-0.09777(12)	0.23473(11)	0.07017(10)	0.042(1)
O(2)	0.75090(12)	0.25282(12)	-0.17294(10)	0.043(1)
N(1)	0.05824(14)	0.59512(13)	0.28262(12)	0.038(1)
N(2)	0.00483(13)	0.37548(13)	0.12322(11)	0.035(1)
C(1)	0.0204(2)	0.68794(17)	0.37525(16)	0.045(1)
C(2)	-0.1116(2)	0.6493(2)	0.40998(18)	0.056(1)
C(3)	-0.2080(2)	0.5137(2)	0.34641(18)	0.056(1)
C(4)	-0.17262(18)	0.41800(18)	0.25154(16)	0.045(1)
C(5)	-0.03591(16)	0.46268(15)	0.22275(13)	0.034(1)
C(6)	0.17771(15)	0.35521(15)	0.15008(13)	0.032(1)
C(7)	0.20254(16)	0.28388(17)	0.02437(13)	0.037(1)
C(8)	0.38128(17)	0.26653(17)	0.04643(14)	0.039(1)
C(9)	0.41666(17)	0.21052(16)	-0.07533(14)	0.038(1)
C(10)	0.52519(17)	0.29203(16)	-0.10271(14)	0.039(1)
C(11)	0.57923(18)	0.25562(18)	-0.21617(15)	0.044(1)
C(12)	0.23965(17)	0.27593(18)	0.25557(14)	0.041(1)
C(13)	0.3420(2)	0.3533(2)	0.36914(16)	0.058(1)
C(14)	0.1825(3)	0.1134(2)	0.2285(2)	0.074(1)
C(15)	0.3237(3)	0.0616(2)	-0.1574(2)	0.068(1)
C(16)	0.1288(2)	0.83671(19)	0.43803(19)	0.060(1)
C(17)	0.3019(3)	0.8237(2)	0.50860(19)	0.068(1)
H(1)	-0.1519	0.2270	-0.0075	0.050
H(2)	0.7950	0.3059	-0.2078	0.051
H(2A)	-0.1350	0.7152	0.4764	0.067
H(3A)	-0.2997	0.4860	0.3683	0.067
H(4A)	-0.2390	0.3243	0.2068	0.054
H(6A)	0.2426	0.4574	0.1822	0.038
H(7A)	0.1671	0.3465	-0.0369	0.045
H(7B)	0.1342	0.1847	-0.0142	0.045
H(8A)	0.4498	0.3643	0.0936	0.047
H(8B)	0.4127	0.1965	0.1016	0.047
H(10A)	0.5744	0.3850	-0.0425	0.047
H(11A)	0.5211	0.1573	-0.2729	0.053
H(11B)	0.5534	0.3315	-0.2660	0.053
H(13A)	0.3815	0.3054	0.4366	0.070
H(13B)	0.3761	0.4568	0.3832	0.070

H(14A)	0.2466	0.0746	0.2997	0.111
H(14B)	0.1962	0.0610	0.1486	0.111
H(14C)	0.0672	0.0977	0.2194	0.111
H(15A)	0.3808	0.0178	-0.2146	0.102
H(15B)	0.2143	0.0741	-0.2090	0.102
H(15C)	0.3153	-0.0043	-0.1024	0.102
H(16A)	0.1266	0.8903	0.3711	0.071
H(16B)	0.0846	0.8970	0.4992	0.071
H(17A)	0.3683	0.9225	0.5393	0.102
H(17B)	0.3439	0.7575	0.4505	0.102
H(17C)	0.3068	0.7826	0.5821	0.102

**Table S6.** Anisotropic displacement parameters ( $\text{\AA}^2$ ) for compound **40a**. The anisotropic displacement factor exponent takes the form:  $-2\pi^2[h^2a^{*2}U_{11} + \dots + 2hka^*b^*U_{12}]$

	$U_{11}$	$U_{22}$	$U_{33}$	$U_{23}$	$U_{13}$	$U_{12}$
O(1)	0.0326(5)	0.0429(6)	0.0469(6)	0.0078(5)	0.0141(4)	-0.0043(4)
O(2)	0.0399(6)	0.0493(6)	0.0473(6)	0.0137(5)	0.0255(5)	0.0023(5)
N(1)	0.0391(6)	0.0378(6)	0.0466(7)	0.0132(5)	0.0265(5)	0.0092(5)
N(2)	0.0286(6)	0.0368(6)	0.0419(6)	0.0090(5)	0.0173(5)	0.0020(5)
C(1)	0.0511(9)	0.0424(8)	0.0549(9)	0.0146(7)	0.0323(8)	0.0155(7)
C(2)	0.0616(11)	0.0580(10)	0.0664(11)	0.0148(8)	0.0464(9)	0.0189(8)
C(3)	0.0493(9)	0.0664(11)	0.0730(12)	0.0230(9)	0.0438(9)	0.0128(8)
C(4)	0.0359(8)	0.0517(9)	0.0569(9)	0.0184(7)	0.0259(7)	0.0058(7)
C(5)	0.0309(7)	0.0394(7)	0.0413(7)	0.0174(6)	0.0185(6)	0.0103(6)
C(6)	0.0273(6)	0.0342(7)	0.0374(7)	0.0079(5)	0.0159(5)	0.0033(5)
C(7)	0.0324(7)	0.0463(8)	0.0365(7)	0.0087(6)	0.0170(6)	0.0055(6)
C(8)	0.0333(7)	0.0468(8)	0.0403(8)	0.0070(6)	0.0181(6)	0.0069(6)
C(9)	0.0348(7)	0.0429(8)	0.0398(7)	0.0060(6)	0.0196(6)	0.0055(6)
C(10)	0.0356(7)	0.0396(8)	0.0411(8)	0.0015(6)	0.0177(6)	0.0009(6)
C(11)	0.0379(8)	0.0519(9)	0.0426(8)	0.0064(7)	0.0194(6)	-0.0040(6)
C(12)	0.0360(7)	0.0533(9)	0.0446(8)	0.0189(7)	0.0239(6)	0.0143(6)
C(13)	0.0505(10)	0.0867(13)	0.0422(9)	0.0206(9)	0.0177(8)	0.0231(9)
C(14)	0.0765(14)	0.0625(12)	0.0956(16)	0.0491(12)	0.0277(12)	0.0116(10)
C(15)	0.0754(13)	0.0559(11)	0.0740(12)	-0.0130(9)	0.0516(11)	-0.0161(9)
C(16)	0.0769(13)	0.0433(9)	0.0735(12)	0.0080(8)	0.0495(10)	0.0135(8)
C(17)	0.0788(14)	0.0598(11)	0.0572(11)	-0.0066(9)	0.0287(10)	-0.0011(10)

**Table S7.** Bond lengths [ $\text{\AA}$ ] for compound **40a**.

atom-atom	distance	atom-atom	distance
O(1)-N(2)	1.4250(14)	O(2)-C(11)	1.4298(18)
N(1)-C(5)	1.3397(18)	N(1)-C(1)	1.3531(18)
N(2)-C(5)	1.4013(17)	N(2)-C(6)	1.4815(16)

C(1)-C(2)	1.386(2)	C(1)-C(16)	1.510(2)
C(2)-C(3)	1.375(3)	C(3)-C(4)	1.373(2)
C(4)-C(5)	1.3988(19)	C(6)-C(12)	1.513(2)
C(6)-C(7)	1.5268(18)	C(7)-C(8)	1.5319(18)
C(8)-C(9)	1.5040(19)	C(9)-C(10)	1.329(2)
C(9)-C(15)	1.498(2)	C(10)-C(11)	1.490(2)
C(12)-C(13)	1.323(2)	C(12)-C(14)	1.491(2)
C(16)-C(17)	1.499(3)	O(1)-H(1)	0.8400
O(2)-H(2)	0.8400	C(2)-H(2A)	0.9500
C(3)-H(3A)	0.9500	C(4)-H(4A)	0.9500
C(6)-H(6A)	1.0000	C(7)-H(7A)	0.9900
C(7)-H(7B)	0.9900	C(8)-H(8A)	0.9900
C(8)-H(8B)	0.9900	C(10)-H(10A)	0.9500
C(11)-H(11A)	0.9900	C(11)-H(11B)	0.9900
C(13)-H(13A)	0.9500	C(13)-H(13B)	0.9500
C(14)-H(14A)	0.9800	C(14)-H(14B)	0.9800
C(14)-H(14C)	0.9800	C(15)-H(15A)	0.9800
C(15)-H(15B)	0.9800	C(15)-H(15C)	0.9800
C(16)-H(16A)	0.9900	C(16)-H(16B)	0.9900
C(17)-H(17A)	0.9800	C(17)-H(17B)	0.9800
C(17)-H(17C)	0.9800		

Symmetry transformations used to generate equivalent atoms:

**Table S8.** Bond angles [°] for compound **40a**.

atom-atom-atom	angle	atom-atom-atom	angle
C(5)-N(1)-C(1)	118.61(12)	C(5)-N(2)-O(1)	111.08(10)
C(5)-N(2)-C(6)	117.64(11)	O(1)-N(2)-C(6)	110.28(10)
N(1)-C(1)-C(2)	121.89(15)	N(1)-C(1)-C(16)	116.41(13)
C(2)-C(1)-C(16)	121.69(14)	C(3)-C(2)-C(1)	118.67(15)
C(4)-C(3)-C(2)	120.51(14)	C(3)-C(4)-C(5)	117.94(15)
N(1)-C(5)-C(4)	122.37(13)	N(1)-C(5)-N(2)	115.66(11)
C(4)-C(5)-N(2)	121.85(13)	N(2)-C(6)-C(12)	112.93(11)
N(2)-C(6)-C(7)	109.42(11)	C(12)-C(6)-C(7)	114.38(11)
C(6)-C(7)-C(8)	111.39(11)	C(9)-C(8)-C(7)	113.96(12)
C(10)-C(9)-C(15)	124.80(13)	C(10)-C(9)-C(8)	119.75(13)
C(15)-C(9)-C(8)	115.44(12)	C(9)-C(10)-C(11)	128.02(13)
O(2)-C(11)-C(10)	109.29(12)	C(13)-C(12)-C(14)	122.11(16)
C(13)-C(12)-C(6)	118.92(15)	C(14)-C(12)-C(6)	118.97(14)
C(17)-C(16)-C(1)	113.32(15)	N(2)-O(1)-H(1)	109.5
C(11)-O(2)-H(2)	109.5	C(3)-C(2)-H(2A)	120.7
C(1)-C(2)-H(2A)	120.7	C(4)-C(3)-H(3A)	119.7
C(2)-C(3)-H(3A)	119.7	C(3)-C(4)-H(4A)	121.0
C(5)-C(4)-H(4A)	121.0	N(2)-C(6)-H(6A)	106.5

C(12)-C(6)-H(6A)	106.5	C(7)-C(6)-H(6A)	106.5
C(6)-C(7)-H(7A)	109.3	C(8)-C(7)-H(7A)	109.3
C(6)-C(7)-H(7B)	109.3	C(8)-C(7)-H(7B)	109.3
H(7A)-C(7)-H(7B)	108.0	C(9)-C(8)-H(8A)	108.8
C(7)-C(8)-H(8A)	108.8	C(9)-C(8)-H(8B)	108.8
C(7)-C(8)-H(8B)	108.8	H(8A)-C(8)-H(8B)	107.7
C(9)-C(10)-H(10A)	116.0	C(11)-C(10)-H(10A)	116.0
O(2)-C(11)-H(11A)	109.8	C(10)-C(11)-H(11A)	109.8
O(2)-C(11)-H(11B)	109.8	C(10)-C(11)-H(11B)	109.8
H(11A)-C(11)-H(11B)	108.3	C(12)-C(13)-H(13A)	120.0
C(12)-C(13)-H(13B)	120.0	H(13A)-C(13)-H(13B)	120.0
C(12)-C(14)-H(14A)	109.5	C(12)-C(14)-H(14B)	109.5
H(14A)-C(14)-H(14B)	109.5	C(12)-C(14)-H(14C)	109.5
H(14A)-C(14)-H(14C)	109.5	H(14B)-C(14)-H(14C)	109.5
C(9)-C(15)-H(15A)	109.5	C(9)-C(15)-H(15B)	109.5
H(15A)-C(15)-H(15B)	109.5	C(9)-C(15)-H(15C)	109.5
H(15A)-C(15)-H(15C)	109.5	H(15B)-C(15)-H(15C)	109.5
C(17)-C(16)-H(16A)	108.9	C(1)-C(16)-H(16A)	108.9
C(17)-C(16)-H(16B)	108.9	C(1)-C(16)-H(16B)	108.9
H(16A)-C(16)-H(16B)	107.7	C(16)-C(17)-H(17A)	109.5
C(16)-C(17)-H(17B)	109.5	H(17A)-C(17)-H(17B)	109.5
C(16)-C(17)-H(17C)	109.5	H(17A)-C(17)-H(17C)	109.5
H(17B)-C(17)-H(17C)	109.5		

Symmetry transformations used to generate equivalent atoms:

**Table S9.** Torsion angles [ $^{\circ}$ ] for compound **40a**.

atom-atom-atom-atom	angle	atom-atom-atom-atom	angle
C(5)-N(1)-C(1)-C(2)	0.4(2)	C(5)-N(1)-C(1)-C(16)	-179.02(14)
N(1)-C(1)-C(2)-C(3)	-1.2(3)	C(16)-C(1)-C(2)-C(3)	178.21(17)
C(1)-C(2)-C(3)-C(4)	0.8(3)	C(2)-C(3)-C(4)-C(5)	0.4(3)
C(1)-N(1)-C(5)-C(4)	0.8(2)	C(1)-N(1)-C(5)-N(2)	176.68(13)
C(3)-C(4)-C(5)-N(1)	-1.2(2)	C(3)-C(4)-C(5)-N(2)	-176.83(14)
O(1)-N(2)-C(5)-N(1)	177.34(11)	C(6)-N(2)-C(5)-N(1)	48.93(16)
O(1)-N(2)-C(5)-C(4)	-6.74(18)	C(6)-N(2)-C(5)-C(4)	-135.14(14)
C(5)-N(2)-C(6)-C(12)	62.28(15)	O(1)-N(2)-C(6)-C(12)	-66.50(14)
C(5)-N(2)-C(6)-C(7)	-169.07(11)	O(1)-N(2)-C(6)-C(7)	62.14(13)
N(2)-C(6)-C(7)-C(8)	177.64(11)	C(12)-C(6)-C(7)-C(8)	-54.53(16)
C(6)-C(7)-C(8)-C(9)	-174.44(12)	C(7)-C(8)-C(9)-C(10)	121.23(16)
C(7)-C(8)-C(9)-C(15)	-59.22(19)	C(15)-C(9)-C(10)-C(11)	-0.6(3)
C(8)-C(9)-C(10)-C(11)	178.85(14)	C(9)-C(10)-C(11)-O(2)	-120.37(17)
N(2)-C(6)-C(12)-C(13)	-107.49(15)	C(7)-C(6)-C(12)-C(13)	126.48(15)
N(2)-C(6)-C(12)-C(14)	72.10(17)	C(7)-C(6)-C(12)-C(14)	-53.92(18)
N(1)-C(1)-C(16)-C(17)	-62.6(2)	C(2)-C(1)-C(16)-C(17)	117.94(19)

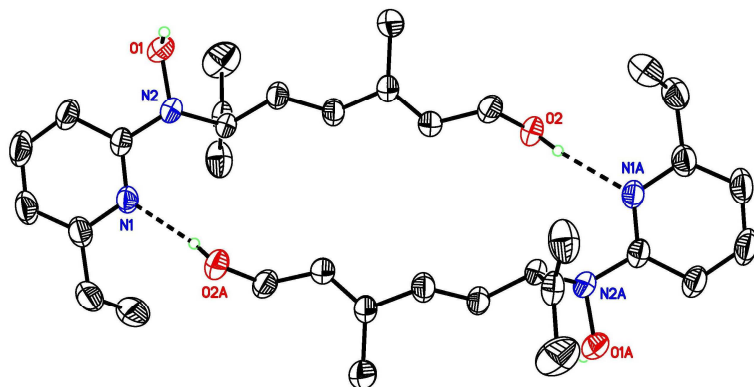
Symmetry transformations used to generate equivalent atoms:

**Table S10.** Hydrogen bonds for compound **40a** [Å and °].

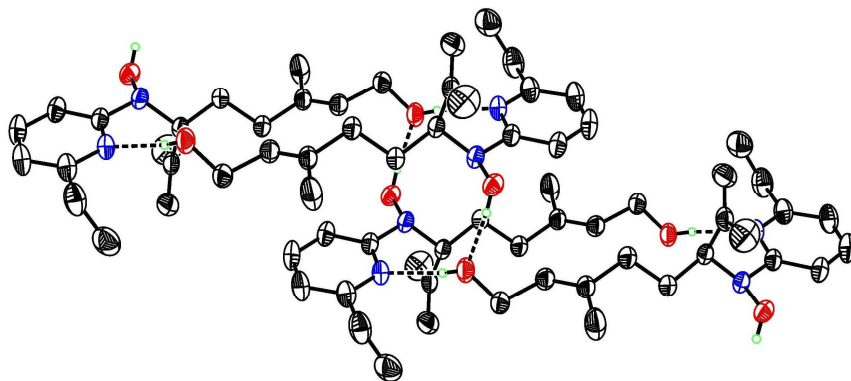
D-H...A	d(D-H)	d(H...A)	d(D...A)	<(DHA)
O(1)-H(1)...O(2)#1	0.84	1.87	2.6969(15)	167.3
O(2)-H(2)...N(1)#2	0.84	2.02	2.8482(15)	167.0

Symmetry transformations used to generate equivalent atoms:

#1  $x-1, y, z$ ; #2  $-x+1, -y+1, -z$



**Figure S1.** Dimer of compound **40a** displayed with 50% probability ellipsoids. Hydrogen atoms (except those involved in H-bonding) omitted for clarity.



**Figure S2.** Stacking of dimers of compound **40a** displayed with 50% probability ellipsoids. Hydrogen atoms (except those involved in H-bonding) omitted for clarity.

## VIII. References

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