

OsWRKY45 alleles play different roles in abscisic acid signaling and salt stress tolerance but similar roles in drought and cold tolerance in rice

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Supplementary Data

OsWRKY45-1	GCAGGCTAGTTGCATGGTCTAACGACAACACTAGGGATGATACATCTCAAAGTATGAAGAGAAACAATGCAGTCATAATA	-1421		
OsWRKY45-2	GCATGCTAGTTGCATGGTCTAACGACAACACTAGGGATGATACATCTCAAAGTATGAAGAGAAACAATGCAGTCATAATA	-1425		
	LTR			
OsWRKY45-1	ATCAAGCAATTATACATTTTCAGGATGGTAGATGCCCTAGATAATATAATCTAGCAGGACAATTAAAGCAAGCAGC	-1341		
OsWRKY45-2	ATCAAGCAATTACACATTTTCAGGATGGTAGATGCCCTAGATAATATAATCTAGCAGGACAATTAAAGCAAGCAGC	-1345		
	MYB			
OsWRKY45-1	TCCCCGTACACCACCCCTCCCTCCCACCAGCATCCCTCTCCCTCTCTTCTCACCACTCAACAGTGACAACTAGATCC	-1261		
OsWRKY45-2	TCCCCGTACACCACCCCTCCCTCCCACCAGCATCCCTCTCCCTCTCTTCTCACCACTCAACAGTGACGACTAGATCC	-1265		
	GCC	GCC	MYB	
OsWRKY45-1	ACAAGAAGGTGACACAGAGCTATAGATAGGCCAGGGCTCAAGGCCGGCGGCCGAGCTGTGCGGGCGATGGTAGTG	-1181		
OsWRKY45-2	ACAAGAAGGTGACATCAGAGCTATAGATAGGCCAGGGCTCAAGGCCGGCGGCCGAGCTGTGCGGGCGACGGTGCTG	-1185		
	GCC	MYB		
OsWRKY45-1	GACACCCACCCCCGTCAAGCTCGCCCTCTTGTCCCTAGCTATGCCGTGCCGGCACAAACCCAAGAGCTCCAA	-1101		
OsWRKY45-2	GACACCCACCCCCGTCAAGCTCGCCCTCTTGTCCCTAGCTATGCCGTGCCGGCACAGCCCAAGAGCTCCAA	-1105		
	DRE			
OsWRKY45-1	GCACGGCGCTGGTACCCCTCCATGCCGACGGTTCTTCTCCCTCTCCCTCTAGCGGCACAGGGCACGAGCGTGG	-1021		
OsWRKY45-2	GCACGGCGCTGGTACCCCTCCATGCCGACGGTTCTTCTCCCTCTCCCTCTAGCGGCACAGGGCACGAGCGTGG	-1025		
	LTR			
OsWRKY45-1	GCCAGTCCTTAAATTGACAAAGAACATTGTTAACCTGGTCCACGATCTTATGGGACAACAAGAACAGAGA	-941		
OsWRKY45-2	GCCAGTCCTTAAATTGACAAAGAACATTGTTAACCTGGTCCACGATCTTATGGGACAACAAGAACAGAGA	-945		
	MYB	MYB		
OsWRKY45-1	CATGGCATGCAATGTCGACCCGGCTTGTGTAAGCTGAACCTACATCTGCCACTGTAAAAGCCCTAGGAGGACAT	-861		
OsWRKY45-2	CCTGGCATGTAATGTCGACCCGGCTTGTGTAAGCTGAACCTACATCTGCCACTGTAAAAGCCCTAGGAGGACAT	-865		
	ABRE	LTR	MYC	MYB
OsWRKY45-1	CAAGGAGGCAAAGGCCATCTGATTCCGAAGTTAAATTATAGTACCACTGTTCTACTCCATGATTCTCAACTGTTG	-781		
OsWRKY45-2	CAAGGAGGCAAAGGCCATCTGATTCCGAAGTTAAATTATAGTACCACTGTTCTACTCCATGATTCTCAACTGTTG	-785		
	MYC	MYB		
OsWRKY45-1	GTTGTCACACGCCACGTICCACCAGGGATCCGATCAAACGATTCCCTCCCTCCCGCGTCCAACCCATCCGCT	-701		
OsWRKY45-2	GTTGTCACACGCCACGTICCACCAGGGATCCGATCAAACGATTCCCTCCCTCCCGCGTCCAACCCATCCGCT	-705		
	ABRE		MYB	
OsWRKY45-1	GCATAAAAATTCCACAAATGACCGGAGCAGCACACCACACCACCCCTCTCATCCTCATTTGCCCCCAACCGGC	-621		
OsWRKY45-2	GCATAAAAATTCCACAAATGACCGGAGCAGCACACCACACCACCCCTCTCATCCTCATTTGCCCCCAACCGGC	-625		
	MYC	MYC	MYB	
OsWRKY45-1	ATCGATCACCTCGTGACGTCGCCCGTGAAGCTCATCCGTTCTCGTCCCCCTCTCCTCGGGATGCTT	-541		
OsWRKY45-2	ATCGATCACCTCGTGACGTCGCCCGTGAAGCTCATCCGTTCTCGTCCCCCTCTCCTCGGGATGCTT	-545		
	ABRE	MYB		
OsWRKY45-1	CCCCCTTGGCCGTACTCCCATGCCCCACCAAGAAGGAACCTGCAACTCAAGTTTTTATTTTTAAAAATAATCT	-461		
OsWRKY45-2	CCCCCTTGGCCGTACTCCCATGCCCCACCAAGAAGGAACCTGCAACTCAAGTTTTTATTTTTAAAAATAATCT	-465		
	MYC	MYC		
OsWRKY45-1	CTCGTGGCCCCCACCACCAAAACTCCCCCGAATTCTCGTACCGCGTCCACTGAGCCGGGGCCACCCCTCCAGGAC	-382		
OsWRKY45-2	CTCGTGGCCCCCACCACCAAAACTCCCCCGAATTCTCGTACCGCGTCCACTGAGCCGGGGCCACCCCTCCAGGAC	-385		
	ABRE			
OsWRKY45-1	TGGCCCACATGTCGCTGATCTCTCCGTGGTTCCCGCGGGTACCGTCACTGTCGCTGGGGCGGTGTAAAGCC	-302		
OsWRKY45-2	TGGCCCACATGTCGCTGATCTCTCCGTGGTTCCCGCGGGTACCGTCACTGTCGCTGGGGCGGTGTAAAGCC	-305		
	MYC	MYB		
OsWRKY45-1	CCCGCCCCCCCCCCCCCCCCCGCTGGCCCACTCACCTCGTGATTCACACCGGATGGGGATTGGTACGCTCGGTTCAT	-222		
OsWRKY45-2	CCCGCCCCCCCCCCCCCCCCCGCTGGCCCACTCACCTCGTGATTCACACCGGATGGGGATTGGTACGCTCGGTTCAT	-225		
	MYB			
OsWRKY45-1	GAGGTTCAACTCGCCGTTACCGCGAGTCACCGCGTGGGGCCACAAATTGGTAGACCGGGTCTATGGTCTCGCGTG	-142		
OsWRKY45-2	GAGGTTCAACTCGCCGTTACCGCGAGTCACCGCGTGGGGCCACAAATTGGTAGACCGGGTCTATGGTCTCGCGTG	-145		
	MYC	MYB	ABRE	GCC
OsWRKY45-1	GTGCCGAGCCGCTCCCTTTCCCATGCCAACGCCAGCGAGGCCGGCCAGTCATGGAAACCCCTGAGGCCGTCACT	-62		
OsWRKY45-2	GTGCCGAGCCGCTCCCTTTCCCATGCCAACGCCAGCGAGGCCGGCCAGTCATGGAAACCCCTGAGGCCGTCACT	-65		
	MYC	MYB	ABRE	GCC
OsWRKY45-1	GGCGTAGCCGACGGTCCCACCTGGCATTGACCGTACCGTCATGACGGTAGACCGC	-1		
OsWRKY45-2	GGCGTAGCCGACGGTCCCACCTGGCATTGACCGTACCGTCATGACGGTAGACCGCGGG	-1		

Figure S1. Alignment of the promoter regions of *OsWRKY45-1* and *OsWRKY45-2*. The nucleotide immediately upstream of the transcription initiation site are numbered as “-1”. The putative ABRE, DRE, GCC, LTRE, MYB, and MYC *cis*-elements are underlined.

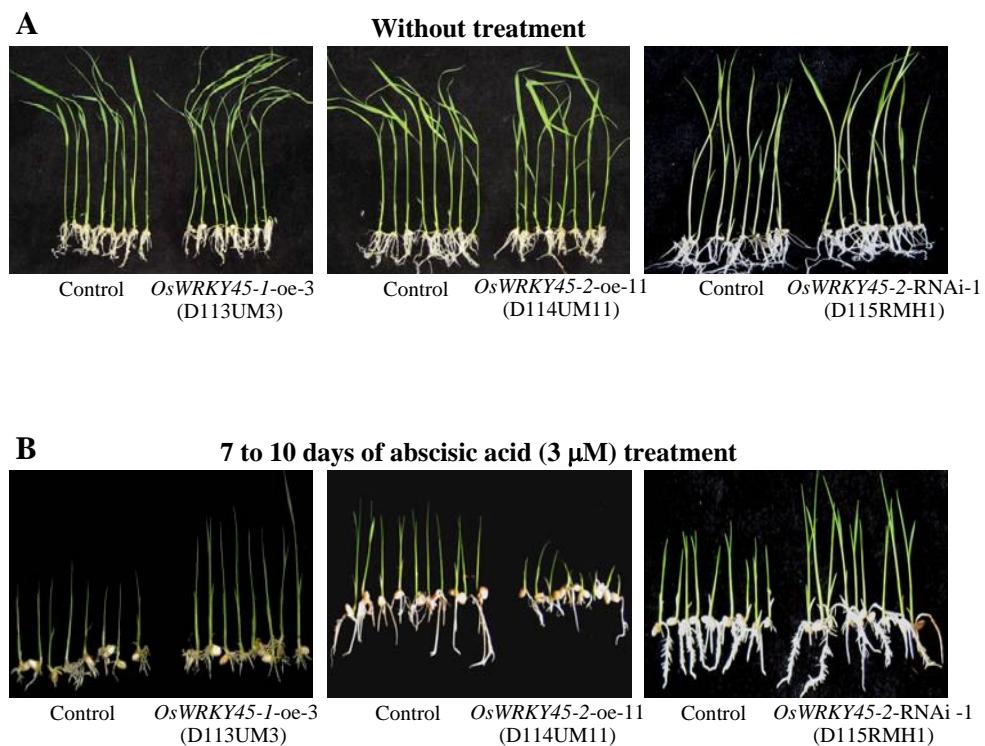


Figure S2. Modulating *OsWRKY45-1* and *OsWRKY45-2* expression influenced rice response to abscisic acid (ABA) treatment. Control, negative siblings from corresponding transgenic segregating populations; oe, overexpression; RNAi, RNA interference. (A) Rice seedlings before ABA treatment. Plant grown on MS medium for 10 d. (B) Phenotypes of transgenic plants after ABA treatment. Rice seedlings grown on MS medium containing ABA for 7 to 10 d (until the transgenic and control plants showed marked differences in growth rate) and then phenotypes were recorded. Because of the variant lengths of ABA-treatment time (7 to 10 d) to different groups of plants, the control plants with the same genetic background in the *OsWRKY45-1-oe* and *OsWRKY45-2-oe* groups exhibited different lengths of shoot and root.

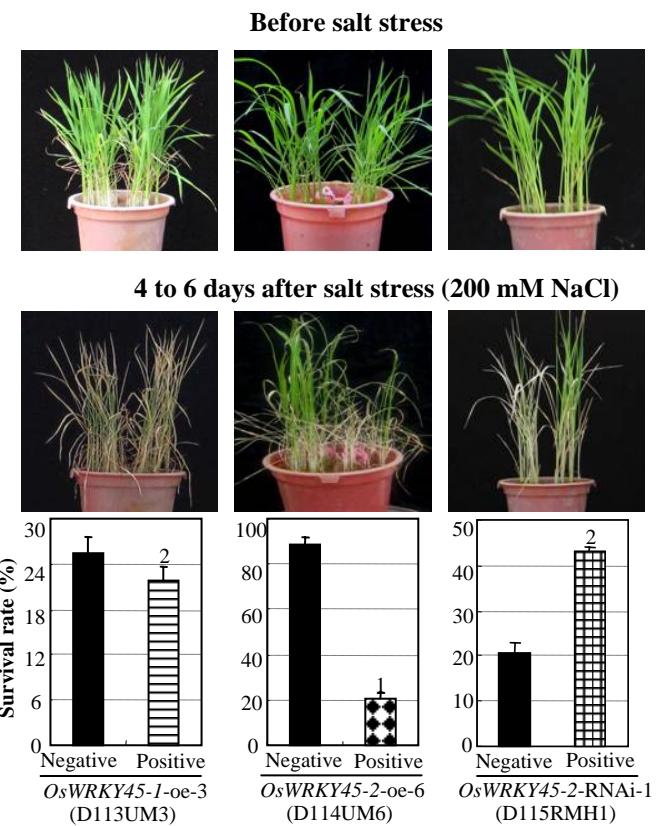


Figure S3. Modulating the expression of *OsWRKY45-2* but not *OsWRKY45-1* markedly influenced rice response to salt stress. Rice seedlings were irrigated with the solution containing 200 mM NaCl at four-to-five leaf stage. After 4 to 6 d of stress (until almost all the leaves of one group in the pot lost their green color and some leaves died), the survival rates were recorded. Negative, negative siblings from corresponding transgenic segregating populations; positive, positive transgenic plants; oe, overexpression; RNAi, RNA interference. Bars represent mean (3 technical replicates with each replicate containing 16 to 20 plants) \pm standard deviation. The “1” or “2” indicates a significant difference was detected between a positive transgenic line and its negative control at $P < 0.01$ or $P < 0.05$, respectively. Because of the variant lengths of salt-stress time (4 to 6 d) to different groups of plants, the control plants with the same genetic background in the *OsWRKY45-1-oe* and *OsWRKY45-2-oe* groups exhibited different survival rates.

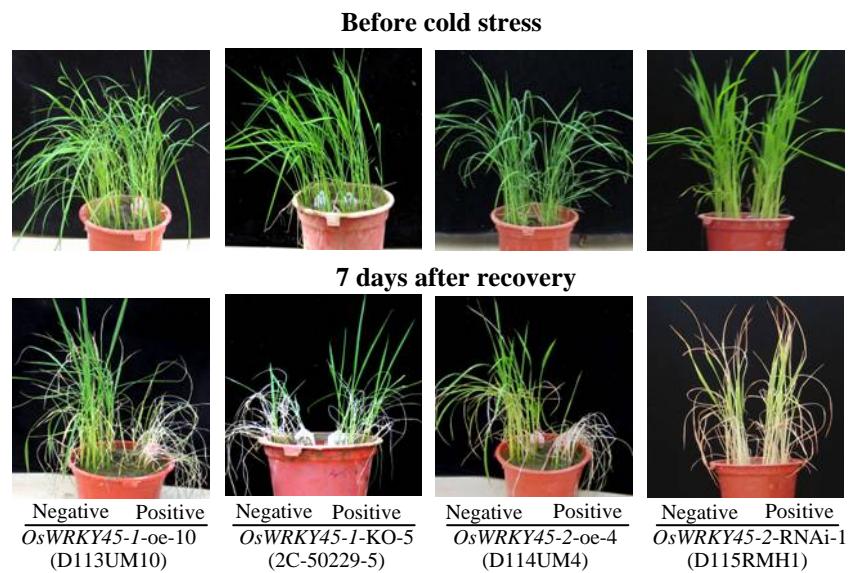


Figure S4. Modulating *OsWRKY45-1* and *OsWRKY45-2* expression influenced rice response to cold stress. Rice seedlings were kept at 4°C for 4 to 6 days (until almost all the leaves of one group in the pot became completely rolled and some leaves died) and then transferred to room temperature (25 to 28°C) for recovery. Negative, negative siblings from corresponding transgenic segregating populations; positive, positive transgenic plants; oe, overexpression; KO, knockout; RNAi, RNA interference.

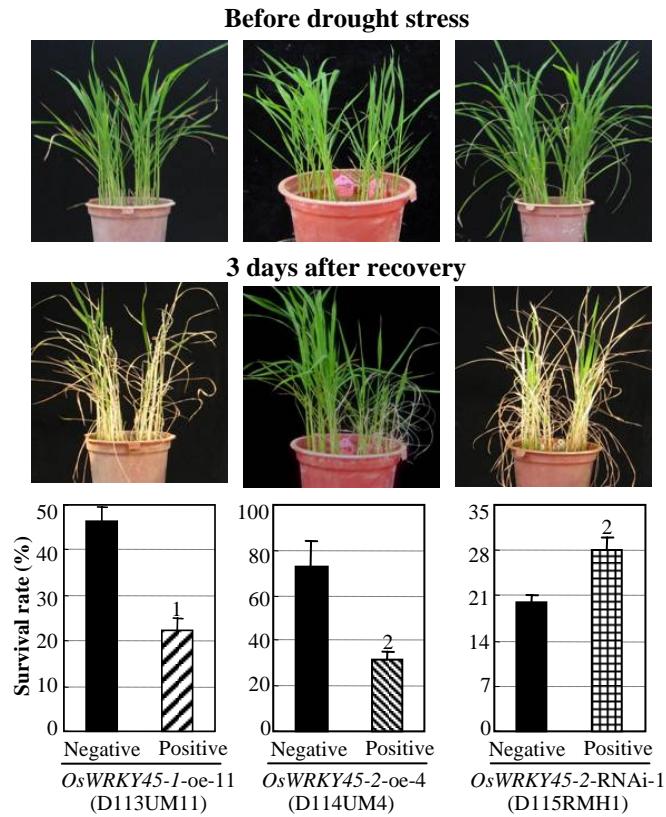


Figure S5. Modulating *OsWRKY45-1* and *OsWRKY45-2* expression influenced rice response to drought stress. Rice seedlings were withheld from water at four-to-five leaf stage for 3 to 5 d (until almost all the leaves of one group in the pot became completely rolled). After three days of recovery, the survival rates were recorded. Negative, negative siblings from corresponding transgenic segregating populations; positive, positive transgenic plants; oe, overexpression; RNAi, RNA interference. Bars represent mean (2-3 replicates with each replicate containing 16 to 20 plants) \pm standard deviation. The “1” or “2” indicates a significant difference was detected between a positive transgenic line and its negative control at $P < 0.01$ or $P < 0.05$, respectively. Because of the variant lengths of water-stress time (3 to 5 d) to different groups of plants, the control plants with the same genetic background in the *OsWRKY45-1-oe* and *OsWRKY45-2-oe* groups exhibited different survival rates.

Table S1. Primers used for PCR amplification

Gene (GenBank accession no.)	Primer name	Forward primer (5'-3')	Reverse primer (5'-3')	Product size (nt)	Use
<i>OsWRKY45</i> (GQ331927, GQ331930)	w45F4/w4 5R4	ATGGTACCGCCTACGCATC ATCTCCTTC ^a	CAGGATCCTTATGGCACA ACATTAGCA ^b	1778-2293	Amplifying genomic fragment of different <i>OsWRKY45</i> alleles and sequencing
<i>OsWRKY45</i>	w45F6/w4 5R6	ATCACAAAGCATAGCATCA TCT	CTCAGCACCTCCTCCTGG TCGG		Sequencing of <i>OsWRKY45</i>
<i>OsWRKY45</i>	w45F/R	TTCCTTGTGATGTGTCGT CTCA	CCCCCAGCTCATAATCAA GAAC	80	Analyzing <i>OsWRKY45</i> expression by RT-PCR
<i>NCED2</i> (AK120176) ^c	NCED2F/ R	TCCGTTGCCAAGATCAA G	CGTCCAACCGTGCAATCA C	75	Analyzing <i>NCED2</i> expression by RT-PCR
<i>NCED4</i> (AK119780)	NCED4F/ R	GATTGCACGGCACCTCA TT	CTCTGTAATTGATTTTC ACTGGCTAAT	98	Analyzing <i>NCED4</i> expression by RT-PCR
<i>NCED5</i> (AK107649)	NCED5F/ R	CCCAGCTTGAAGCTTTG CT	ACAACACTGCAACTATCC CTATCACT	97	Analyzing <i>NCED5</i> expression by RT-PCR
<i>ABI5</i> (AK070998)	ABI5 F/R	AGTGGCACACTCATCCTG CTT	TGAAAGAAGACTTAATGT GCAAATCC	83	Analyzing <i>ABI5</i> expression by RT-PCR
<i>Rab16D</i> (AK109096)	RAB16D F/R	CGGGTAAACAATAAGTC GTGATG	GCGCACTTACATACAGTG CTACGT	74	Analyzing <i>RAB16D</i> expression by RT-PCR
<i>Rab21</i>	RAB21	CACACCACAGCAAGAGCT	TGGTGCTCCATCCTGCTT	101	Analyzing <i>RAB21</i>

(AK121952)	F/R	AAGTG	AAG		expression by RT-PCR
<i>RD22</i>	RD22 F/R	ATCTTGGCGCCTGATCGA	AAAGAACCAAATTGCGTG	120	Analyzing <i>RD22</i>
(AK065358)		G	CAT		expression by RT-PCR
<i>SNAC1</i>	SNAC1	GGGTGCTGTGTCGGCTGT	CGACTGCGACGTAACCAT	100	Analyzing <i>SNAC1</i>
(AK067690)	F/R	A	GT		expression by RT-PCR
<i>OsDREB1B</i>	OsDREB1	ACAGAGTAGGCAATGAG	TTACAGGAATTCTATTGAC	120	Analyzing <i>OsDREB1B</i>
(AY166833)	B F/R	ACTGAGGAT	TGCACAT		expression by RT-PCR
<i>OsDREB1A</i>	OsDREB1	TTACCACACTCGAGCAGA	CCGCTCATCTCCTGCTTG	100	Analyzing <i>OsDREB1A</i>
(AY345233)	A F/R	GCAA	AT		expression by RT-PCR
<i>Actin</i>	Actin-F/R	TGTATGCCAGTGGTCGTAC	CCAGCAAGGTGAGACG	121	Analyzing <i>actin</i> expression
(X15865)		CA	AA		by RT-PCR

^aThe underlined nucleotides are the digestion site of *Kpn*I.

^bThe underlined nucleotides are the digestion site of *Bam*HI.

Table S2. The numbers of known abiotic stress-responsive *cis*-acting elements in the promoters of *OsWRKY45-1* and *OsWRKY45-2*

<i>cis</i> -elements	P _{<i>OsWRKY45-1</i>}	P _{<i>OsWRKY45-2</i>}	Stress	Reference
ABRE	3	5	ABA, drought, salt, cold	6, 9
DRE	1	1	drought, salt, cold	5, 7
GCC	4	4	ABA, drought	8
LTRE	3	3	ABA, drought, cold	3
MYB	13	13	ABA, drought, cold	1, 2
MYC	8	9	ABA, drought, cold	1, 2, 4

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