



Supplementary information, Figure S4. Characterization of ShO-iPS generated from MEFs by treatment of Shh and retroviral transduction with Oct4. (A) RT-PCR analysis and qPCR analysis of ES cell marker genes in mES cells, MEFs, and ShO-iPS cells. (B) Western blot (left) and FACS (right) analyses of ShO-iPS cells. Protein levels of Oct4, Sox2, Nanog, c-Myc, and SSEA1 were similar to those in mES cells. Antibody control, blue line; SSEA1 and Oct4, green line. (C) Bisulfite genomic sequencing of the Oct4 and Nanog promoters from ShO-iPS cells. Open and filled circles indicate unmethylated and methylated CpG dinucleotides, respectively. (D) ChIP analysis of Oct4, Sox2, and Nanog promoters for diMeK9H3 and AcH3 in mES cells, MEFs and ShO-iPS cells. *P < 0.05 compared to MEFs. (E) PCR of genomic DNA to detect integration of exogenous Oct4 and Bmi1 genes in MEFs and ShO-iPS cells. hBmi1/Oct4 was used as positive controls using expression plasmid DNA (pBabe hBmi1 and pBabe Oct4). (F) RT-PCR to detect expression of exogenous Oct4 and Bmi1 transcripts in MEFs and ShO-iPS cells. (G) Scatter plots of the global gene expression comparing ShO-iPS cells with either mES cells or MEFs as described above. (H) In-vitro differentiation of ShO-iPS cells. Micrographs show EBs generated from mES cells (left) and ShO-iPS cells (right). In-vitro differentiation of ShO-iPS cells into ectodermal, mesodermal, and endodermal cell types was confirmed by RT-PCR and immunoreactivity to typical markers BIII Tubulin (Tuj1), Brachyury and SMA, and GATA4, respectively. Scale bars, 200 µm. (I) Induction of Bmi1 by shh in the course of reprogramming.