

Supplementary information, Figure S5. Generation and characterization of PO-iPS cells and OxyO-iPS cells. (**A**) Transdifferentiation of MEFs into neural stem cell-like cells by purmorphamine or oxysterol. MEFs were cultured in DMEM containing 1 μ M purmorphamine or 0.5 μ M 25-hydroxycholesterol (oxysterol) for 3 days. mRNA levels of Gli1, Bmi1, p16^{lnk4a}, p19^{Arf}, and N-Myc were analyzed by RT-PCR (left panel) and real-time PCR (right panel). GAPDH was used as a control. Scale bars, 200 μ m. (**B**) Phase contrast images of vehicle- (middle) and purmorphamine- or oxysterol-treated (right) MEFs cultured in proliferation medium (left) and in NSC medium (right). Purmorphamine- or oxysterol-treated MEFs cultured in NSC medium for 7 days rapidly changed morphology, resulting in bipolar cells (right) and expanded neurosphere-like cells (right). Scale bars, 200 μ m. (**C**) Characterization of purmorphamine- or oxysterol-treated MEFs was performed by AP staining,

immunocytochemistry and RT-PCR for Sox2, Nestin, SSEA1 (left, panel), Musashi1 and CD133 (right, panel). Timeline for PO- and OxyO-iPS cell induction using purmorphamine or oxysterols treatment with retroviral transduction of Oct4 (right, panel). (D) The efficiency of BO-, PO-, OxyO-, and POxyO-iPS cells in reprogramming MEFs. Reprogramming efficiency was calculated by quantifying the percentage of Nanog positive colonies. (E) RT-PCR and qPCR analysis of ES cell marker genes in mES cells, MEFs, and OxyO-iPS clones. (F) Western blot (left) and FACS (right) analyses of OxyO-iPS-1 cells. Protein levels of Oct4, Sox2, Nanog, c-Myc, and SSEA1 were similar to those in mES cells. Antibody control, blue line; SSEA1 and Oct4, green line. (G) Phase contrast images showing ESC-like morphology of OxyO-iPS clones on feeder cells and characterization of OxyO-iPS clones. AP staining, as well as SSEA1, Oct4, Sox2 and Nanog immunoreactivity, was detected in OxyO-iPS clones. Scale bars, 200 µm. (H) Genomic DNA PCR and RT-PCR to detect expression and integration of exogenous Oct4 and Bmi1 genes (upper panels) and transcripts (lower panels), respectively, in MEFs, PO-iPS clones, and OxyO-iPS clones. hBmi1/Oct4 was used as positive controls using expression plasmid DNA (pBabe hBmi1 and hOct4) (I) Bisulfite genomic sequencing of Oct4 and Nanog promoters in mES cells, MEFs, and OxyO-iPS clones. Open and filled circles indicate unmethylated and methylated CpG dinucleotides, respectively. (J) ChIP analysis of Oct4 and Sox2 promoters for diMeK9H3 and AcH3 status in mES cells, MEFs, and OxyO-iPS-1 cells. *P < 0.05 compared to MEFs. (K) Scatter plots of the global gene expression comparing OxyO-iPS cells with either mES cells or MEFs as described previously. (L) In vitro differentiation of OxyO-iPS cells. Micrographs show EBs generated from OxyO-iPS cells (right). In vitro differentiation of OxvO-iPS cells into ectodermal, mesodermal, and endodermal cell types was revealed by immunoreactivity to the tissue-specific markers Nestin, SMA, and GATA4, respectively. Scale bars, 200 µm. (M) The in vivo developmental potential of OxyO-iPS clones. Teratomas of OxyO-iPS clones differentiated into epithelium (endoderm; left), muscle and fat (mesoderm; middle), and neural rosettes (ectoderm; right). Hematoxylin and eosin-stained sections of teratomas derived from OxyO-iPS cells in a nude mouse host after 8-10 weeks are shown. Scale bars, 200 µm.