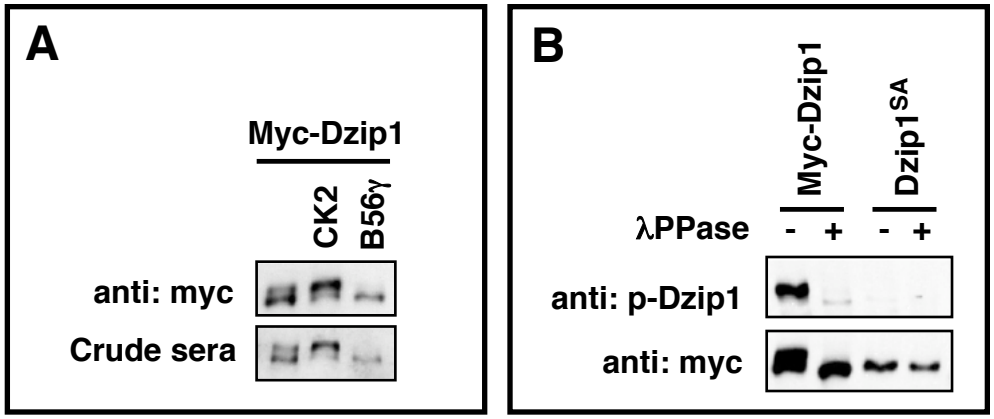
Supplemental figure legends

S-Figure 1 Specificity of anti-Dzip1 antibody.
(**A**) Crude sera. NIH3T3 cells were transfected with Dzip1, Dzip1/CK2, or Dzip1/B56γ. Western blot was performed using an anti-myc antibody or crude anti-sera raised against a KLH-coupled Dzip1 peptide (Ac-CAVK(pS)DTDWTEG(pS)EMD-amide). Both un-phosphorylated and phosphorylated Dzip1 were detected by crude anti-sera. (**B**) Phosphopeptide affinity purified antibody. NIH3T3 cells were transfected with Dzip1 or Dzip1^{SA}. Western blot was performed using an anti-myc antibody or phosphopeptide affinity purified anti-Dzip1 antibody (p-Dzip1). The affinity purified anti-p-Dzip1 antibody was specific to the slower migrating form of Dzip1. It did not cross-react with the faster migrating form of Dzip1 or Dzip1^{SA}. In addition, the immunoreactivity was abolished by treating cell lysate with λPPase. Thus, the anti-p-Dzip1 antibody specifically recognizes Dzip1 that is phosphorylated on Ser706/714.

S-Figure 2 Dzip1-dependent Gli turnover and Dzip1 dependent-ciliogenesis can be uncoupled.
(**A**) Various doses of DMOs were injected into fertilized eggs. Subsequently, a mixture of Gli2 (1 ng) and myc-EGFP (50 pg) RNAs were injected. Embryos were harvested at the late-gastrula stage and subjected to western blot analysis. As low as 10 ng of DMO was sufficient to induce Gli stabilization. (**B**) Immunofluorescence showing primary cilia (arrowheads) of floor plate (top) and epidermis motile cilia (bottom) in control (left) and embryos injected with 10 ng of DMOs (right). Cilia were labeled with an anti-acetylated Tubulin antibody. Dashed lines outline the border of the lumen of the neural tube.

Supplemental Figure 1



Supplemental Figure 2