SUPPLEMENTARY DATA

FIG. S1: Preparation of H1 depleted donor chromatin. The soluble chromatin was supplemented with 0.6 M NaCl after MNase treatment and fractionated over Sephacryl S400 equilibrated with H1 depletion buffer (15 mM Tris pH 7.5, 0.5 M NaCl, 1 mM EDTA, 0.2 m M PMSF, 15 mM BME and protease inhibitors). The figure shows separation of histones on 15 % SDS PAGE.
Lane 1: H1 containing chromatin.
Lane 2: H1 depleted chromatin.

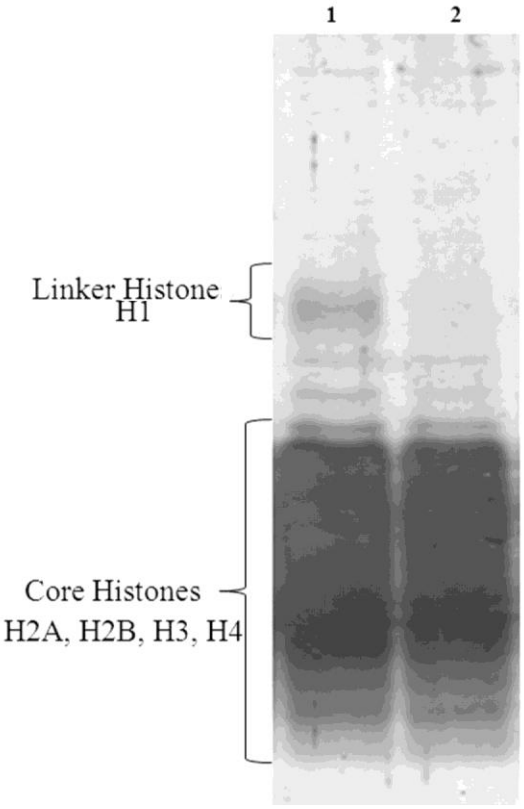
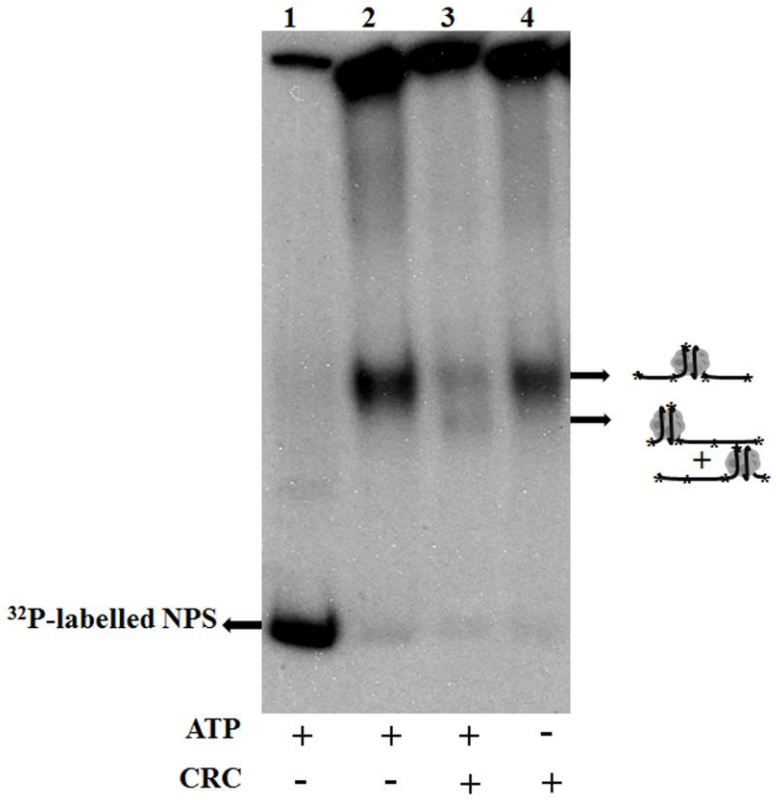


FIG. S2: Histone octamer *cis*-transfer on NPS. ATP-dependent chromatin remodeling activity observed as octamer transfer in *cis*. This activity was monitored as electrophoretic mobility shift on 5 % native PAGE after incubation of the assay mixture at 37 °C for 30 min by using nucleosomes reconstituted on labelled NPS by the salt jump method and purified as described in the Materials and Methods. The assay conditions were the same as that for *trans*-transfer.
Lane 1: ^{32}P-labelled nucleosome positioning sequence (NPS) in the assay mixture.
Lane 2: Nucleosomes assembled by the salt jump method.
Lane 3: Histone octamer mobility in *cis* in the presence of chromatin remodelling complex (CRC) purified from wheat nuclei and ATP.
Lane 4: Reaction carried out in absence of ATP.
The reaction conditions are indicated at the bottom of the lanes. The arrows on the right corresponding to the positions of the bands indicate the polar position of the octamer on NPS, as shown in the diagrams, due to its *cis*-mobility on NPS as described by Eberharter *et al.* (2004).



Reference

Eberharter A, Langst G, Becker P. 2004. A nucleosome sliding assay for chromatin remodeling factors. In: Allis DC, Wu C. eds. *Methods in enzymology*, vol. 377. Elsevier/Academic Press, pp. 344–353.