Mucosal CD8 memory T Cells are selected in the periphery

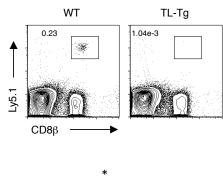
by an MHC Class I molecule

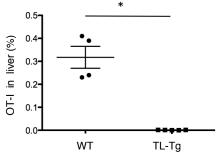
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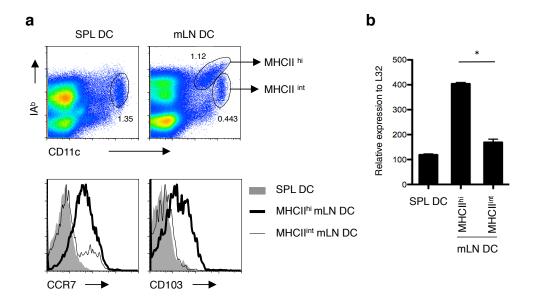
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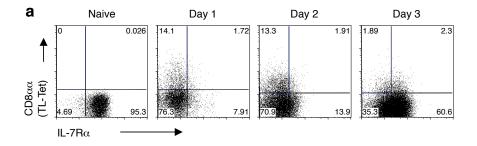


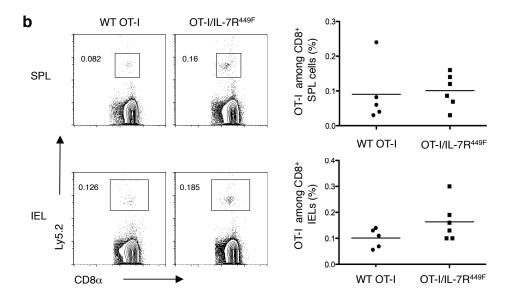


Supplementary Figure 1. TL deletes activated CD8 $\alpha\beta^+$ T cells. 1×10^6 naïve Ly5.1⁺OT-I cells were transferred into WT or TL-Tg recipients. One day after transfer, mice were orally infected with 1×10^9 ActA⁻ Lm-OVA. Memory CD8⁺ OT-I cells in the livers were analyzed 2 month p.i.. Graph depicts pooled data \pm s.e.m.. * P < 0.001 (unpaired t-test). Data are representative of two independent experiments

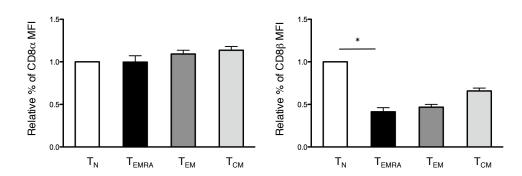


Supplementary Figure 2. MHC class II^{hi}, CD11c⁺ MLN DC have the phenotype of CCR7⁺ CD103⁺ migratory DC. (a) mLN contain a distinct MHC class II^{hi} DC subset that is absent in the spleen (dot plots). Surface expression of CCR7 and CD103 were analyzed on CD11c ⁺MHC II⁺ DCs from SPL and CD11c⁺MHC II^{hi} or CD11c⁺MHC II^{int} DCs from mLN (histograms). Data are representative from at least three independent experiments. (b) Spleen or mLN DCs were sorted based on the CD11c and MHC II expression and used for mRNA quantification for TL expression by qRT-PCR. * P < 0.05 (unpaired t-test). Data are representative from three independent experiments.

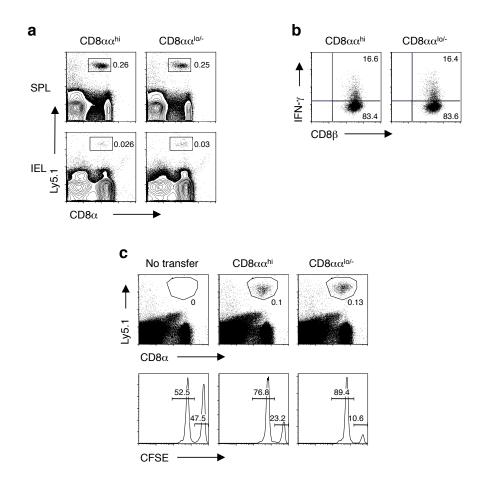




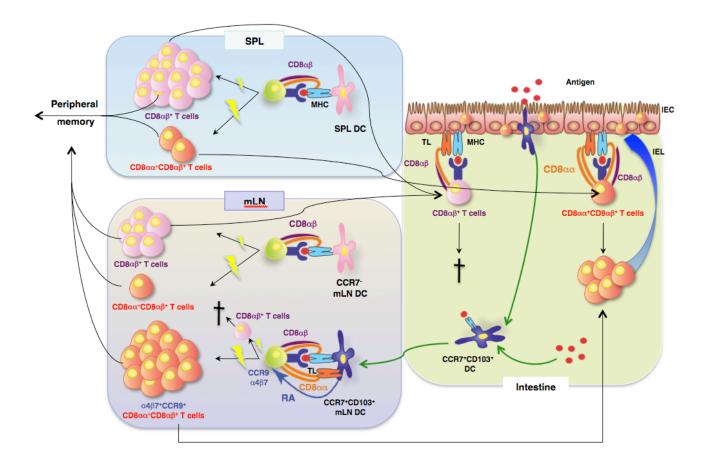
Supplementary Figure 3. Affinity-based selective programming of memory precursor cells does not require IL-7R signals. (a) Naïve CD8+ OT-I cells were cultured in the presence of APC (MEC.B7.SigOVA). CD8αα and IL-7Rα expression on OT-I cells was determined by flow cytometry at different time points. (b) 1×10^4 naïve CD8+ WT OT-I cells or IL-7Ra^{449F} OT-I cells were adoptively transferred into Ly5.1+ WT recipient mice. 1 d after transfer, mice were orally infected with 1×10^9 ActA- Lm-OVA. Donor OT-I cells were tracked in the spleens and IEL 2 months p.i.. Data are representative of two independent experiments.



Supplementary Figure 4. Expression of CD8 α and CD8 β on human CD8⁺ T cell subsets. The bar graphs show the relative percentage of CD8 α and CD8 β mean fluorescence intensity (MFI) in human CD8⁺ T cell subsets (n = 9). * P < 0.01 (unpaired t-test).



Supplementary Figure 5. Characterization of *in vitro* activated CD8αα^{hi} and CD8αα^{lo/-} CD8αβ⁺ T cells. (a) Naïve Ly5.1⁺ CD8⁺ OT-I cells were cultured in the presence of APC (MEC.B7.SigOVA). After two days' culture, CD8αα^{hi} and CD8αα^{lo/-} OT-I cells were sorted and cultured for 3 more days *in vitro*. Then 0.5×10^6 CD8αα^{hi} or CD8αα^{lo/-} were adoptively transferred into B6 recipients. 3 d after transfer, effector OT-I cell were tracked in the spleen and IEL. (b) Sorted CD8αα^{hi} and CD8αα^{lo/-} OT-I cells were directly stimulated with OVA₂₅₇₋₂₆₄ peptide for 5 h and intracellular staining for IFN-γ was performed. (c) 3 d after transfer of CD8αα^{hi} or CD8αα^{lo/-}, *in vivo* cytotoxicity assay was performed in recipient mice. B6 splenocytes labeled as CFSE^{hi} and pulsed with OVA₂₅₇₋₂₆₄ peptide were used as target cells and splenocytes labeled as CFSE^{low} and pulsed with the control peptide were used as control target cells. Data are representative from three independent experiments.



Supplementary Figure 6. Proposed model for the roles of TL and CD8αα on selection of memory CD8αβ+ T cells. CD8αα expression is selectively induced on high affinity/avidity primary effector CD8αβ+ T cells and further enhanced by RA released by mucosal migratory (CCR7+) DC, which also promote gut-tropism (α4β7+CCR9+) of the effector T cells. Activated migratory DC express the CD8αα high affinity ligand, TL, which when interacting with CD8αβ on activated T cells leads to TICD. High affinity primary effector cells that induce CD8αα, escape TICD by sequestering TL away from CD8αβ via CD8αα leading to affinity-based selective survival. TL constitutively expressed on the intestinal epithelial cells (IEC) mediates affinity maturation of the mucosal T_{EM} and eliminates low affinity/avidity primary and secondary effector cells that home to the gut and fail to induce CD8αα.