

# Supporting Information

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## SI Methods

The PCR array data provided by Qiagen regarding the primers used for the *TACR1* transcripts in the array are as follows:

Transcript	RefSeq	Catalog no.	Amplicon size	Reference position
fl- <i>TACR1</i>	NM_001058	PPH68645A	58	4264–4285
tr- <i>TACR1</i>	NM_015727	PPH68646A	80	1581–1604

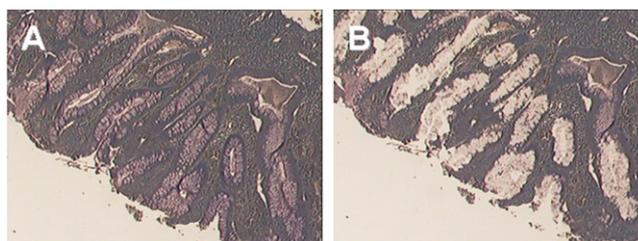


Fig. S1. Colonic epithelium stained with Paradise Plus stain before (A) and after (B) microdissection by laser capture microscopy showing selective removal of epithelium.

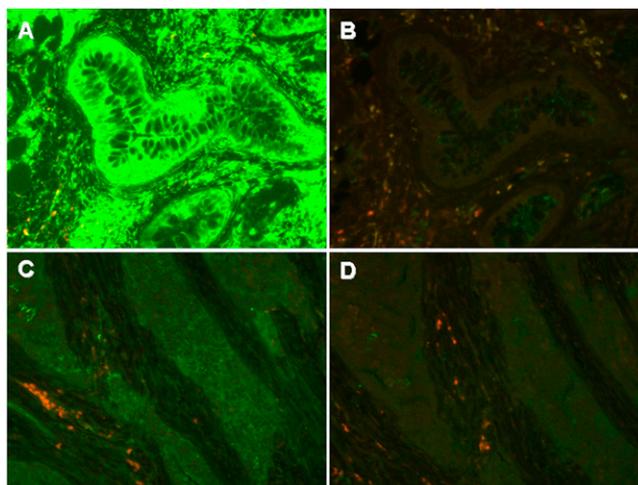


Fig. S2. Specificity of the two NK-1R antibodies: fluorescent microscopy images showing the antibodies binding to human colon carcinoma tissue with and without preincubation with blocking peptides. Antibody targeting the internal epitope of NK-1R binds colon sections without (A) but not with (B) preincubation with blocking peptide. Antibody targeting the C-terminal epitope of NK-1R binds colon sections without (C) but not with (D) blocking peptide. Exposure time = 500 ms. Orange tissue autofluorescence is evident in some sections and does not represent binding.

**Table S1. PCR array genes that did not show significant changes among groups, with fold changes and *P* values**

Gene	QC vs. HGD		QC vs. CA		HGD vs. CA	
	Fold change	<i>P</i> value	Fold change	<i>P</i> value	Fold change	<i>P</i> value
<i>ALOX5</i>	0.85	0.88	2.00	0.37	2.36	0.44
<i>BAD</i>		NA		NA		NA
<i>BCL2</i>	0.44	0.26	0.52	0.35	1.18	0.80
<i>BCL2L1</i>	0.56	0.11	1.28	0.51	2.30	0.07
<i>CAT</i>	0.60	0.37	0.91	0.83	1.52	0.23
<i>CCL2</i>	1.77	0.49	4.11	0.26	2.33	0.42
<i>CCNB1</i>	1.43	0.71	2.27	0.35	1.58	0.63
<i>CCND1</i>		NA		NA		NA
<i>CD44</i>		NA		NA		NA
<i>CD74</i>	2.60	0.30	2.68	0.45	1.03	0.98
<i>CDH1</i>	1.65	0.51	3.16	0.45	1.92	0.58
<i>CDKN1A</i>	0.68	0.31	0.99	0.98	1.45	0.49
<i>CHUK</i>	0.99	0.99	2.71	0.31	2.75	0.13
<i>CXCL10</i>	0.34	0.29	0.47	0.06	1.36	0.70
<i>CXCL12</i>	0.37	0.06	0.79	0.68	2.14	0.30
<i>CXCR4</i>	1.79	0.64	1.49	0.64	0.84	0.84
<i>CYP24A1</i>		NA		NA		NA
<i>CYP27B1</i>	0.34	0.32	1.97	0.66	5.82	0.16
<i>EGFR</i>	6.59	0.19	2.36	0.46	0.36	0.42
<i>EPAS1</i>		NA		NA		NA
<i>EPCAM</i>	1.47	0.79	0.93	0.97	0.63	0.64
<i>GPX1</i>		NA		NA		NA
<i>HIF1A</i>	2.64	0.29	3.73	0.19	1.41	0.77
<i>ICAM1</i>	0.55	0.43	1.12	0.87	2.03	0.53
<i>IGF1</i>	0.33	0.23	0.51	0.41	1.56	0.49
<i>IKBKB</i>	0.55	0.30	1.39	0.73	2.53	0.30
<i>IKBKE</i>	0.38	0.36	1.65	0.35	4.35	0.20
<i>IL1B</i>	0.51	0.28	0.50	0.37	0.97	0.97
<i>IL2</i>	0.22	0.16	0.58	0.44	2.60	0.50
<i>IL6</i>	0.30	0.37	1.13	0.92	3.73	0.34
<i>IL6R</i>	0.33	0.07	0.34	0.13	1.06	0.93
<i>IL8</i>	0.86	0.85	1.36	0.76	1.58	0.35
<i>IL8RA</i>		NA		NA		NA
<i>IL8RB</i>		NA		NA		NA
<i>MCL1</i>	2.33	0.53	0.56	0.78	0.24	0.46
<i>MIF</i>	1.45	0.56	2.46	0.22	1.69	0.51
<i>MMP1</i>		NA		NA		NA
<i>MMP13</i>	2.39	0.47	4.35	0.24	1.82	0.49
<i>MMP2</i>	0.71	0.44	1.92	0.34	2.71	0.20
<i>MMP7</i>	1.04	0.92	8.46	0.26	8.11	0.26
<i>MMP9</i>	1.65	0.37	3.12	0.14	1.89	0.46
<i>MSH2</i>	2.36	0.35	4.66	0.14	1.97	0.47
<i>NFKB1</i>		NA		NA		NA
<i>NFKBIA</i>	2.08	0.21	6.32	0.12	3.03	0.14
<i>NOS2</i>	1.09	0.93	0.93	0.94	0.86	0.86
<i>PDGFA</i>	2.08	0.59	4.06	0.29	1.95	0.57
<i>PDGFB</i>	1.65	0.08	5.13	0.10	3.12	0.19
<i>PDGFRA</i>	0.46	0.18	0.71	0.79	1.54	0.67
<i>PDGFRB</i>		NA		NA		NA
<i>PTGS2</i>	4.66	0.39	8.94	0.09	1.92	0.64
<i>RAF1</i>		NA		NA	1.36	0.72
<i>RELA</i>	1.39	0.38	1.07	0.95	0.77	0.80
<i>RELB</i>	0.28	0.09	0.51	0.44	1.82	0.26
<i>SELL</i>		NA		NA		NA
<i>SMAD2</i>	1.07	0.94	3.94	0.48	3.68	0.25
<i>SMAD4</i>		NA		NA		NA
<i>SOD1</i>	1.82	0.39	1.03	0.99	0.57	0.70
<i>SOD2</i>	9.45	0.10	13.93	0.09	1.47	0.36
<i>STAT3</i>	2.36	0.25	6.68	0.08	2.83	0.22
<i>TAC1</i>		NA		NA		NA

