Supporting Information

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Fig. S1. Examples of hydroxyurea (HU)-induced copy number variants. Illumina SNP array intensity data (log2R) and B allele frequency (B Freq) are at the top and bottom of each panel, respectively. Each dot represents a single probe on the array. Blue dots represent data generated from the untreated control population. Red dots represent data from an HU-treated clone. (*A*) A 335.3-kb deletion at 9q22.33 in clone H3A11 is easily detected by a reduction in the log2R intensity and loss of heterozygous B allele frequencies, shifting from 0.5 to 0 or 1. (*B*) A 389.8-kb duplication at 5q12.3 in clone H2C11 can be identified by an increase in the log2R values and a shift in the B allele frequency from 0.5 to 0.33 or 0.67.



Fig. 52. Overlap of copy number variant (CNV) regions with genes. Histogram (blue bars) shows the frequency of iterations in a 10,000-iteration simulation that had different percentages of simulated regions crossing RefSeq genes, fitted to a Gaussian distribution (red line). Black line shows the percentage of observed copy number variant regions that overlap genes. Details and discussion in main text (*Materials and Methods*).

DNAS

H1A11 2	p22.2 deletion HU)	36,479,534 I	$\frac{H^2}{C}$
Bkpt#1	CATACTTAATAAATAGTCACGTCG	TTAA TAGTGACAGAGACTTGGAATGCT	B
Del Der +#0	CATACTTAATAAATAGTCACGTCC	TTAAACTTGATAAGTCTTACTAGTTTA	De
BKpt#2	TIGIGGGGGCCAGGACTIAGIGI	TTAAACTTGATAAGTCTTACTAGTTTA	B
		36,487,221	
H2A31 3	q13.31 deletion	117,635,151	H
(200µM			(3
Del	CAGTATGAATAACCAGTTTTAGTC	AAAGAACTCCACGTCTGTGTTACCTCT	De
Bkpt#2	GTTTTACAAGTTTCTGTCTTTGAG	AAAGAACTCCACGTCTGTGTTACCTCT	B
		l 117,889,426	
H2C41 5	p15.2 deletion	9,128,809	N
(200µM	HU)	1	(1
Bkpt#1	ACGGGTATCAGGGAGGTGTTGCAA	AAGGAAATGAGGAAATGGGTTGATGGA	B
Bkpt#2	AGGCAGTCTCTGTGGCTACTGGTC	AAGTTAGCTGAACAGCAGCCATAGATG	B
		<u> </u>	
		9,363,325	
H2A31 1	3q21.1 deletion	54,416,275	N'
(200µM	HU)		(1
Bkpt#1 Del	CTGGAAGTCGCTCTGGGTAAGTCA CTGGAAGTCGCTCTGGGTAAGTCA	CTGAGTGAGTGGCGAGTGCTTGTGAAG	De
Bkpt#2	TTTTCTTTTTTGATATGCCTTTG	CTGGTTTTGGTATCAGGGTAATACTGG	B
		l 54,556,077	
1103 61 0		506 221	
(200uM	HU) I	5,506,551	1
Bkpt#1	TGGTGACAAATTCCATTATCAG	A-CATTCATCAGAGGCTCAGATGAG	B
Del Dimt#0	TGGTGACAAATTCCATTATCAG	A-CATTGCTGTACTTTGGCCATCACCA	De
BKpt#2	CCAGTCCATGTAGAACAATATT <mark>CA</mark>	MACATT GCTGTACTTTGGCCATCACCA	В
	30	5,589,228	
H2A61 1	8g22 1 deletion	64 194 028	N
(200µM	HU)	1	(1
Bkpt#1	GAAAAGTGTTTATCATTTAATTCT	TGCA TTTAGAACAAGTAATTTGAGTCA	B
Del Bkpt#2	GAAAAGTGTTTATCATTTAATTCT TGCAATTACAATGAGCCTGGAAG2	TGCAGTGCTGGAAATAGTTGACTAAGT	De
Dirponz			2.
		64,220,275	
H2B11 1	0q21.1 deletion	53,638,925	H:
(200µM	HU)		(:
BKpt#1 Del	TGGAGTAGGGGGGTCCATAAACTTT TGGAGTAGGGGGGTCCATAAACTTT	TTCTGCAGTGAGCTTTGTAGTGAATAT TGGATTGGGAAGGGGACGATTGAGTAG	D1
Bkpt#2	TTTAGGAGCCTAGGATTGGAGGAC	AGGATTGGGAAGGGGACGATTGAGTAG	B
		 E2 671 022	
		53,671,033	
H2C21 3	q13.31 deletion	118,084,102	H
(200µM Bkpt#1			() B1
Del	TCAGTGTAAAATTGAGAACTATAA	AAATAGACATGAGCCACCATGCCTGGC	Di
Bkpt#2	CCTCAGTCTCCCAAAGTGCTGGGA	ATTATAGACATGAGCCACCATGCCTGGC	B
		l 118,135,927	
H2C21 7	all 22 deletion	68.890.457	N
(200µM	dii.zz derecion		1
Bkpt#1	HU)	1	(1
Del	HU) GCATATTTTTGAATAACCTTCACT	I	B
Bkpt#2	HU) GCATATTTTTGAATAACCTTCACT GCATATTTTTGAATAACCTTCACT ACTGTTAGAATCAATATTTTTTG	I I <mark>AGCATTTATTATAGTAGTTATTTATT IAG</mark> AAGCCAAAGTTTATAGGTAAAGGCA IG <mark>G</mark> AAGCCAAAGTTTATAGGTAAAGGCA	BI
Bkpt#2	HU) GCATATTTTTGAATAACCTTCACT GCATATTTTTGAATAACCTTCACT ACTGTTAGAATCAATATTTTTTGI	 	BI Di BI
Bkpt#2	HU) GCATATTTTTGAATAACCTTCACT GCATATTTTTGAATAACCTTCACT ACTGTTAGAATCAATATTTTTTGT	I I <mark>AGCATTTATTATAGTAGTTATTTTATT</mark> IAGAAGCCAAAGTTTATAGGTAAAGGCA I 69,202,211	Bl Di Bl
Bkpt#2 <u>H2</u> C31 7	911.12 deletion GCATATTTTTGAATAACCTTCACT GCATATTTTTGAATAACCTTCACT ACTGTTAGAATCAATATTTTTTGT 921.3 deletion	I AGCATTTATTATAGTAGTTATTTTATT AGAAGCCAAAGTTTATAGGTAAAGGCA GAAGCCAAAGTTTATAGGTAAAGGCA I 9, 202, 211 7, 996, 602	BI Di BI
Bkpt#2 <u>H2C31 7</u> (200µM	911.12 deletion HU) GCATATTTTTGAATAACCTTCACT GCATATTTTTGAATAACCTTCACT ACTGTTAGAATCAATATTTTTTGT 921.3 deletion HU)	I INAGCATTTATTATAGTAGTTATTTTATT INGAAGCCAAAGTTTATAGGTAAAGGCA GAAGCCAAAGTTTATAGGTAAAGGCA I 69,202,211 7,996,602 I	BI Di BI
Bkpt#2 <u>H2C31 7</u> (200µM Bkpt#1 Del	P21.3 deletion HU D21.3 deletion HU TATTTTAGAAAATAAGCATAATG TATTTTAGAAAATAAAGCATAATG	AGCATTTATATAAGTAGTTATTTATT AGAAGCCAAAGTTTATAGGTAAAGGCA GAAGCCAAAGTTTATAGGTAAAGGCA I 69,202,211 7,996,602 I CTATCCTCCAATATGTTATGACACTTT CTTTTCTTTAATACTGTGGATAATGA	BI Du BI
Bkpt#2 H2C31 7 (200µM Bkpt#1 Del Bkpt#2	PULLS deletion HU) GCATATTTTTGAATAACCTTCACT GCATATTTTTGAATAACCTTCACT ACTGTTAGAATCAATATTTTTTGT P21.3 deletion HU) TATTTTAGAAAATAAAGCATAATC TATTTTAGAAAATAAAGCATAATC CTAGACAGTTAAGTCTGGAGTTCT	AGCATTTATATAGTAGTTATTTTATT AGAAGCCAAAGTTTATAGGTAAAGGCA GAAGCCAAAGTTTATAGGTAAAGGCA I 69,202,211 7,996,602 I CTATCCTCCAATATGTTATGACACTTT CTTTTCTTTAATACTGTGGATAATTGA GTTTTCTTTAATACTGTGGATAATTGA	BI Di BI
Bkpt#2 H2C31 7 (200µM Bkpt#1 Del Bkpt#2	911.121 GETETION HU) GCATATTTTTGAATAACCTTCACT ACTGTTAGAATCAATAACCTTCACT ACTGTTAGAATCAATAATTTTTTGT 'p21.3 deletion HU) TATTTTAGAAAATAAAGCATAATG TATTTTAGAAAATAAAGCATAATG CTAGACAGTTAAGTCTGGAGTTCT	AGCATTTATATAGTAGTTATTTTATT AGAAGCCAAAGTTTATAGGTAAAGGCA GGAAGCCAAAGTTTATAGGTAAAGGCA I 69,202,211 7,996,602 I CTATCCTCCAATATGTTATGACACTTT CTTTTCTTTAATACTGTGGATAATTGA I 8,072,022	BI Di BI

2C41 12p13.33 deletion 1,095,921 200µM HU) kpt#1 TGGAATCTGTTAAAGTTCTAGATAG<mark>T</mark> CTAATTTTAAACCACAGATTATTAG TGGAATCTGTTAAAGTTCTAGATAGT el kpt#2 CCTCGGCCTCCCTGAGTGCTGGGATTATAGGCGTGAGCCGCTGCGCCCAGC 1,463,394 3B11 3q13.31 deletion 118,095,904 300µM HU) kpt#1 TTAATAAATCAAGTTTTCAATCAA<mark>AGA</mark>CAAATAAAATCTGAAATACTGCAG TTAATAAATCAAGTTTTCAATCAA<mark>AGA</mark>ATGTCACTTACAATGTCCTAGACC el kpt#2 CAATACTAGATGAAAATTAGGCTC<mark>AGA</mark>ATGTCACTTACAATGTCCTAGACC 118,200,900 58,263,350 TB31 14q23.1 deletion (TI TGTTCAGAGGCAGAGGTCTCGGCT<mark>GCA</mark>GTCACAAAAACTAACAGAAAGAGA kpt#1 TGTTCAGAGGCAGAGGTCTCGGCT<mark>GCA</mark>TCATAACAGCATGCAGCTCCTGAC e1 TCTGTTTCCCCACCTGTAACATGGGCATCATAACAGCATGCAGCTCCTGAC kpt#2 58,309,487 TC21 3q13.31 deletion NT) 117,153,488 CAAAAGAAGATATACAAATGGCCA<mark>GCA</mark>AACATGAAAAATGCTCAACATCAC kpt#1 CAAAAGAAGATATACAAATGGCCA<mark>GCA</mark>CATGCCTGTAATTCCAGCTACTTG el kpt#2 ATACAAAAATTGGCTGGGCATGTG<mark>GCA</mark>CATGCCTGTAATTCCAGCTACTTG 117,814,247 TG51 7p15.1 deletion 31,756,287 (TI 1 kpt#1 AGTGGAAATTTGGAAGAATAAAGCAATTCAAAAAGATCTCCACAGCACATT el kpt#2 GCTCAAAGATTCCCAAACAGATA<mark>CAATT</mark>CAAAAAGATCTCCACAGCACATT 31,874,852 IG51 2q32.3 deletion 196,823,287 NT) GACAGCTCTGCAAATCAGGGTTCGTATGTTCCAGTTGTTAATGGTTCAAAT kpt#1 GACAGCTCTGCAAATCAGGGTTCGTAATGCTGGGAGTCTCTGGATCTTAAC el kpt#2 AGGCTCTGAAAAGGAAAGGTCTCA<mark>TA</mark>ATGCTGGGAGTCTCTGGATCTTAAC . 197,031,352 A51 3q27.3 duplication 189,205,577 100µM HU) TGTCAGAATGCCTGGTGGATGCCTGGGTGCGACACCTGCCAAGGCATACAG kpt#1 TGTCAGAATGCCTGGTGGATGCCTGGTAGCTGCTAGGCTCTCTATCCTGGG up kpt#2 TTTTAAATAATGCCTTTTTGAGGAACTAGCTGCTAGGCTCTCTATCCTGGG 188,457,031 2B41 17p12 duplication 14,348,849 200µM HU) kpt#1 CCAGCCCTGGCTTCAATGCTCTGG<mark>TT</mark>CACTTCCGTCAGACTGGTCTCTCTC up 14,349,353 TA11 10q24.13 duplication 102,832,429 (TI GTAAGGGATCTCTTCCAGGTTGGAT TCCCAAATAGATGAGACCCTCAAT kpt#1 **GTAAGGGATCTCTTCCAGGTTGGATTTT**TATTATTGAGTTGTATGTTCTGG ap kpt#2 TATTTTTGAATATAGTTATTTGTCTTTTTTTTTTGAGTTGTATGTTCTGG 102,710,396

Fig. S3. Copy number variant breakpoint junctions from hydroxyurea (HU)-treated and untreated clones. Breakpoints from 12 deletions and 2 duplications from hydroxyurea-treated cells and 4 deletions and 1 duplication from untreated cells were sequenced, revealing microhomologies and blunt ends. Sequences from the left and right breakpoint regions are red and blue, respectively. Regions of perfect homology at the junction are underlined and highlighted in yellow.



Fig. S4. Comparison of breakpoint sequences in hydroxyurea- and aphidicolin-induced copy number variants (CNVs) in vitro and germline CNVs in vivo. The frequencies of CNV breakpoints with 0–10 bp of microhomology are compared between all replication stress-induced CNVs from this and previous studies (red line) (1, 2) and germline CNVs seen in vivo (blue line) (3, 4). The expected frequency if microhomologous junctions occurred randomly is also shown (gray line).

1. Arlt MF, et al. (2009) Replication stress induces genome-wide copy number changes in human cells that resemble polymorphic and pathogenic variants. Am J Hum Genet 84:339–350.

2. Arlt MF, et al. (2011) Comparison of constitutional and replication stress-induced genome structural variation by SNP array and mate-pair sequencing. Genetics 187:675-683.

3. Vissers LE, et al. (2009) Rare pathogenic microdeletions and tandem duplications are microhomology-mediated and stimulated by local genomic architecture. Hum Mol Genet 18: 3579–3593.

4. Conrad DF, et al. (2010) Mutation spectrum revealed by breakpoint sequencing of human germline CNVs. Nat Genet 42:385-391.

	Observed CNVs		Simulation mean			
CNVs in region (N)	Regions (R _{obs})	CNVs	Regions (λ _N)	CNVs	<i>P</i> (>0)	P(R _{obs})
1	103	103	193	193	1.0	1.0
2	11	22	6.7	13.4	1.0	0.08
3	5	15	0.28	0.84	0.2	0.00001
4	3	12	0.011	0.044	0.01	0.0000002
6	1	6	0.0001	0.0006	0.0001	0.0001
7	1	7	0	0	<0.0001	<0.0001
11	1	11	0	0	<0.0001	<0.0001
31	1	31	0	0	<0.0001	<0.0001

Table S1. Monte Carlo simulation to identify copy number variant (CNV) hotspots

Details in main text (Materials and Methods).

Other Supporting Information Files

Dataset S1 (XLSX)