

SUPPLEMENTARY TABLE 1

Gene name and nucleotide composition for primers used in real-time PCR. Cycling parameters are described in references.

Gene	Primer Sequences
<i>Hprt</i> (1)	For: 5'-AGCCTAAGATGAGCGCAAGT-3' Rev: 5'-TTACTAGGCAGATGGCCACA-3'
<i>Stat1</i> *	For: 5'-CCTGCAACTGAAGGAACAGA-3' Rev: 5'-TCACCACGACAGGAAGAGAG-3'
<i>Stat6</i> (2)	For: 5'-GACTATGGTAGAGGACAGTTGCCTAA-3' Rev: 5'-GGTTCTCCAGGAGAAGCTTGGT-3'
<i>Il4</i> (3)	For: 5'-CCACGGATGCGACAAAATC-3' Rev: 5'-TGTTCTTCGTTGCTGTGAGGAC-3'
<i>Il5</i> (4)	For: 5'-TGACAAGCAATGAGACGATGAGG-3' Rev: 5'-ACCCCCACGGACAGTTTGATTC-3'
<i>Il13</i> (5)	For: 5'-GCAACATCACACAAGACCAGA-3' Rev: 5'-GTCAGGGAATCCAGGGCTAC-3'
<i>Gata3</i> (6)	For: 5'-AGTTCGCGCAGGATGTCC-3' Rev: 5'-AGAACCGGCCCTTATCAA-3'
<i>Il1b</i> (7)	For: 5'-CAACCAACAAGTGATATTCTCCATG-3' Rev: 5'-GATCCACACTCTCCAGCTGCA-3'
<i>Il12p40</i> (3)	For: 5'-AAAGGCTGGGTATCGGTGG-3' Rev: 5'-ACTGGCTGTGCTGGA ACTCC-3'
<i>Tnfa</i> (8)	For: 5'-AAGGGAGAGTGGTCAGTTGCC-3'

Tbet (6) Rev: 5'-CCTCAGGGAAGAGTCTGGAAAGG-3'
For: 5'-TTCTCTGTTTGGCTGGCTGTT-3'
Rev: 5'-GGATTCTGGGGTTTACTTCTT-3'

Inos (9) For: 5'-CACCAAGCTGAACTTGAGCGA-3'
Rev: 5'-CCATAGGAAAAGACTGCACCGA-3'

Arg1 (9) For: 5'-CAGAAGAATGGAAGAGTCAG-3'
Rev: 5'-CAGATATGCAGGGAGTCACC-3'

Chi3l3 [Ym1] (10) For: 5'-CATGAGCAAGACTTGCGTGAC-3'
Rev: 5'-CATGAGCAAGACTTGCGTGAC-3'

Retnla [Fizz1] (10) For: 5'-TCCCAGTGAATACTGATGAGA-3'
Rev: 5'-CCACTCTGGATCTCCCAAGA-3'

Mmp12 (9) For: 5'-TCAATTGGAATATGACCCCCTG-3'
Rev: 5'-ACCAGCAAGCACCCCTTCACTAC-3'

Il4ra (11) For: 5'-GAGTGAGTGGAGTCCTAGCATC-3'
Rev: 5'-GCTGAAGTAACAGAACAGGC-3'

Il13ra1 (11) For: 5'-GCAGCCTGGAGAAAAGTCGTCAAT-3'
Rev: 5'-ACAGCGTCGGCAAGAACACCA-3'

Il13ra2 (12) For: 5'-TGCAGATGATGGAATTTGGA -3'
Rev: 5'-GCTCAATGTGGGTTTCAGGTT-3'

Ifng (13) For: 5' -AGCTCATCCGGTGGTCCAC-3 '
Rev: 5'-AAAATTCAAATAGTGCTGGCAGAA-3'

Cxcl9 For: 5'-TCGAGGAACCCTAGTGATAA-3'
Rev: 5'-ACTGTTTGAGGTCTTTGAGG-3'

Cxcl10 For: 5'-GTCTGAGTGGGACTCAAGG-3'

Irf1 Rev: 5'-TGATTTCAAGCTTCCCTATG-3'
For: 5'- AGAGAGAAAGTCCAAGTCCAGC -3'
Rev: 5'- GTCCATCAGAGAAAGTGTCCG -3'

Ifnb For: 5'- TACGTCTCCTGGATGAACTC -3'
Rev: 5'- TCTTCAAGTGGAGAGCAGTT -3'

Socs1 (14) For: 5'-ACCTTCTTGGTGCGCGAC -3'
Rev: 5'- AAGCCATCTTCACGCTGAGC -3'

Rantes For: 5'- GGAGTATTTCTACACCAGCAGC -3'
Rev: 5'- TCTTGAACCCACTTCTTCTCTG -3'

Fasl For: 5'- GGTGCTAATGGAGGAGAAGAGG -3'
Rev: 5'- CACTGGTAAGATTGAATACTGCC -3'

Fas For: 5'- ACTTCTACTGCGATTCTCCTGG -3'
Rev: 5'- TGTATTGCTGGTTGCTGTGC -3'

Tnfr1 For: 5'- CCCCAAATGGAAATGTGCTATG -3'
Rev: 5'- GGATGCTACAGATGCGGTGG -3'

Ccl3 For: 5'-CCAATTCATCGTTGACTATTT-3'
Rev: 5'-CTTGGACCCAGGTCTCTT-3'

Ccl4 For: 5'-CCTGCTGTTTCTCTTACACC-3'
Rev: 5'-TTTGGTCAGGAATACCACAG-3'

Mcp1 For: 5'- TCTGCCCTAAGGTCTTCAGCAC -3'
Rev: 5'- GCTTGAGGTGGTTGTGGAAAAG -3'

Il17a(6) For: 5'- CAGACTACCTCAACCGTTCC -3'
Rev: 5'- AGCATCTTCTCGACCCTGAA -3'

*Purchased from Real Time Primers, LLC (Elkins Park, PA)

SUPPLEMENTARY TABLE 2

Correlation coefficient matrix for 10 genes from Figure 9A and C in a cohort of 45 patients with biliary atresia.

		CCR4	IL18	CD74	CD86	AREG/AREGB-1	IL4R	BCL3	BCL6	IL1RL1	IL6
CCR4	Pearson's r R Std Error t p-value	1.									
IL18	Pearson's r R Std Error t p-value	-0.094 0.023 -0.623 0.535	1.								
CD74	Pearson's r R Std Error t p-value	0.165 0.022 1.100 0.277	0.450 0.018 3.311 0.001	1.							
CD86	Pearson's r R Std Error t p-value	0.264 0.021 1.801 0.078	0.341 0.020 2.380 0.021	0.634 0.013 5.381 <0.001	1.						
AREG/AREGB-1	Pearson's r R Std Error t p-value	0.384 0.019 2.727 0.009	0.316 0.020 2.186 0.034	0.140 0.029 0.928 0.358	0.535 0.016 4.153 <0.001	1.					
IL4R	Pearson's r R Std Error t p-value	0.124 0.022 0.824 0.414	0.117 0.022 0.778 0.440	-0.087 0.023 -0.573 0.569	0.153 0.022 1.019 0.313	0.612 0.014 5.078 <0.001	1.				
BCL3	Pearson's r	0.331	0.143	0.075	0.331	0.646	0.866	1.			

	R Std Error	0.021	0.022	0.0231	0.020	0.013	0.005			
	t	2.307	0.950	0.494	2.302	5.556	11.385			
	p-value	0.025	0.347	0.623	0.026	<0.001	<0.001			
BCL6	Pearson's r	0.127	0.113	-0.025	0.216	0.549	0.835	0.815	1.	
	R Std Error	0.022	0.022	0.023	0.022	0.016	0.007	0.007		
	t	0.844	0.752	-0.167	1.450	4.310	9.953	9.237		
	p-value	0.402	0.456	0.868	0.154	<0.001	<0.001	<0.001		
IL1RL1	Pearson's r	0.248	0.172	0.094	0.587	0.846	0.712	0.699	0.656	1.
	R Std Error	0.021	0.022	0.0230	0.015	0.006	0.011	0.011	0.013	
	t	1.684	1.151	0.621	4.750	10.431	6.660	6.421	5.711	
	p-value	0.099	0.256	0.537	<0.001	<0.001	<0.001	<0.001	<0.001	
IL6	Pearson's r	0.318	0.131	-0.062	0.373	0.752	0.544	0.647	0.652	0.692
	R Std Error	0.021	0.022	0.023	0.020	0.010	0.016	0.013	0.013	0.012
	t	2.206	0.870	-0.407	2.640	7.493	4.256	5.564	5.639	6.293
	p-value	0.032	0.388	0.685	0.011	<0.001	<0.001	<0.001	<0.001	<0.001

SUPPLEMENTARY TABLE 3

Correlation coefficient matrix for 10 genes from Figure 9A and C in a cohort of 7 normal controls.

		CCR4	IL18	CD74	CD86	AREG/AREGB-1	IL4R	BCL3	BCL6	IL1RL1	IL6
CCR4	Pearson's r	1.									
	<i>R Std Error</i>										
	<i>t</i>										
	<i>p-value</i>										
IL18	Pearson's r	-0.537	1.								
	<i>R Std Error</i>	0.142									
	<i>t</i>	-1.425									
	<i>p-value</i>	0.214									
CD74	Pearson's r	0.748	-0.310	1.							
	<i>R Std Error</i>	0.088	0.181								
	<i>t</i>	2.521	-0.729								
	<i>p-value</i>	0.053	0.499								
CD86	Pearson's r	0.503	0.271	0.275	1.						
	<i>R Std Error</i>	0.149	0.185	0.185							
	<i>t</i>	1.301	0.631	0.641							
	<i>p-value</i>	0.250	0.556	0.550							
AREG/AREGB-1	Pearson's r	0.032	0.050	0.258	-0.447	1.					
	<i>R Std Error</i>	0.200	0.199	0.187	0.160						
	<i>t</i>	0.072	0.113	0.596	-1.118						
	<i>p-value</i>	0.945	0.915	0.577	0.315						
IL4R	Pearson's r	0.431	-0.272	0.770	-0.059	0.421	1.				
	<i>R Std Error</i>	0.163	0.185	0.081	0.199	0.165					

	<i>t</i>	1.068	-0.631	2.701	-0.131	1.038				
	<i>p-value</i>	0.334	0.556	0.043	0.901	0.347				
BCL3	Pearson's r	0.001	0.114	0.585	-0.047	0.062	0.741	1.		
	<i>R Std Error</i>	0.200	0.197	0.132	0.200	0.199	0.090			
	<i>t</i>	0.003	0.256	1.612	-0.104	0.139	2.470			
	<i>p-value</i>	0.998	0.808	0.168	0.921	0.895	0.057			
BCL6	Pearson's r	-0.025	0.145	0.472	-0.337	0.456	0.323	0.554	1.	
	<i>R Std Error</i>	0.200	0.196	0.155	0.177	0.158	0.179	0.139		
	<i>t</i>	-0.056	0.327	1.198	-0.800	1.145	0.762	1.489		
	<i>p-value</i>	0.958	0.757	0.285	0.460	0.304	0.480	0.197		
IL1RL1	Pearson's r	-0.457	0.745	-0.468	0.260	-0.216	-0.171	0.096	-0.205	1.
	<i>R Std Error</i>	0.158	0.089	0.156	0.186	0.191	0.194	0.198	0.192	
	<i>t</i>	-1.149	2.499	-1.183	0.603	-0.495	-0.388	0.215	-0.467	
	<i>p-value</i>	0.303	0.055	0.290	0.573	0.642	0.714	0.838	0.660	
IL6	Pearson's r	0.411	-0.164	0.316	0.439	-0.573	0.302	0.419	-0.044	0.300
	<i>R Std Error</i>	0.166	0.195	0.180	0.161	0.134	0.182	0.165	0.200	0.182
	<i>t</i>	1.007	-0.371	0.746	1.094	-1.564	0.709	1.032	-0.098	0.703
	<i>p-value</i>	0.360	0.726	0.489	0.324	0.179	0.510	0.349	0.926	0.514

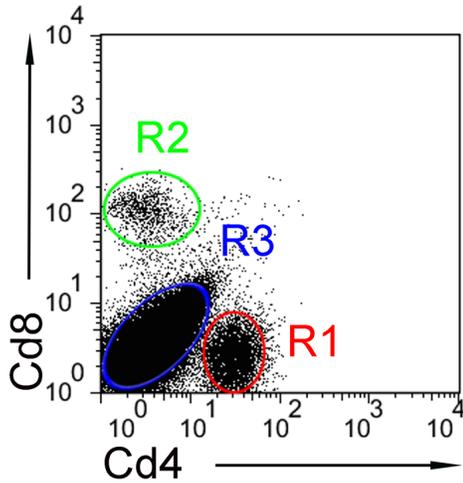
REFERENCES

1. Fontenot, J.D., Gavin, M.A., and Rudensky, A.Y. 2003. Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. *Nat Immunol* 4:330-336.
2. Morimoto, M., Zhao, A., Madden, K.B., Dawson, H., Finkelman, F.D., Mentink-Kane, M., Urban, J.F., Jr., Wynn, T.A., and Shea-Donohue, T. 2006. Functional importance of regional differences in localized gene expression of receptors for IL-13 in murine gut. *J Immunol* 176:491-495.
3. Shivakumar, P., Campbell, K.M., Sabla, G.E., Miethke, A., Tiao, G., McNeal, M.M., Ward, R.L., and Bezerra, J.A. 2004. Obstruction of extrahepatic bile ducts by lymphocytes is regulated by IFN-gamma in experimental biliary atresia. *J Clin Invest* 114:322-329.
4. Ramalingam, T.R., Pesce, J.T., Sheikh, F., Cheever, A.W., Mentink-Kane, M.M., Wilson, M.S., Stevens, S., Valenzuela, D.M., Murphy, A.J., Yancopoulos, G.D., et al. 2008. Unique functions of the type II interleukin 4 receptor identified in mice lacking the interleukin 13 receptor alpha1 chain. *Nat Immunol* 9:25-33.
5. Moisan, J., Grenningloh, R., Bettelli, E., Oukka, M., and Ho, I.C. 2007. Ets-1 is a negative regulator of Th17 differentiation. *J Exp Med* 204:2825-2835.
6. Villarino, A.V., Artis, D., Bezbradica, J.S., Miller, O., Saris, C.J., Joyce, S., and Hunter, C.A. 2008. IL-27R deficiency delays the onset of colitis and protects from helminth-induced pathology in a model of chronic IBD. *Int Immunol* 20:739-752.
7. Ishigame, H., Kakuta, S., Nagai, T., Kadoki, M., Nambu, A., Komiyama, Y., Fujikado, N., Tanahashi, Y., Akitsu, A., Kotaki, H., et al. 2009. Differential roles of interleukin-17A and -17F in host defense against mucoepithelial bacterial infection and allergic responses. *Immunity* 30:108-119.
8. Shivakumar, P., Sabla, G., Mohanty, S., McNeal, M., Ward, R., Stringer, K., Caldwell, C., Chougnet, C., and Bezerra, J.A. 2007. Effector role of neonatal hepatic CD8+ lymphocytes in epithelial injury and autoimmunity in experimental biliary atresia. *Gastroenterology* 133:268-277.

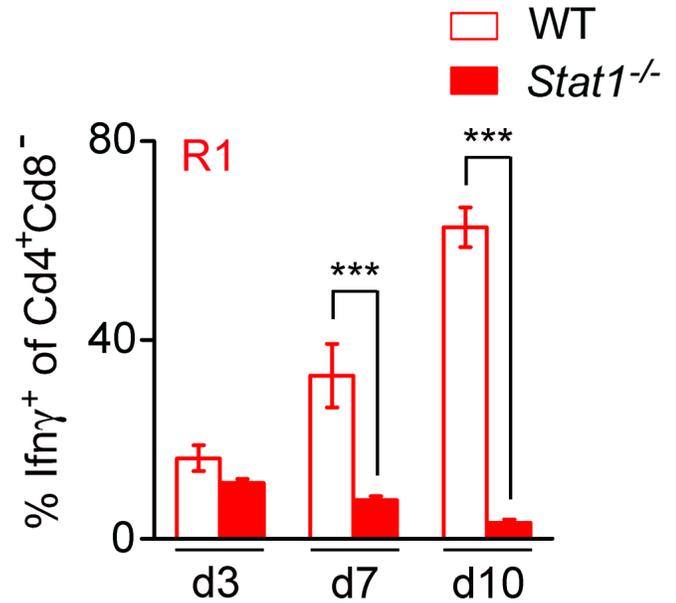
9. Kahnert, A., Seiler, P., Stein, M., Bandermann, S., Hahnke, K., Mollenkopf, H., and Kaufmann, S.H. 2006. Alternative activation deprives macrophages of a coordinated defense program to *Mycobacterium tuberculosis*. *Eur J Immunol* 36:631-647.
10. Sandler, N.G., Mentink-Kane, M.M., Cheever, A.W., and Wynn, T.A. 2003. Global gene expression profiles during acute pathogen-induced pulmonary inflammation reveal divergent roles for Th1 and Th2 responses in tissue repair. *J Immunol* 171:3655-3667.
11. Chiaramonte, M.G., Mentink-Kane, M., Jacobson, B.A., Cheever, A.W., Whitters, M.J., Goad, M.E., Wong, A., Collins, M., Donaldson, D.D., Grusby, M.J., et al. 2003. Regulation and function of the interleukin 13 receptor alpha 2 during a T helper cell type 2-dominant immune response. *J Exp Med* 197:687-701.
12. Jaruga, B., Hong, F., Sun, R., Radaeva, S., and Gao, B. 2003. Crucial role of IL-4/STAT6 in T cell-mediated hepatitis: up-regulating eotaxins and IL-5 and recruiting leukocytes. *J Immunol* 171:3233-3244.
13. Zheng, Y., Danilenko, D.M., Valdez, P., Kasman, I., Eastham-Anderson, J., Wu, J., and Ouyang, W. 2007. Interleukin-22, a T(H)17 cytokine, mediates IL-23-induced dermal inflammation and acanthosis. *Nature* 445:648-651.
14. Yu, C.R., Mahdi, R.M., Ebong, S., Vistica, B.P., Chen, J., Guo, Y., Gery, I., and Egwuagu, C.E. 2004. Cell proliferation and STAT6 pathways are negatively regulated in T cells by STAT1 and suppressors of cytokine signaling. *J Immunol* 173:737-746.

Supplementary Figure 1

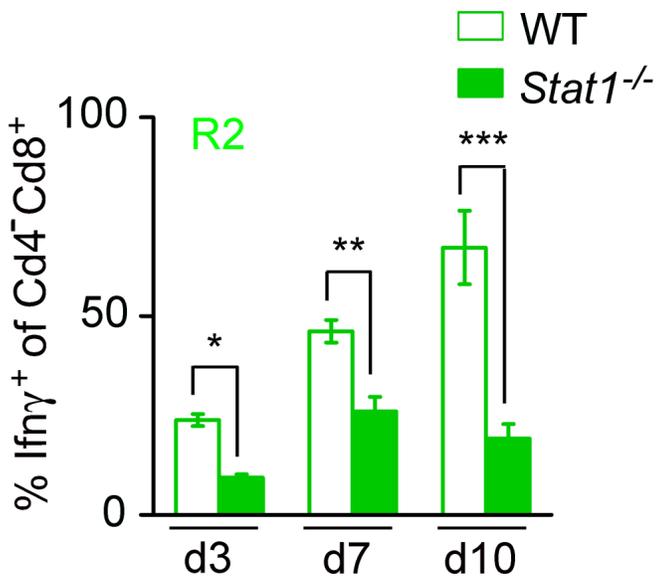
A



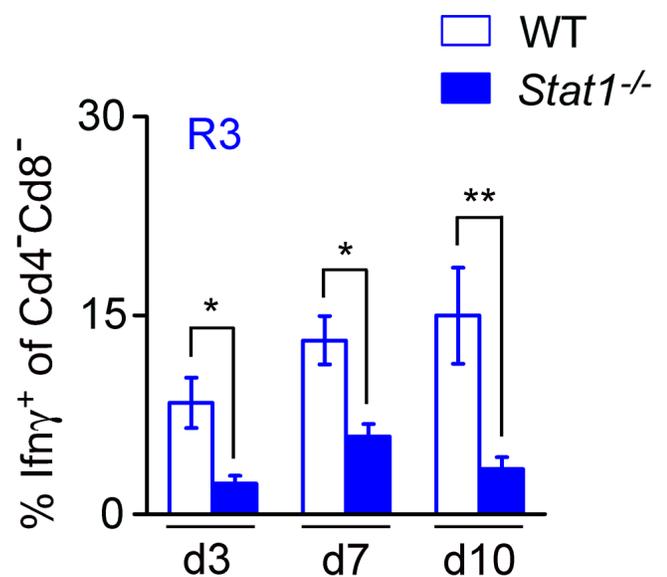
B



C

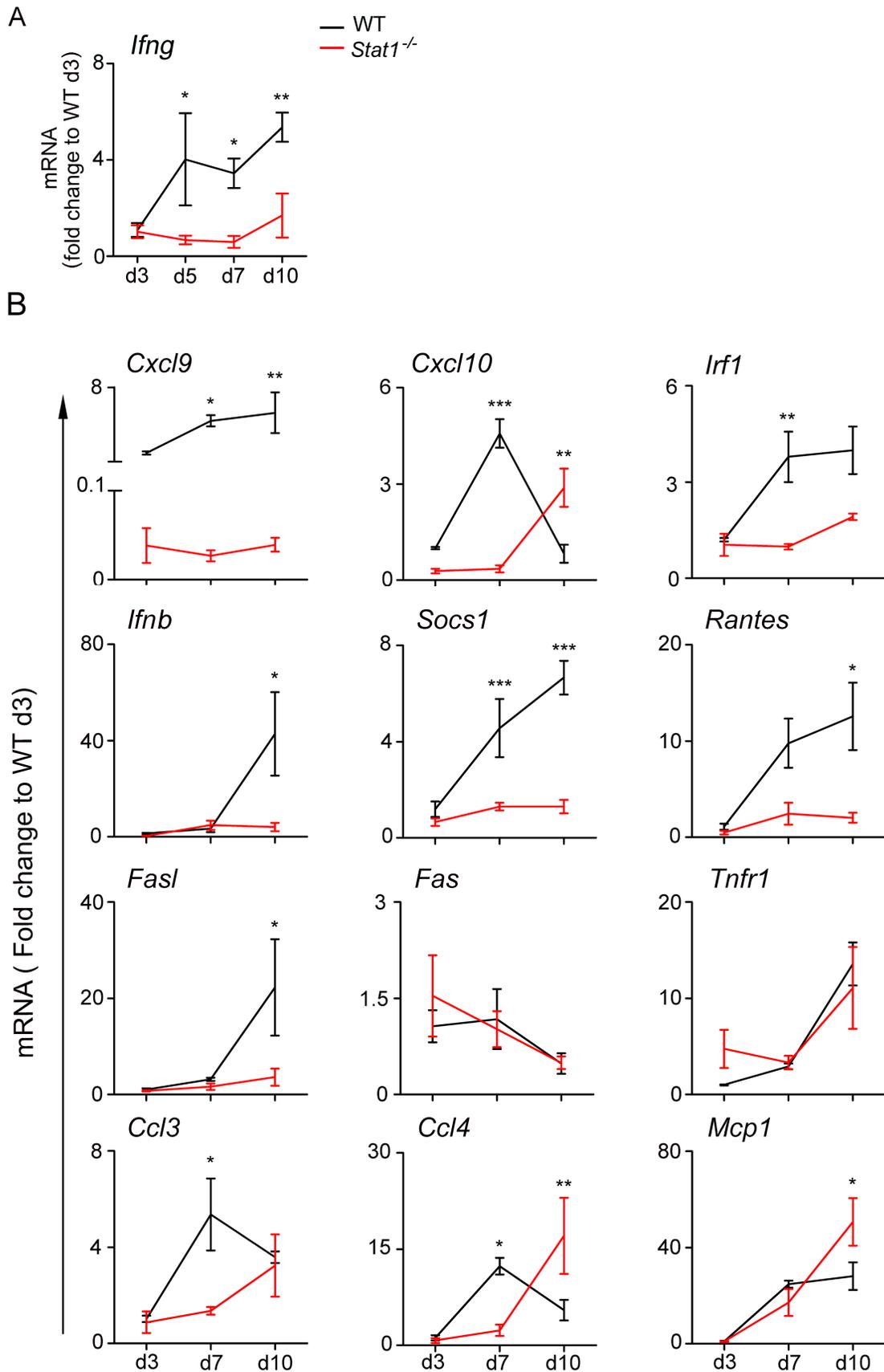


D



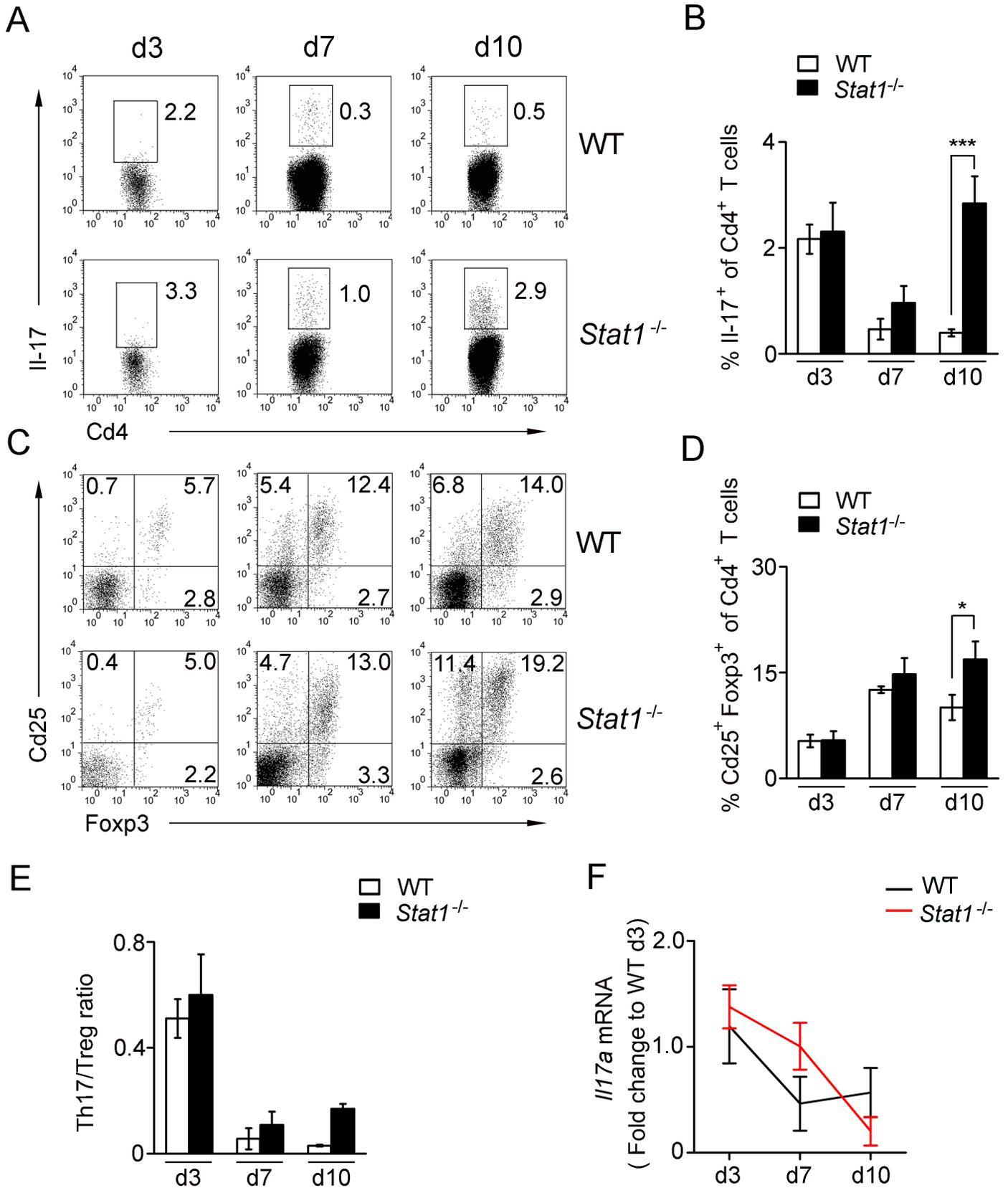
Decrease in Ifn γ production in *Stat1*^{-/-} mice after RRV. Gating strategy for Cd4⁺ (R1), Cd8⁺ (R2) and Cd4⁻Cd8⁻ (R3) hepatic mononuclear cells (A). The quantification of the percent of these cells expressing Ifn γ at days 3, 7 and 10 after RRV infection is higher in WT than *Stat1*^{-/-} mice. *P<0.05, **P<0.01, ***P<0.001. Note that panel B is also in Figure 3B, but it is shown here to demonstrate the gating strategy to quantify these cells.

Supplementary Figure 2



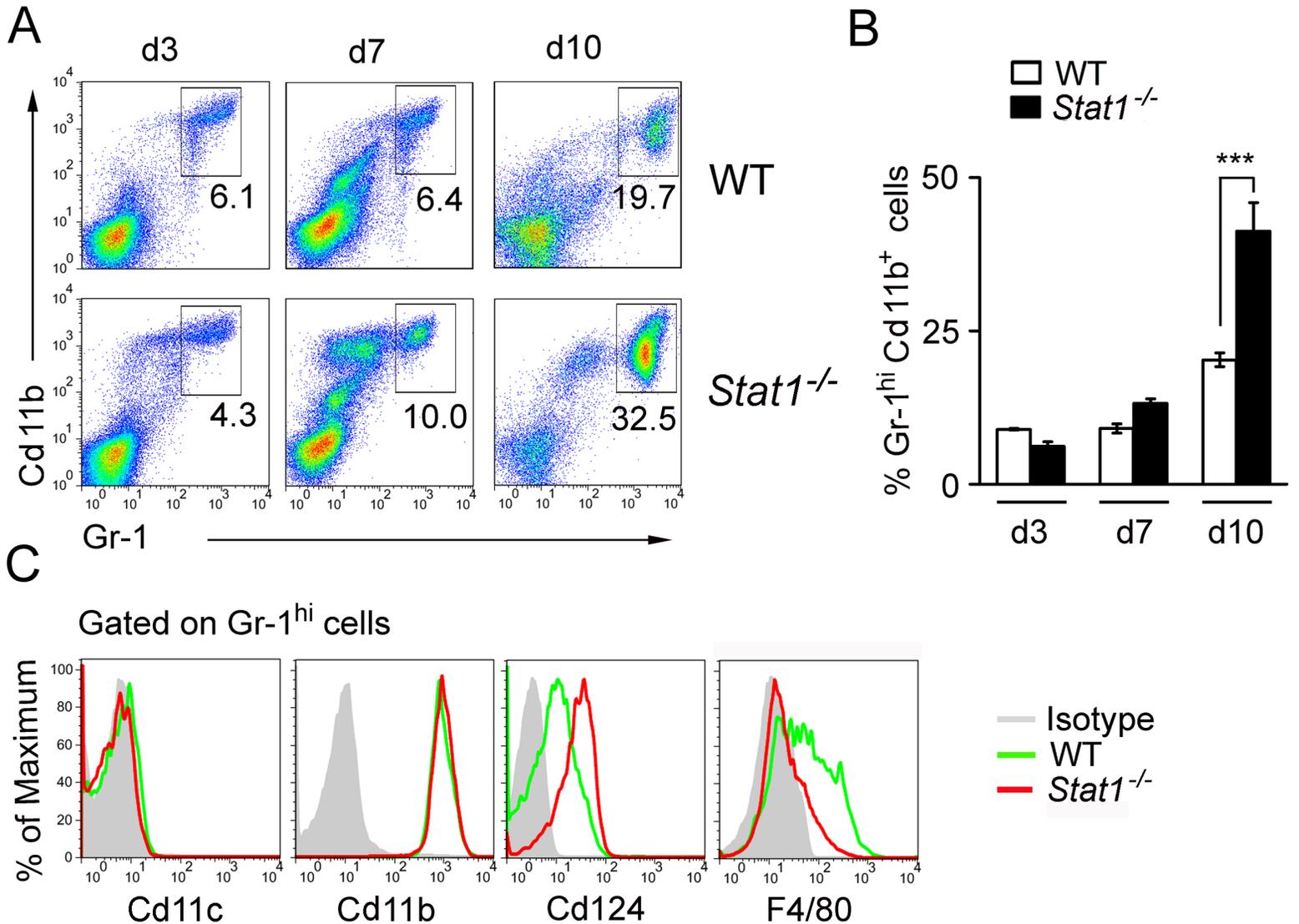
mRNA expression for *Ifng* and target genes. Individual graphs display the hepatic mRNA expression for *Ifng* (A) and *Ifng*-target genes (B) in WT and *Stat1*^{-/-} mice at different time points after RRV challenge. mRNA expression for most genes is decreased in *Stat1*^{-/-} livers when compared to WT. mRNA expression was first normalized to internal *Hprt* control and then normalized to the value from WT mice at day 3. N=3-4 livers per group and time point; *P<0.05, **P<0.01, ***P<0.001.

Supplementary Figure 3



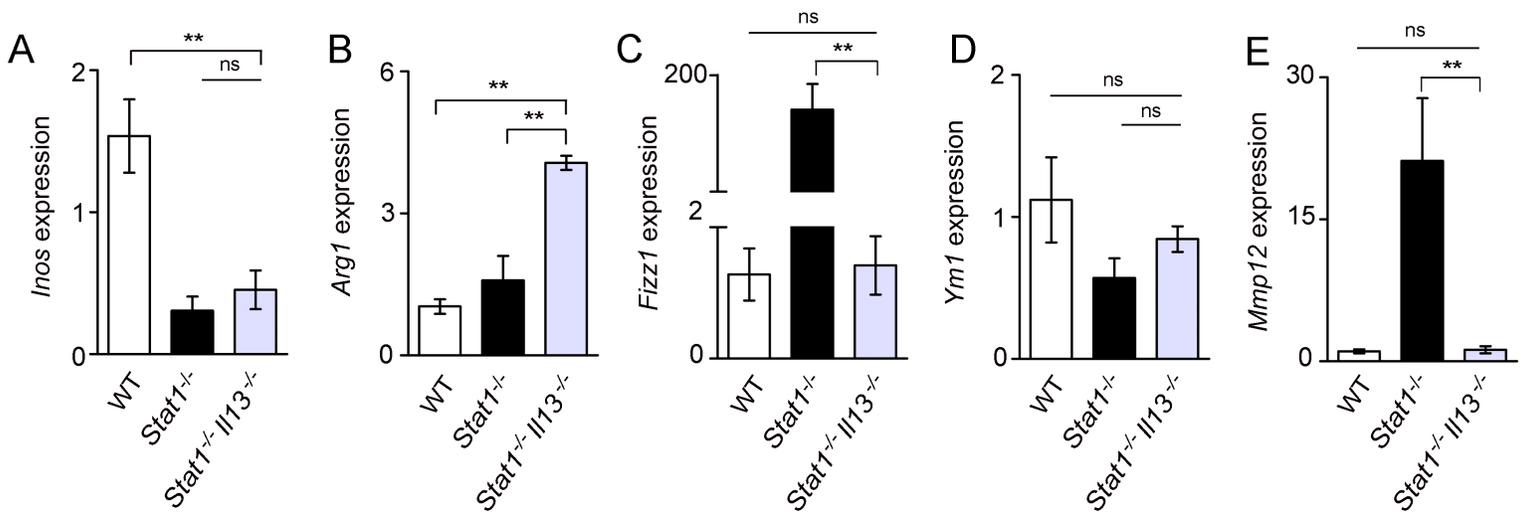
Th17 expression in *Stat1*^{-/-} mice after RRV. Flow cytometric analysis showing the expression of IL-17 (A,B) or Cd25⁺Foxp3⁺ (C,D) by hepatic Cd4⁺ cells in WT and *Stat1*^{-/-} mice at different time points after RRV. Panel E shows a Th17:Treg ratio below 1, and panel F shows a similar pattern of hepatic *Il17a* mRNA expression in both groups after RRV. Data are shown as representative dot plots (A,C) and mean±SEM (B,D); N=3 livers per group per time point; *P<0.05, ***P<0.001.

Supplementary Figure 4



Expression of $IL4R\alpha$ and F4/80 in Gr-1^{hi} cells in *Stat1*^{-/-} livers after RRV. Flow cytometry analyses show similar levels of Gr-1^{hi} cells in WT and *Stat1*^{-/-} livers 3 and 7 days after RRV, which increase in *Stat1*^{-/-} at day 10 (A,B). Expression of markers linked to the myeloid-derived suppressor cell (MDSC) phenotype shows an increased number of Gr-1^{hi} cells co-expressing Cd124 ($IL4R\alpha$) in *Stat1*^{-/-} livers but only few express F4/80⁺ 10 days after RRV (C). N=6-10 livers per group and time point; experiments repeated twice; ***P<0.001.

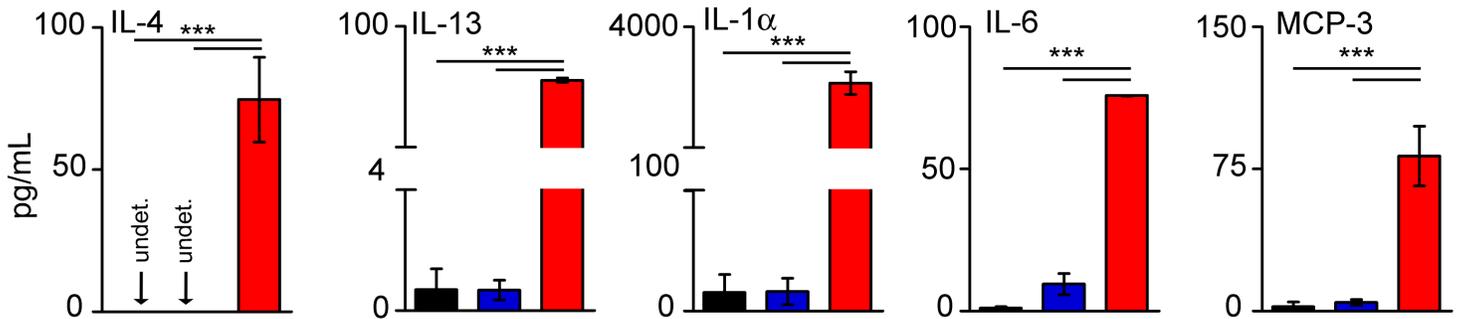
Supplementary Figure 5



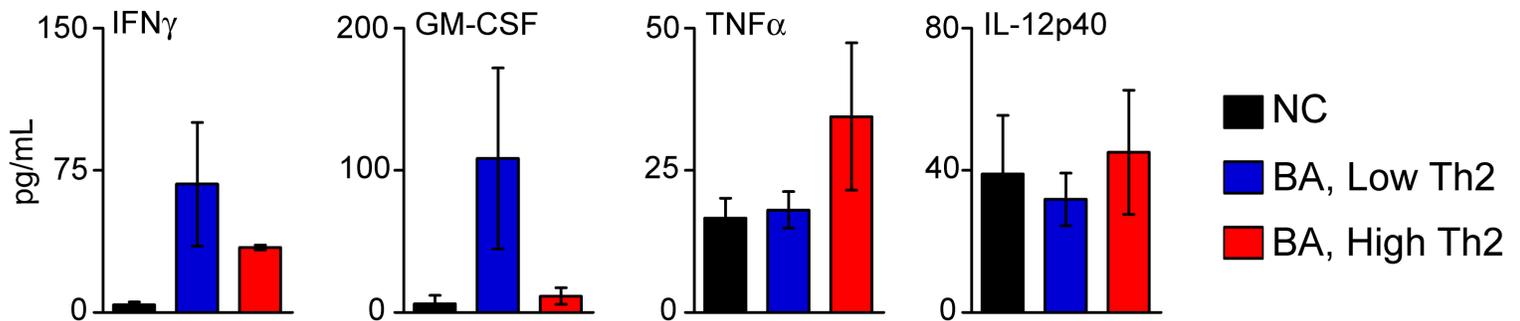
Decreased M2 activation after depletion of Il-13 in *Stat1*^{-/-} mice. Hepatic mRNA expression for M1 and M2 genes in WT, *Stat1*^{-/-} and *Stat1*^{-/-}/*Il13*^{-/-} mice 7 days after RRV. Values are expressed as a ratio to *Hprt* and then normalized to WT; N=3-5 livers per genotype; **P<0.01; ns=no significant difference.

Supplementary Figure 6

A



B



Serum levels of Th1 and Th2 cytokines/chemokines at diagnosis of biliary atresia. Mean (\pm S.E.) serum concentration of Th1 and Th2 cytokines/chemokines for infants at the time of diagnosis of biliary atresia (BA; age < 4 months) and age-matched normal infants (normal controls=NC). Data in panel A are shown in three groups: NC (N=3), BA Low Th2 group (N=9) and BA High Th2 group (N=2). ***P<0.001 (one-way ANOVA).