Supplemental data

MULTIVALENT BINDING OF FBP21-TANDEM-WW DOMAINS FOSTERS PROTEIN RECOGNITION IN THE PRE-SPLICEOSOME.

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SF3B3HNRPDassembly-relatedSFRS3SF3B4HNRPFassembly-relatedSFRS6SF3B5HNRPG (RBMX)DDX15SFRS8SF3B14HNRPHhPrp19SFRS15SMN-complexHNRPMHRMT1L5SKIV2L2HNRPQSF1SRPK1Gemin3HNRPRSRm300SR140Gemin5HNRPUL1SRPK2STIPPRMT5WBP11VBP11	snRNPD3 snRNPE snRNPF snRNPG U1 snRNP70 U1C U2 SF3A1 SF3A2 SF3A3 SF3B1 SF3B2 SF3B3 SF3B4 SF3B5 SF3B4 SF3B5 SF3B4 SF3B5 SF3B14 SF3B5 SF3B14 SMN-complex Gemin3 Gemin4 Gemin5	U4/U6.U5 U4/U6.U5 snRNP 110 kDa HSPC006 U5 U5 snRNP 100 kDa (DDX23) U5 snRNP 102 kDa U5 snRNP 116 kDa U5 snRNP 200 kDa U5 snRNP 200 kDa U5 snRNP 220 kDa (PrpF8) hnRNPs HNRPC HNRPD HNRPF HNRPG (RBMX) HNRPH HNRPM HNRPQ HNRPR HNRPU HNRPU	RBM17 RNPS1 RNA-processing CPSF25 CPSF6 CPSF7 DDX1 DDX3 DDX30 DDX30 DDX36 MATR3 MOV10 PAB1 PABPC4 assembly-related DDX15 hPrp19 HRMT1L5 SF1 SRm300 SRPK2	CHERP CREAP1 DDX5 DDX46 LSM11 NONO PTBP1 PUF60 RBM10 RBM14 RBM25 RBM39 RED SF1 SF4 SFPQ SFR33 SFRS6 SFRS8 SFRS75 SKIV2L2 SRPK1 SR140 SRM2 STIP LUDE5
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Figure S1. Spliceosomal proteins interacting with FBP21-tandem-WW as identified by SILAC-MS.

SILAC experiments were performed as described in experimental procedures. Molecules that belonged to the hundred most highly enriched proteins in these pulldown experiments are depicted in black while proteins which were found in the pulldown but were only moderately enriched are depicted in gray.



spotsequence (N to C)

Figure S2. Peptide SPOT experiments to classify relative affinities of the FBP21 single domains and tandem domain along the PRS containing SmB-tail region.

Peptides with an overlapping sequence of 15-18 amino acids derived from the SmB tail (aa 163-231) were spotted onto a cellulose membrane and incubated either with GST fusions of WW1, WW2 or tandem-WW. In case of the tandem-WW domains we observed moderate to strong binding for all peptides except peptide 4 (no binding) and 9 (weak binding). Considering that the spotted sequences contain an overlap of 10-13 amino acids we can exclude a single preferred binding site. Qualitatively, relative affinities for individual sequences of the SmB-tail are comparable for WW1, WW2 and tandem-WW indicating that all three constructs exert similar binding specificity (control motifs: B/binder; N/non-binder).



Figure S3. Substitution analysis of a bivalent SmB ligand in interaction with FBP21tandem-WW by SPOT analysis.

Single amino acid replacements of each residue of a shortened SmB-2 peptide (SmB aa 218-231) were probed by GST-FBP21-tandem-WW as described in the experimental procedures. Most strikingly, substitution of the arginine leads to a prominent reduction in signal intensity indicating a reduced affinity (red horizontal mark). On the other hand the binding strength could be enhanced by introducing an additional arginine residue at different positions (red vertical mark). The importance of the positive charge is underlined by the finding, that the introduction of a negative charge (aspartic acid, glutamic acid) within the ligand reduces the affinity (green mark).



MRPPMGPPMGIPPGRGTPMGMPPPGMRPPPGMRGLL

Figure S4. Chemical shift perturbations and K_D values (ITC) of the FBP21-tandem-WW domains upon binding SmB ligands with a different number of PRS. SmB-4 was divided into four non-overlapping peptides (SmB-1A-D). Affinities were measured by ITC and KD values and stoichiometries determined when possible . The addition of three residues ("MRP") at the N-terminus of SmB-1D enhances the affinity by a factor of two as shown for SmB-1. The stoichiometry is approximately two arguing that the two WW domains in the tandem-WW construct each interact with an individual peptide. SmB-2 comprises two PRS motifs and allows for a bivalent interaction (N=1) while SmB-4 possesses four potential PRM motifs (see also Figure 2 in the main text). The higher ligand valency in SmB-4 leads to an additional affinity enahcnement while the stoichiometry remains N=1. Residues affected by ligand binding are mainly located within the three beta-sheets of WW1 and WW2 and are part of the known interface for monovalent WW:PRS interactions.

SmB-4: MRPPMGPPMGIPPGRGTPMGMPPPGMRPPPGMRGLL



Figure S5. SPOT analysis of truncation experiment of bivalent SmB ligands derived from aa 195-231 of SmB. Bivalent SmB peptides were shortened by individual amino acids starting from either the N or C terminus and incubated with the GST-FBP21-tandem-WW domain. The truncation of one PRS motif leads to a decreased signal intensity implicating a loss of affinity based on the transition from a bivalent to a monovalent interaction. A similar pattern was observed in a substitution analysis where peptide residues where replaced by alanine-serine (Ala₄Ser) stretches (data not shown). Positive and negative controls are shown below the dotted line (from top to bottom: non-binder, non-binder, binder).



Figure S6. Chemical shift perturbations of the FBP21-tandem-WW domains upon binding ligands with a different number of PRS.

Figure S6 shows a mapping of weighted chemical shifts for NH backbone resonances after saturation with SmB (1, 2 or 4 motifs) and SF3B4 (1 or 2 motifs) ligands. Residues affected by ligand binding are mainly located within the three beta-sheets of WW1 and WW2 and are part of the known interface for monovalent WW:PRS interactions. Only subtle differences are observed between ligands of different valency.



Figure S7. ¹⁵N Relaxation rates for the FBP21-tandem-WW domain in complex with SmB-2 (left) and SmB-4 (right). ¹⁵N-¹H R1 and R2 rates were measured at 750 MHz indicate a slightly higher flexibility and mobility for the linker region compared to the WW-domains in both complexes. Correlation times of the tandem-WW domain in complex with SmB-2 (8 ± 0.3 ns) and SmB-4 (9 ± 0 ns) were calculated from T1/T2 ratios and are in agreement with the molecular weight of a 1:1 binding mode in both cases (tandem-WW + SmB-2: 11 kDa, tandem-WW + SmB-4: 12.5 kDa).