

ONLINE SUPPLEMENTAL MATERIALS

Investigation of Unanticipated Alkylation at the N(π) Position of a Histidyl Residue Under Mitsunobu Conditions and Synthesis of New Orthogonally Protected Histidine Analogues

Wenjian Qian, Fa Liu[§] and Terrence R. Burke, Jr.*

Chemical Biology Laboratory, Molecular Discovery Program, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Frederick, MD 21702, USA

*Corresponding author:

Terrence R. Burke, Jr., Ph.D.

National Cancer Institute

National Institutes of Health

Building 376 Boyles St., NCI-Frederick

Frederick, MD 21702

U. S .A.

Phone: (301) 846-5906; Fax: (301) 846-6033

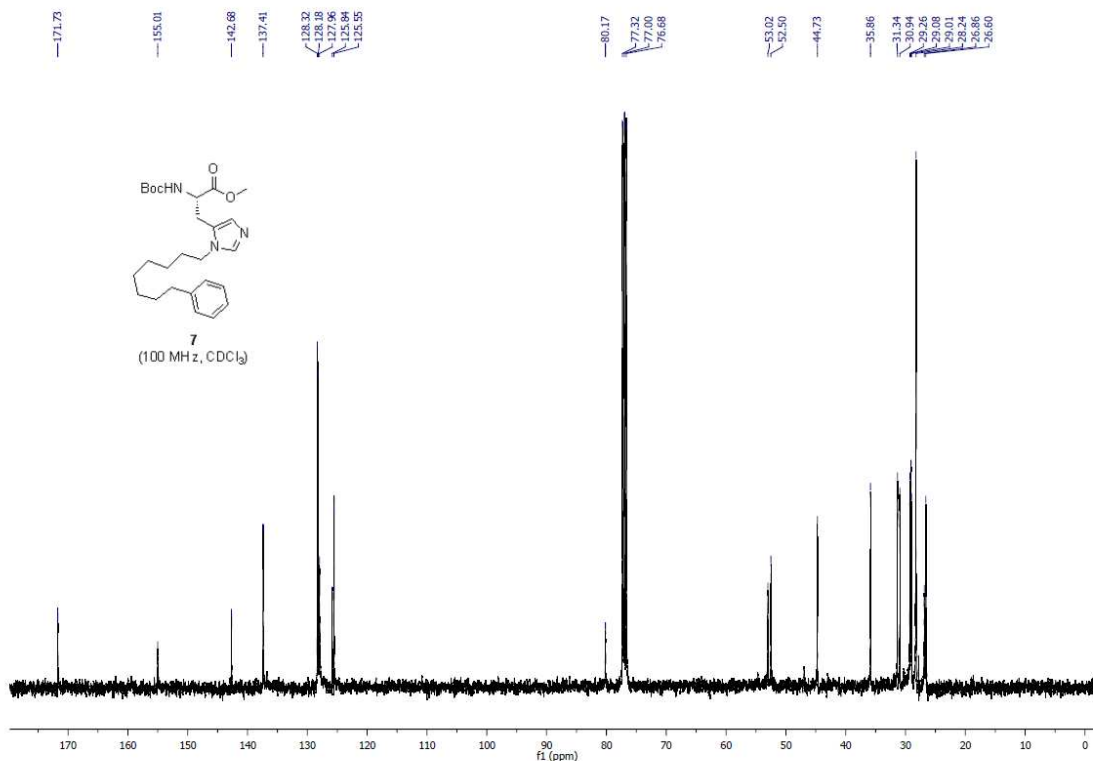
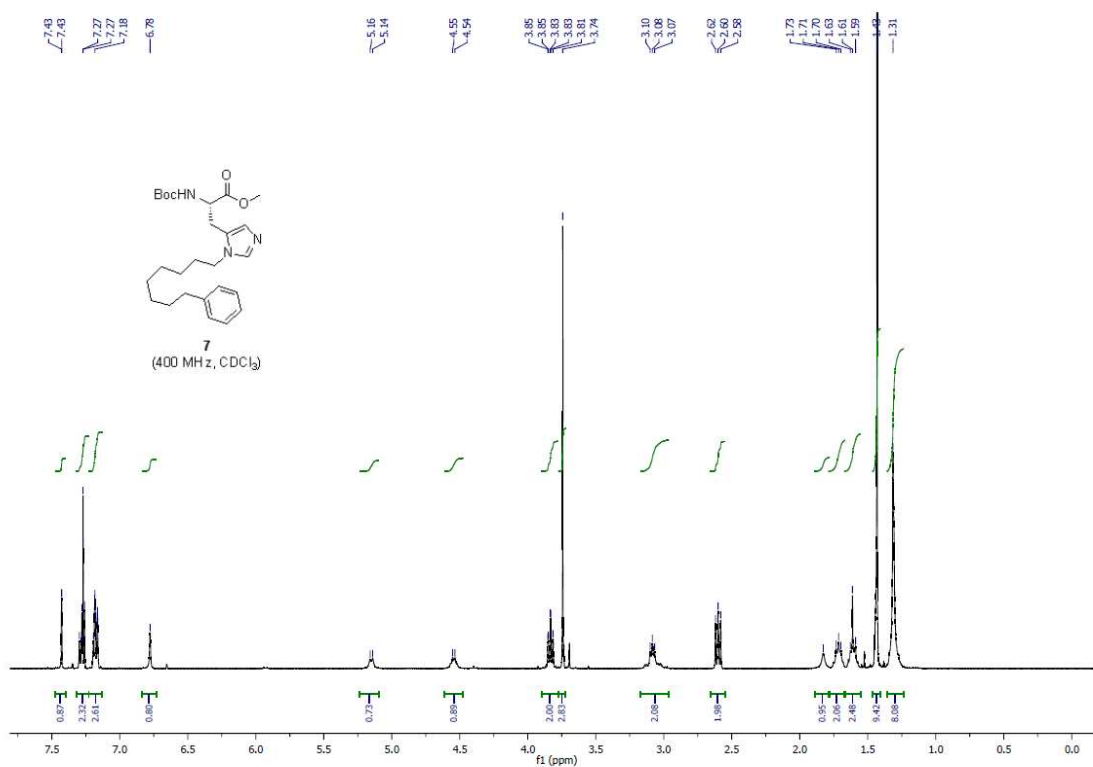
E-mail: tburke@helix.nih.gov

[§]Currently with Lilly Research Laboratories, Indianapolis, IN 46285

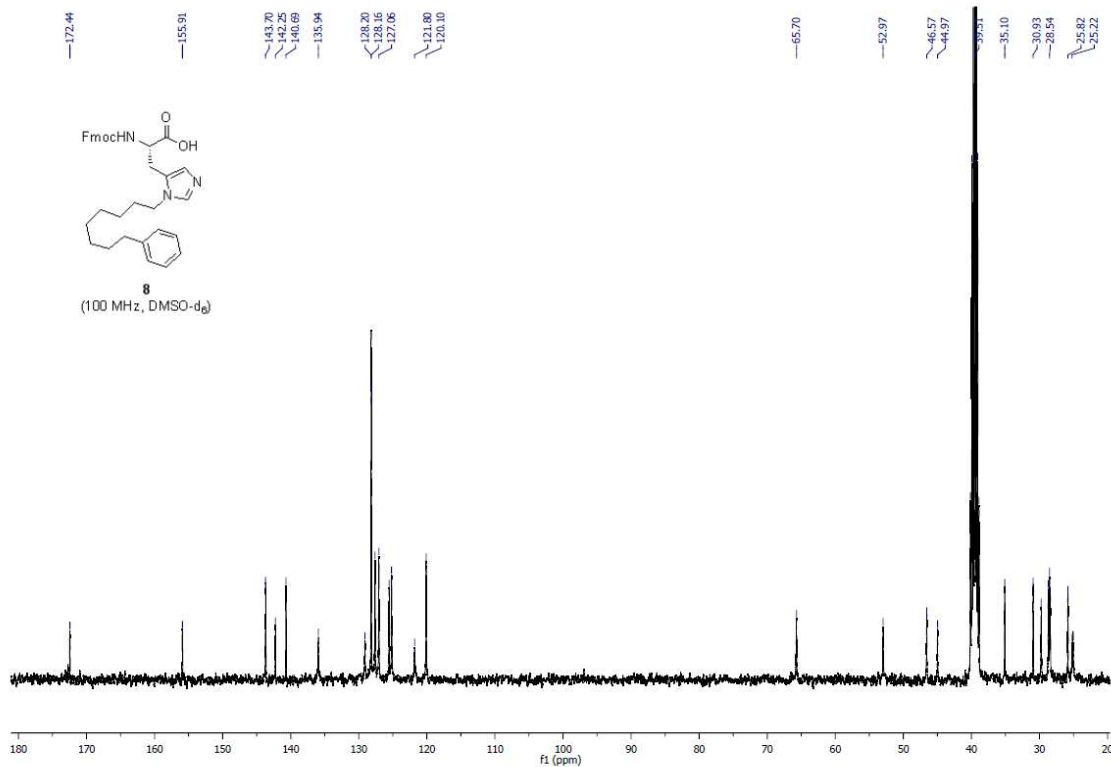
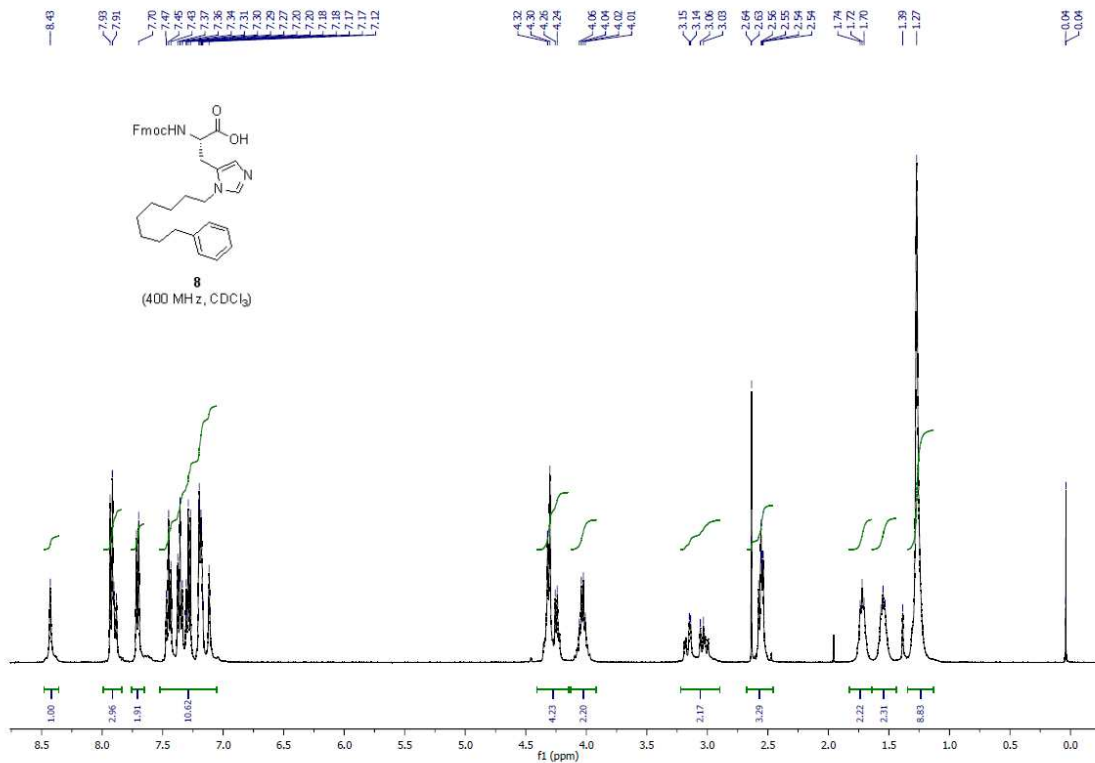
Contents

¹H and ¹³C NMR spectra of **7**, **8**, **9**, and **11** and 1D NOESY assignment of N(π)-alkylation in **11**, HPLC charts of peptide **2**, **3**, **4**, **5**, **13**.

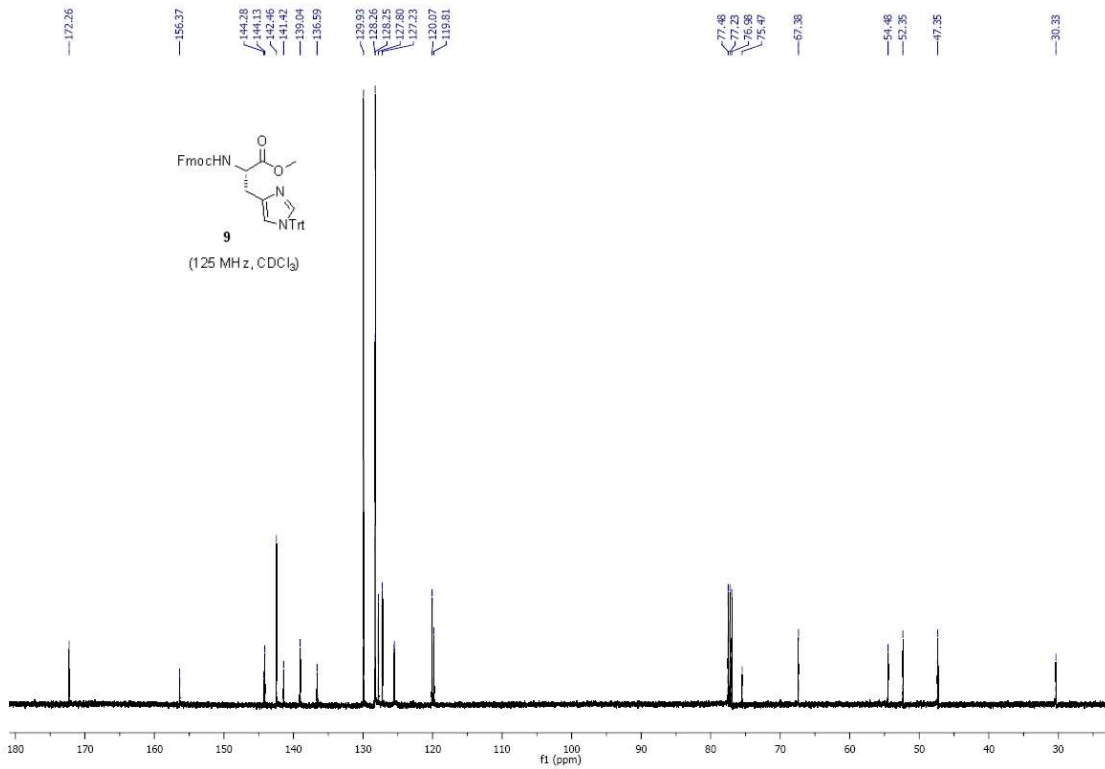
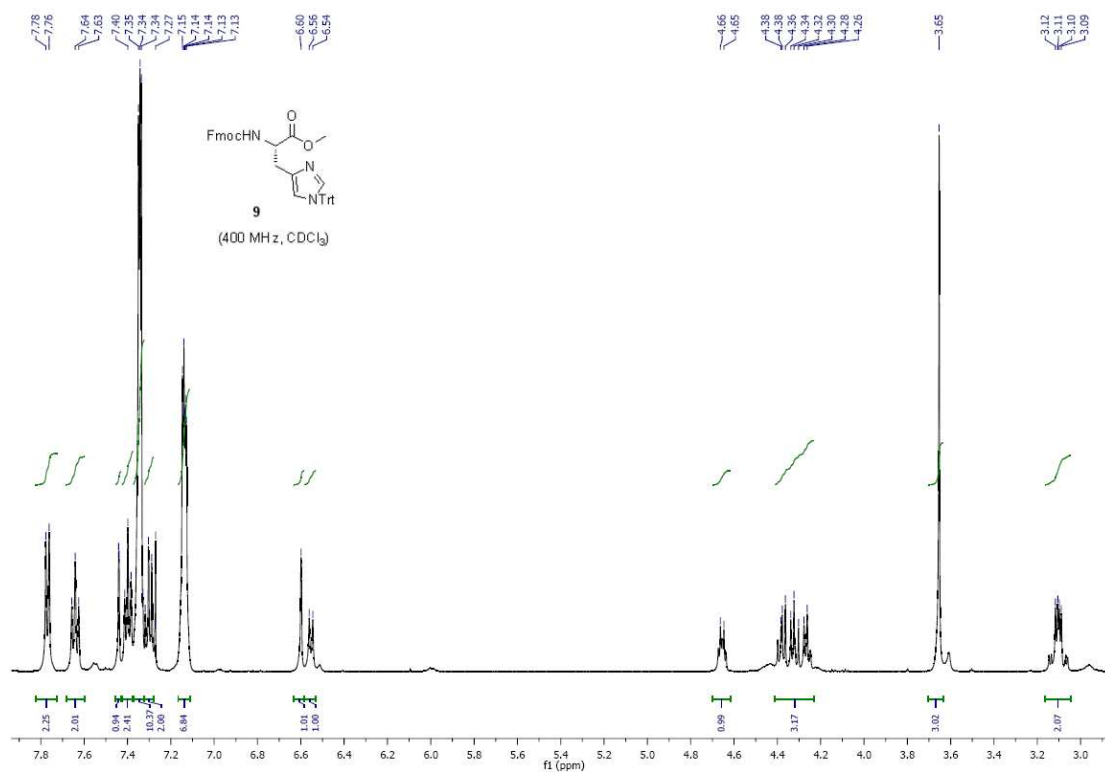
^1H and ^{13}C NMR spectra of 7



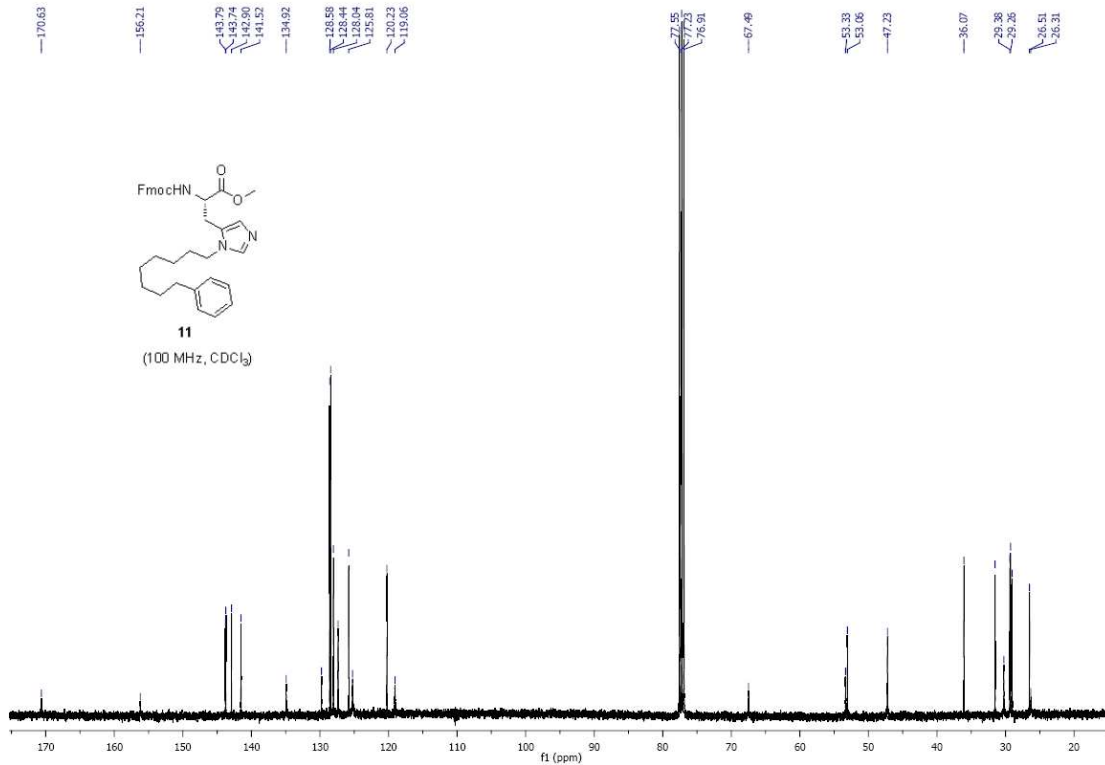
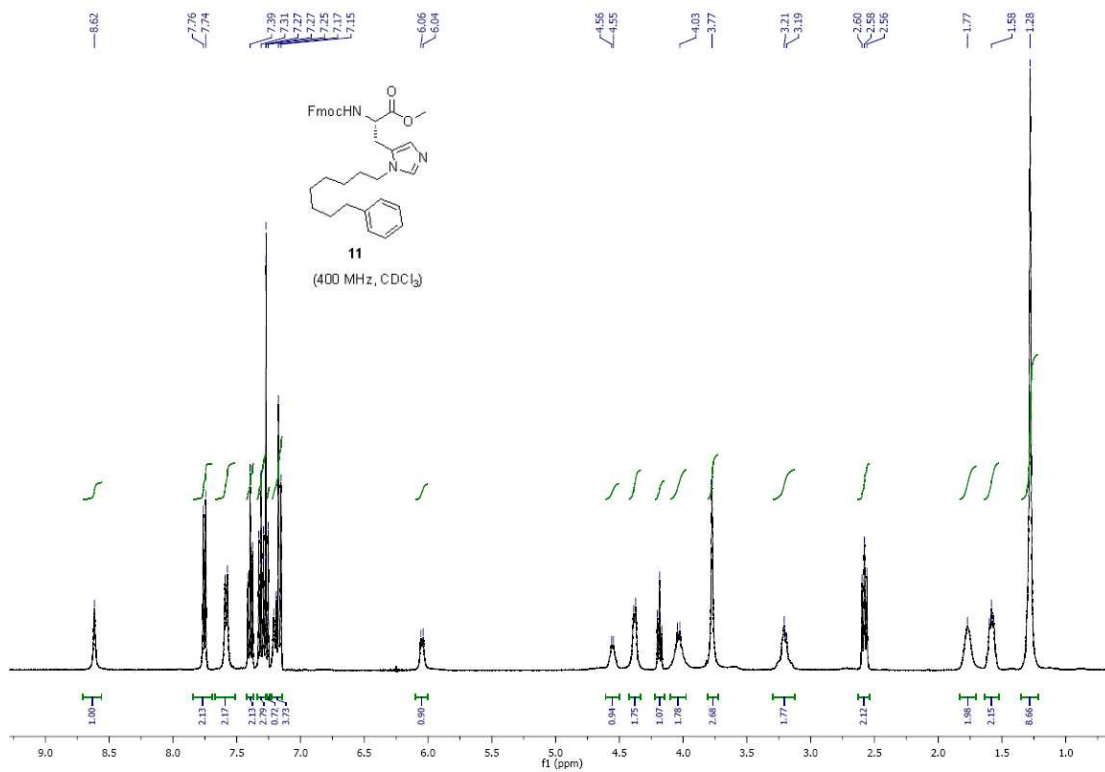
¹H and ¹³C NMR spectra of 8



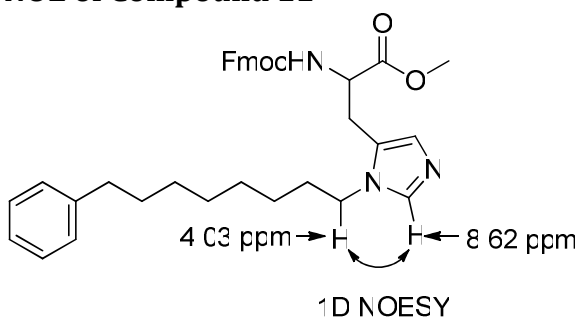
¹H and ¹³C NMR spectra of 9



¹H and ¹³C NMR spectra of 11



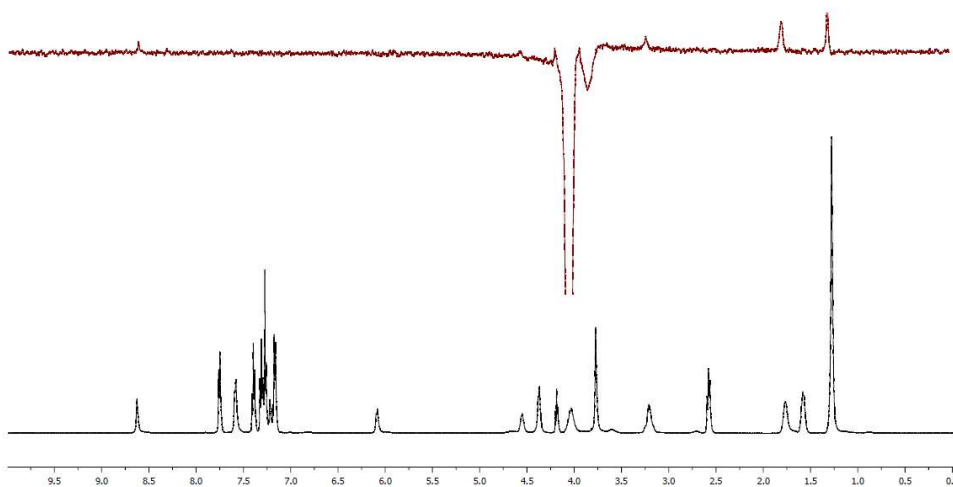
NOE of Compound 11



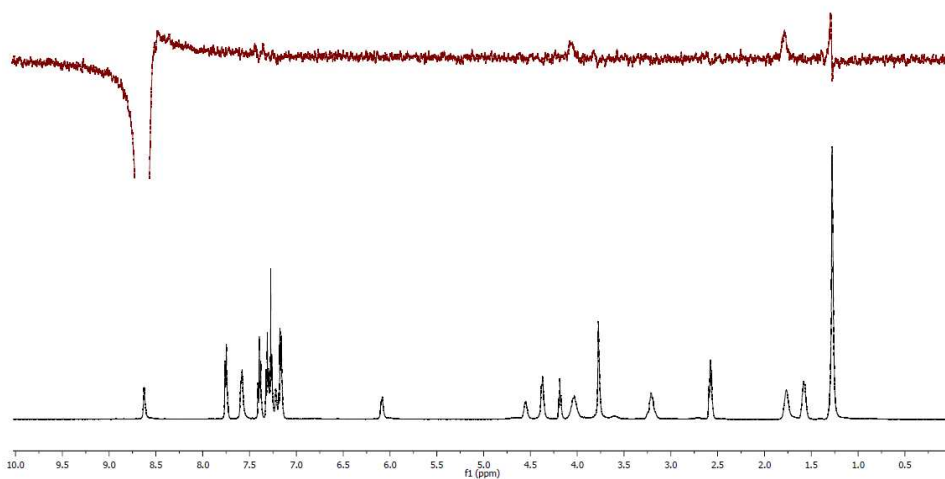
1D NOESY

500 MHz, relax. Delay 1.0 sec, mixing time 500 ms

Selective band center: 4.03 ppm



Selective band center: 8.62 ppm



HPLC Charts for Peptide 2, 3, 4, 5, 13

Table 1. MS and HPLC retention time

No	Expect (M + H) ⁺	Observed (M + H) ⁺	Retention time/ min	HPLC method ^a
2	863.4	863.4	25.2	II
3	863.4	863.4	24.3	II
4	1051.6	1051.5	29.8	I
5	1037.6	1037.4	29.2	I
13	861.5	861.4	23.3	I

^a HPLC Method:

$\lambda = 220$ nm, flow rate: 10 mL/min

Solvent A: H₂O, 0.1% TFA; Solvent B: MeCN, 0.1% TFA

I:

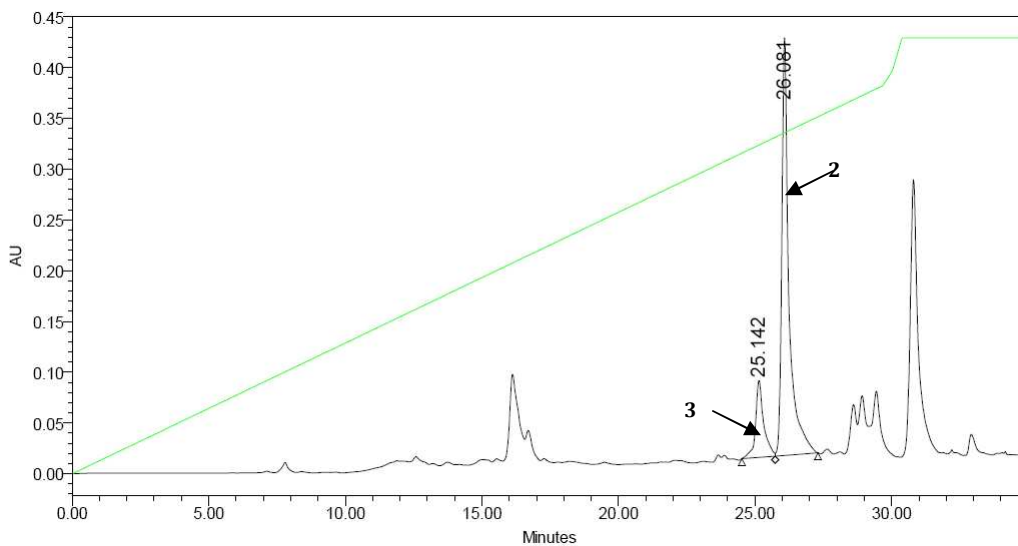
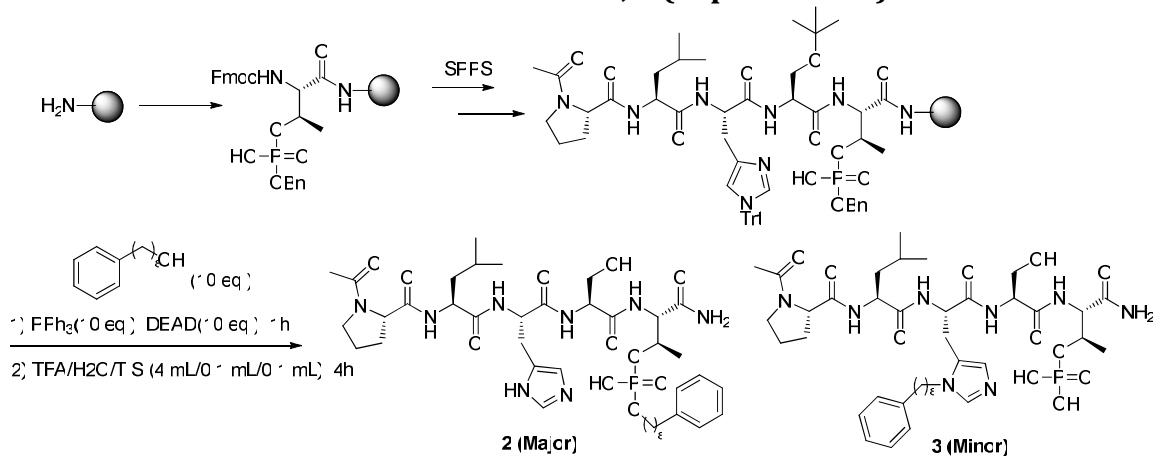
Time	0	30	30.1	35
B%	0	100	100	100

II:

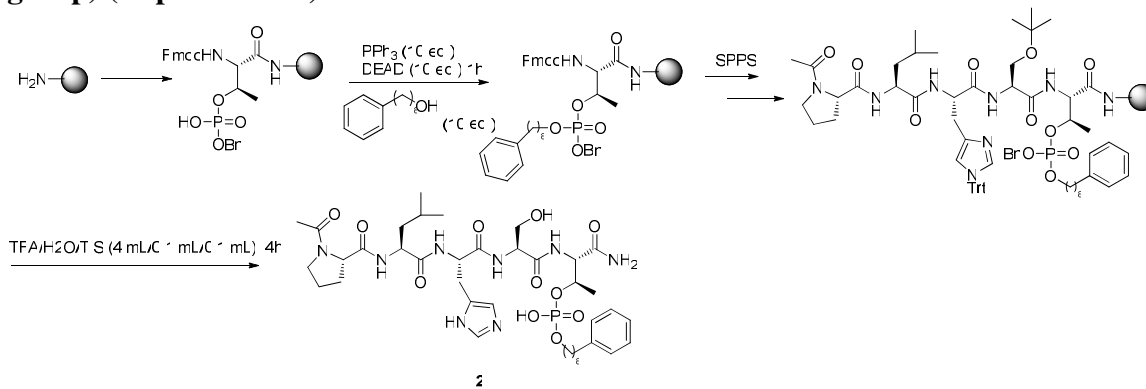
Time	0	30	30.1	35
B%	0	90	100	100

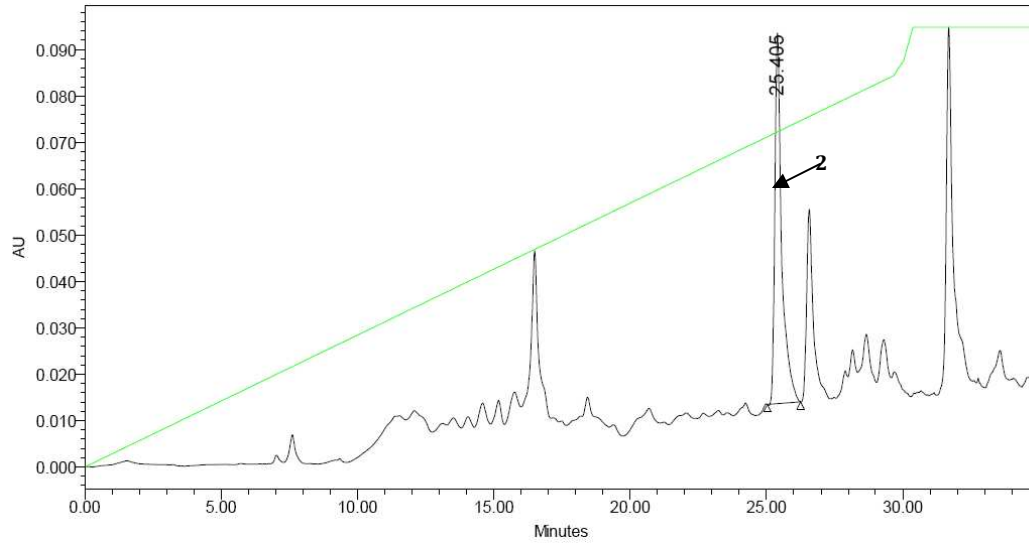
HPLC spectra:

1. on-resin mistunobu reaction to afford 2, 3 (Experiment A)

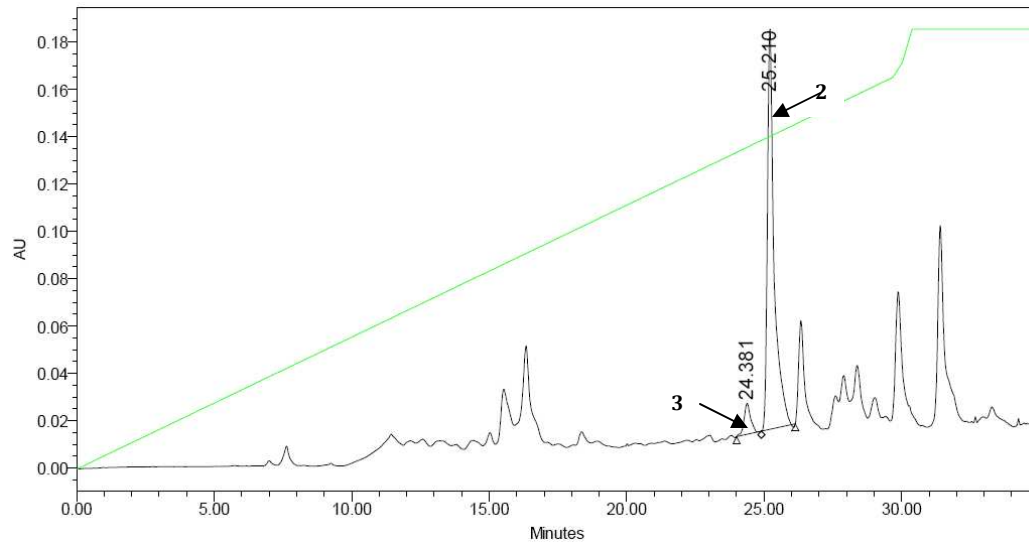


2. Test of Mechanism I (Intramolecular alkyl transfer of the phosphoryl ester group) (Experiment B)



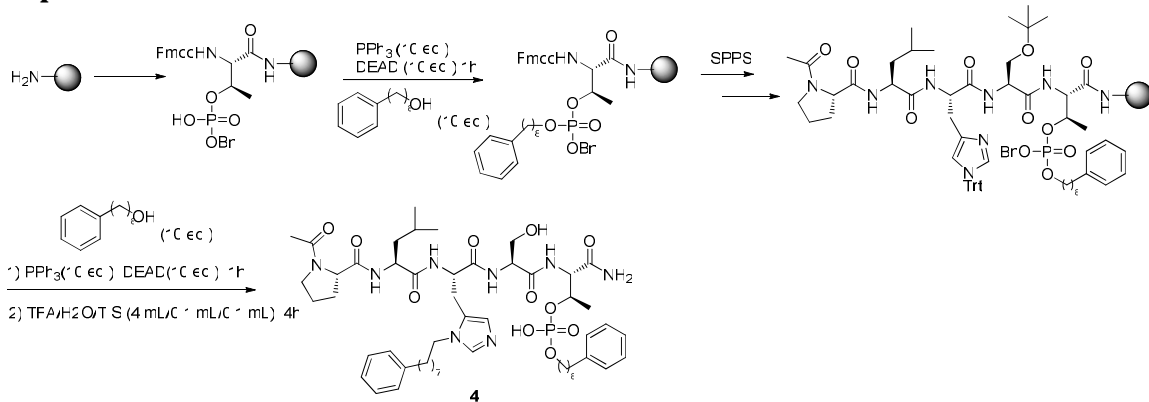


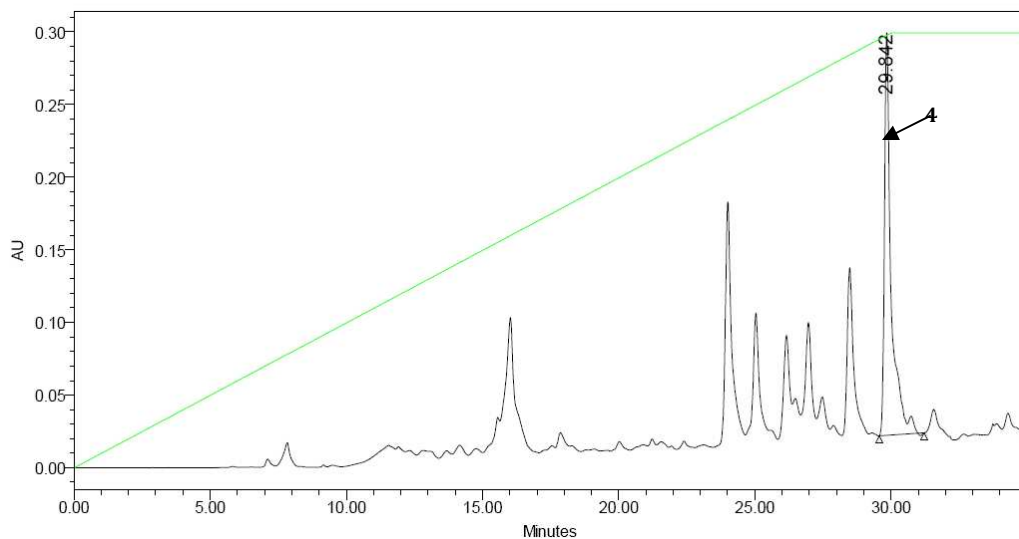
3. Mixture Test (Experiment A mixed with Experiment B)



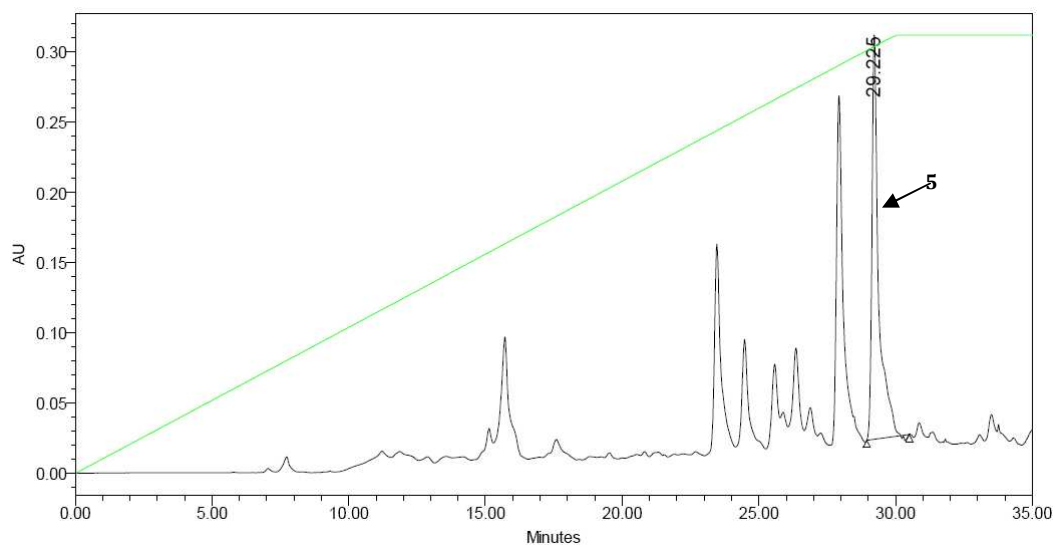
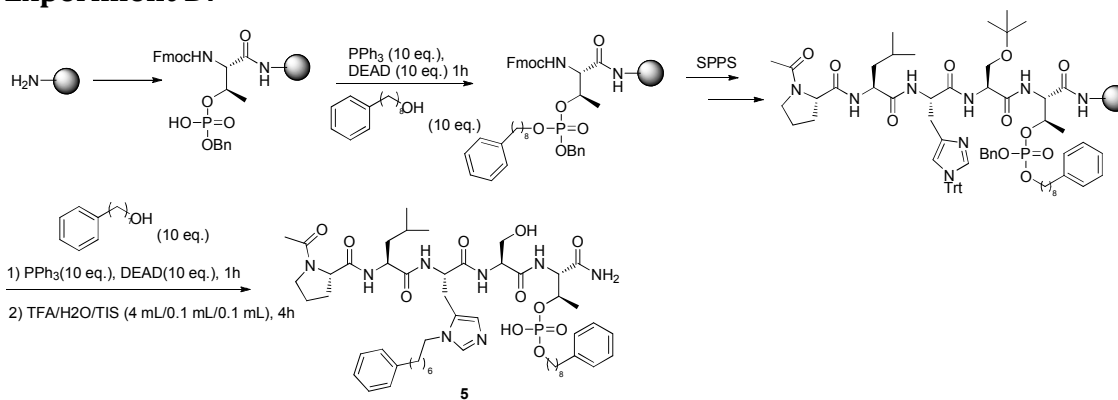
4. Test of Mechanism II (Experiment C and D)

Experiment C:

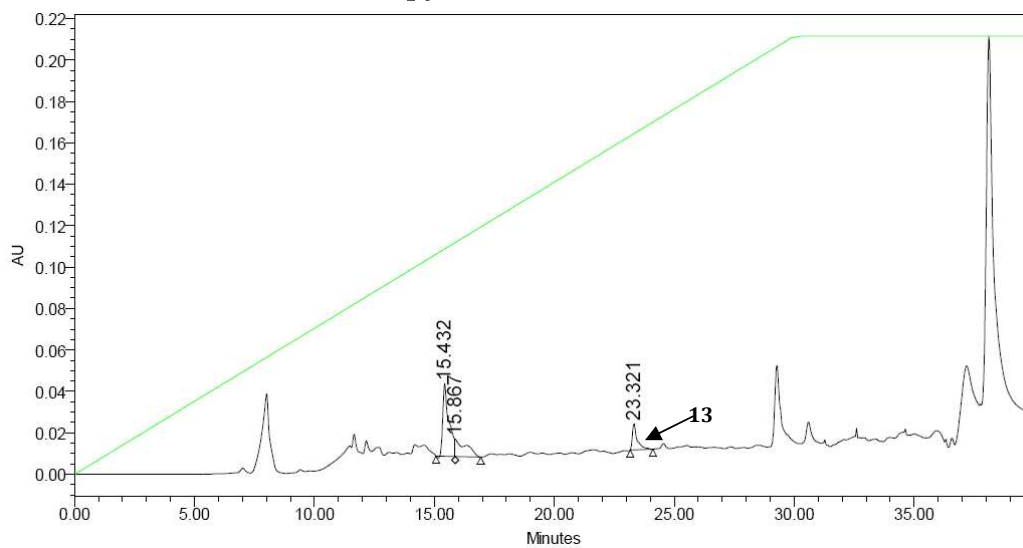
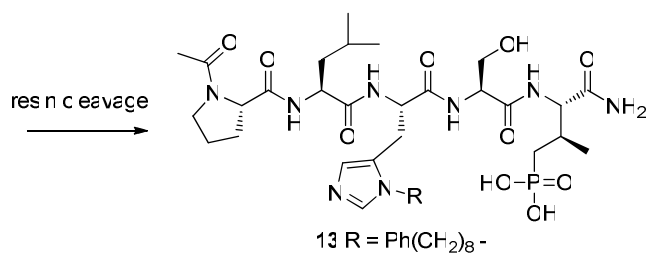
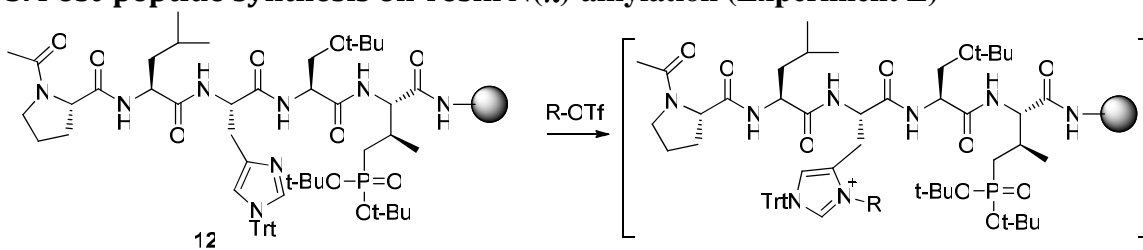




Experiment D:



5. Post-peptide synthesis on-resin N(π)-alkylation (Experiment E)



Purity of Peptide 2, 3, 4, 5, 13

Table 2. Purity of Peptides

No	2	3	4	5	13
Purity	97%	96%	98%	98%	100%

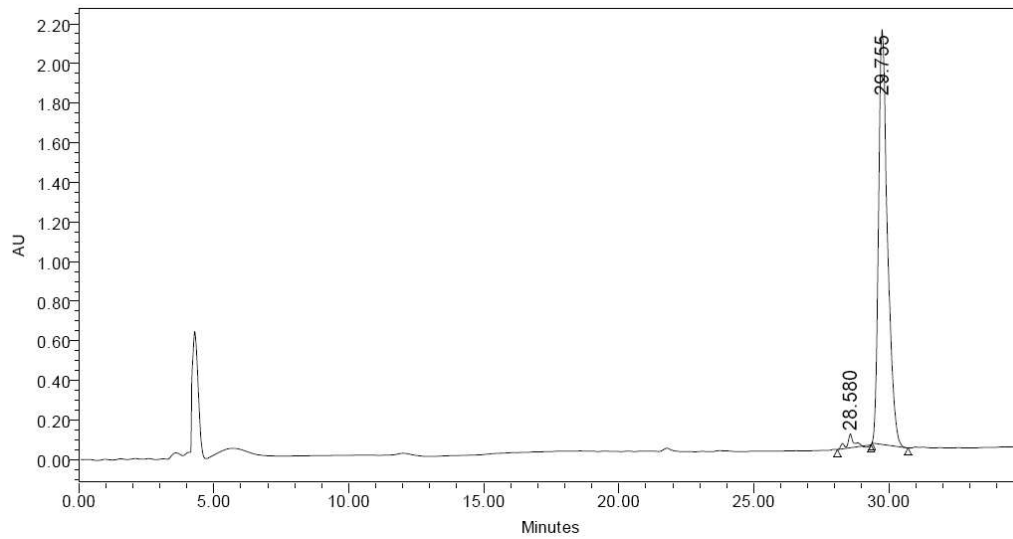
HPLC Method:

$\lambda = 220$ nm, flow rate: 1.0 mL/min

Solvent A: H₂O, 0.1% TFA; Solvent B: MeCN, 0.1% TFA

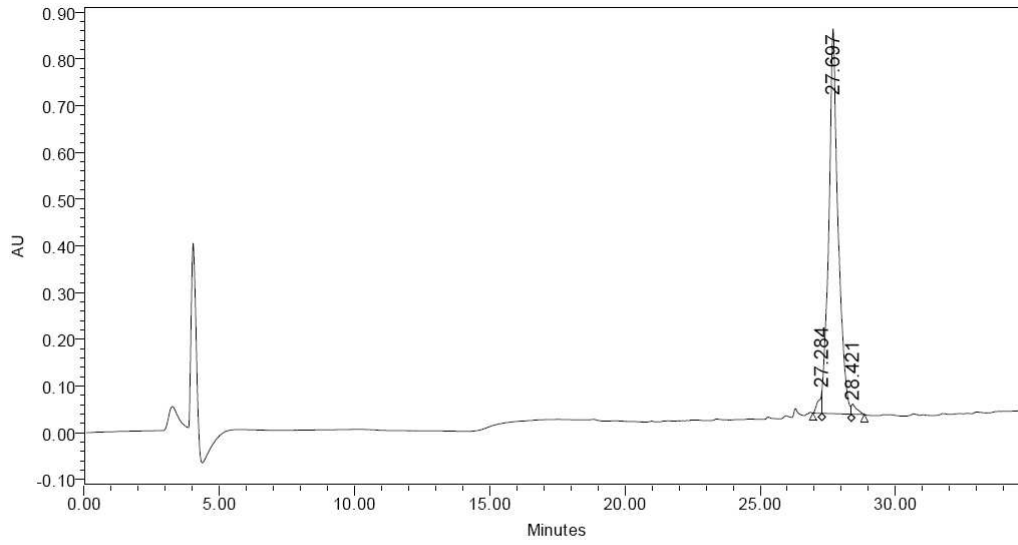
Time	0	30	30.1	35
B%	0	100	100	100

Peptide 2:



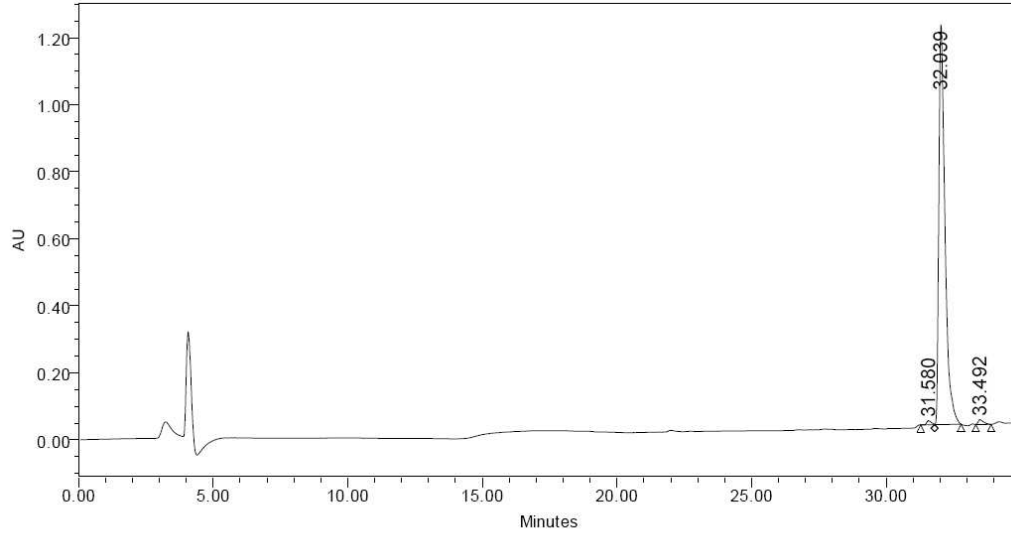
	RT	Area	% Area	Height
1	28.580	1298495	2.62	69683
2	29.755	48329081	97.38	2088105

Peptide 3



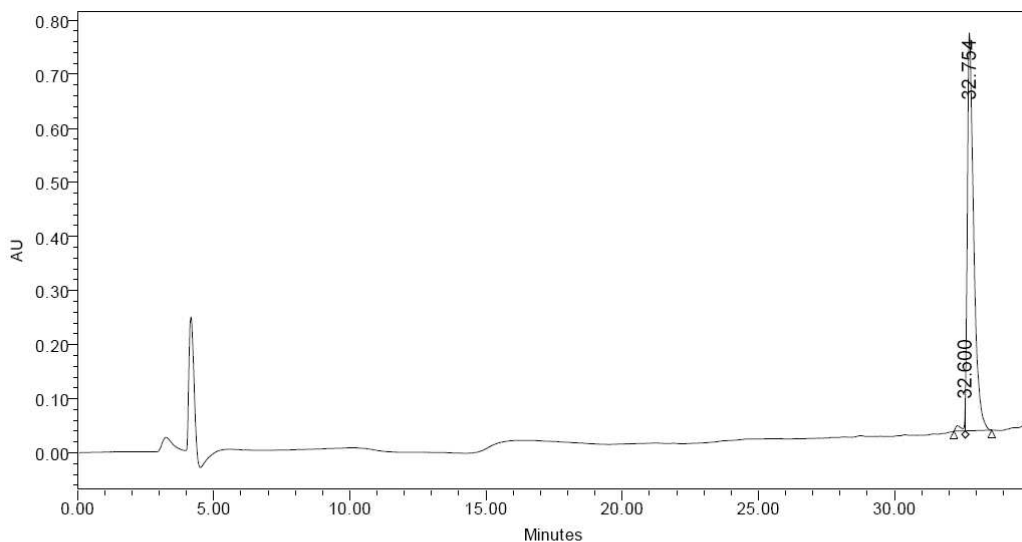
	RT	Area	% Area	Height
1	27.284	403588	1.98	47621
2	27.697	19721340	96.58	823302
3	28.421	293918	1.44	21107

Peptide 4



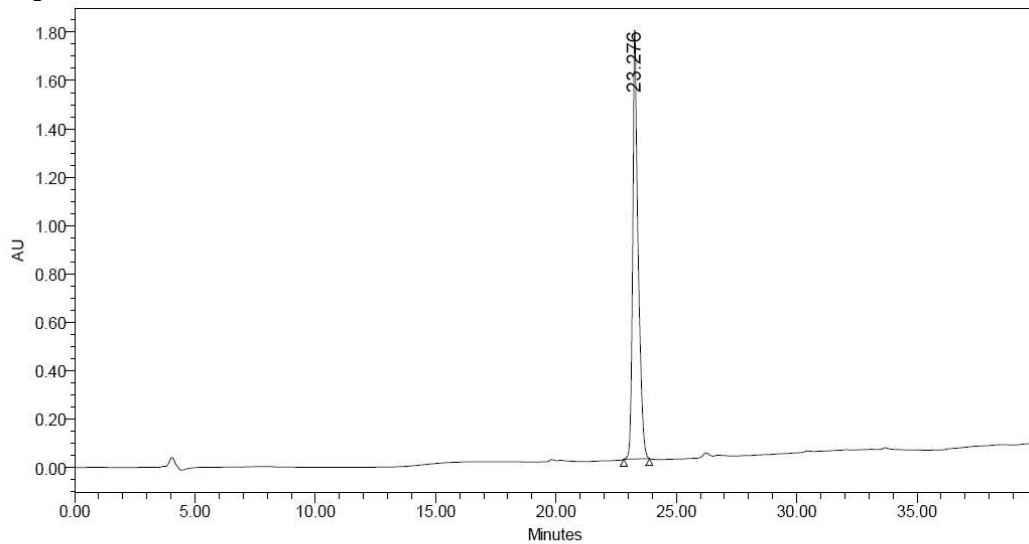
	RT	Area	% Area	Height
1	31.580	149603	0.78	11971
2	32.039	18959805	98.22	1195612
3	33.492	193813	1.00	14291

Peptide 5



	RT	Area	% Area	Height
1	32.600	201622	1.66	51715
2	32.754	11917745	98.34	736127

Peptide 13



	RT	Area	% Area	Height
1	23.276	27639992	100.00	1781543