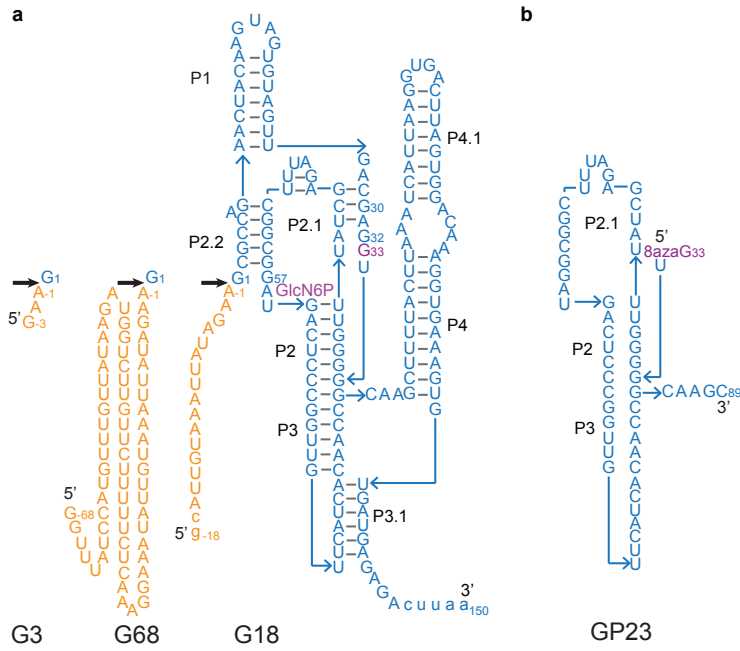


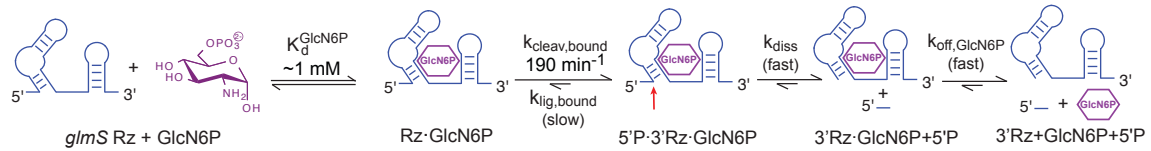
An Active Site Guanine Participates in *glmS* Ribozyme Catalysis in its Protonated State

Júlia Viladoms, Lincoln G. Scott, and Martha J. Fedor

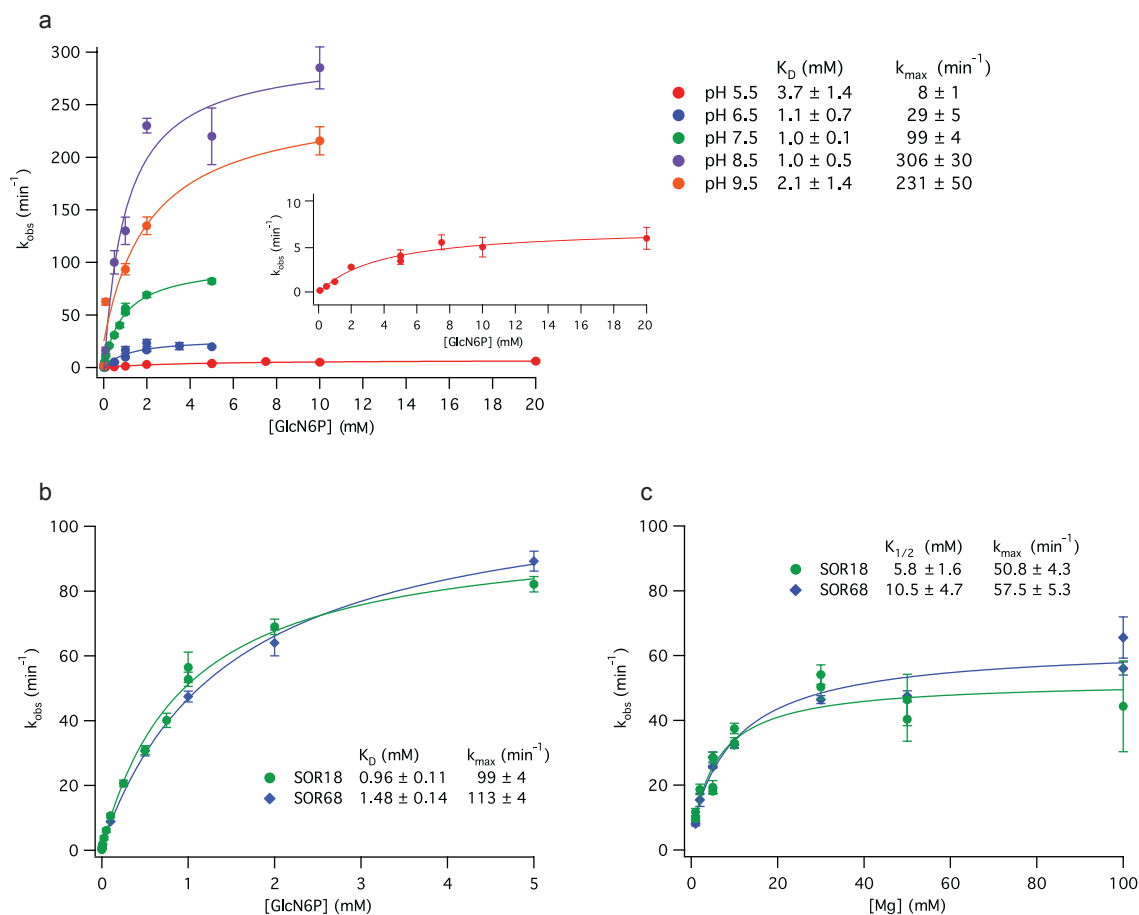
SUPPLEMENTARY FIGURES



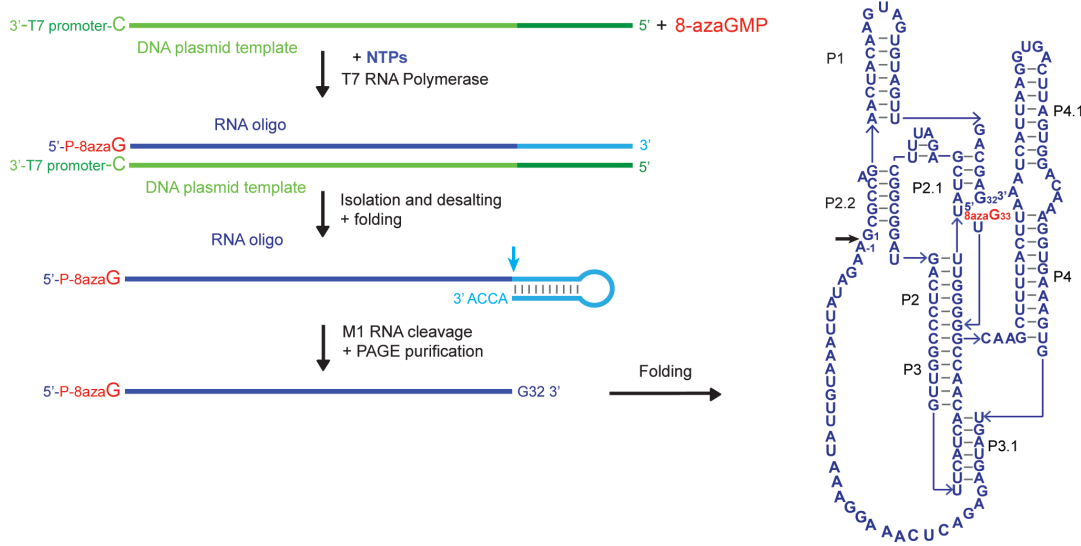
Supplementary Figure S1. RNAs used for kinetic and fluorescence experiments. **a)** G18 ribozyme used for kinetic studies, G68 ribozyme used to investigate ligation activity and G3 ribozyme used to evaluate product dissociation kinetics. **b)** GP23 is a 57 nt unstructured RNA used as a control for fluorescence studies.



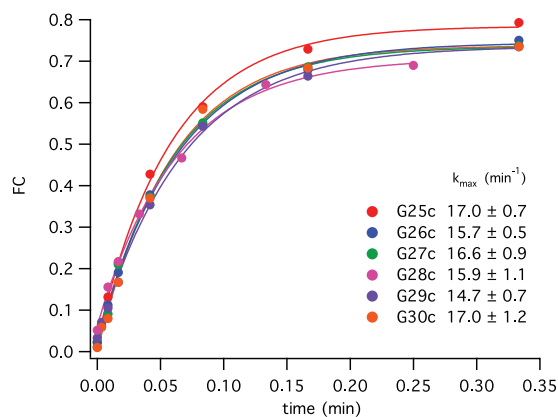
Supplementary Figure S2. Minimal kinetic mechanism of the *glmS* ribozyme reaction that includes binding of the cofactor GlcN6P, self-cleavage and product dissociation.



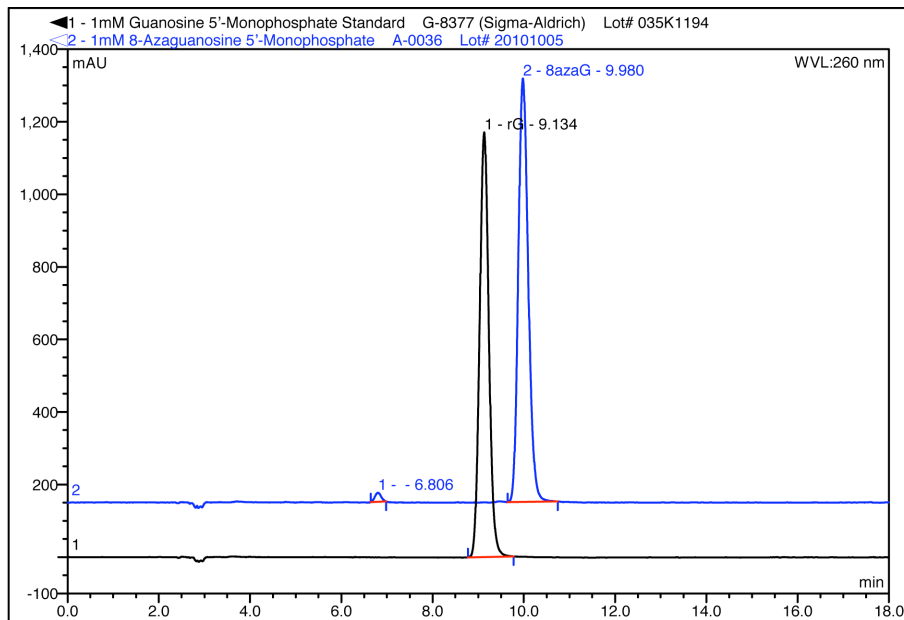
Supplementary Figure S3. *glmS* ribozyme affinity for GlcN6P and Mg^{2+} . **a)** GlcN6P concentration dependence of G18 binding at different pH values (50 mM buffer, 0.1 mM EDTA, 50 mM $MgCl_2$, 25 °C). **b)** Comparison of GlcN6P binding curves for G18 and G68 (50 mM HEPES pH 7.5, 0.1 mM EDTA, 50 mM $MgCl_2$, 25 °C). **c)** Comparison of the Mg^{2+} dependence of G18 and G68 cleavage kinetics (50 mM HEPES pH 7.5, 0.1 mM EDTA, 25 °C, 1 mM GlcN6P).



Supplementary Figure S4. Schematic representation of the strategy used to prepare the circular permuted G28c with GMP or 8azaGMP at the 5'-end and uniform 3'-terminus through M1 RNA processing.



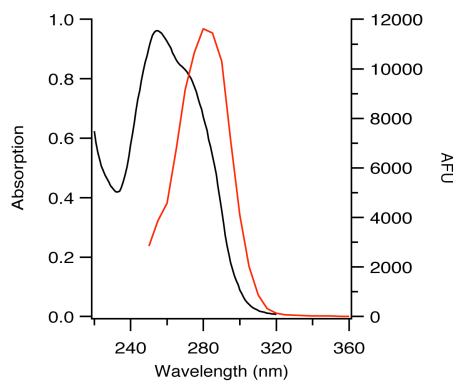
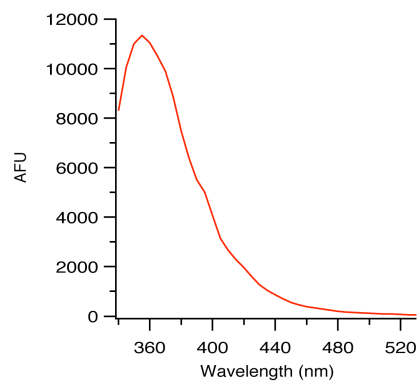
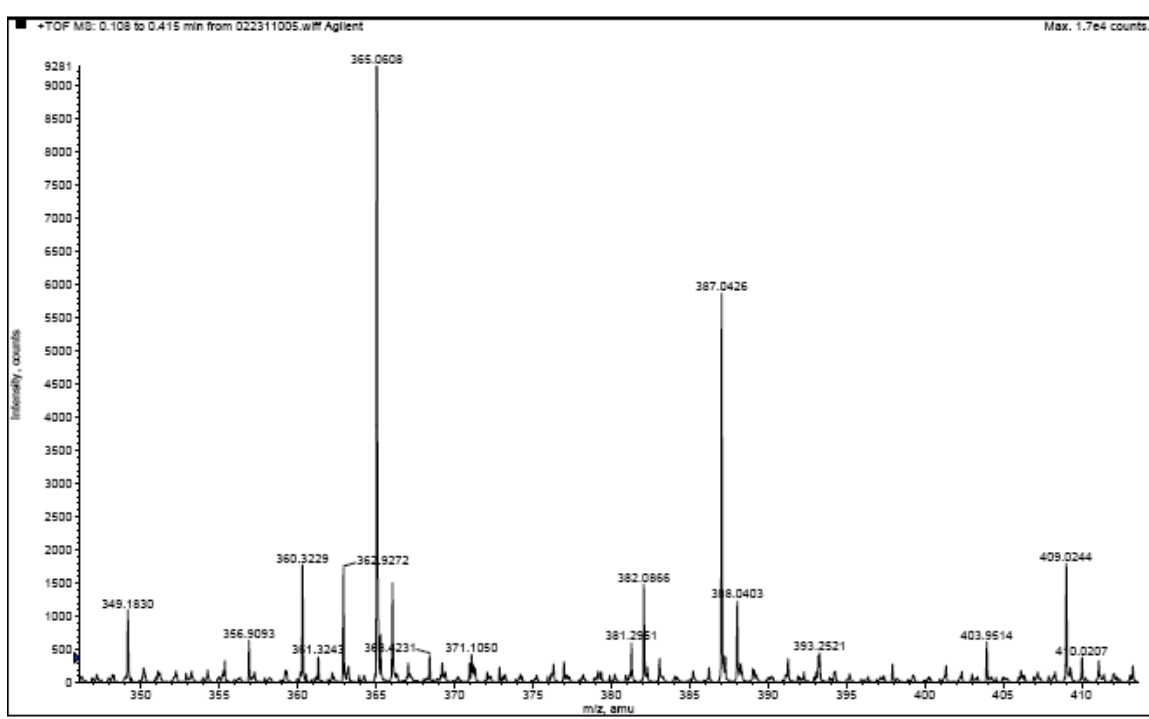
Supplementary Figure S5. Cleavage kinetics at pH 7.5 (50 mM HEPES, 0.1 mM EDTA, 50 mM MgCl₂, 10 mM GlcN6P) for the *glmS* circular permutants with different chain lengths.



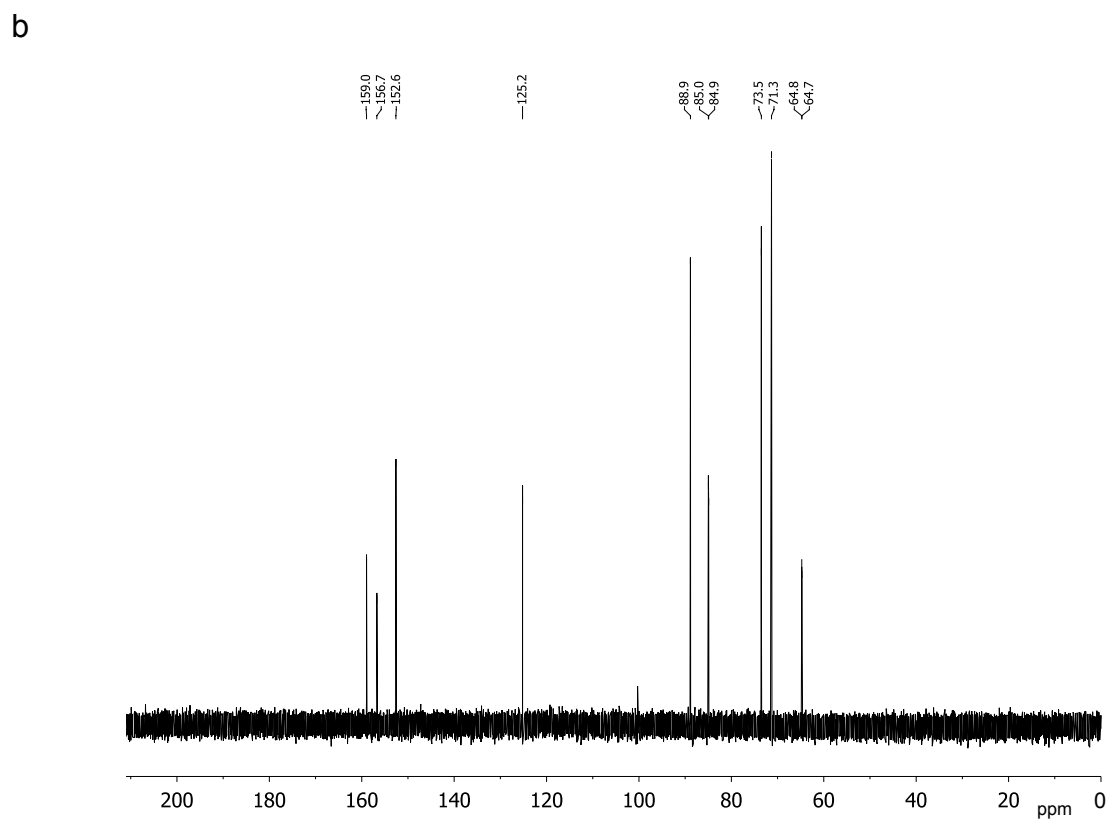
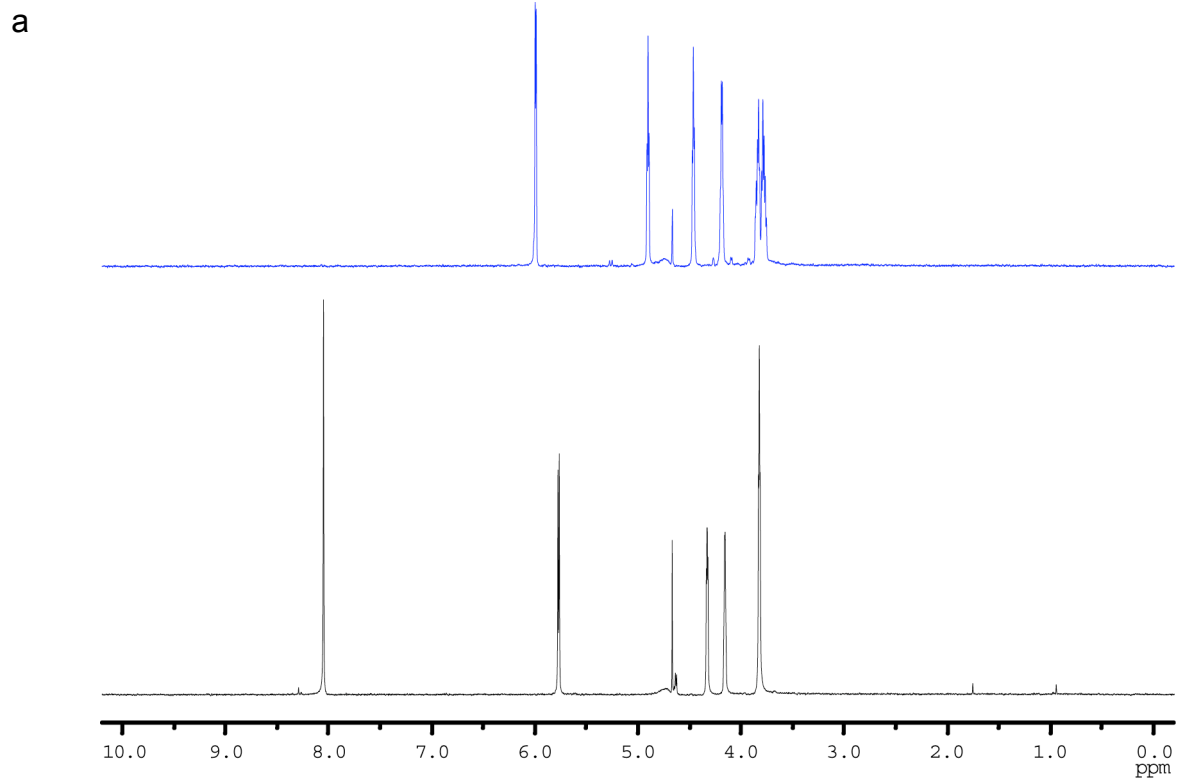
| No. | Ret.Time min | Peak Name | Height mAU | Area mAU*min | Rel.Area % | Amount | Type |
|---------------|-----------------|-----------|---------------|-----------------|---------------|--------|--------------------|
| 1 | 6.81 | | 22.058 | 3.556 | 1.34 | n.a. | BMB ⁺ ^ |
| 2 | 9.98 | 8azaG | 1037.775 | 261.834 | 98.66 | n.a. | BMB ⁺ ^ |
| Total: | | | 1059.833 | 265.389 | 100.00 | 0.000 | |

| No. | Ret.Time min | Peak Name | Height mAU | Area mAU*min | Rel.Area % | Amount | Type |
|---------------|-----------------|-----------|---------------|-----------------|---------------|--------|--------------------|
| 1 | 9.13 | rG | 1170.297 | 275.165 | 100.00 | n.a. | BMB ⁺ ^ |
| Total: | | | 1170.297 | 275.165 | 100.00 | 0.000 | |

Supplementary Figure S6. HPLC analysis of 8-Azaguanosine 5'-Monophosphate. Quality control reversed phase HPLC chromatogram. Column: Higgins HAISIL 100 C18. Conditions: 83.3 mM Triethylammonium Phosphate, 10% Methanol (v/v), pH 6.0 (w/ Phosphoric Acid). Isocratic, 30 min. UV detection at 260 nm. T = 20 °C.¹

a**b****c**

Supplementary Figure S7. UV and MS analysis of 8-Azaguanosine 5'-Monophosphate. **a)** 1mM 8azaGMP pH 7.6 UV spectrum (black) and excitation spectrum (red). **b)** 1mM 8azaGMP pH 7.6 emission spectrum. **b)** HRMS (ESI(+)-TOF) of 8azaGMP showing $[M+H]^+$ (365.0608) and $[M+Na]^+$ (387.0426).



Supplementary Figure S8. NMR characterization of 8-azaguanosine 5'-monophosphate. **a)** ^1H -NMR spectra (D_2O) of 5 mM 8azaGMP (top) and guanosine 5'-monophosphate (bottom). **b)** ^{13}C NMR spectrum (D_2O) of 10 mM 8azaGMP.

SUPPLEMENTARY REFERENCES

(1) Batey, R. T.; Battiste, J. L.; Williamson, J. R. In *Methods in Enzymology*; Thomas, L. J., Ed.; Academic Press: 1995; Vol. Volume 261, p 300.