Supporting information for

Light-Initiated Hydroxylation of Lauric Acid Using Hybrid P450 BM3 Enzymes

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- Fig. S1: FPLC chromatogram showing the separation of Ru-Q397C-BM3 and Q397C-BM3 proteins after labeling reaction using stepwise elution gradient (Solvent A: 10 mM Tris pH = 8.0, Solvent B: 10 mM Tris, 300 mM NaCl, HiTrap Q column).
- Fig. S2: Mass Spectra of the Q397C-BM3 mutant (blue) and the Ru-Q397C-BM3 (red) displaying a mass increase of 646, indication of the covalent attachment of the Ru(II) photosensitizer to BM3 mutants.
- Fig. S3: UV-Vis spectra of the Q397C-BM3 mutant (blue), Ru-Q397C-BM3 (red) and Ru(II) photosensitizer (green) at the same concentration of 3 μM.
- Fig. S4: Steady state emission quenching of the Ru(II)-Q397C-BM3 with increased concentration of the reductive quencher DTC ($\lambda_{ex} = 455$ nm).
- Fig. S5: Representative curves showing formation of products over time for the Ru-Q397C-BM3 mutants (3 µM) with 0.1, 0.5 and 1 mM lauric acid concentrations under constant light irradiation.
- Fig. S6: Lineweaver-Burk plot for the Ru-Q397C-BM3 and Ru-K97C-BM3 hybrid enzymes.
- Table S1: Initial rates of reaction for the different enzymatic systems determined in triplicates after one-minute reaction with 1.5 mM lauric acid.
- Fig. S7: Difference spectrum for the BM3-WT and Ru-BM3 enzymes in the presence of CO under photoreductive conditions (100 mM DTC and constant light irradiation from a mercury lamp with UV- and IR-cutoff filters).
- Fig. S8: Absorption spectra showing the Ru-Q397C-BM3 protein decay over the course of the reaction (dashed lines) and with 10 μ M catalase (solid lines). Inset: Single exponential fit of the protein decay (rate = 0.02 min⁻¹ with 10 μ M catalase).
- Fig. S9: Representative curves showing formation of products over time with 3 μ M BM3-WT and 10 mM H₂O₂ (black), 3 μ M Ru-Q397C-BM3 and 10 mM H₂O₂ (red) and 100 mM DTC + light (blue).

Fig. S1:

FPLC chromatogram showing the separation of Ru-Q397C-BM3 and Q397C-BM3 proteins after labeling reaction using stepwise elution gradient (Solvent A: 10 mM Tris pH = 8.0; Solvent B: 10 mM Tris, 300 mM NaCl; HiTrap Q column).



Fig. S2:

Mass Spectra of the Q397C-BM3 mutant (blue) and the Ru-Q397C-BM3 (red) displaying a mass increase of 646, indication of the covalent attachment of the Ru(II) photosensitizer to BM3 mutants.



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Fig. S3:

UV-Vis spectra of the Q397C-BM3 mutant (blue), Ru-Q397C-BM3 (red) and Ru(II) photosensitizer (green) at the same concentration of 3 μ M.





Steady state emission quenching of the Ru(II)-Q397C-BM3 with increased concentration of the reductive quencher DTC ($\lambda_{ex} = 455$ nm).



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Fig. S5:

Representative curves showing formation of products over time for the Ru-Q397C-BM3 mutants (3 μ M) with 0.1, 0.5 and 1 mM lauric acid concentrations under constant light irradiation.







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Table S1:

Initial rates of reaction for the different enzymatic systems determined in triplicates after one-minute reaction with 1.5 mM lauric acid.

| Enzymatic system | Initial reaction rate (mol product/mol enzyme/min) |
|--|--|
| $10 \ \mu M \ WT + 10 \ mM \ H_2O_2$ | 0.68 ± 0.05 |
| $10 \ \mu M \ WT + 100 \ \mu M \ Ru(bpy)_3^{2+}$ | 0.11 ± 0.05 |
| 5 μM Ru-K97C-BM3 + 100 mM DTC + hv | 1.41 ± 0.13 |
| 5 μM Ru-K97C-BM3 + 100 mM DTC + hv +10 | 1.90 + 0.09 |
| μM catalase | 1.80 ± 0.08 |
| 5 µM Ru-Q397C-BM3 + 100 mM DTC + hv | 1.59 ± 0.16 |
| 5 μM Ru-Q397C-BM3 + 100 mM DTC + hv + | 265 + 0.14 |
| 10 μM catalase | 2.05 ± 0.14 |

Fig. S7:

Difference spectrum for the BM3-WT and Ru-BM3 enzymes in the presence of CO under photoreductive conditions (100 mM DTC and constant light irradiation from a mercury lamp with UV- and IR-cutoff filters).



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Fig. S8:

Absorption spectra showing the Ru-Q397C-BM3 protein decay over the course of the reaction (dashed lines) and with 10μ M catalase (solid lines). Inset: Single exponential fit of the protein decay (rate = 0.02 min^{-1} with 10μ M catalase).



Fig. S9:

Representative curves showing formation of products over time with 3 μ M BM3-WT and 10 mM H₂O₂ (black), 3 μ M Ru-Q397C-BM3 and 10 mM H₂O₂ (red) and 100 mM DTC + light (blue).

