

**Hyperthermic seizures and aberrant cellular homeostasis in *Drosophila*  
dystrophic muscles**

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## Supplementary Information

Supplementary Table 1: ROS levels seen in *Dys*, *Dg* and *Cam* mutants

Genotype	ROS ([AU]/ $\mu$ g/ml protein)	P-Value
<i>OregonR</i> <sup>2</sup>	1.00 $\pm$ 0.26	-
<i>DysDf/+</i>	1.25 $\pm$ 0.83	0.091
<i>Dg</i> <sup>O86</sup> /+	1.04 $\pm$ 0.14	0.37
<i>Cam</i> <sup>n339</sup> /+	0.88 $\pm$ 0.10	0.22
<i>DysDf</i>	4.66 $\pm$ 2.54	3.0X10 <sup>-8</sup> ***
<i>Dg</i> <sup>O86/O55</sup>	0.61 $\pm$ 0.37	0.0074**
<i>Dg</i> <sup>O86</sup> /+; <i>DysDf</i> /+	0.66 $\pm$ 0.22	0.0034**
<i>Cam</i> <sup>n339</sup> /+; <i>DysDf</i> /+	0.49 $\pm$ 0.17	2.0X10 <sup>-5</sup> ***
<i>cora</i> <sup>k08713</sup> /+; <i>DysDf</i> /+	0.86 $\pm$ 0.07	0.18

<sup>1</sup> P-Values are relative to control animals of genotype *OregonR*,

\*P  $\leq$  0.05, \*\*P  $\leq$  0.01, \*\*\*P  $\leq$  0.001

<sup>2</sup> All values are normalized relative to *OregonR*

<sup>3</sup> errors reported represent the standard deviation

Supplementary Table 2: Decrease in *Dystrophin* mRNA level

Genotype	<i>Dys</i> Average C <sub>T</sub>	<i>RpL32</i> Average C <sub>T</sub>	$\Delta C_T$ <i>Dys</i> – <i>RpL32</i> <sup>1</sup>	$\Delta\Delta C_T$ ( $\Delta C_T$ – $\Delta C_{T,control}$ <sup>2</sup> )	Average <i>Dys</i> relative to control <sup>3</sup>	<i>Dys</i> mRNA fold reduction relative to control <sup>4</sup>
<i>tub-Gal4/+</i>	22.40±0.09	18.35±0.20	4.05±0.22	0.00±0.32	1.00±0.22	1.00±0.22
<i>tub-Gal4::dsDys/+</i>	22.26±0.06	16.35±0.04	5.91±0.08	1.86±0.24	0.28±0.05	3.63±0.60

<sup>1</sup> the  $\Delta C_T$  value is determined by subtracting the average *RpL32* C<sub>T</sub> value from the average *Dys* C<sub>T</sub> value. The standard deviation of the difference is calculated from the standard deviation of the *Dys* and *RpL32* values using the formula “ $s=\sqrt{(s_1^2+s_2^2)}$ ”, where s=stdev

<sup>2</sup> the calculation of the  $\Delta\Delta C_T$  involves subtraction by the  $\Delta C_T$  calibrator value. This standard deviation is determined the same as in ‘1’

<sup>3</sup> the range given for *Dys* relative to Control is determined by evaluating the expression:  $2^{-\Delta\Delta C_T}$  where the error is determined using regression analysis

<sup>4</sup> the fold reduction given for *Dys* relative to Control is determined by evaluating the expression:  $2^{\Delta\Delta C_T}$  where the error is determined using regression analysis

Supplementary Table 3: Decrease in *Dystrophin* mRNA level using *tub-Gal80<sup>ts</sup>* system

Genotype	<i>Dys</i> Average C <sub>T</sub>	<i>RpL32</i> Average C <sub>T</sub>	$\Delta C_T$ <i>Dys</i> – <i>RpL32</i> <sup>1</sup>	$\frac{\Delta \Delta C_T}{\Delta C_{T,cont}^2}$ ( $\Delta C_T - \Delta C_{T,cont}^2$ )	T Index <sup>3</sup>	T corrected <i>Dys</i> mRNA fold reduction relative to control <sup>4</sup>
<i>tub-Gal80<sup>ts</sup>/+;</i> <i>tub-Gal4/+</i> pupae at 18°C	24.46±0.16	16.61±0.03	7.86±0.17	0.00±0.24	-	1.00±0.16
<i>tub-Gal80<sup>ts</sup>/+;</i> <i>tub-Gal4/+</i> pupae 3d. at 29°C	21.87±0.02	16.53±0.27	5.34±0.27	-2.52±0.32	5.72	1.00±0.31
<i>tub-Gal80<sup>ts</sup>/+;</i> <i>tub-Gal4::dsDys/+</i> pupae at 18°C	23.47±0.17	16.20±0.06	7.27±0.17	0.00±0.25	-	1.00±0.17
<i>tub-Gal80<sup>ts</sup>/+;</i> <i>tub-Gal4::dsDys/+</i> pupae 3d. at 29°C	22.60±0.12	16.28±0.13	6.32±0.17	-0.95±0.25	5.72	2.97±0.27
<i>tub-Gal80<sup>ts</sup>/+;</i> <i>tub-Gal4::dsDys/+</i> flies at 18°C	23.88±0.17	18.19±0.29	5.70±0.34	0.00±0.48	-	1.00±0.33
<i>tub-Gal80<sup>ts</sup>/+;</i> <i>tub-Gal4::dsDys/+</i> flies 3d. at 29°C	24.76±0.23	17.28±0.03	7.48±0.24	1.78±0.41	-	3.44±0.99

<sup>1</sup> the  $\Delta C_T$  value is determined by subtracting the average *RpL32* CT value from the average *Dys* C<sub>T</sub> value. The standard deviation of the difference is calculated from the standard deviation of the *Dys* and *RpL32* values using the formula “ $s = \sqrt{(s_1^2 + s_2^2)}$ ”, where s=stdev

<sup>2</sup> the calculation of the  $\Delta \Delta C_T$  involves subtraction by the  $\Delta C_T$  calibrator value. This standard deviation is determined the same as in ‘1’

<sup>3</sup> the temperature dependent scaling factor was determined because expression levels of *Dys* in control pupae varied greatly depending on the temperature. Where applicable this value is calculated as  $2^{-\Delta \Delta C_T}$ . This scaling factor is then used to determine the relative expression levels in mutant animals of the same life stage at 29°C relative to the same animals at 18°C (Supplementary Figure 2)

<sup>4</sup> the fold reduction given for *Dys* relative to Control is determined by evaluating the expression:  $2^{\Delta \Delta C_T}$  where the error is determined using regressional analysis. The temperature related scaling index has been multiplied by this value where applicable

Supplementary Table 4: Frequency of muscle degeneration in *Dys RNAi* mutants with and without developmentally restricted expression of *RNAi*

Genotype	<i>Dys</i> down regulated	Condition <sup>1</sup>	Age, days	n	% of muscle degeneration	P-Value <sup>2</sup>	Normalized <sup>3</sup> % of muscle degeneration	P-Value <sup>2</sup>
<i>tub-Gal4/+</i>	Throughout lifetime	25°C	16	121	9.1	0.021*	2.4	0.11
<i>tub-Gal4:dsDys</i>				73	21.9			
<i>tub-Gal80<sup>ts</sup>/+; tub-Gal4/+</i>	As adult	18°C development 29°C adulthood	19	84	21.4	0.0081 **	2.0	
<i>tub-Gal80<sup>ts</sup>/+; tub-Gal4:dsDys/+</i>				108	42.6			

<sup>1</sup> Note that higher temperature (29°C) accelerates aging and muscle degeneration.

<sup>2</sup> P-value determined from the  $\chi^2$  statistic

<sup>3</sup> Normalized to the control value under the same conditions

Supplementary Table 5: Relative Dg immunofluorescence intensities

Genotype	Relative Intensity	P-Value <sup>1</sup>
<i>OregonR</i> <sup>2</sup>	1.00 ± 0.37 <sup>3</sup>	-
<i>DysDf</i>	0.49 ± 0.27	0.042*
<i>Dg</i> <sup>O55</sup>	0.02 ± 0.02	1.2 X 10 <sup>-4</sup> ***
<i>cora</i> <sup>k08713</sup>	0.11 ± 0.11	1.4 X 10 <sup>-3</sup> **
<i>Dg</i> <sup>O86/+</sup> ; <i>DysDf</i> /+	0.06 ± 0.05	1.9 X 10 <sup>-5</sup> ***
<i>cora</i> <sup>k08713/+</sup> ; <i>DysDf</i> /+	0.07 ± 0.06	1.1 X 10 <sup>-4</sup> ***
<i>cora</i> <sup>k08713</sup> / <i>Dg</i> <sup>O86</sup>	0.10±0.12	9,9 X 10 <sup>-5</sup> ***

<sup>1</sup> P-Values are relative to control animals of genotype *OregonR*,

\*P ≤ 0.05, \*\*P ≤ 0.01, \*\*\*P ≤ 0.001

<sup>2</sup> All values are normalized relative to *OregonR*

<sup>3</sup> errors reported represent the average deviation

Supplementary Table 6: Decrease in *Dystroglycan* mRNA level

Genotype	<i>Dg</i> Average $C_T$	<i>RpL32</i> Average $C_T$	$\Delta C_T$ <i>Dg</i> - <i>RpL32</i> <sup>1</sup>	$\Delta\Delta C_T$ ( $\Delta C_T$ - $\Delta C_{T,control}$ ) <sup>2</sup>	Average <i>Dg</i> relative to control <sup>3</sup>	<i>Dg</i> mRNA fold reduction relative to control <sup>4</sup>
<i>tub-Gal4/+</i>	23.33±0.07	18.35±0.20	4.98±0.21	0.00±0.30	1.00±0.21	1.00±0.21
<i>tub-Gal4::dsDg/+</i>	25.94±0.13	18.37±0.11	7.57±0.18	2.59±0.28	0.17±0.03	6.02±1.16
<i>tub-Gal80<sup>ts</sup>/+;</i> <i>tub-Gal4::dsDg/+</i> flies at 18°C	20.97±0.05	16.09±0.05	4.87±0.07	0.00±0.10	1.00±0.07	1.00±0.07
<i>tub-Gal80<sup>ts</sup>/+;</i> <i>tub-Gal4::dsDg/+</i> flies 3d. at 29°C	25.26±0.07	19.09±0.03	6.17±0.08	1.29±0.11	0.41±0.03	2.45±0.18

<sup>1</sup> the  $\Delta C_T$  value is determined by subtracting the average *RpL32*  $C_T$  value from the average *Dg*  $C_T$  value. The standard deviation of the difference is calculated from the standard deviation of the *Dg* and *RpL32* values using the formula " $s=\sqrt{(s_1^2+s_2^2)}$ ", where  $s$ =stdev

<sup>2</sup> the calculation of the  $\Delta\Delta C_T$  involves subtraction by the  $\Delta C_T$  calibrator value. This standard deviation is determined the same as in '1'

<sup>3</sup> the range given for *Dg* relative to Control is determined by evaluating the expression:  $2^{-\Delta\Delta C_T}$  where the error is determined using regression analysis

<sup>4</sup> the fold reduction given for *Dg* relative to Control is determined by evaluating the expression:  $2^{\Delta\Delta C_T}$  where the error is determined using regression analysis

Supplementary Table 7: Impact of Ca<sup>2+</sup> channel blockers on seizure activity of dystrophic animals (*DysDf*)

Drug	n	%Seized <sup>‡</sup>	Avg. T <sub>s</sub> ±sd (°C)*	Avg. A <sub>max</sub> ±sd (mV)*	Avg. Area±sd (pixel x 10 <sup>-3</sup> )*	Median S <sub>i</sub> ** (25 <sup>th</sup> /75 <sup>th</sup> percentiles)	P-Value
Sucrose	5	80	37.8±2.1	6.6±5.7	2.4 ±1.9	1.9 (1.8/2.1)	-
3.6 mM Nifedipine	5	100	37.8±3.3	3.1±2.2	1.2 ± 0.5	1.8 (1.8/1.9)	0.50
0.5 mM 2-APB	5	60	35.7±5.7	2.8±1.2	0.8 ± 0.4	1.5 (0.0/1.7)	0.21
20 μM Ryanodine	6	33	34.4±0.6	2.8±0.6	0.9 ±0.2	0.0 (0.0/1.4)	0.078

n = number of animals fed the indicated drug

‡ percent of animals measured that had a seizure

\* Calculated using data from animals that seized only

\*\* Index calculated by integrating the area of the graph of voltage vs. temperature during a seizure, taking the natural logarithm of this number, dividing by the temperature that the seizure started, and then multiplying times ten.



## Supplementary Figure Legends

### Supplementary Figure 1:

Control animals do not have temperature-sensitive seizures as can be seen by monitoring the output voltage from the IFMs as the temperature increases. b) All tested *Dys* mutant alleles exhibit hyperthermic seizures.

### Supplementary Figure 2:

Average *Dys* expression relative to appropriate controls in *RNAi* knock-down mutants that are under the control of the *tub-Gal80<sup>ts</sup>* driver. a) Control pupae showed a vast increase in *Dys* expression upon being shifted from 18°C to 29°C indicating a natural increase in protein expression with temperature increase. Thus, to determine adequately the effectiveness of *RNAi* down-regulation this change in expression had to be taken into consideration using a temperature index (Supplementary Table 5). b) Average *Dys* expression levels relative to control in pupae (after the temperature index was considered) and in adults after being shifted to 29°C for four days.

### Supplementary Figure 3:

Seizure indices showing that animals with *RNAi* directed against muscle (*24B-Gal4*) and mononeuron (*D42-Gal4*) *Dg* mRNA do not have temperature-sensitive seizures. Endogenous (*tub-Gal4*) expression results in a reduced amount of seizures that are not significant over controls, but this is reversed when *Dg* is down-regulated after development. Refer to Table 1 for exact P-values compared to control animals.

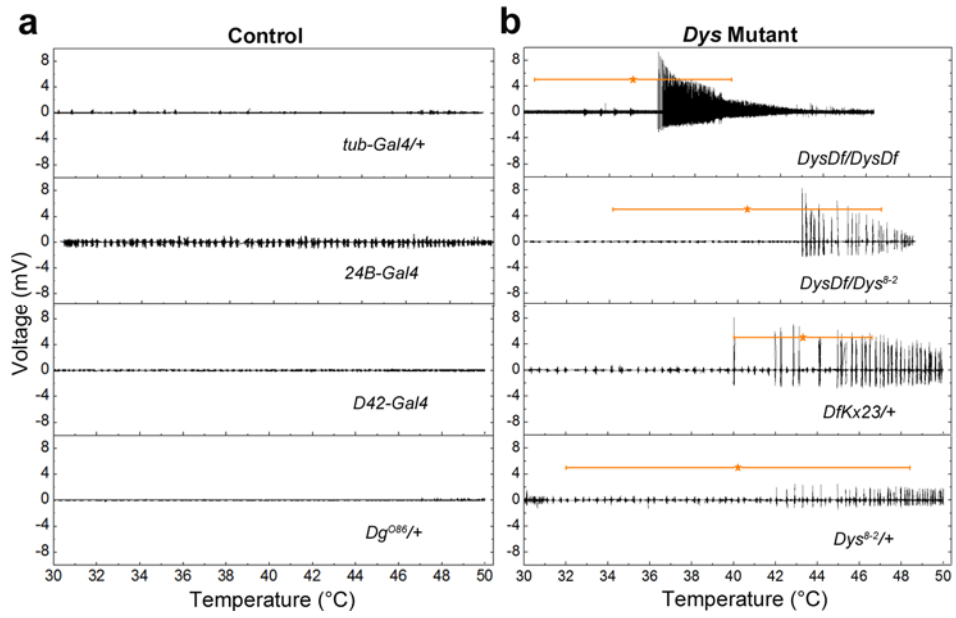
#### Supplementary Figure 4:

a)  $\beta_{PS}$  integrin subunit loss of function allele heterozygotes (*mys*<sup>1/+</sup>) had seizures when heated, most likely due to compromised stability of the sarcolemma and/or NMJ. Introducing a mutant copy of *Dg* into the genome of these animals does not alter the seizure character. b) Seizure indices of *mys*<sup>1/+</sup> heterozygous and *mys*<sup>1/+</sup>; *Dg*/+ transheterozygous animals where a Kruskal-Wallis test show that there are no significances in the indices ( $\chi^2 = 2.93$ ,  $P = 0.231$ ). The following P-values were determined upon comparison to wild type animals: *mys*<sup>1/+</sup> (\*\* $P = 2.0 \times 10^{-4}$ ), *mys*<sup>1/+</sup>; *Dg*<sup>086/+</sup> (\* $P = 0.013$ ), *mys*<sup>1/+</sup>; *Dg*<sup>055/+</sup> (\* $P = 0.082$ ) using a one-tailed Mann-Whitney U-test. 100% of *mys*<sup>1/+</sup> animals tested had seizures (n=8) with  $T_s = 39.2 \pm 2.6$ ,  $A_{max} = 3.6 \pm 1.7$  and Avg. Area =  $1.5 \pm 1.1 \times 10^3$ . 80% of *mys*<sup>1/+</sup>; *Dg*<sup>086/+</sup> animals tested had seizures (n=5) with  $T_s = 36.0 \pm 0.7$ ,  $A_{max} = 3.8 \pm 2.4$  and Avg. Area =  $1.2 \pm 0.5 \times 10^3$ . 83% of *mys*<sup>1/+</sup>; *Dg*<sup>055/+</sup> animals tested had seizures (n=6) with  $T_s = 40.3 \pm 3.7$ ,  $A_{max} = 4.3 \pm 3.0$  and Avg. Area =  $1.2 \pm 1.0 \times 10^3$ .

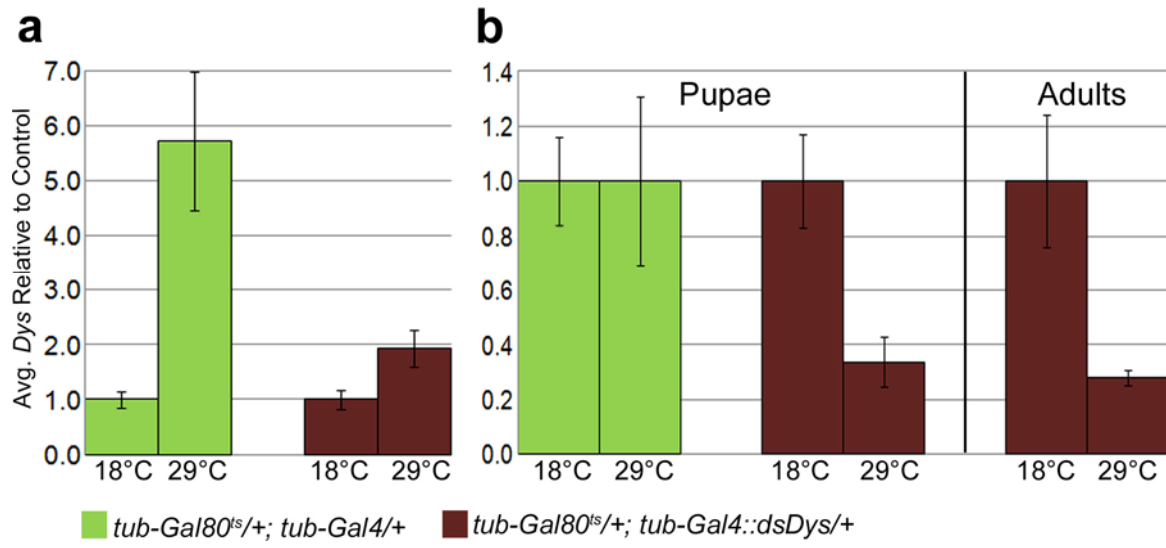
#### Supplementary Figure 5:

a) TGF- $\beta$  pathway mutants, *tkv* (type-I receptor) and *Mad* (receptor-regulated Smad) have seizures similar to what is seen in *Dys* mutants, however; when animals are missing one copy of *Dys* and one copy of each of these genes there is no significant increase in seizures over what is seen in *Dys*. Down-regulation of *tkv* after NMJ development (*tub-Gal80<sup>ts</sup>*) does not alleviate seizures implying that there is an additional need for *tkv* after development is complete. Animals that express *RNAi* against *tkv* prior to pupae formation die supporting an important role for TGF- $\beta$  signaling during pupation. Transheterozygous animals when compared to *Dys* heterozygous

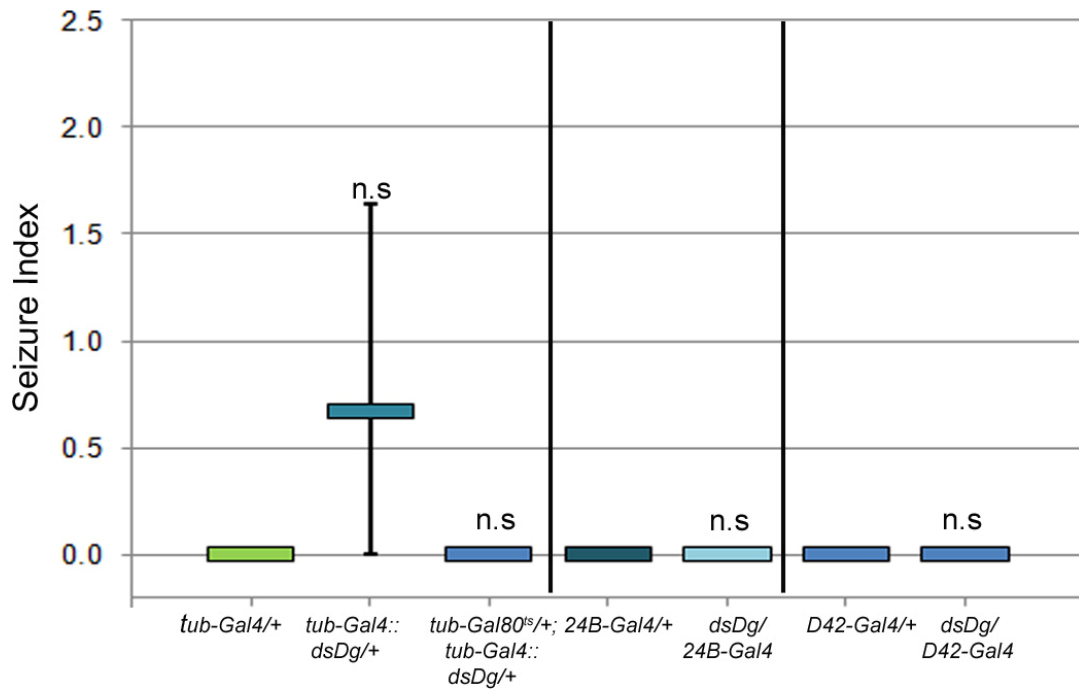
animals using a Mann-Whitney U-test had P-values of 0.25 and 0.46 for *tkv/Dys* and *Mad/Dys* respectively and P-values of 0.099 and 0.040 when compared to control animals respectively. Animals of genotypes *tkv<sup>1</sup>*, *tub-Gal80<sup>ts</sup>/act-Gal4;tkv<sup>RNAi</sup>* and *Mad<sup>12</sup>/+* have P-values of  $1.9 \times 10^{-3}$ , 0.012 and 0.072 when compared to controls respectively. 100% of *tkv<sup>1</sup>* animals tested had seizures (n=9) with  $T_s = 39.4 \pm 4.7$ ,  $A_{max} = 4.8 \pm 3.6$  and Avg. Area =  $1.3 \pm 0.8 \times 10^3$ . 60% of *tkv<sup>1</sup>/+;DysDf/+* animals tested had seizures (n=5) with  $T_s = 44.2 \pm 2.1$ ,  $A_{max} = 7.4 \pm 1.3$  and Avg. Area =  $1.9 \pm 1.3 \times 10^3$ . 67% of *Mad<sup>12</sup>/+;DysDf/+* animals tested had seizures (n=6) with  $T_s = 42.2 \pm 4.0$ ,  $A_{max} = 15 \pm 8.7$  and Avg. Area =  $2.0 \pm 1.0 \times 10^3$ . 80% of *Mad<sup>12</sup>/+* animals tested had seizures (n=5) with  $T_s = 42.8 \pm 3.2$ ,  $A_{max} = 8.8 \pm 6.3$  and Avg. Area =  $1.7 \pm 1.4 \times 10^3$ . 100% of *tub-Gal80<sup>ts</sup>/act-Gal4; tkv<sup>RNAi</sup>/+* animals tested had seizures (n=5) with  $T_s = 40.0 \pm 3.4$ ,  $A_{max} = 7.3 \pm 2.8$  and Avg. Area =  $4.4 \pm 2.8 \times 10^3$ . b) Electrical output from IFM vs. temperature of TGF- $\beta$  pathway mutants exhibiting seizures. \*P  $\leq$  0.05, \*\*P  $\leq$  0.01 and \*\*\*P  $\leq$  0.001



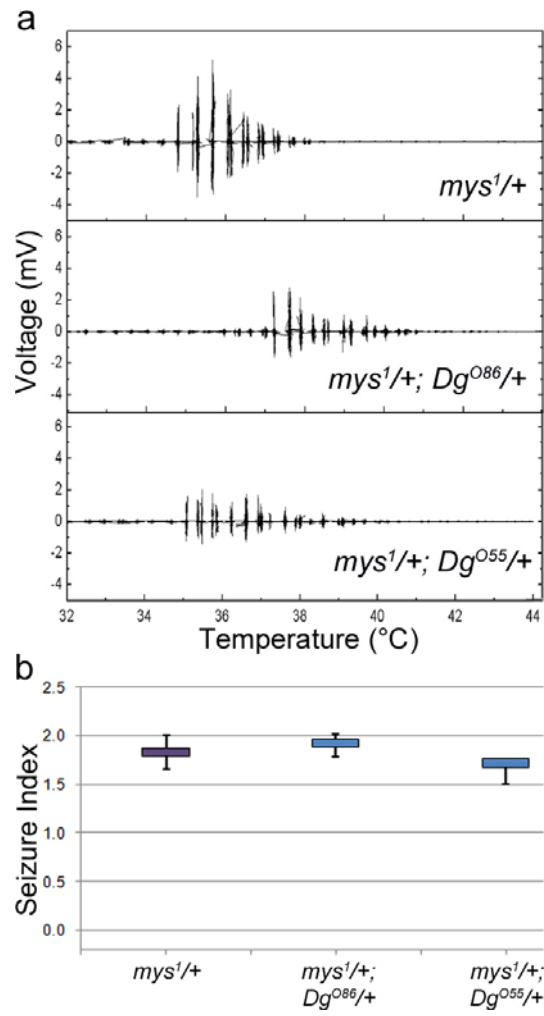
Supplementary Figure 1



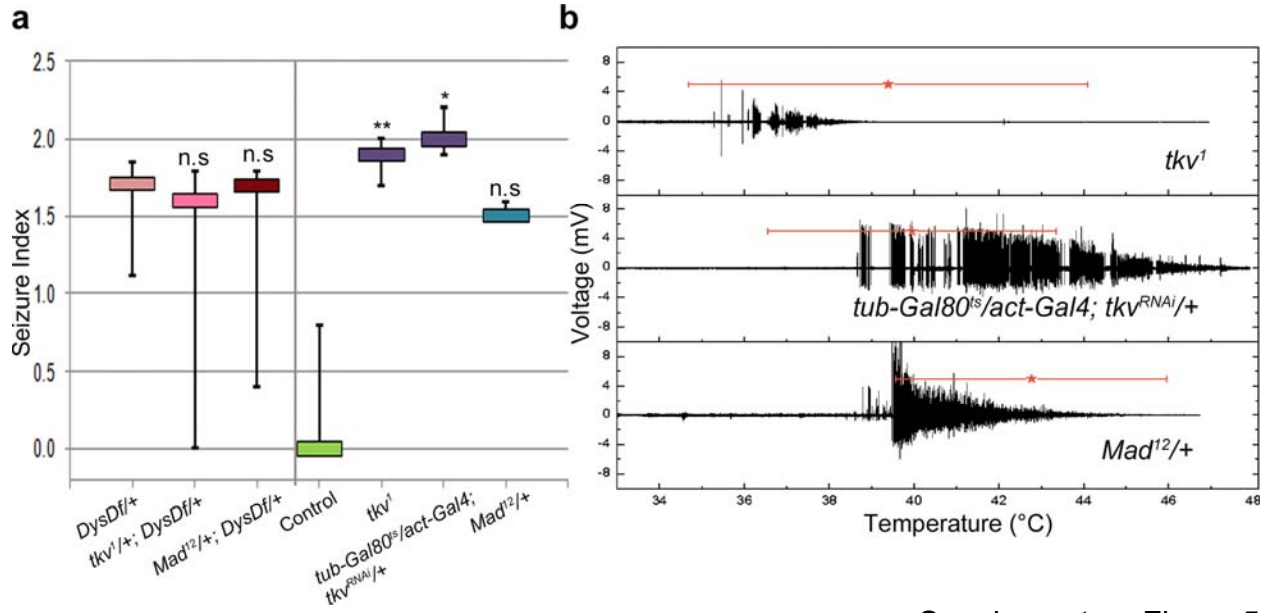
Supplementary Figure 2



Supplementary Figure 3



Supplementary Figure 4



Supplementary Figure 5