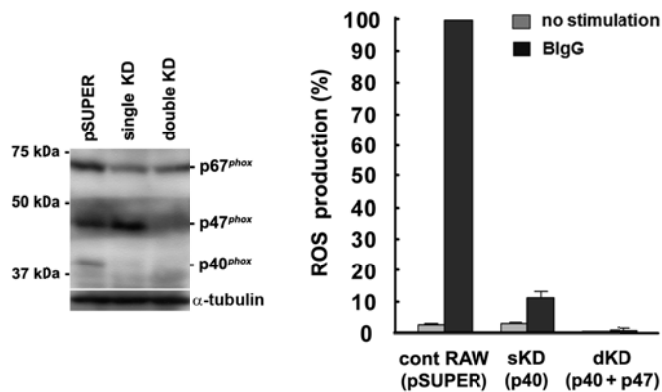


**Supplemental Fig. 1, Establishment of HEK293<sup>Nox2/FcγRIIa</sup> cells.**

A, Stable expression of human Nox2 and human FcγRIIa in HEK293<sup>Nox2/FcγRIIa</sup> cells is confirmed by immunoblotting (mAb against Nox2, pAb against FcγRIIa, mAb against p22<sup>phox</sup>). Stable expression of human Nox2 greatly enhances detection of endogenous p22<sup>phox</sup>.

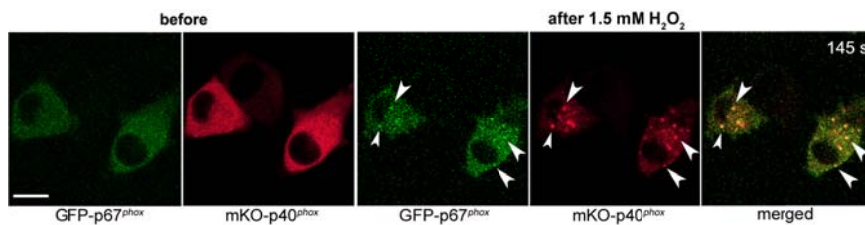
B, Endogenous p22<sup>phox</sup>, which is localized at ER in the absence of Nox (1), is localized at the PM (arrow) and dot-like structures (arrowheads) in HEK293<sup>Nox2/FcγRIIa</sup> cells (left), in agreement with findings in CHO cells and RAW 264.7 macrophages (2). PM localization of FcγRIIa in HEK293<sup>Nox2/FcγRIIa</sup> is confirmed (right). Bar: 10 μm

C, ROS measurements stimulated by BigG using HEK293<sup>Nox2/FcγRIIa</sup> cells transfected with cytoplasmic phox proteins show p40<sup>phox</sup> dependency in assays using luminol + HRP [with or without catalase (1000U/ml, Sigma) + SOD (10U/ml, Sigma)], luminol without HRP or isoluminol + HRP. Catalase and SOD were preincubated for 10 min with cells before assays. ROS detected by luminol + HRP with catalase + SOD and ROS detected by luminol without HRP, representing intracellular ROS production (3), were 15.2 ± 2.0 % and 5.7 ± 1.1 % of that detected by luminol + HRP without catalase + SOD, respectively. ROS production stimulated by PMA detected by luminol + HRP with catalase + SOD and by luminol without HRP were 4.5 ± 0.7 % and 0.86 ± 0.06 % of that detected by luminol + HRP without catalase + SOD, respectively. The results indicate that ROS measurements stimulated by BigG detected by luminol + HRP was the sum of extracellular ROS and intracellular ROS, but predominantly represented extracellular ROS. Comparable expression of p47<sup>phox</sup>, p67<sup>phox</sup>, p40<sup>phox</sup> and p40<sup>phox</sup> (105K) is confirmed by immunoblotting (pAbs against p47<sup>phox</sup>, p67<sup>phox</sup> and p40<sup>phox</sup>).



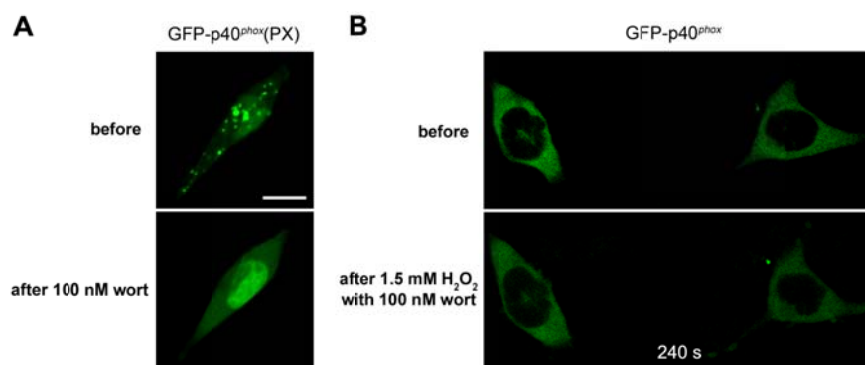
**Supplemental Fig. 2, Establishment of RAW264.7<sup>p40KD</sup> and RAW264.7<sup>p40/p47KD</sup> cells.**

Establishment of RAW264.7 macrophages with stable knockdown of p40<sup>phox</sup> (in sKD: RAW264.7<sup>p40KD</sup>) or stable knockdown of both p40<sup>phox</sup> and p47<sup>phox</sup> (in dKD: RAW264.7<sup>p40/p47KD</sup>) is confirmed by immunoblotting (pAb against p47<sup>phox</sup>, mAb against p67<sup>phox</sup>, pAb against mouse p40<sup>phox</sup>) and by ROS measurements using luminol + HRP. Note the decreased expression of p67<sup>phox</sup> in both sKD (RAW264.7<sup>p40KD</sup>) and dKD (RAW264.7<sup>p40/p47KD</sup>).



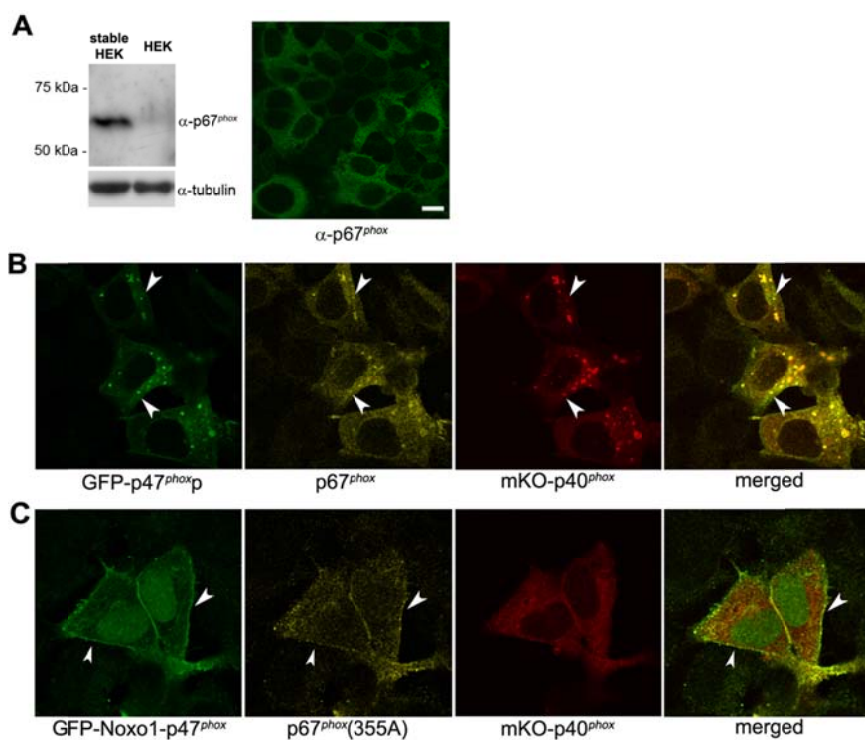
**Supplemental Fig. 3, H<sub>2</sub>O<sub>2</sub> stimulates co-translocation of GFP-p67<sup>phox</sup> and mKO-p40<sup>phox</sup> in RAW 264.7 macrophages.**

After stimulation with 1.5 mM H<sub>2</sub>O<sub>2</sub> (145 sec), cytoplasmic GFP-p67<sup>phox</sup> translocates to dot-like structures (arrowheads) when co-expressed with mKO-p40<sup>phox</sup>. Bar: 10 μm



**Supplemental Fig. 4, Wortmannin inhibits the targeting/translocation of GFP-p40<sup>phox</sup> (PX) and GFP-p40<sup>phox</sup> in RAW 264.7 macrophages.**

Treatment with wortmannin (100 nM for 15 min) abolishes the dot-like localization pattern of GFP-p40<sup>phox</sup>(PX) (A) and the translocation of GFP-p40<sup>phox</sup> to dot-like structures in response to 1.5 mM H<sub>2</sub>O<sub>2</sub> (240 sec) (B) (compare Supplemental Fig. 4). Bar: 10 μm

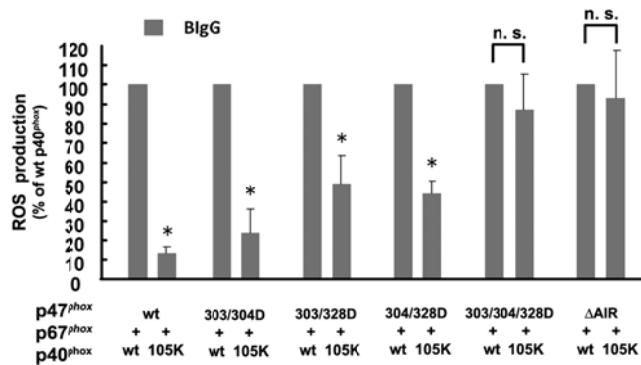


**Supplemental Fig. 5, Establishment of HEK293<sup>p67<sup>phox</sup></sup> cells.**

A, Stable expression of human p67<sup>phox</sup> in HEK293<sup>p67<sup>phox</sup></sup> cells is confirmed by immunoblotting and immunostaining. Bar: 10 μm

B, GFP-p47<sup>phox</sup>p, p67<sup>phox</sup> and mKO-p40<sup>phox</sup> are co-localized at dot-like structures (arrowheads) in HEK293<sup>p67<sup>phox</sup></sup> cells.

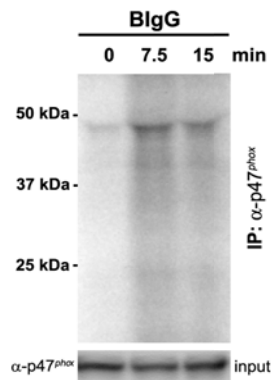
C, The dot-like and PM localizations of mKO-p40<sup>phox</sup> disappear when p67<sup>phox</sup>(355K) is co-expressed instead of p67<sup>phox</sup> in wt HEK293 cells. In contrast, GFP-Noxo1-p47<sup>phox</sup> and p67<sup>phox</sup>(355K) are still co-localized on PM (arrowheads). B and C; fixed cells were stained using polyclonal Ab against p67<sup>phox</sup> and Cy5-conjugated secondary Ab.



**Supplemental Fig. 6, The phosphorylation status of p47<sup>phox</sup> affects dependency of PI(3)P binding of p40<sup>phox</sup> in intracellular ROS production in HEK293<sup>Nox2/FcγRIIa</sup> cells.**

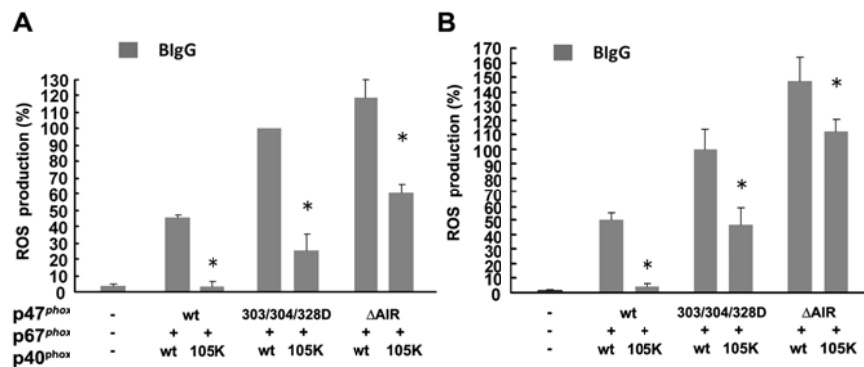
BlgG-stimulated ROS production detected by luminol without exogenous HRP in HEK293<sup>Nox2/FcγRIIa</sup> cells, which reflects intracellular ROS production, shows no PI(3)P binding dependency of p40<sup>phox</sup> in p47<sup>phox</sup>(S303/304/328D) and p47<sup>phox</sup>(ΔAIR), but PI(3)P-binding dependency in 2 sites phosphorylation-mimicking mutants of p47<sup>phox</sup> (303/304D, 303/328D, 304/328D). asterisk: p < 0.01

n. s.: not statistically significant



**Supplemental Fig. 7, Phosphorylation of wt p47<sup>phox</sup> in response to B1gG in HEK293<sup>Nox2/FcγRIIa</sup> cells using <sup>32</sup>P label**

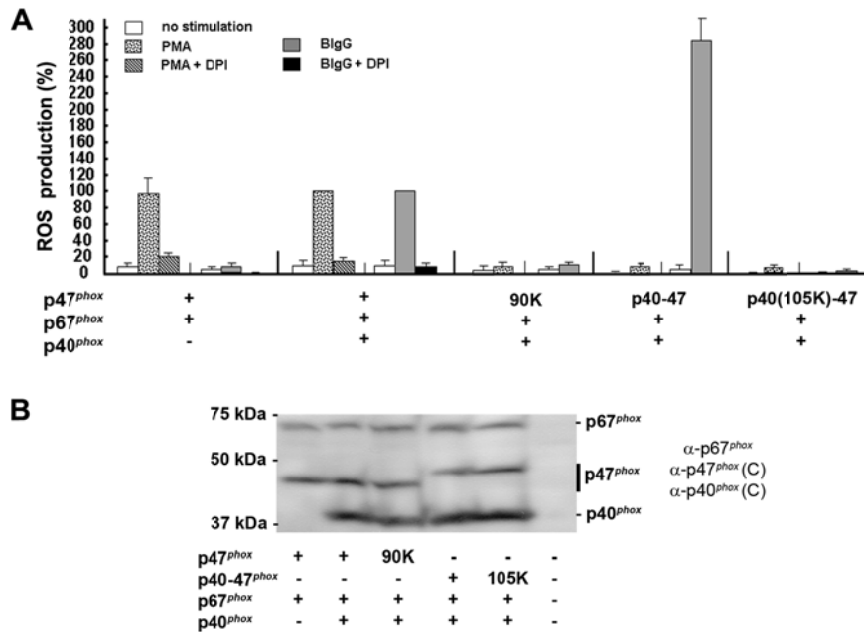
HEK293<sup>Nox2/FcγRIIa</sup> cells transfected with p47<sup>phox</sup> + p67<sup>phox</sup> + p40<sup>phox</sup> were labeled for 1 h with [<sup>32</sup>P]phosphoric acid (0.33 mCi/ml) in phosphate-free DMEM (Invitrogen) containing 0.1% fatty acid-free BSA. After extensively washing, the cells were subsequently stimulated with B1gG (five targets per cell) in phosphate-free DMEM (0.1% BSA) for indicated times at 37°C. Cells were then harvested in homogenate buffer (4) in the presence of 0.25 % Triton and phosphatase inhibitor cocktail (Nacalai Tesque), and settled for 10 min on ice. Cells were lysed by sonication and settled for 1h on ice. After centrifugation at 13,000 × g for 15 min at 4°C, the supernatants were incubated with pAb against p47<sup>phox</sup> for 2 h at 4 °C and then with protein G-Sepharose 4B for additional 2 h at 4 °C. The precipitates were subjected to SDS-PAGE and the phosphorylated proteins were visualized by autoradiography using BAS2500. Phosphorylated bands, which are strongest at 7.5 min after stimulation with B1gG, are detected under 50 kDa.



**Supplemental Fig. 8, The phosphorylation status of p47<sup>phox</sup> affects dependency on p40<sup>phox</sup> PI(3)P binding for both intracellular and extracellular ROS production in RAW264.7<sup>p40/p47KD</sup> cells.**

PI(3)P-dependency is shown for reconstituted ROS production by wt p47<sup>phox</sup>, p47<sup>phox</sup>(S303/304/328D) or p47<sup>phox</sup>(ΔAIR), detected both by luminol without exogenous HRP, which reflects intracellular ROS production (A) and by isoluminol with exogenous HRP, which reflects extracellular ROS production (B).

asterisk: p < 0.01



**Supplemental Fig. 9, PMA-responsive ROS production requires the PX domain of p47<sup>phox</sup>, but not the PX domain of p40<sup>phox</sup> in HEK293<sup>Nox2/FcγRIIa</sup> cells.**

A, PMA-stimulated, but not BIgG-stimulated, ROS production is not affected by presence of p40<sup>phox</sup>. R90K mutation in the PX domain of p47<sup>phox</sup> shows almost no ROS production in response to PMA or BIgG. In the case of p40-47<sup>phox</sup> + p67<sup>phox</sup> + p40<sup>phox</sup>, ROS production stimulated by PMA dramatically decreased ( $10.1 \pm 1.1$  %); in sharp contrast, that stimulated by BIgG increased by about 280 %, compared with p47<sup>phox</sup> + p67<sup>phox</sup> + p40<sup>phox</sup>. R105K mutation in the PX domain of p40-p47<sup>phox</sup> chimeric protein shows almost no ROS production in response to PMA or BIgG.

B, Comparable expression of p47<sup>phox</sup>, p47<sup>phox</sup> mutant, p47<sup>phox</sup> chimeras, p67<sup>phox</sup>, p40<sup>phox</sup> and p40<sup>phox</sup> mutant is confirmed by immunoblotting using pAb against p67<sup>phox</sup> and mAbs against C-terminal of p47<sup>phox</sup> and p40<sup>phox</sup>.

**References**

1. Ueyama, T., Geiszt, M., and Leto, T. L. (2006) *Mol Cell Biol* **26**, 2160-2174
2. Casbon, A. J., Allen, L. A., Dunn, K. W., and Dinayer, M. C. (2009) *J Immunol* **182**, 2325-2339
3. Dahlgren, C., and Karlsson, A. (1999) *Journal of Immunological Methods* **232**, 3-14
4. Ueyama, T., Eto, M., Kami, K., Tatsuno, T., Kobayashi, T., Shirai, Y., Lennartz, M. R., Takeya, R., Sumimoto, H., and Saito, N. (2005) *J Immunol* **175**, 2381-2390

**Supplemental Video 1**, GFP-p40<sup>phox</sup>p localized at EE accumulates on phagosomes during ingestion of B1gG (total time: 675 sec). In supplemental videos (1-5), only indicated GFP-tagged protein is transiently expressed in HEK293<sup>Nox2/FcγRIIa</sup> cells.

**Supplemental Video 2**, GFP-p40<sup>phox</sup> weakly accumulates on phagosomes during ingestion of B1gG (total time: 395 sec).

**Supplemental Video 3**, No accumulation of GFP-FYVE-p40<sup>phox</sup> on phagosomes during ingestion of B1gG (total time: 665 sec).

**Supplemental Video 4**, GFP-PH(TAPP1)-p40<sup>phox</sup> accumulates on phagosomal cup and nascent phagosomes during ingestion of B1gG (total time: 445 sec).

**Supplemental Video 5**, GFP-FYVE-p40<sup>phox</sup>p localized at EE accumulates on phagosomes during ingestion of B1gG (total time: 665 sec)