SUPPLEMENTAL DATA

<u>Suppl. Fig. 1.</u> β c receptors in TF1 cells and primary EOS traffic through the recycling pathway. Human TF1 cells and EOS were treated with or without 40 μ M monensin for 30 minutes. β c cell surface levels were measured by flow cytometry after labeling with anti- β c-PE antibodies and are expressed as mean MFI±SEM minus isotype control MFI in the absence or presence of inhibitor; TF1 (N=3) MFI: 0' (-Mon) = 777 ± 166; 0' (+Mon) = 468 ± 38; 30' (-Mon) = 169 ± 19; 30' (+Mon) = 102 ± 34. EOS (N=2): 0' (-Mon) = 97 ± 35; 0' (+Mon) = 48 ± 13; 30' (-Mon) = 78.5 ± 45; 30' (+Mon) = 18.5 ± 12.

<u>Suppl. Fig. 2.</u> Ubiquitination-deficient β c K16R receptors show impairment in IL-5-stimulated endocytosis. Cy3-IL-5 (50 ng/ml) was added to cells grown on cover slips expressing WT β c and β c K16R receptors for 20 min on ice. Unbound Cy3-IL-5 was removed and cells were moved to 37°C for 1 hour. Cells were not acid washed to visualize the amount of Cy-3IL-5 remaining on the cell surface after internalization. Images were acquired and displayed as described in Fig. 2. Note reduced Cy3IL-5 internalization in β c K16R-expressing cells (white arrow, right panel) as compared to the vesicular Cy3-IL-5 pattern in cells expressing WT β c.

<u>Suppl. Fig. 3.</u> Ubiquitination-deficient β c receptors bind Cy3-IL-5. *A.* Pre-chilled and stablytransduced HEK293 cells expressing either WT β c, β c K16R, or β c K(1-3)R were incubated with 10 or 50 ng/ml Cy3-labeled IL-5 for cell surface binding as described in Experimental Procedures. Cell surface bound Cy3-IL-5 was measured by flow cytometry and is expressed as mean MFI±SEM minus unlabeled control MFI (N=3 for all, except for β c K(1-3)R at 10 ng/ml is N=2). For labels with 10 ng/ml: WT β c = 61 ± 16; β c K16R = 64 ± 0.5; β c K(1-3)R = 62 ± 5. For labels with 50 ng/ml: WT β c = 244 ± 43; β c K16R = 210 ± 48; β c K(1-3)R = 292 ± 50. *B.* Same as in (A) except pre-chilled cells grown on cover slips were incubated with 50 ng/ml Cy3-IL-5, washed extensively, and analyzed by deconvolution fluorescence microscopy. Shown are single middle Z-stack images to show outline of cell surface bound Cy3-IL-5.

<u>Suppl. Fig. 4.</u> β c K(1-3)R receptors accumulate on the cell surface similar to β c K16R. *A*. HEK293 cells stably expressing either WT β c, β c K16R, β c K(1-3)R, or β c Δ Box1 receptors were grown on cover slips and either left unstimulated (-IL-5) or stimulated with 10 ng/ml IL-5 for 1h (+IL-5). Cells were stained with anti- β c antibodies (BD Biosciences) followed by labeling with AlexaFluor-488 secondary antibody, and analyzed by deconvolution fluorescence microscopy as described in Figure 2. Note how cells expressing Ub-deficient β c K(1-3)R receptors accumulate on the cell surface like β c K16R, whereas β c Δ Box1 mutant receptors have an intermediate phenotype. *B*. Same as in (A) except β c cell surface levels were measured by flow cytometry after labeling unstimulated HEK293 cells with anti- β c-PE antibodies and are expressed as mean MFI±SEM (N=3) minus isotype control MFI: WT β c =

29,514 ± 16376; β c K16R = 64,467 ± 17908; β c K(1-3)R = 104,783 ± 9344; and β c Δ Box1 = 50,579 ± 5894. *C*. β c endocytosis assay is the same as described in Figure 6A except that β c K(1-3)R-expressing cells were analyzed in this figure. The MFI of immunoreactive β c receptors in both cell lines at 0' IL-5 (unstimulated) is represented as 100% and the loss of immunoreactivity (MFI) was plotted for each time point (N=2, except β c K16R N=1). MFIs: WT β c, 5'= 35%±4; β c K(1-3)R, 5'= 67%±0.2; β c K16R, 5'= 76%; WT β c, 10'= 22%±4; ; β c K(1-3)R, 10'= 55%±8; β c K16R, 10'= 54%; WT β c, 15'= 19%±3; ; β c K(1-3)R, 15'= 49%±5; β c K16R, 15'= 46%.