Table S1. Primers

pGADT7-TRIM28	*Eco*RI from PCR-amplified cDNA fragment using primers TTAGAATTCTTGCGTGATAGTGGCAGTAAGG and TAAGAATTCTGGTTCTACCAGCACAGCAG
pGBKT7-DBD-ZFP568 (full length)	*Eco*RI from PCR-amplified cDNA fragment using primers AATTGTCGACACCCAGCCTTGAAATACCAG and AATTGTCGACTGTTATCCACCACAGGGTTTT
pGBKT7-DBD-ZFP568 (KRAB domains)	*Eco*RI from PCR-amplified cDNA fragment using primers ATTGTCGACACCCAGCCTTGAAATACCAG and AATTGTCGACCTCACTGGCCTTTGCCTTAC
pGBKT7-DBD-ZFP568 (ZF domains)	*Eco*RI from PCR-amplified cDNA fragment using primers AATTGTCGACGTAAGGCAAAGGCCAGTGAG and AATTGTCGACTGTTATCCACCACAGGGTTTT
pAcGFP-ZFP568	*Bam*HI-*Xho*I from PCR-amplified cDNA fragment using primers TCAGATCTCGAGATGGAGCGCTTGTCCCAGATG and ACCGGTGGATCCCGTTCACTCCTCCGTCCTGTATG
pCMV-Myc-TRIM28	*Eco*RI from PCR-amplified cDNA fragment using primers TTAGAATTCGTCCGGCTGCTTCCTCAG and TAAGAATTCGTGGTTCTACCAGCACAGCAG
chatwo site directed mutagenesis	CCTGGCCCTGTTCTGGAATGAACCATGCCGTC and GACGGCATGGTTCATTCCAGAACAGGGGGAGG were used on pGADT7-TRIM28, pCDNA3-Flag-TRIM28, pcDNA3.1-Gal4DBD-TRIM28, pCMV-Myc-TRIM28 to generate *chatwo* mutant versions
D7CU11	CATGTCTATATGAGCATCCCAAGA and GAGTCTACTATGCAACAAGTGTCTTT
D7CU14	TGAGCCTACACGAGACACCA and GTCTAACCTGGGCCACACAG
chatwo-RFLP with *Bsl*I	ATGTGTTGTGTGGCCCAGTA and ACCTCTTGGCACCTGCAAC
Trim28 cDNA for in situ hybridization probe	TOPO cloning of PCR-amplified cDNA fragment with primers GTGGAGCCTCATGGTGAGAT and TCCAAGCCTGAGCTGGTACT
Trim28 for qRT-PCR	GTGGAGCCTCATGGTGAGAT and TACTTCCATGGGCTGGCTAC
Zfp568 for qRT-PCR	TGACCCAGCCTTGAAATACC and AGGCCTGGTCCTGTCTTCTT (Shibata and Garcia-Garcia, 2011)
IAP 5′UTR for qRT-PCR	CGGGTCGCGGTAATAAAGGT and ACTCTCGTTCCCCAGCTGAA (Rowe et al., 2010)
Gapdh for qRT-PCR	ACTGCCACCCAGAAGACTGT and GATGCAGGGATGATGTTCTG