

SUPPORTING INFORMATION:

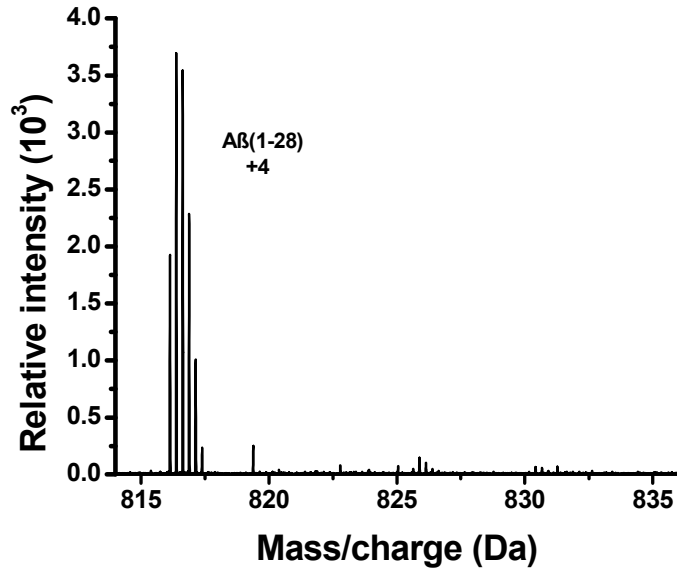


Figure S1. A portion of a positive-ion ESI-FTICR mass spectrum of A β (1-28) showing the quadruply charged ion of A β (1-28).

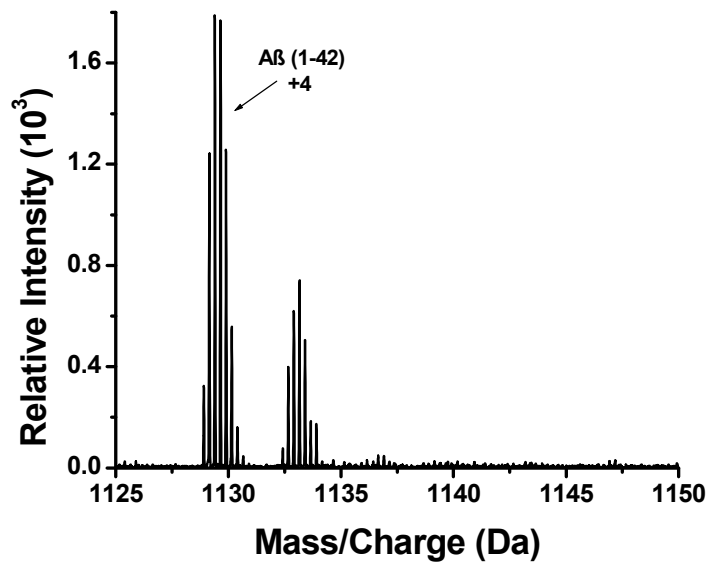


Figure S2. A portion of a positive-ion ESI-FTICR mass spectrum showing the quadruply charged ion of A β (1-42).

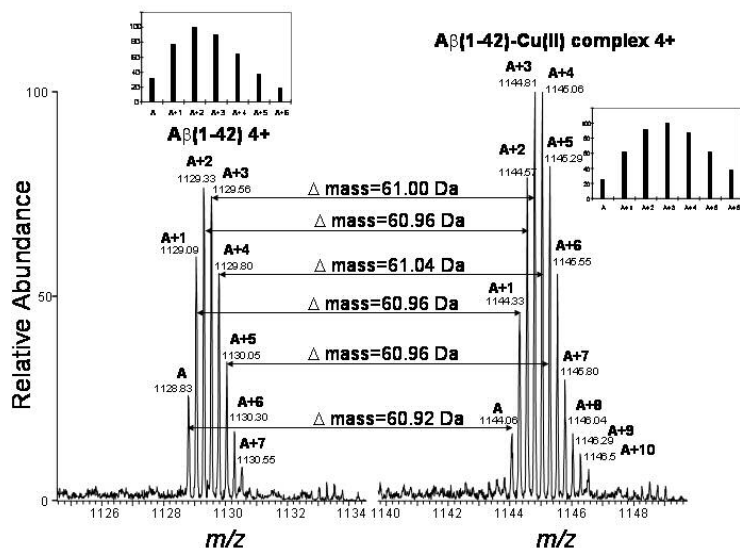


Figure S3. High-resolution mass spectrum of quadruply charged $A\beta(1-42)$ and $A\beta(1-42)-Cu(II)$ complex ions acquired on a linear-ion trap mass spectrometer operated in the high resolution, “ultra zoom scan” mode. A solution with an $A\beta(1-42)/Cu(II)$ molar ratio of 1:10 was employed for the measurement. Inset: theoretical isotopic distribution profiles of quadruply charged ions of $A\beta(1-42)$ and $A\beta(1-42)-Cu(II)$ complex.

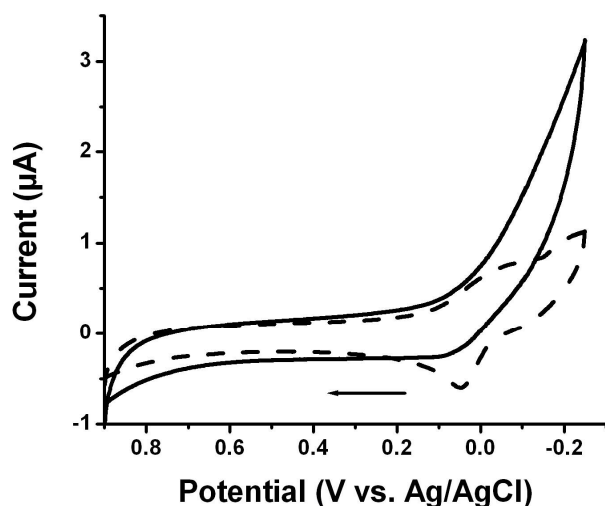


Figure S4. CVs of 100 μM $Cu(II)$ in an air-saturated 10 mM phosphate buffer solution (pH 7.4) containing 0.1 M Na_2SO_4 (solid curve) and a N_2 -purged solution (dashed curve). A glassy carbon electrode with a diameter of 3 mm was used. The scan rate was 0.020 V/s. The arrow indicates the initial scan direction.

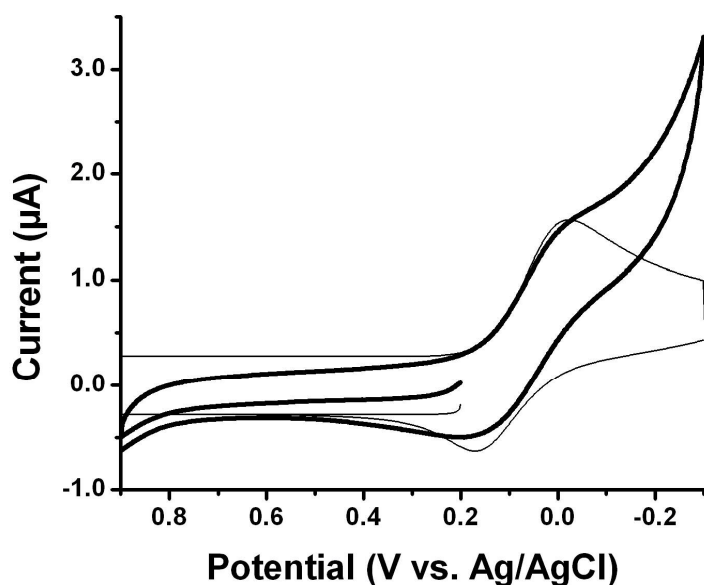


Figure S5. Simulated (thin solid curve) and experimental (thick solid curve) CVs of A β (1-16)-Cu(II) in the presence of O₂. The rate constant used for the one-electron reduction of A β (1-16)-Cu(II) complex was $5 \times 10^{-4} \text{ s}^{-1}$ and the redox reaction rate constant for the follow-up O₂ reduction to H₂O₂ and regeneration of A β (1-16)-Cu(II) was $1 \times 10^3 \text{ M}^{-1}\text{s}^{-1}$.

Simulation of the electron transfer process in Figure 3 was conducted with a CH simulation program. In performing the simulation, diffusion coefficients of $3 \times 10^{-6} \text{ cm}^2\text{s}^{-1}$ were used for the A β (1-16)-Cu(II) and A β (1-16)-Cu(I) complexes, based on the measured diffusion coefficient of a similar A β peptide.(*cf. Reference 1*) A diffusion coefficient of $1 \times 10^{-6} \text{ cm}^2/\text{s}$ was used for O₂. The concentration of A β (1-16)-Cu(II) was estimated to be *ca.* 0.2 mM by using the complexation constant of $1 \times 10^7 \text{ M}^{-1}$ (*cf. Reference 2*). The electrode surface area is 0.071 cm^2 , and the double-layer capacitance of the electrode is $14 \text{ }\mu\text{F}$. The O₂ concentration used is 0.26 mM, which is a value commonly found in an air-saturated aqueous solution.

In carrying out the simulation, we ignored the electrolytic reduction of O₂ at the electrode and the possible electroreduction of any uncomplexed Cu²⁺ in the solution. Both of these reductions occur right after the A β -Cu(II) reduction and might have contributed to the rising portion of the reduction current beyond 0.0 V. Despite these complications, the simulated and experimental voltammograms are in reasonable agreement, confirming the proposed mechanism (equations 1-2 in the text).

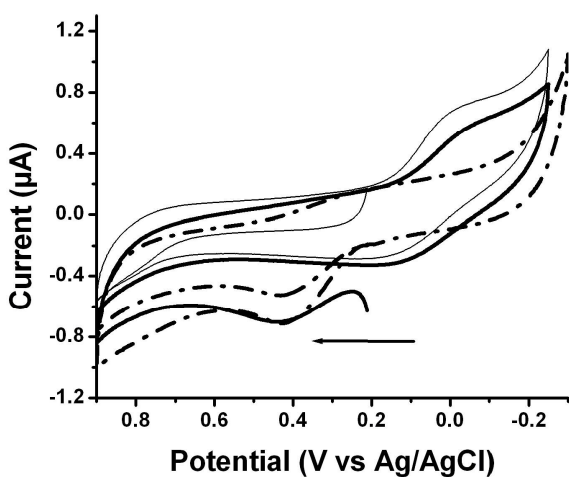


Figure S6. CVs of 100 μM ascorbic acid (dash-dot-dash curve), a mixture of 200 μM $\text{A}\beta(1-16)$, 200 μM Cu(II) , and 200 μM ascorbic acid (thin solid curve), and a solution containing 200 μM $\text{A}\beta(1-16)$, 200 μM Cu(II) , and 400 μM ascorbic acid (thick solid curve) in 10 mM phosphate buffer containing 0.1 M Na_2SO_4 (pH 7.4). A glassy carbon electrode with a diameter of 3 mm was used. The scan rate was 0.020 V/s. The arrow indicates the initial scan direction.

In acquiring the CVs, the potential was scanned from 0.2 V to 0.9 V to initiate the possible oxidation of ascorbic acid and continued to the cathodic direction to observe the reduction of $\text{A}\beta(1-16)\text{-Cu(II)}$ complex. Notice that the ascorbic acid oxidation peak disappeared during the second potential scan in the thick solid curve, whereas the ascorbic acid oxidation peak was well-defined and substantial when the $\text{A}\beta(1-16)\text{-Cu(II)}$ complex was absent (see the second potential scan in the dash-dot-dash curve).

Table S1: Extracting the copper oxidation state from mass spectral peaks of the A β (1-42)-Cu(II) complex. A solution with an A β (1-42)/Cu(II) molar ratio of 1:10 was used for ESI-MS measurement.

A β (1-42): Cu ²⁺ 1:10				A β (1-42)-Cu(II)			A β (1-42)-Cu(I)		
	Meas. <i>m/z</i>	Meas. Rel. Abund. (%)		Calcd. <i>m/z</i>	Calcd. Rel. Abund. (%)	Dev. (ppm)	Calcd. <i>m/z</i>	Calcd. Rel. Abund. (%)	Dev. (ppm)
4+ Ion	A	1144.06	16	1144.05	25	6	1144.31	25	215
	A+1	1144.30	49	1144.30	62	4	1144.56	62	198
	A+2	1144.55	86	1144.55	92	4	1144.81	92	207
	A+3	1144.80	100	1144.81	100	5	1145.06	100	216
	A+4	1145.04	99	1145.06	87	14	1145.31	87	216
	A+5	1145.29	79	1145.31	62	14	1145.56	62	234
	A+6	1145.53	50	1145.56	37	23	1145.81	37	226
	A+7	1145.78	27	1145.81	20	24	1146.06	20	226
	A+8	1146.04	18	1146.06	9	15	1146.31	9	235
	A+9	1146.29	10	1146.31	4	16	1146.56	4	235

Table S2. The reduction peak currents recorded at different A β /Cu(II) molar ratios. The A β concentration was kept at 200 μ M while varying the Cu(II) concentration. All the solutions were air-saturated.

A β (1-16)/Cu(II) molar ratio	I _{pc} (μ A)
4:1	0.31
2:1	0.65
4:3	0.92
1:1	1.25

References used to select some of the parameters for the simulation:

- (1) Mansfield, S. L., Jayawickrama, D. A., Timmons, J. S., and Larive, C. K. (1998) Measurement of peptide aggregation with pulsed-field gradient nuclear magnetic resonance spectroscopy. *Biochim. Biophys. Acta* 1382, 257-265.
- (2) Syme, C. D., Nadal, R. C., Rigby, S. E. J., and Viles, J. H. (2004) Copper binding to the amyloid-beta peptide associated with Alzheimer's disease. *J. Biol. Chem.* 279, 18169–18177.