

# Evolution and organization of the human protein C gene

(thrombosis/factor IX/coagulation)

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**ABSTRACT** We have isolated overlapping phage genomic clones covering an area of 21 kilobases that encodes the human protein C gene. The gene is at least 11.2 kilobases long and is made up of nine exons and eight introns. Two regions homologous to epidermal growth factor and transforming growth factor are encoded by amino acids 46-91 and 92-136 and are precisely delimited by introns, as is a similar sequence in the genes for coagulation factor IX and tissue plasminogen activator. When homologous amino acids of factor IX and protein C are aligned, the positions of all eight introns correspond precisely, suggesting that these genes are the product of a relatively recent gene duplication. Nevertheless, the two genes are sufficiently distantly related that no nucleic acid homology remains in the intronic regions and that the size of the introns varies dramatically between the two genes. The similarity of the genes for factor IX and protein C suggests that they may be the most closely related members of the serine protease gene family involved in coagulation and fibrinolysis.

Protein C is a two-chain vitamin K-dependent serine protease that plays a fundamental role in hemostasis by preventing coagulation and promoting fibrinolysis (1). The anticoagulant effects of protein C are achieved through cleavage of factors Va and VIIIa of the intrinsic pathway (2), while clot lysis appears to result from interaction of protein C with the inhibitor of plasminogen activator (3). First isolated from bovine plasma in 1976 by Stenflo (4), the human protein (5) has since been shown to be a 62-kDa dimer with a heavy chain that contains the active serine site and a light chain that contains, at its amino terminus,  $\gamma$ -carboxylglutamic acid residues, which are highly conserved among the vitamin K-dependent factors (6). The protein circulates as a zymogen (7), is activated by thrombin coupled with an endothelial cofactor, thrombomodulin (8), and is inactivated by a specific plasma protease inhibitor (9). Heterozygous deficiency of protein C was first identified by Griffin *et al.* (10) and has been shown by several workers to be an autosomally dominant disorder manifested by a markedly increased tendency to clot (11). Homozygous protein C-deficient patients, with no detectable antigenic levels of protein C, suffer from massive venous thrombosis as neonates (12). A deficiency of protein S, the cofactor for protein C-mediated inactivation of factor Va, has been reported as also causing increased thrombosis (13). Partial cDNAs for protein C were characterized by Foster and Davie (14), and a full length cDNA giving the complete amino acid sequence of the human protein has been isolated in our laboratory (36). We report the isolation of overlapping genomic clones of the protein C gene, analysis of the gene's organization, and relationships to other serine protease genes.

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## MATERIALS AND METHODS

**Isolation and Mapping of Genomic Clones.** Two human genomic libraries constructed at the *EcoRI* site of Charon 4a and Charon 28 were screened by the Benton and Davis technique (15) using a protein C cDNA (36) corresponding to amino acids 225-419 plus the 3' nontranslated region. Approximately  $6 \times 10^5$  phage plaques were screened and three overlapping fragments of human genomic DNA, each of  $\approx 20$  kilobase pairs (kbp), were mapped using the restriction enzymes *BamHI*, *EcoRI*, *HindIII*, *Pst I*, and *Sma I*.

**DNA Sequencing.** Genomic subfragments (see Fig. 1) from isolated  $\lambda$  phage were ligated into the appropriate restriction sites of plasmid pBR322, and the resulting chimeric plasmids were grown in *Escherichia coli* strain RR1 using standard procedures. Plasmid DNA was purified by CsCl banding as described elsewhere (36).

The strategy utilized to locate the desired regions for sequencing (intron-exon junctions and all exonic segments) was detailed Southern mapping (16) with specific radiolabeled protein C cDNA subfragments which represented the entire cDNA. By digesting with several different enzymes, small ( $\leq 600$  bp) hybridizing fragments were identified as suitable for direct sequencing. For the purpose of identifying fragments for sequencing, the modified bi-directional Southern transfer described by Smith and Summers (17) was used. Isolated DNA fragments were sequenced by the chemical modification method of Maxam and Gilbert (18) as outlined (36).

**Southern Blotting.** Total human genomic DNA was isolated as described (19) and subjected to restriction endonuclease digestion. The DNA fragments were then separated on agarose gels, transferred to nitrocellulose, and hybridized as described by Southern (16), using cDNA probes corresponding to several regions of the protein C precursor.

**Primer Extension.** One microgram of a synthetic oligonucleotide [5' d(GGGGCGGGTCGTGGAGATACTCG)] corresponding to nucleotides 30-53 as shown in Fig. 2 was hybridized with 3  $\mu$ g of human liver poly(A)-selected RNA in 60% (vol/vol) formamide, 0.4 M NaCl, 20 mM Pipes (pH 6.4), and 2 mM EDTA, by heating to 85°C for 5 min and allowing the water bath ( $\approx 1$  liter) to come to room temperature over a period of about 4 hr. The hybrids were recovered and primer extension reactions carried out and analyzed on 8% sequencing gels as described (20).

## RESULTS

**Organization of the Protein C Gene.** From the  $6 \times 10^5$  phage genomic clones screened, three phage clones, designated  $\lambda$ pc4,  $\lambda$ pc14, and  $\lambda$ pc17, hybridized to our full length cDNA probe. By restriction endonuclease mapping, these clones were found to overlap as illustrated in Fig. 1. The region mapped includes the entire gene of 11.2 kbp, as well as 6 kbp of 5' flanking DNA, and 4 kbp of 3' flanking DNA.

Abbreviation: bp, base pair(s).

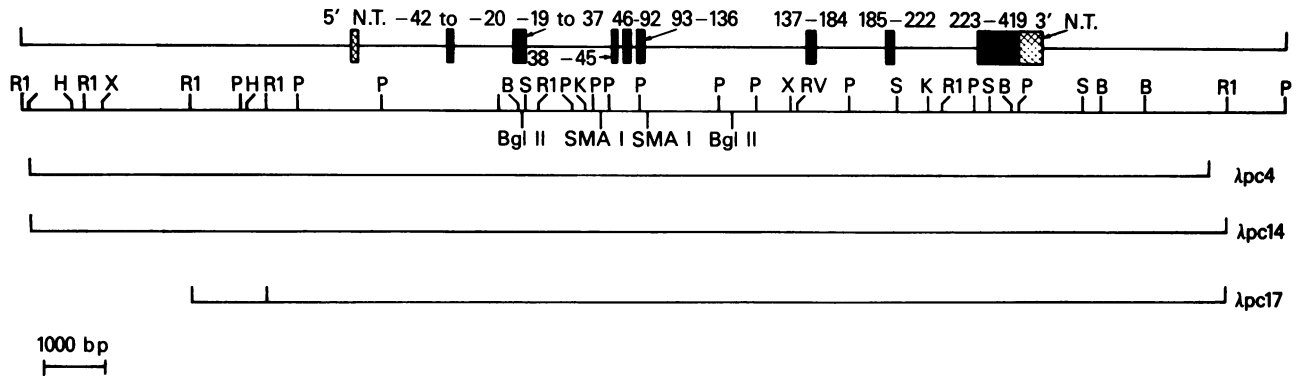


FIG. 1. Organization of the human protein C gene. The first line shows the positions of exons as rectangles on the chromosomal DNA. Numbers above the line indicate the amino acids at which intron/exon junctions occur. Nontranslated regions are shown as cross-hatched areas and coding areas are in black. The second line gives the positions of restriction endonuclease recognition sites. The straight lines below indicate the regions of overlap of the three phage clones: λpc4, λpc14, and λpc17. For the purpose of DNA sequencing, the 4.6-kbp *HindIII/BamHI*, 8.3-kbp *BamHI* and 1.3-kb *BamHI* fragments were inserted into plasmid pBR322. The abbreviations used are as follows: X, *Xba* I; H, *HindIII*; R<sub>1</sub>, *EcoRI*; P, *Pst* I; B, *BamHI*; S, *Sst* I; K, *Kpn* I; R, *EcoRv*. Note from Fig. 2 that a small 5' noncoding region is present in the second exon.

The locus mapped in Fig. 1 represents the only locus closely homologous to protein C in the human genome since analysis of total human genomic DNA by Southern blotting of DNA digested with several restriction enzymes reveals

only those restriction fragments expected from the locus shown in Fig. 1 (data not shown).

**Sequence Analysis.** The sequence of the exons and their flanking DNA is shown in Fig. 2. The coding sequence and

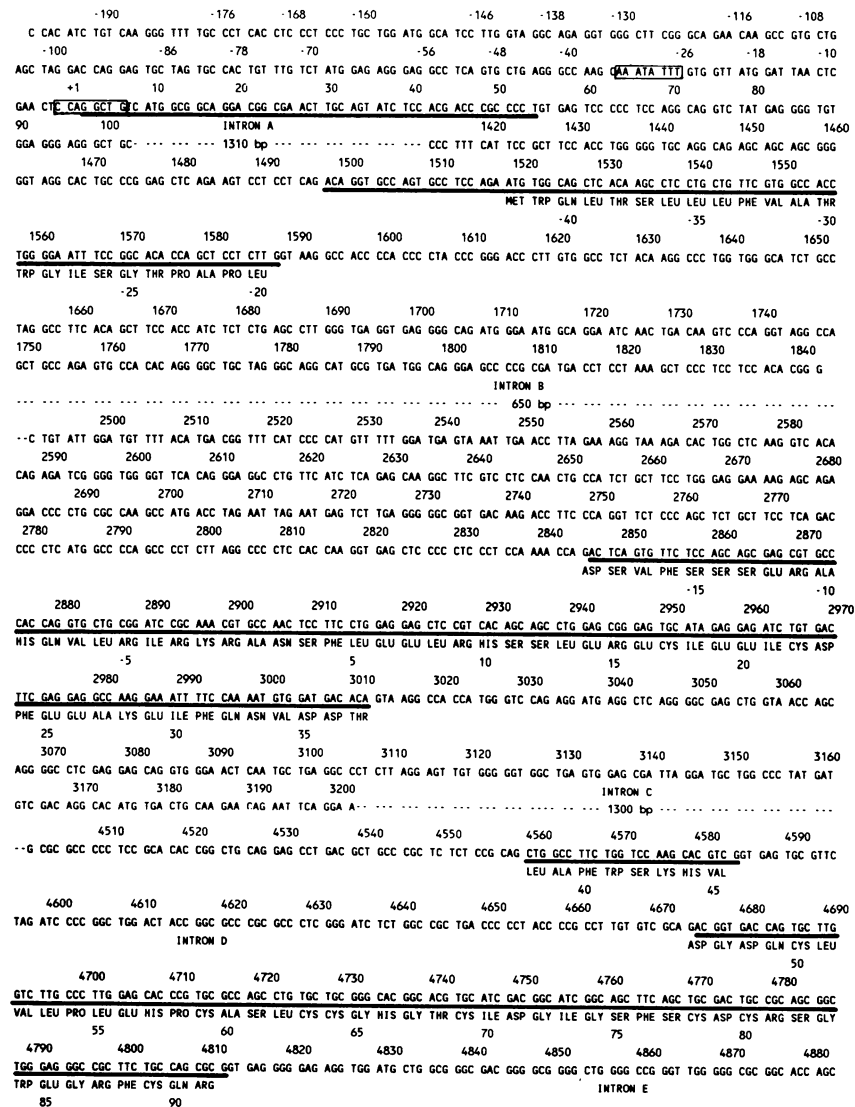


FIG. 2. (Figure continues on the opposite page.)

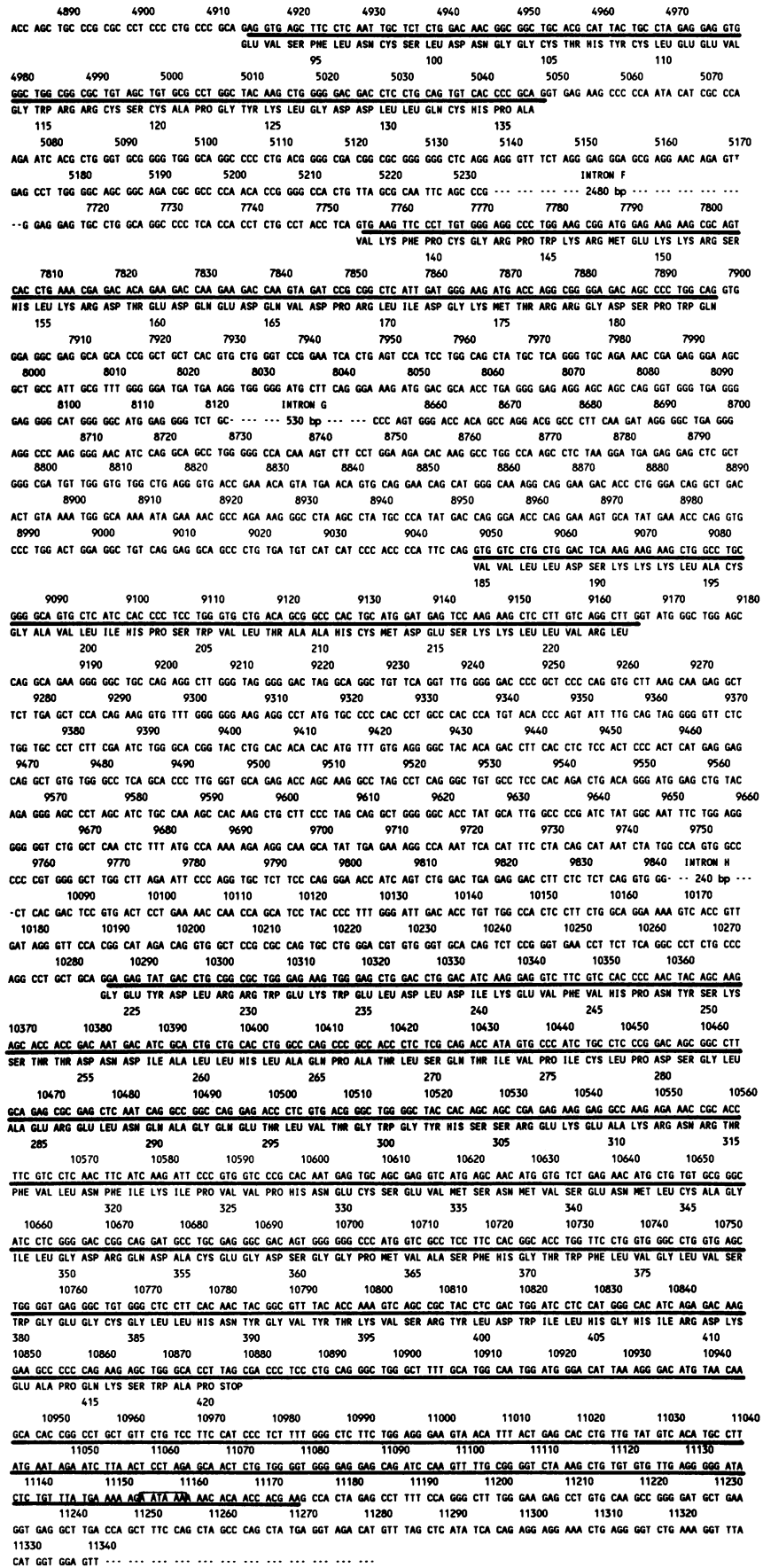


FIG. 2. Nucleic acid sequence of the protein C gene. Bases are numbered relative to the proposed transcription initiation site. Exons are underlined and amino acids are numbered from the amino-terminal residue in the plasma protein. Gaps in the sequence are shown as dashed lines, with the approximate length of the gap noted. Regions corresponding to a TATA box (-34 to -25), a transcriptional start site (-2 to +6) and a polyadenylation recognition site (11,155 to 11,160) are boxed.

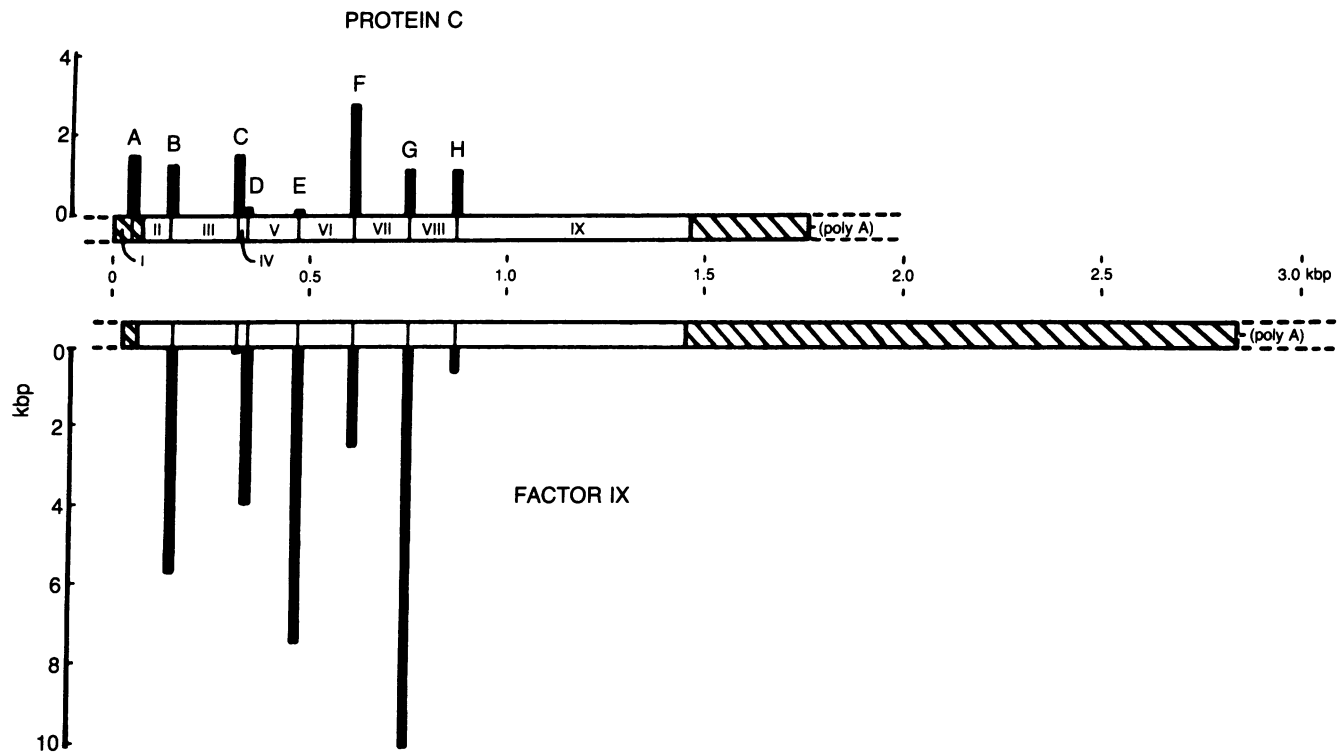


FIG. 3. Comparison of the size of introns and exons in the protein C gene. The length in bp of exons is shown by the horizontal bars while the length of each intron is proportional to the height of each vertical bar. The horizontal bars have been aligned on the basis of amino acid sequence homology. Hatched horizontal regions represent noncoding regions of the mRNAs.

5' and 3' nontranslated regions are divided into nine exons by eight introns. There is perfect agreement between the sequence of the exons and the full length cDNA sequence (36). This again indicates that only a single copy of the protein C gene is present in the human genome. Analysis of the sequences at the borders of introns is shown in Fig. 3 and reveals that in every case the GT/AG rule (21) is observed. The entire length of the gene is 11.2 kbp; hence 83% of the gene consists of intronic sequences. The 3' nontranslated region contains a polyadenylation signal (AAUAAA) 21 nucleotides from the termination codon and polyadenylation occurs on either an adenosine or on the preceding guanosine.

**Transcription Initiation Site.** Comparison of the total length of the cDNA sequence with the mRNA size based upon RNA

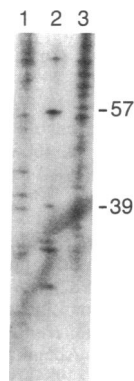


FIG. 4. The start-site for transcription of the protein C gene. Primer extension was carried out. Lanes 1 and 3, M13 sequence ladders used to obtain size markers. The figures shown on the right are the length, in bp, from the end of the oligonucleotide used for primer extension. The cluster of fainter bands at the top of the gel correspond to 72–85 nucleotides from the end of the oligonucleotide.

blot hybridization (36) suggests that the cDNA is very close to full length. Analysis of the genomic sequence immediately upstream from the region corresponding to the mRNA sequence (underlined in Fig. 2) reveals one potential transcriptional start site, one base upstream from the cloned cDNA sequence. This proposed start site is based upon sequence similarity with the transcriptional start site consensus sequence (PyCAPyPyPyPyPy) reported by Corden *et al.* (22), and its position 30 bases downstream from a potential "TATA" box (21). Alternatively, the adjacent upstream genomic sequence could be a part of a second intron in the 5' untranslated region.

Analysis of transcripts produced by primer extension using an oligonucleotide (Fig. 4) corresponding to positions 30–53 of the protein C sequence indicated a start site 54–56 nucleotides from the end of the primer corresponding to the nucleotides about 1 bp upstream of the cDNA sequence shown in Fig. 2. In addition, several weaker bands corresponding to longer and shorter transcripts were present on the gel (Fig. 4), and it is unclear if these longer transcripts represent additional sites of initiation of transcription or crosshybridization to other mRNAs.

## DISCUSSION

**Comparison of the Gene Structures of the Serine Proteases.** Protein C, like many of the blood coagulation proteins, is a serine protease and exerts its role in blood coagulation by virtue of an active-site serine in the heavy chain. Neurath *et al.* (23) have found that each of these genes is likely to be derived from the same primitive gene, and chymotrypsin, trypsin, and elastase are generally felt to be the archetypal serine proteases. The trypsin gene family can be traced to prokaryotes by virtue of sequence homology between mammalian, invertebrate, and prokaryotic trypsins. In this evolutionary pathway, invertebrate trypsin from crayfish repre-

sents the link, having homology to mammalian and prokaryotic trypsins (24).

Evolutionary relatedness between proteins is commonly established by comparing amino acid homology between the proteins. However, with the ability to study the structure of the genes encoding these proteins, it is possible to provide an independent test of these estimates of relatedness, in that the number and positions of introns are likely to be a relatively immutable feature and provide an independent estimate of the relatedness of two proteins. Several groups have explored the similarities of serine proteases (see ref. 25) and with the same objective we have studied the positions of introns in the genes of some of the members of the serine protease family. Remarkably, a comparison of the gene region corresponding to the serine protease domain of protein C with the rat genes (25) for trypsin (three introns), chymotrypsin (five introns), and elastase (six introns) reveals that the sole protein C intron position within this domain aligns only with the first intron of elastase. In contrast, when homologous amino acids are aligned, the position of all eight introns of factor IX (26) and protein C correspond (Fig. 3). This fact attests to the close evolutionary relationship between the two genes and suggests that they are products of a relatively recent duplication.

Despite the conservation of exon size and sequence between protein C and factor IX, the sizes of the introns have diverged remarkably (Fig. 3). Furthermore, no recognizable homology exists between intron sequences of the two genes among the 3 kbp of intronic sequences presented in this paper [compare Fig. 2 of this manuscript with Fig. 4 of Anson *et al.* (26)].

Banjai *et al.* (27) have noted a region within several coagulation factors with close homology to epidermal growth factor (28) and, more recently, to the transforming growth factors produced by retroviral-infected cells (29). Interestingly, these regions (amino acids 46–91 and 92–136 in the protein C sequence) is sharply delimited by exons in protein C, factor IX, factor X, and tissue plasminogen activator (30). The conservation of these regions and its presence on a single exon in such diverse proteins as tissue plasminogen activator and protein C suggests that these regions were incorporated into the structure of the genes before the original duplication and have maintained a remarkable degree of homology to transforming growth factor and epidermal growth factor through several hundred million years of evolution. In particular, the position of each half cysteine bond remains the same in this region, suggesting that each of these primary structures has a similar tertiary structure. Thus it is possible that these regions have some conserved function, possibly related to that of the growth factors.

Prothrombin, another vitamin K-dependent serine protease coagulation factor, does not contain the growth factor domain but has in its place two "kringle" structures, first described by Magnusson *et al.* (31). Prothrombin, however, does have clear homology with other vitamin K-dependent coagulation factors in the leader peptide,  $\gamma$ -carboxylglutamate or GLA region and serine protease domains (6, 32, 36).

Congenital deficiency of protein C is an autosomally dominant disease characterized by superficial and deep venous thrombosis (11) occurring during childhood or early adulthood. The heterozygous form of the disease has an incidence of 1 per 16,000 individuals (33). Individuals that inherit two defective protein C alleles have catastrophic thrombotic events as neonates that are invariably fatal without treatment (13). Because individuals with a defective protein C allele do not always manifest the disease or have abnormally low levels of the protein (34), a precise genetic test for the defective allele would be useful. With the organization of the normal protein C gene defined, the

feasibility of genetic testing for hereditary protein C deficiency can now be determined.

**Note Added in Proof.** Since submission of this manuscript, an additional sequence for the human protein C gene has been published (35).

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