Supporting Information

Synthesis, Photophysical, Electrochemical, Tumor-Imaging and Phototherapeutic Properties of Purpurinimide-N-substituted Cyanine Dyes Joined with Variable Length of Linkers

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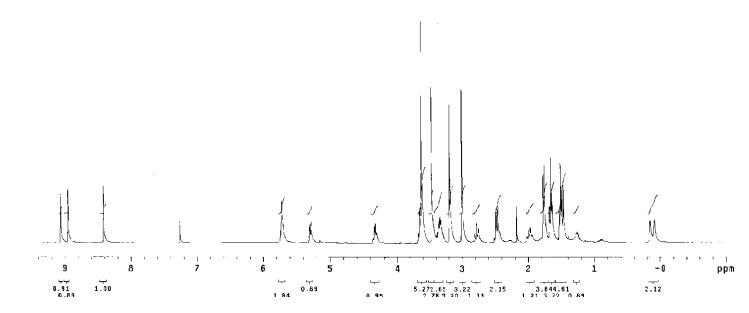
Page S11-S12: HPLC chromatogram of conjugates 7-9

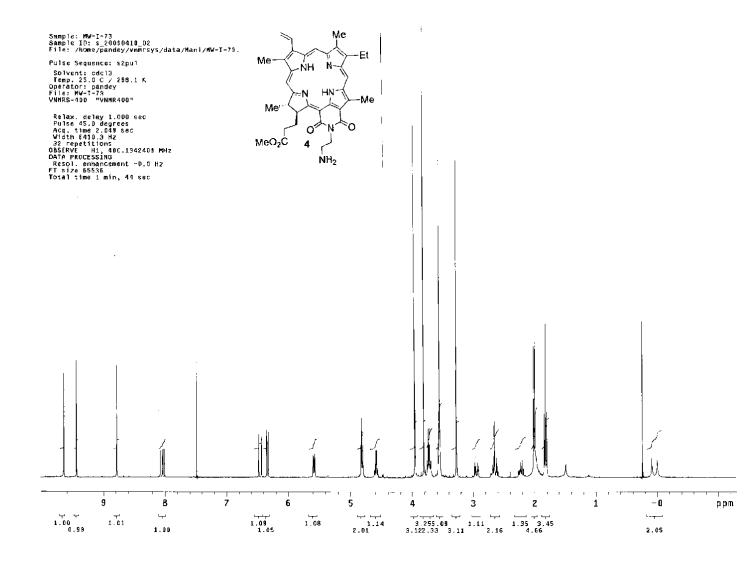
In Vitro Phototoxicity Assay (MTT Assay): To determine the in vitro efficacy of the compounds the following procedure was performed. Colon-26 cells were grown in RPMI 1640 with 10% FCS, 2mM L-Glutamine, 50 IU penicillin/50 µg/ml streptomycin. Cells were maintained in 5 % CO₂ and 95% air at 100% humidity. For phototoxicity studies, Colon-26 cells were plated in 96 well plates at a density of 3500 cells/well in complete medium. 4 h later, compounds were added at concentrations ranging from 0.003 to 1 µM. After 24 h incubation in the dark at 37 °C, the cells were irradiated with laser light from an argon pumped dye laser using fluences of 0-2 J/cm² at a dose rate of 3.2 mW/cm² with light adjusted to the specific maximum wavelength of each drug. The light treatment is shown in Figure 13. After irradiation, the cells were incubated for 48h. After 48 h, 15 µL/well of 4 mg/ml 3-(4,5-dimethylthiazol-2-yl)-2,5diphenyltetrazolium bromide MTT in PBS was added, and cells were incubated for a further 4 h. MTT is a yellow water soluble tetrazolium salt that is taken up by cells, reduced to water insoluble purple formazan crystals by metabolically active (viable) cells. The MTT-containing media was removed, and 100µl DMSO was added to solubilize the formazan crystals. The absorbance of the wells was read on a microtiter plate reader at a wavelength of 560 nm. The results were plotted as percent growth of control (untreated) cells vs. concentration range for each individual drug.

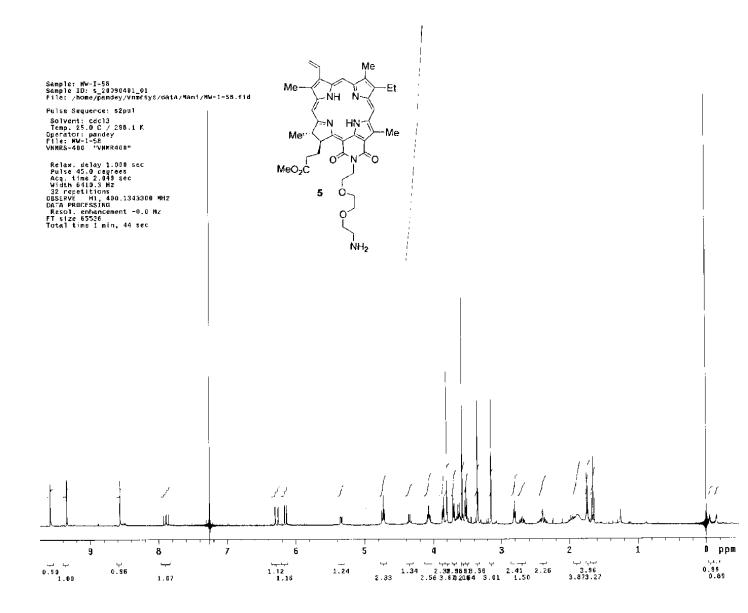
In Vivo Reflectance Spectroscopy: In vivo reflectance spectroscopy is a non-invasive method of following the pharmacokinetics and measuring the concentration of a photosensitizer in an experimental animal. By measuring the light diffusely reflected from tissue containing the dye, the time at which the concentration is at its maximum can be determined and the light treatment can be delivered at the optimal tumor to normal tissue ratio, for the greatest selectivity. As described in reference 9, monochromatic light was delivered through a quartz fiber which was placed perpendicular to and touching the tissue. At a measured distance (3-5 mm) from the delivery fiber, a pick up fiber is also placed in contact with the tissue. Light is delivered at optical power levels of about 1μ W to avoid any potential PDT effects during measurement. Light

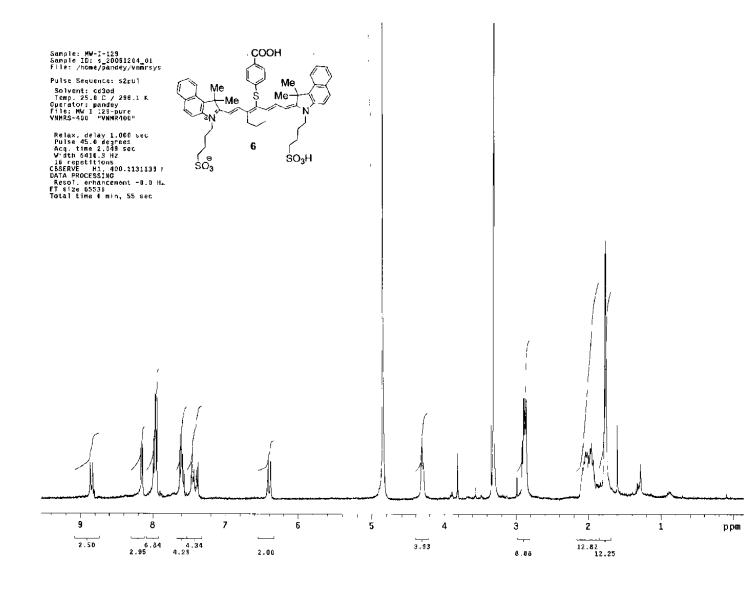
$$C = \frac{1}{2 * \varepsilon} \left\{ \left\{ -\sqrt{\mu_s^2 + \frac{4}{3}\alpha_0^2} + \sqrt{\mu_s^2 + \frac{4}{3}\left(\frac{OD}{r} + \alpha_0^2\right)^2} \right\}$$
 Equation 1

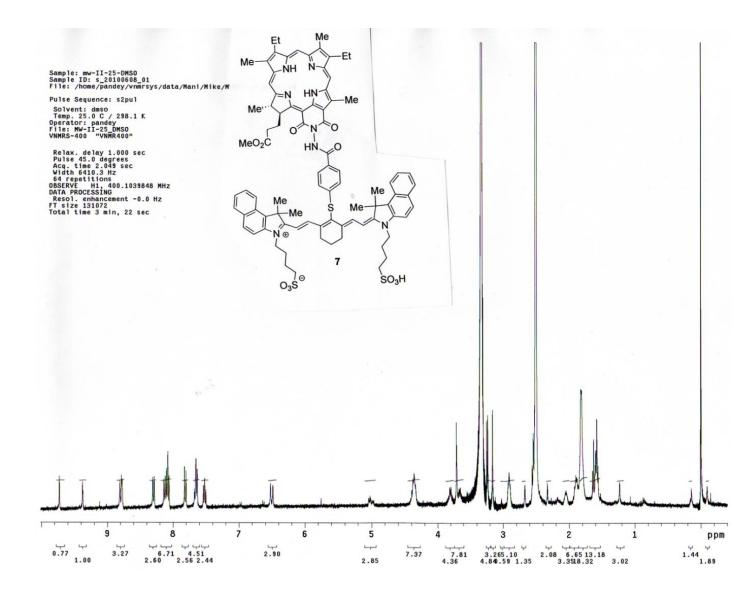
that enters the pickup fiber was conducted to a silicon photodiode detector that measured the photocurrent linear in power over 7-8 orders of magnitude. The wavelength was varied from 650-1000 nm and a spectrum of diffusely scattered photons was recorded. The concentration C was calculated using the formula in equation 1. Where ε is the molar extinction coefficient, $\alpha 0$ is the intrinsic tissue coefficient, OD is diffuse optical density, μ_3 is the reduced scattering coefficient, r is the distance from the detector to the pickup fiber. Molar extinction coefficient (ε) is measured using the Beer- Lambert law ($A=\varepsilon$ lc).

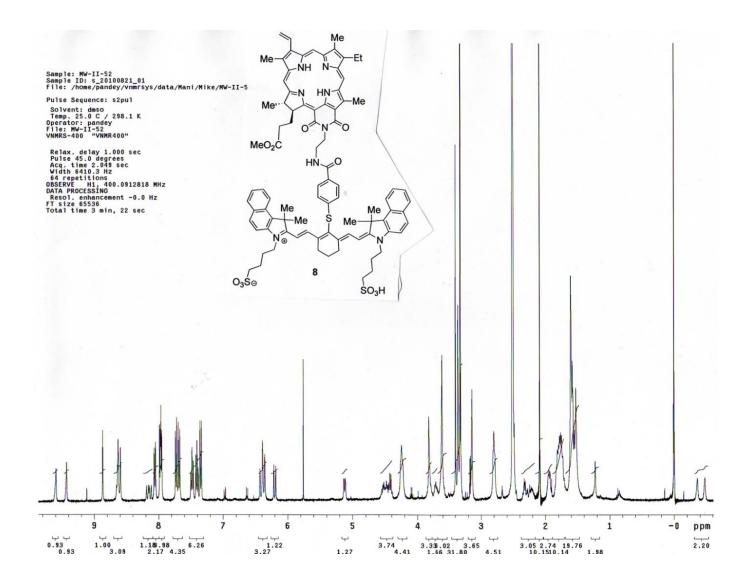


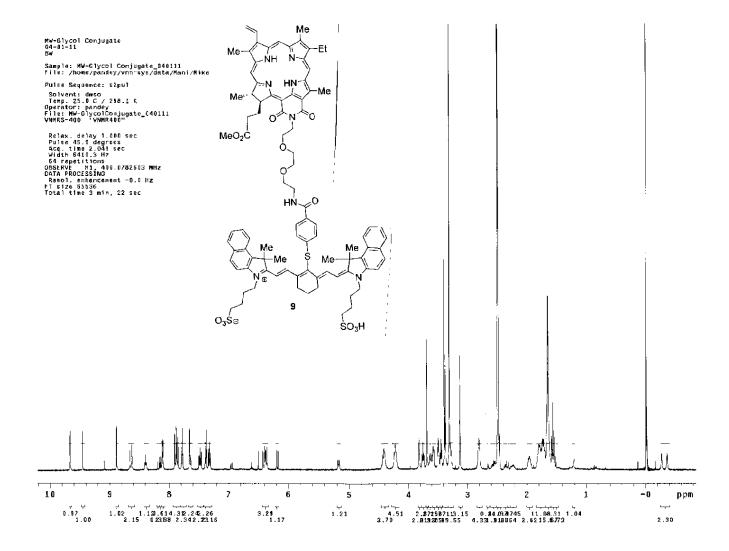












HPLC Analysis

Method: HPLC analysis of conjugates # 7, # 8, and # 9 was carried out using a Waters Delta 600 System consisting of the 600 Controller, 600 Fluid Handling Unit and 996 Photodiode Array Detector equipped with a Waters Symmetry C18 column, 5 micron, with dimensions 4.6 x150mm. The mobile phase flow program was as follows: flow rate of 1.0 ml / min; mobile phase composition - 75% Methanol/ 25% water from 0 to 20 minutes, then a linear gradient to 100% Methanol from 20 minutes to 30 minutes, maintained at 100% Methanol from 30 minutes on.. Data collected and spectrally monitored from 300 nm to 799 nm. Percent purity of compounds based on 416 nm channel.

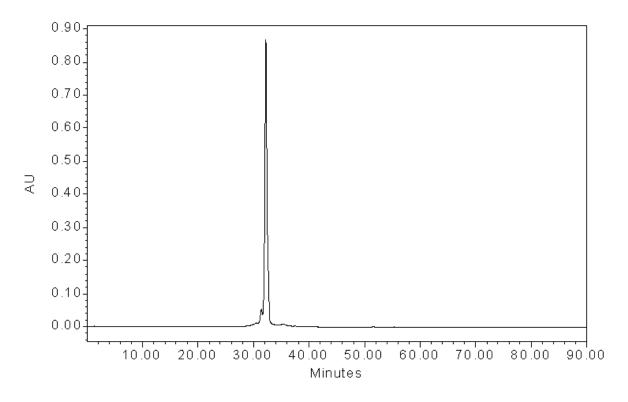


Figure S1: HPLC Chromatogram of Conjugate # 7

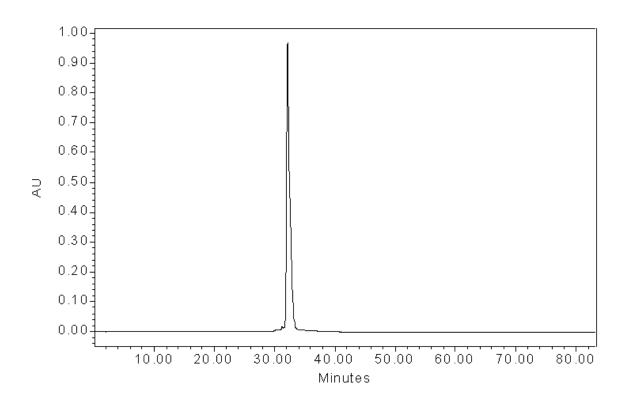


Figure S2: HPLC Chromatogram of Conjugate #8

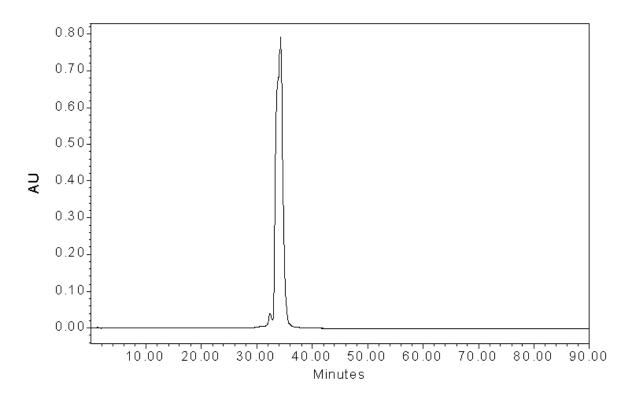


Figure S3: HPLC Chromatogram of Conjugate # 9