Supplemental Data

Structure function analysis of an ADP-ribosyltransferase Type III Effector and its RNA-Binding Target in Plant Immunity

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| Structure refinement statistics | | |
|---------------------------------------|-------------|--|
| Resolution range (Å) | 50.0-2.70 | |
| $R_{ m work}/R_{ m free}~(\%)^{ m b}$ | 17.87/21.18 | |
| R.M.S. Deviation from ideality | | |
| Bonds, (Å) | 0.010 | |
| Angles, (°) | 1.319 | |
| Average B factor, (Å ²) | 27.49 | |
| Ramachandran plot statistics | | |
| Most favored regions (%) | 89.7 | |
| Allowed regions (%) | 8.9 | |
| Generously allowed regions (%) | 1.4 | |

Table S1. Refinement statistics of HopU1 structure^a

^a the Crystallization and preliminary crystallographic analysis has been published in Lin et al.

(2010. Acta Crystallogr Sect F Struct Biol Cryst Commun 66: 932-934).

 ${}^{b}R_{free} = \sum_{Test} ||F_{obs}| - |F_{calc}|| / \sum_{Test} |F_{obs}|$, where "Test" is a test set of about 5% of the total

reflections randomly chosen and set aside prior to refinement for the complex.

Figure S1. Structure comparison of HopU1 and selected ADP-ribosyltransferases. (A) Structure of the HopU1 ADP-RT is shown as ribbon representation and colored in purple blue and L1 and L4 loops are colored yellow (**B-D**) Structures of the rat Ecto-ADP-RT ART2.2 (1GXZ.PDB), the *Clostridium botulinum* C3bot2 ADP-RT (1R45.PDB), and the *L Clostridium limosum* C3 exoenzyme ADP-RT (3BW8.PDB) are shown as ribbon representation and colored in green, purple, and red, respectively.(**E**) Superimposition of the four ADP-RT structures. (**F**) PN loop and ARTT loop are indicated in compared structures. (**E-F**) The structures are colored as in Fig. S1A-D.

Figure S2. (A) Structures of HopU1 separately indicating the location of mutations M5-M9 and then summarized in central square. The altered residues are highlighted and shown as stick representations. (B) Isothermal titration calorimetry of wild type HopU1 and HopU1 mutants M5-M9 with GRP7, which indicates the extent that HopU1 derivatives can interact with GRP7. (C) The electrospray ionization-mass spectrometry results of ADP-RT activity for wild type HopU1 and HopU1 derivatives are shown. The peak of substrates and products of each enzymatic reaction is indicated. The molecular mass of GRP7 (1-90, GPHM after 3C cleavage at N terminus) is 11,260 Dalton and GRP7 with an ADP ribose modification is 11,800 Dalton. (D) Result summary of the ADP-RT activity and substrate binding affinity for wild type HopU1 and HopU1 mutants M1-M9.

Figure S3. MS/MS spectrum of doubly charged ion of m/z 1029.92 in the tryptic digest of GRP7.

The spectrum contains a nearly complete series of y-type ions. Additional ions supporting ADP ribosylation of this peptide observed in the MS/MS spectrum are protonated adenine (m/z 136), AMP (m/z 348) and ADP (m/z 428).

Figure S4. Electrophoretic mobility shift assay of an RNA probe with differing amounts of GRP7-GST after treatment with HopU1 or its catalytic inactive mutant HopU1_{DD}. Standard ADP-ribosylation reactions were performed with varying concentrations of GRP7-GST in the presence of HopU1 or HopU1_{DD}, and a ³²P-labeled probe (ATGRP7 UTR WT) was added to each reaction mix. These were run on native polyacrylamide gels, and exposed to X-ray films.

Figure S5. Expression of GRP7 in different transgenic *A. thaliana* **lines.** Immunoblot using anti-HA antibodies showing the expression of GRP7-HA and GRP7_{R49K}-HA in *A. thaliana* Col-0 and the Col-0 grp7-1 mutant.

Figure S6. Superimposed HopU1 and AvrPphF structures. Two views (**A**, **B**) of the HopU1 structure (blue) superimposed with the AvrPphF structure (light green). The root mean square deviation (RMSD) of 2.9 angstrom for 64 C α atoms indicating that the structures did not superimpose well. The N-termini of both proteins are quite different. The C-terminal catalytic domains of both proteins share a similar fold found in ADP-RTs.



Supplemental Figure 1



Supplemental Figure 2



Supplemental Figure 3



Supplemental Figure 4



Supplemental Figure 5



Supplemental Figure 6