

Supporting Information

Selective Molecular Sequestration with Concurrent Natural Product Functionalization and Derivatization: From Crude Natural Product Extracts to a Single Natural Product Derivative in One Step

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General Information

Commercially available ACS grade solvents were used without further purification. Peppercorns were purchased from Pepper-Passion Inc. (Issaquah, WA, www.pepper-passion.com), Allspice, cloves and ground cloves were obtained from Martin's supermarket, South Bend, IN, USA. The Rink resin (100-200 mesh, 1% DVB, 0.75 mmol/g) and Wang resin (100-200 mesh, 1% DVB, 1.0 mmol/g) were from Advanced ChemTech (Louisville, KY, www.peptide.com), the hydroxymethyl resin (100-200 mesh, 1% DVB, 0.98 mmol/g) from NovaBiochem (San Diego, CA, www.emdbiosciences.com/html/NBC/home.html). Synthesis was carried out on Domino Blocks in disposable polypropylene reaction vessels. A Labquake Tube Rotator was used for gentle but efficient tumbling of resin slurry.

All NMR spectra were recorded using 600 MHz or 800 MHz spectrometers. ^1H and ^{13}C chemical shifts, δ_{H} and δ_{C} , are reported relative to the corresponding residual solvent signal (CD_3OD $\delta_{\text{H}}=3.31$ ppm, $\delta_{\text{C}}=49.00$ ppm; CD_3CN $\delta_{\text{H}}=1.94$ ppm, $\delta_{\text{C}}=1.39$ ppm).

All reactions were carried out at ambient temperature (~ 21 °C) unless stated otherwise. The volume of wash solvent was 10 mL per 1 g of resin. For washing, resin slurry was shaken with the fresh solvent for at least 1 min before changing the solvent. After adding a reagent solution, the resin slurry was manually vigorously shaken to break any potential resin clumps. Resin-bound intermediates were dried by a stream of nitrogen for prolonged storage and/or quantitative analysis.

For the LC/MS analyses, a sample of resin (~ 5 mg) was treated with TFA in dichloromethane (DCM), the cleavage cocktail was evaporated by a stream of nitrogen, and cleaved compounds extracted into 0.5 mL of MeOH. Alternatively, a sample of resin-bound product on a silicon-based linker was cleaved by 0.1 M TBAF in THF for 30 min, the solution was diluted with MeOH and analyzed.

Types of peppercorn and amounts of materials extracted.

Table 1. Total extracts and content of piperine and piperettine in peppercorns.

Entry	Peppercorn	Total extract	piperine	piperettine
1	Lampong Black	292 mg	116 mg	19.7 mg
2	Malabar Black	241 mg	77 mg	14.5 mg
3	Madagascar Black	248 mg	89 mg	16.6 mg
4	Sarawak Black	204 mg	61 mg	12.6 mg
5	Tellicherry Black	248 mg	71 mg	14.8 mg
6	Talamanca Del Caribe Black	252 mg	80 mg	8.7 mg
7	Vietnamese Black	223 mg	63 mg	11.0 mg
8	Pohnpei peppercorns	278 mg	82 mg	15.1 mg
9	Muntok White	213 mg	68 mg	12.3 mg
10	Talamanca Del Caribe White	249 mg	87 mg	9.3 mg
11	Sarawak Extra Fancy White	227 mg	60 mg	12.3 mg
12	Freeze Dried Green peppercorns	315 mg	68 mg	15.8 mg

Table 2. The effect of spice, its quantity and freshness on yield.

Entry	Spice	Quantity	Age	Yield
1	Lampong Black	50 mg	freshly ground	2 %
2	Lampong Black	100 mg	freshly ground	7 %
3	Lampong Black	200 mg	freshly ground	19 %
4	Lampong Black	400 mg	freshly ground	37 %
5	Lampong Black	800 mg	freshly ground	46 %
6	Lampong Black	800 mg	freshly ground	45 %
7	Lampong Black	800 mg	1 day old ^a	21 %
8	Lampong Black	800 mg	2 days old ^a	20 %
9	Lampong Black	800 mg	2 days old ^a	18 %
10	Lampong Black	800 mg	2 days old ^b	36 %
11	Lampong Black	800 mg	4 days old ^a	6 %

12	Allspice	800 mg	freshly ground	20 %
13	cloves	800 mg	freshly ground	69 %
14	ground cloves	800 mg	freshly opened	65 %

Note: Yield was estimated from UV response on LC traces and calculated with respect to the initial resin loading; ^aexposed to open air; ^bin a sealed container

The LC/MS analyses were carried out using a 3 x 50 mm C18 reverse phase column. Mobile phases: 10 mM ammonium acetate in HPLC grade water (A) and HPLC grade acetonitrile (B). A gradient was formed from 5% to 80% of B in 10 min at 0.7 mL/min. The MS electrospray source operated at capillary voltage 3.5 kV and a desolvation temperature 300 °C.

Purification was carried out on a C18 column (19 x 100 mm, 5 µm particles). A solvent gradient was formed from 10 mM aqueous ammonium acetate and acetonitrile with a flow rate of 15 mL/min.

General Procedure for Agar Diffusion Antibiotic Susceptibility Assay

Antibacterial activity of the compounds was determined by an agar diffusion assay.¹ All test organisms used in this study were from sources provided in **Table 3**. Overnight cultures of test organisms were grown in LB broth for 18-24 hours and standard suspensions of $\sim 1.5 \times 10^6$ cfu/mL were prepared in sterile saline solution (0.9% NaCl) according to a BaSO₄ 0.5 McFarland Standard (bioMérieux, Inc).^{2,3} This standardized suspension (0.1 mL) was added to 34 mL of sterile, melted, and tempered (47-50 °C) Mueller-Hinton No. 2 agar (HiMedia Laboratories). After gentle mixing, the inoculated melted agar was poured into a sterile petri dish (145 mm x 20 mm, Greiner Bio-One) and allowed to solidify. Wells of 9 mm diameter were cut from the petri dish agar and filled with 50 µL of the test sample solution. The petri dish was incubated at 37 °C for 18-24 hours and the inhibition zone diameters were measured (mm) with an electronic caliper after 24-48 hours. The results are summarized in **Table 4**.

Table 3. Test organisms used in this research.

Strain	Marker	Origin/Reference
Gram-positive bacteria		
<i>Bacillus subtilis</i> ATCC 6633	Wild Type	American Type Culture Collection
<i>Staphylococcus aureus</i> SG511	Wild Type	Hans Knöll Institute, Jena, Germany
<i>Mycobacterium vaccae</i> IMET 10670	Wild Type	Hans Knöll Institute, Jena, Germany
<i>Mycobacterium smegmatis</i> mc ² 155	Wild Type	Snapper et al. 1990 ⁴
<i>Micrococcus luteus</i> ATCC 10240	Wild Type	American Type Culture Collection
Gram-negative bacteria		
<i>Pseudomonas aeruginosa</i> KW799/WT	Wild Type	Zimmermann 1980 ⁵
<i>Pseudomonas aeruginosa</i> KW799/61	Antibiotic Susceptible Penetration Mutant	Zimmermann 1980 ⁵
<i>Escherichia coli</i> X580	β-Lactam Hypersensitive	Eli Lilly & Co.

Table 4. Antibacterial activity of compounds in the agar diffusion assay

compd ^a	Gram-positive bacteria					Gram-negative bacteria		
	<i>B. subtilis</i> 6633 ATCC	<i>S. Aureus</i> SG511	<i>M.smegmatis</i> MC155	<i>M. vaccae</i> 10670 IMET	<i>M. luteus</i> 10240 ATCC	<i>P. aeruginosa</i> KW799/WT	K799/61	<i>E. coli</i> X580
4	14	15	15P ^c	15	20	12	13	14
12	18p ^b	19P	15ps	19	17	11	13	0
13	12/14p	13/16P	17s	19	21	16	18p	15
16	14P	18P	28s	17	26s	11	12	13P
17	0	12	15ps	19	38s	15	0	17
Cipro	26/30	-	18/27P	25	0	22	33	31

^a 50 µL of a 2.0 mM solution in DMSO : MeOH (1 : 9) of each compound was filled in 9 mm wells in agar media (Standard I Nutrient Agar, Serva or Mueller Hinton II Agar, Becton, Dickinson and Company). Inhibition zones read after incubation at 37 °C for 24 h. Cipro (ciprofloxacin) was dissolved in H₂O to give a 5 mg/mL solution. ^b p, partially clear inhibition zone/colonies in the inhibition zone. ^c P, unclear inhibition zone/many colonies in the inhibition zone. ^d s, indicates single colonies in the inhibition zone.

Table 5. ^1H and ^{13}C NMR data for compounds **4**, **12**, **13** in CD_3OD at 298 K.

4				12				13			
Substructural unit	δ_{C} [ppm]	δ_{H} [ppm]	Proton multiplicity ^a $J(\text{H}_i, \text{H}_j)$ [Hz]	Substructural unit	δ_{C} [ppm]	δ_{H} [ppm]	Proton multiplicity ^a $J(\text{H}_i, \text{H}_j)$ [Hz]	Substructural unit	δ_{C} [ppm]	δ_{H} [ppm]	Proton multiplicity ^a $J(\text{H}_i, \text{H}_j)$ [Hz]
C-1	170.7			CH ₂ -1	61.6	3.68	t, 5.8	CH ₂ -1	61.5	3.68	t, 5.8
C-2	120.1			CH ₂ -2	43.3	3.47	t, 5.8	CH ₂ -2	43.1	3.47	t, 5.8
CH-3	148.4	8.57	dd, 0.7, 2.4	C-3	169.0			C-3	168.5		
C-4	165.3			C-4	120.9			C-4	120.6		
CH-5	108.1	7.10	dd, 2.4, 9.0	CH-5	148.5	8.60	dd, 2.5, 0.7	CH-5	147.9	8.55	d, 2.2
CH-6	138.3	8.01	dd, 0.7, 9.0	C-6	165.3			C-6	165.3		
CH-7	68.5	4.50	dd, 4.7, 12.6	CH-7	108.4	7.04	dd	CH-7	107.8	7.10	d, 9.0
CH ₂ -8	24.0	2.41	mt, $\Sigma J=43.4$	CH-8	137.5	7.96	dd, 8.9	CH-8	137.6	7.99	d, 9.0
		1.58	mt	CH-9	62.5	5.14	dd, 3.8, 11.9	CH-9	68.6	4.49	dd, 4.7, 12.6
CH ₂ -9	40.2	1.48	mt, $\Sigma J=20.4$	CH ₂ -10	30.5	2.30	mt, $\Sigma J=34.0$	CH ₂ -10	23.6	1.57	mt
		1.35	ddd 4.2, 13.1, 13.2			1.60	mt			2.40	mt, $\Sigma J=35.5$
C-10	33.9			CH ₂ -11	34.2	2.39	qd, 13.6, 4.8	CH ₂ -11	39.9	1.34	mt
CH ₂ -11	51.6	1.94	dd, 3.0, 13.3			1.90	mt			1.47	mt
			1.26	d, 13.2	C-12	153.6			C-12	33.6	
C-12	77.9			CH-13	47.5	2.34	mt, $\Sigma J=20.6$	CH ₂ -13	51.4	1.26	d, 13.4
CH ₂ -13	37.6	2.28	ddd, 4.1, 7.5, 13.9	CH-14	55.9	1.83	mt			1.94	dd, 2.9, 13.4
		1.37	mt	CH ₂ -15	34.1	1.77	dtd, 14.0, 4.2, 1.0	C-14	77.6		
CH ₂ -14	24.2	1.59	mt			1.47	mt	CH ₂ -15	37.3	1.38	ddd, 4.5, 9.2, 14.4
		1.50	ddd, 3.9, 9.1, 9.3	2.35	dt, 13.6, 4.8	2.29	ddd, 3.9, 7.7, 14.4				
CH-15	48.2	1.71	ddd, 5.5, 8.7, 12.2	CH ₂ -16	38.2	1.88	mt	CH ₂ -16	23.9	1.51	mt

CH-16	41.61	2.12	ddd, 8.0, 11.1, 11.4	C-17	147.26					1.59	mt, $\Sigma J=27.0$
CH ₂ -17	37.4	1.54	dd, 7.9, 9.6	CH ₂ -18	37.8	1.82	mt	CH-17	47.9	1.71	mt, $\Sigma J=26.2$
		1.40	mt					CH-18	41.3	2.12	ddd, 8.0, 10.8, 11.2
C-18	35.5			C-19	33.8			CH ₂ -19	37.2	1.40	t, 8.1
CH ₃ -19	30.9	1.01	s	CH ₃ -20	22.0	1.00	S			1.54	dd, 8.1, 9.6
CH ₃ -20	21.0	1.03	s	CH ₃ -21	30.0	1.02	s	C-20	36.2		
CH ₃ -21	27.0	0.87	s	CH ₂ -22	109.9	4.98	mt	CH ₃ -21	30.6	1.01	s
							4.87	t, 1.3	CH ₃ -22	20.7	1.03
				CH ₂ -23	115.5	5.10	d, 2.0	CH ₃ -23	26.8	0.86	s
							5.09	d, 2.0			

a) Due to overlap of several proton signals the values of coupling constants or ΣJ could not be always determined.

Table 6. ^1H and ^{13}C NMR data for compounds **16** and **17** in CD_3OD at 298 K.

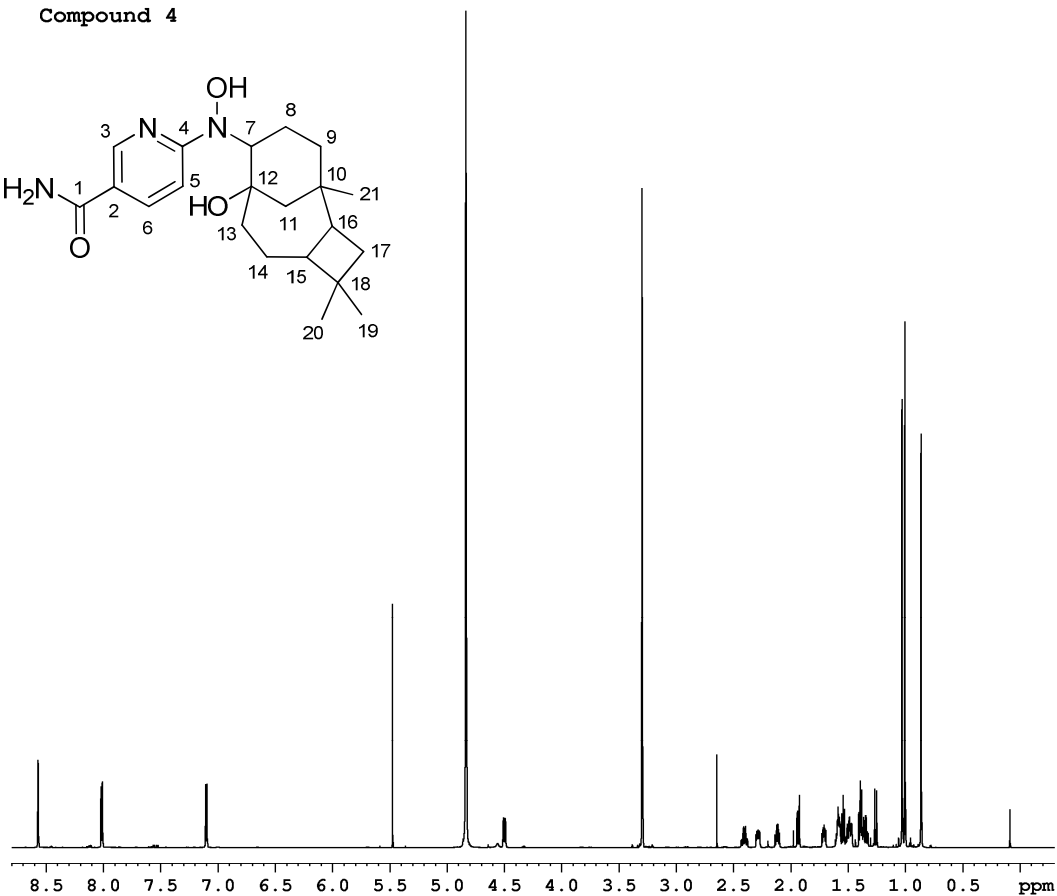
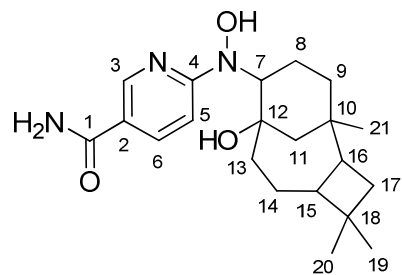
16^a				17			
Substructural unit	δ_{C} [ppm]	δ_{H} [ppm]	Proton multiplicity ^b J(H _i ,H _j) [Hz]	Substructural unit	δ_{C} [ppm]	δ_{H} [ppm]	Proton multiplicity ^b J(H _i ,H _j) [Hz]
CH ₂ -1	61.5	3.610	t, 5.5	CH ₂ -1	61.3	3.70	t, 5.9
CH ₂ -2	43.1	3.425	t, 5.5	CH ₂ -2	43.2	3.49	t, 5.9
C-3	167.9			C-3	169.9		
C-4	134.9			C-4	134.9		
CH-5,9	127.9	7.780	d, 8.3	CH-5,9	128.2	7.82	d, 8.4
CH-6,8	128.2	7.422	d, 8.3	CH-6,8	128.3	7.50	d, 8.4
C-7	140.8			C-7	141.7		
NH		7.233	t, 5.3	CH ₂ -10	67.5	5.23	d, 13.1
CH ₂ -10	67.1	5.583	d, 13.1			5.20	d, 13.1
		5.092	d, 13.1				
C-11	157.3			CH-12	67.5	4.02	dd, 13.1
CH-12	61.8	4.715	bs	CH ₂ -13	24.4	2.32	qd, 13.5, 4.4
CH ₂ -13	24.1	2.340	mt			1.57	mt
		1.679	mt	CH ₂ -14	39.7	1.47	mt
CH ₂ -14	38.5	1.473	mt			1.32	mt
		1.423	mt	C-15	33.7		
C-15	34.1			CH ₂ -16	51.0	1.86	dd, 2.9, 13.4
CH ₂ -16	44.6	2.351	mt			1.16	d, 13.4
		1.952	mt	C-17	77.0		
C-17	97.4			CH ₂ -18	36.7	2.23	ddd, 3.8, 8.1, 14.3
CH ₂ -18	33.9	2.357	mt			1.34	mt
		1.824	mt	CH ₂ -19	23.6	1.56	mt
CH ₂ -19	23.6	1.688	mt			1.44	mt
		1.444	mt	CH-20	47.7	1.69	ddd, 5.7, 8.5, 12.1
CH-20	46.8	1.74	ddd, 5.2, 9.1, 12.0	CH-21	41.1	2.07	ddd, 8.0, 10.8, 11.8
CH-21	40.7	2.019	ddd, 8.1, 10.9, 11.1	CH ₂ -22	37.1	1.52	dd, 8.0, 9.6

CH ₂ -22	36.64	1.518	dd, 8.1, 9.5			1.37	dd, 9.6, 10.6
		1.387	t, 10.0	C-23	35.3		
C-23	34.8			CH ₃ -24	30.6	0.99	s
CH ₃ -24	30.3	0.993	s	CH ₃ -25	20.7	1.00	s
CH ₃ -25	20.6	0.996	s	CH ₃ -26	26.7	0.84	s
CH ₃ -26	26.5	0.867	s				

a) In CD₃CN

b) Due to overlap of several proton signals the values of coupling constants or ΣJ could not be always determined.

Compound 4

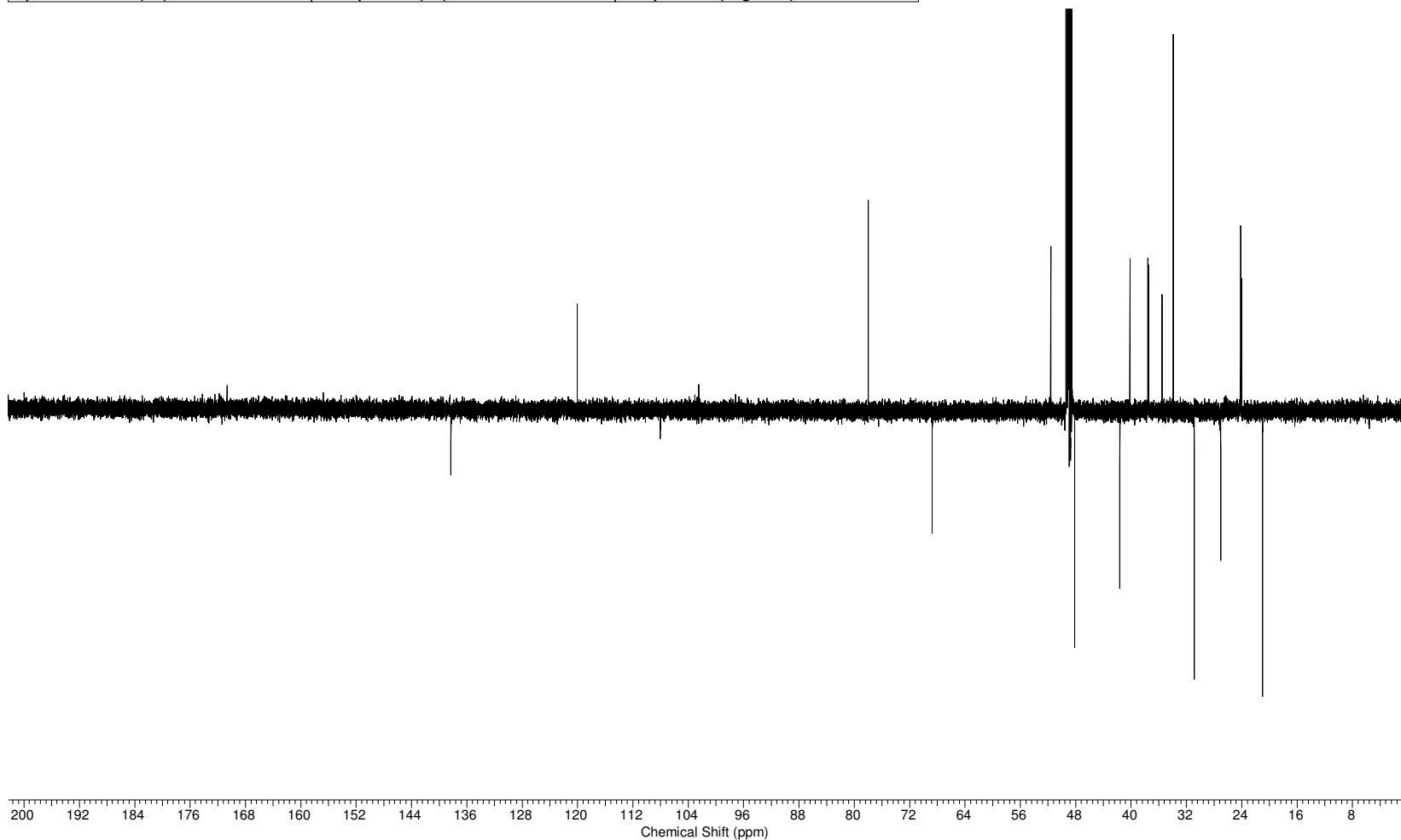


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FIDRES         0.125483 Hz
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D1            1.00000000 sec
TD0           1
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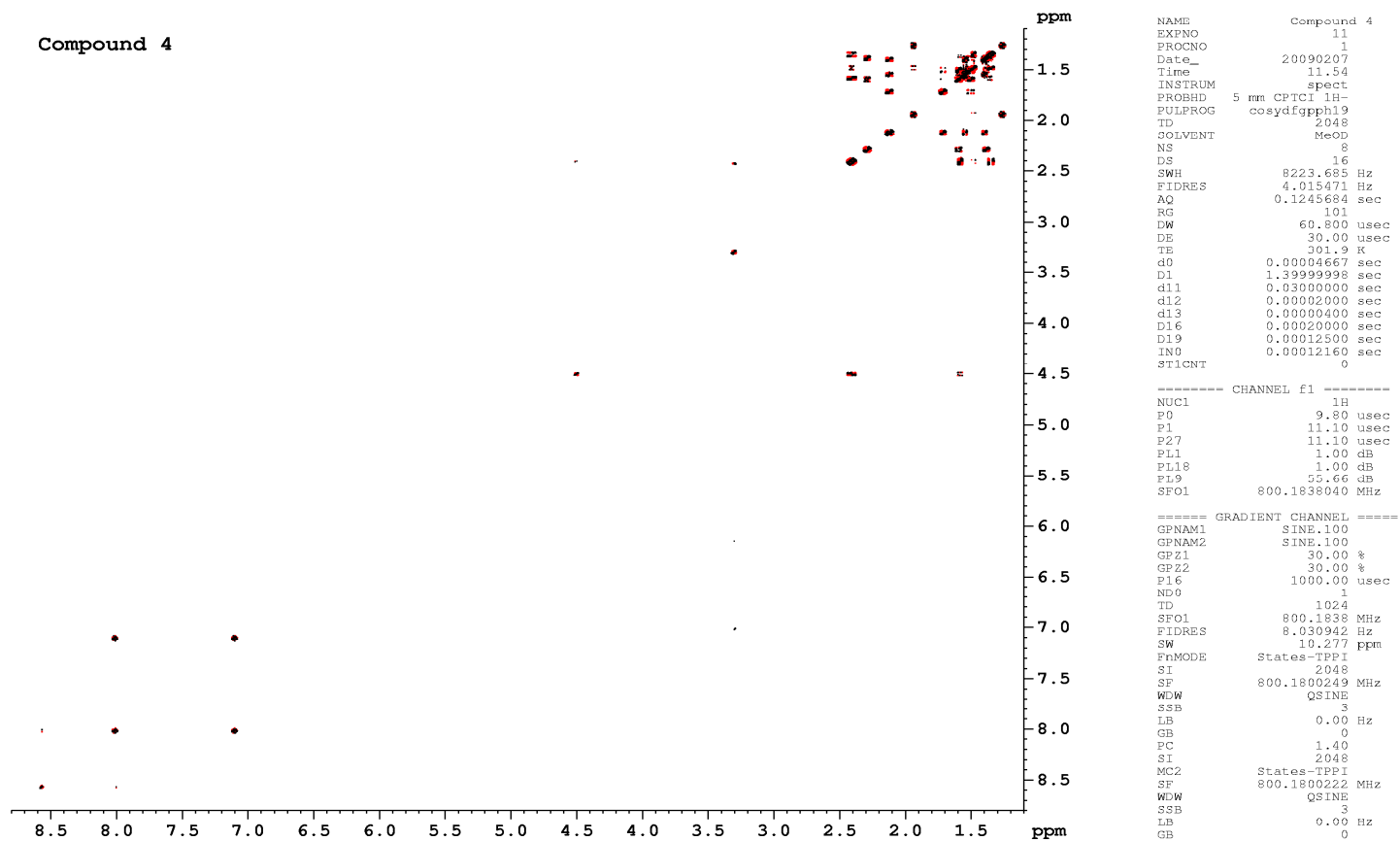
^1H spectrum of 4 in CD_3OD at 25°C .

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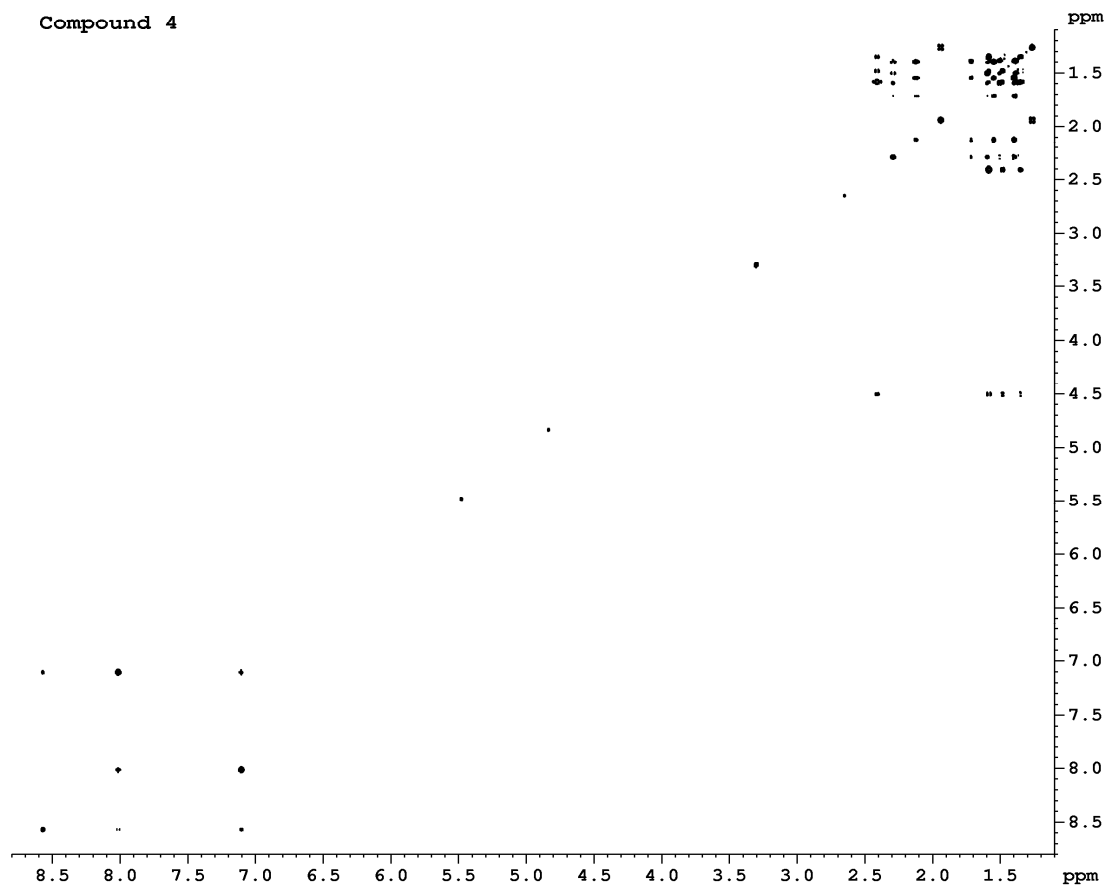
¹³C APT spectrum of **4** in CD₃OD at 25° C.

Compound 4



2D DQF-COSY spectrum of **4** in CD₃OD at 25° C. The black and red contours represent positive and negative peaks, respectively.

Compound 4



```

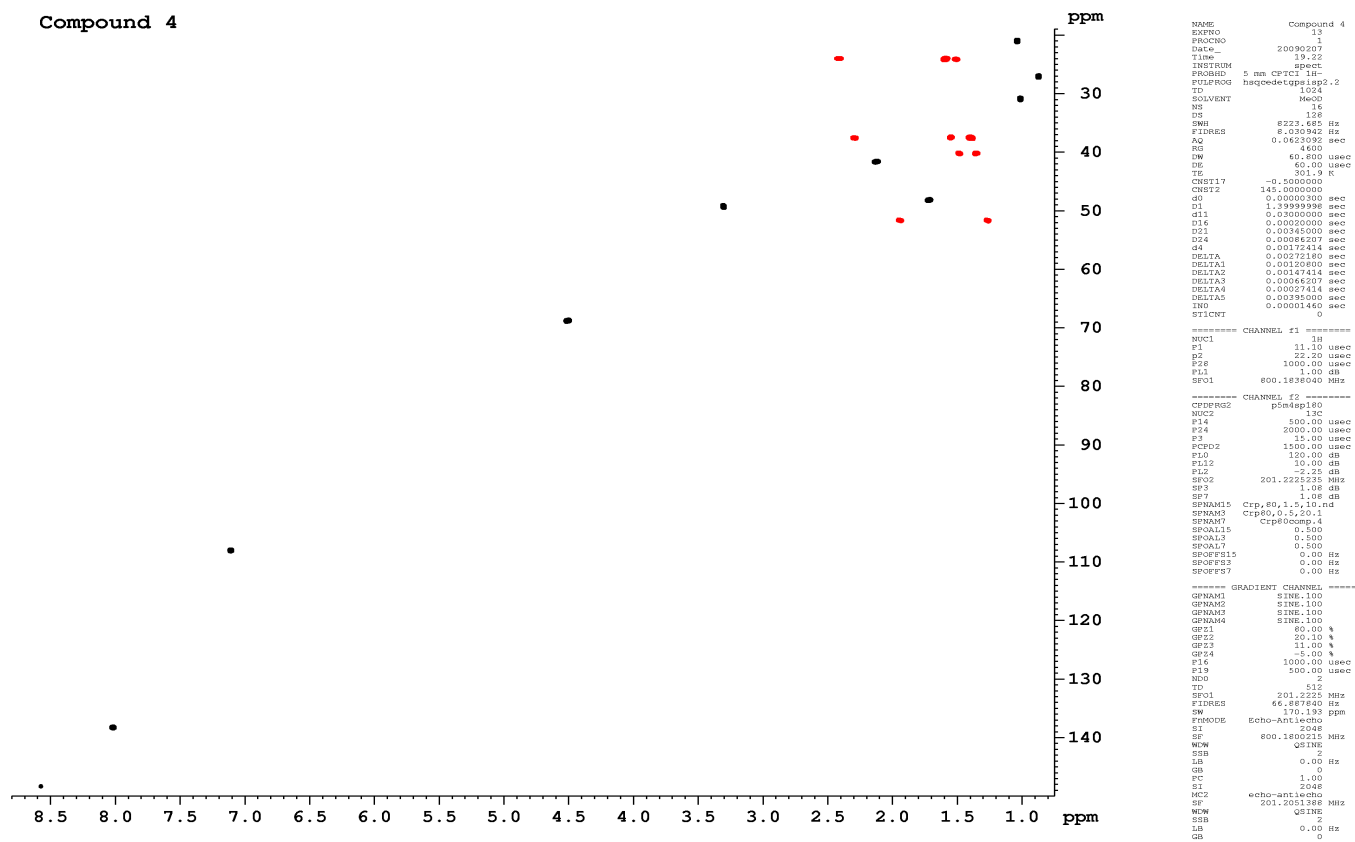
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SOLVENT       MeOD
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FACTOR1       11
IN0           0.00012140 sec
l1            22
STICNT        0

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PL10          7.70 dB
PL18          1.00 dB
PL9           55.66 dB
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----- GRADIENT CHANNEL -----
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GPNAM2        SINE.100
GPNAM3        SINE.100
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GP22          20.00 %
GP23          50.00 %
P16           1000.00 usec
NDO           1
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SSB           2
LB            0.00 Hz
GB            0
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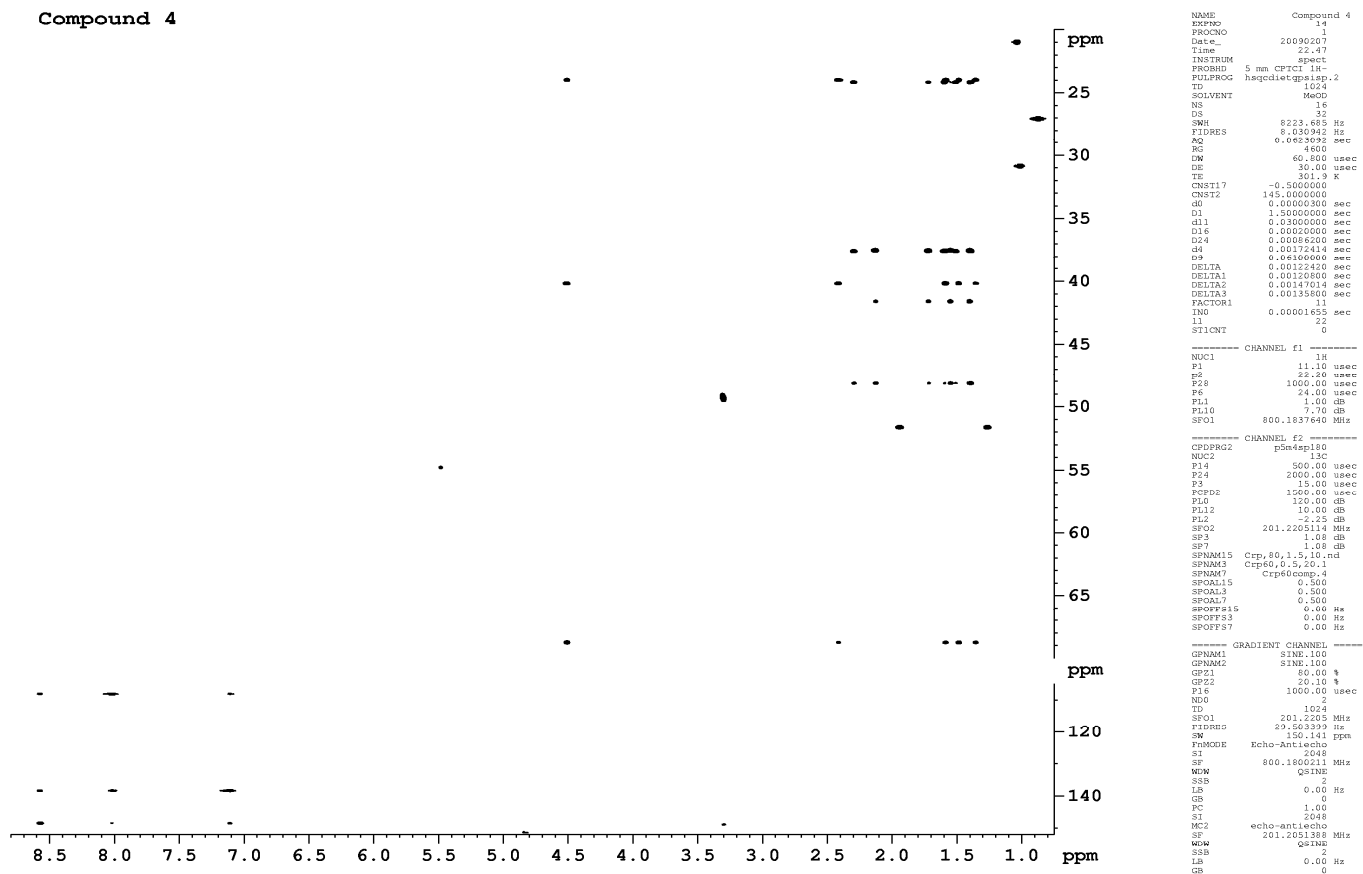
2D TOCSY spectrum of 4 in CD₃OD at 25° C.

Compound 4

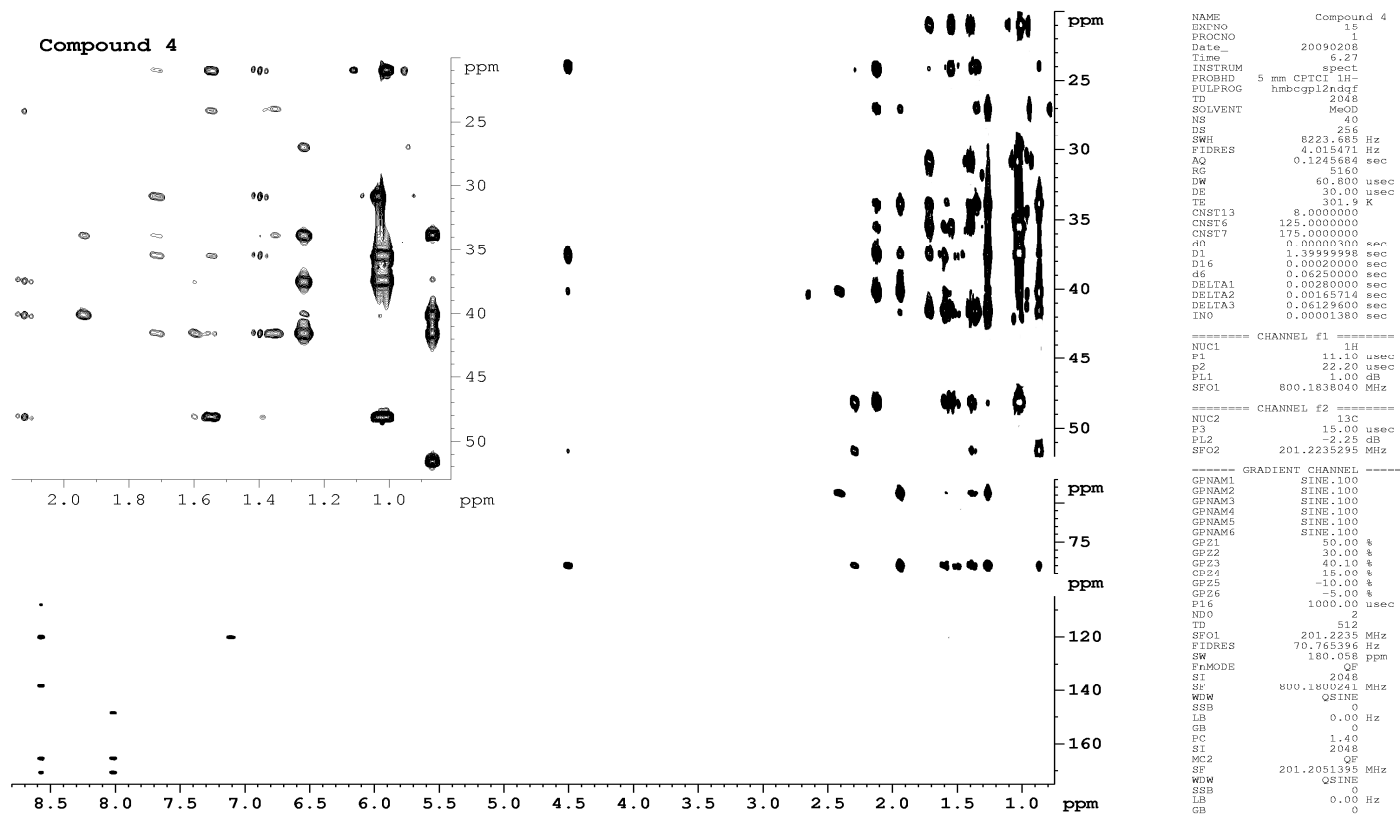


2D ^{13}C -HSQC spectrum of **4** in CD_3OD at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively. The black cross-peaks correspond to CH_3 and CH groups, the red cross-peaks correspond to CH_2 groups.

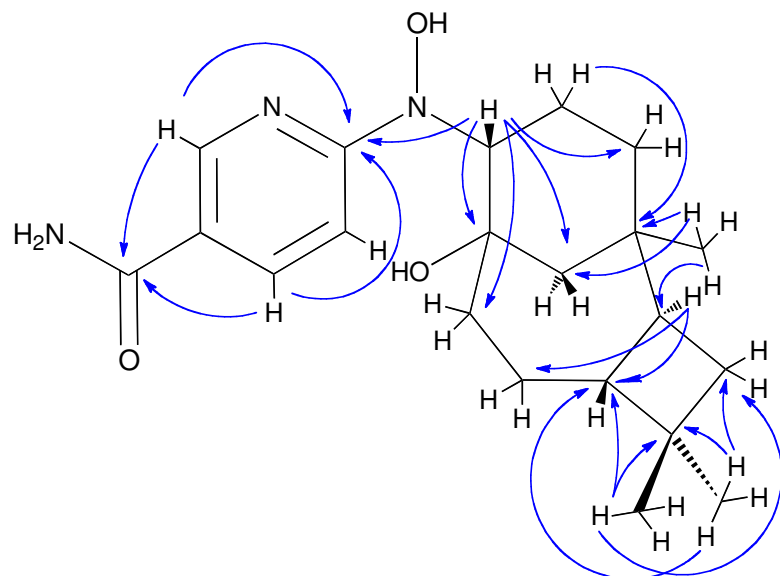
Compound 4



2D ^{13}C -HSQC-TOCSY spectrum of **4** in CD_3OD at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively.

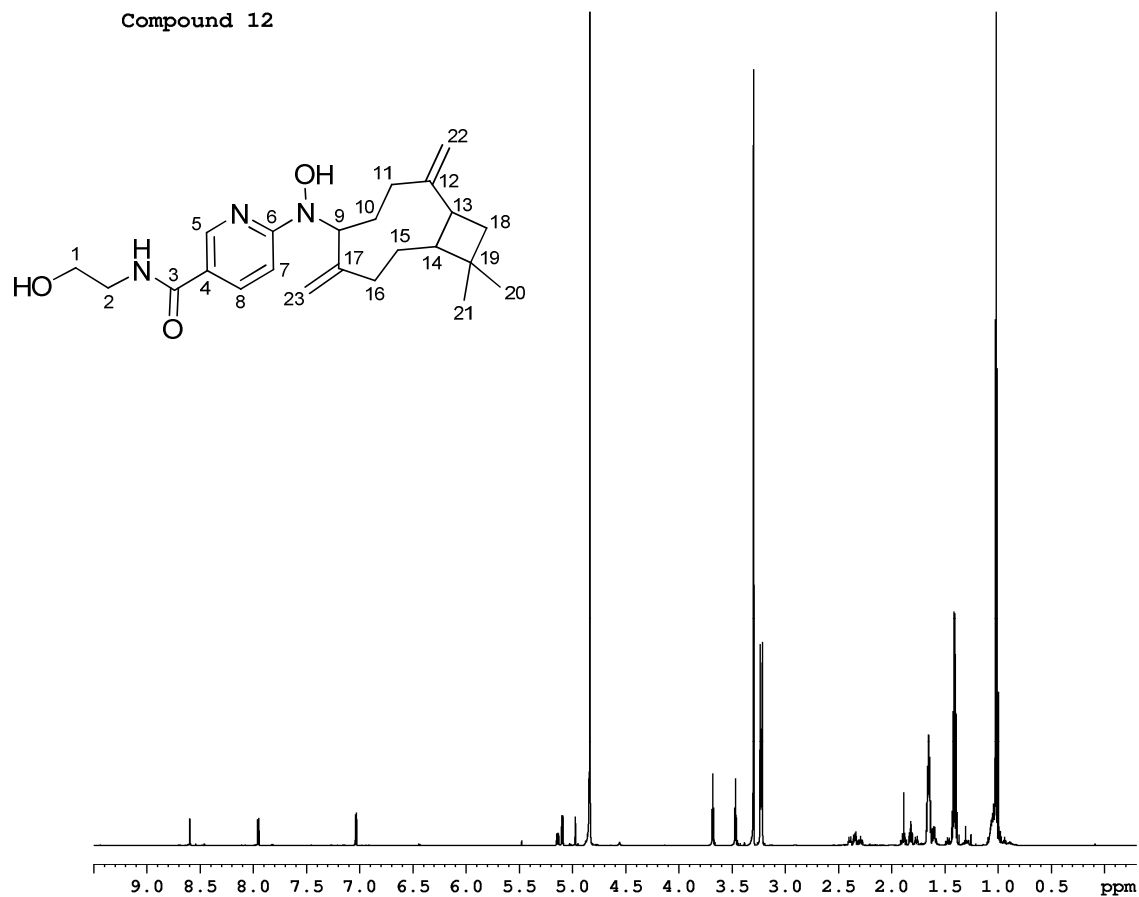


2D ¹³C-HMBC spectrum of **4** in CD₃OD at 25° C. The horizontal and vertical axes display ¹H and ¹³C chemical shifts, respectively. The insert in the upper left corner represents an expansion of the high field region plotted with reduced cross-peaks intensities for a better clarity.



Correlation diagram for compound **4**. The HMBC correlations (in blue), together with proton-proton correlations in DQF-COSY and TOCSY spectra and one bond proton-carbon correlations in the HSQC spectrum unambiguously establish the proposed structure of compound **4**.

Compound 12



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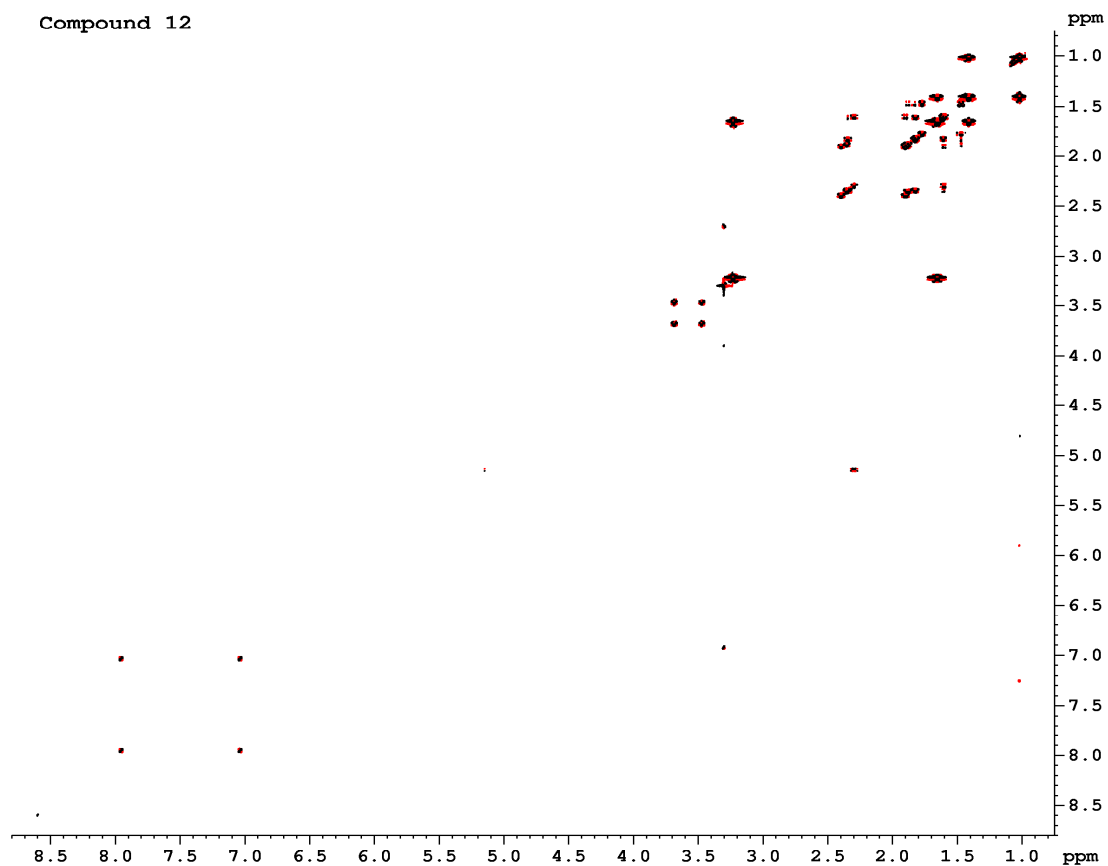
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PROCNO        1
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PULPROG       zg
TD            65536
SOLVENT       MeOD
NS            256
DS            4
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FIDRES        0.125483 Hz
AQ            3.9846387 sec
RG            32
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DE            30.00 usec
TE            301.9 K
D1            0.10000000 sec
TD0           1
    
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P1            10.30 usec
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WDW           no
SSB           0
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¹H spectrum of **12** in CD₃OD at 25° C.

Compound 12



```

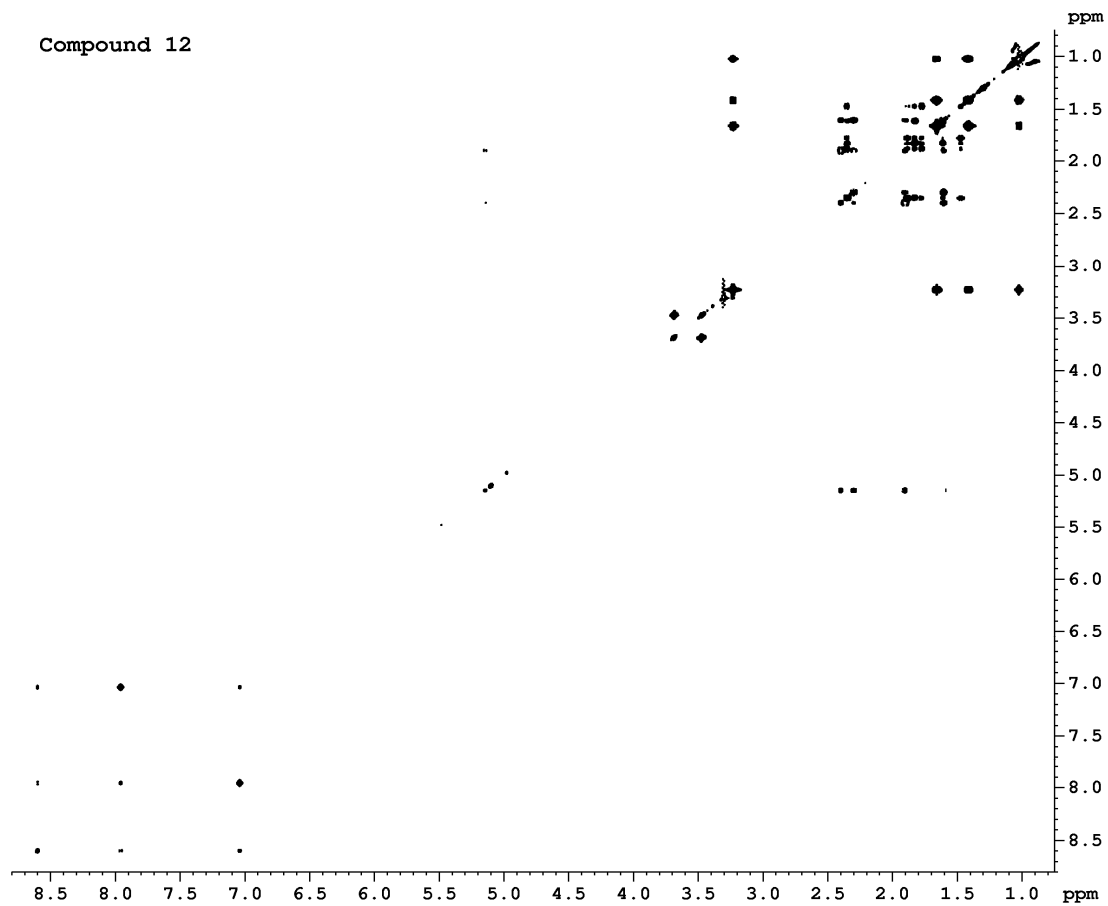
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PROCNO       1
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PULPROG      cosydfgpph19
TD           2048
SOLVENT      MeOD
NS           24
DS           16
SWH          8223.685 Hz
FIDRES       4.015471 Hz
AQ           0.1245684 sec
RG           114
DW           60.800 usec
DE           30.00 usec
TE           301.9 K
d0           0.00004769 sec
d1           1.39999998 sec
d11          0.03000000 sec
d12          0.00020000 sec
d13          0.00004000 sec
D16          0.00020000 sec
D19          0.00012500 sec
IN0          0.00012160 sec
ST1CNT       0

===== CHANNEL f1 =====
NUC1         1H
P0           9.20 usec
P1           10.30 usec
P27          10.30 usec
PL1          0.20 dB
PL18         0.20 dB
PL19         55.66 dB
SFO1         800.1838776 MHz

===== GRADIENT CHANNEL =====
GPNAM1      SINE.100
GPNAM2      SINE.100
GPZ1        30.00 %
GPZ2        30.00 %
P16         1000.00 usec
ND0         1
TD           1024
SFO1         800.1839 MHz
FIDRES       8.030942 Hz
SW           10.277 ppm
F2MODE      States-TFPI
SI           2048
SF           800.1800263 MHz
WDW          QSINE
SSB          3
LB           0.00 Hz
GB           0
PC           1.40
SI           2048
MC2         States-TFPI
SF           800.1800264 MHz
WDW          QSINE
SSB          3
LB           0.00 Hz
GB           0
    
```

2D DQF-COSY spectrum of **12** in CD₃OD at 25° C. The black and red contours represent positive and negative peaks, respectively.

Compound 12



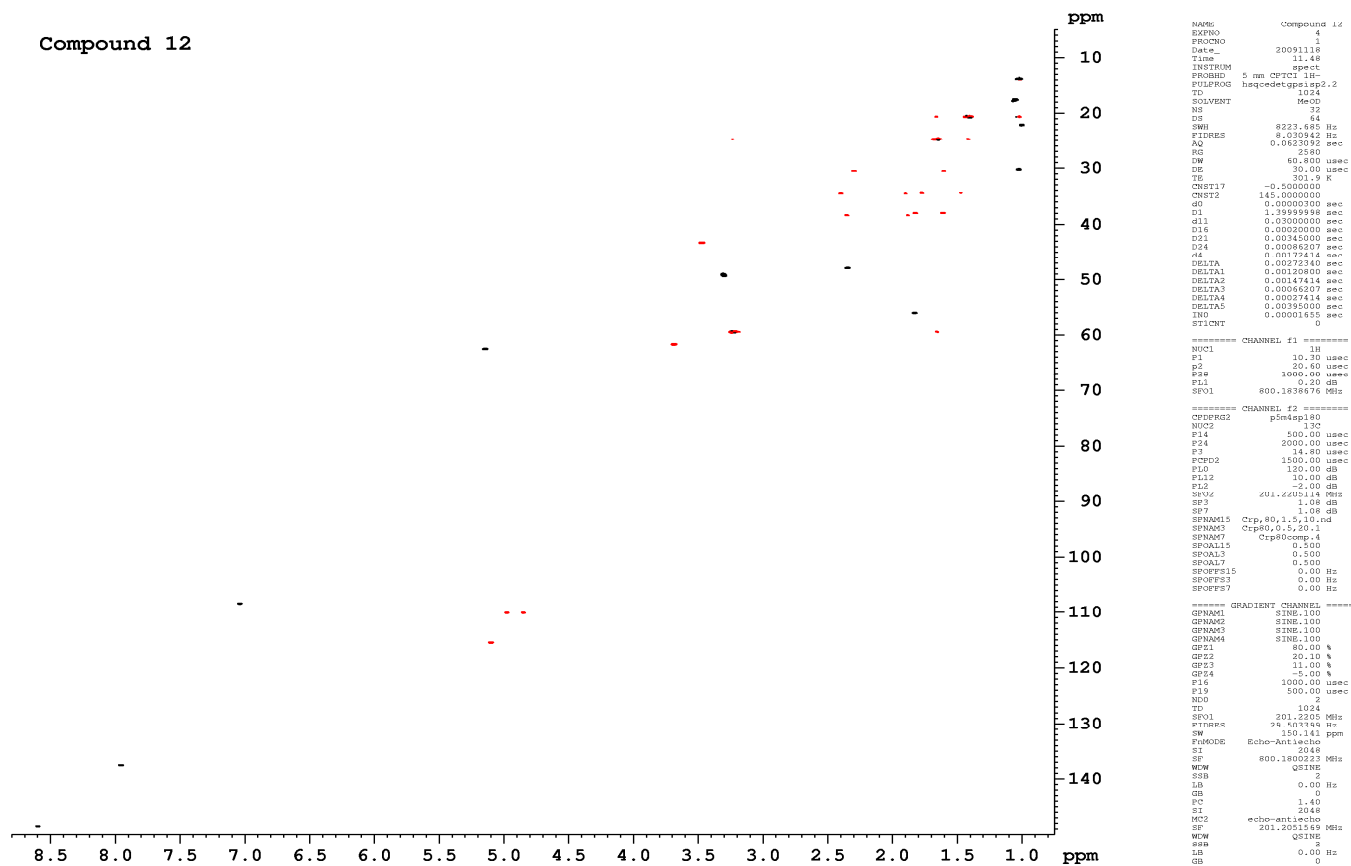
```
NAME          Compound 12
EXPNO         3
PROCNO       1
Date_         20091118
Time         0.28
INSTRUM      spect
PROBHD       5 mm CPTCI 1H-
PULPROG      d1psi2etgpsi19
TD           2048
SOLVENT      MeOD
NS           24
DS           16
SWH          8223.685 Hz
FIDRES       4.015471 Hz
AQ           0.1245684 sec
RG           101
DW           60.800 usec
DE           30.00 usec
TE           301.9 K
d0           0.00000000 sec
d1           1.39999998 sec
d11          0.03000000 sec
d12          0.00002000 sec
d13          0.00000400 sec
d16          0.00020000 sec
d19          0.00012500 sec
d20          0.00001000 sec
d21          0.00001000 sec
D9           0.06000000 sec
DELTA        0.00120300 sec
FACTOR1      11
l1           0.00012180 sec
l1           22
ST1CNT       0

===== CHANNEL f1 =====
NUC1         1H
P0           10.30 usec
P1           10.30 usec
P2           20.60 usec
P27          10.30 usec
P6           24.00 usec
PL1          0.20 dB
PL10         7.55 dB
PL18         0.20 dB
PL9          55.66 dB
SFO1         800.1838676 MHz

===== GRADIENT CHANNEL =====
GPNAM1      SINE.100
GPNAM2      SINE.100
GPNAM3      SINE.100
GPZ1        50.00 %
GPZ2        20.00 %
GPZ3        50.00 %
PI6         1000.00 usec
ND0         1
TD           1024
SFO1         800.1839 MHz
FIDRES       8.030942 Hz
SW           10.277 ppm
FnmODE      Echo-Antiecho
SI           2048
SF           800.1800244 MHz
WDW          QSINE
LB           2
GB           0
PC           1.40
SI           2048
MC2          echo-antiecho
SF           800.1800244 MHz
WDW          QSINE
SSB          2
LB           0.00 Hz
GB           0
```

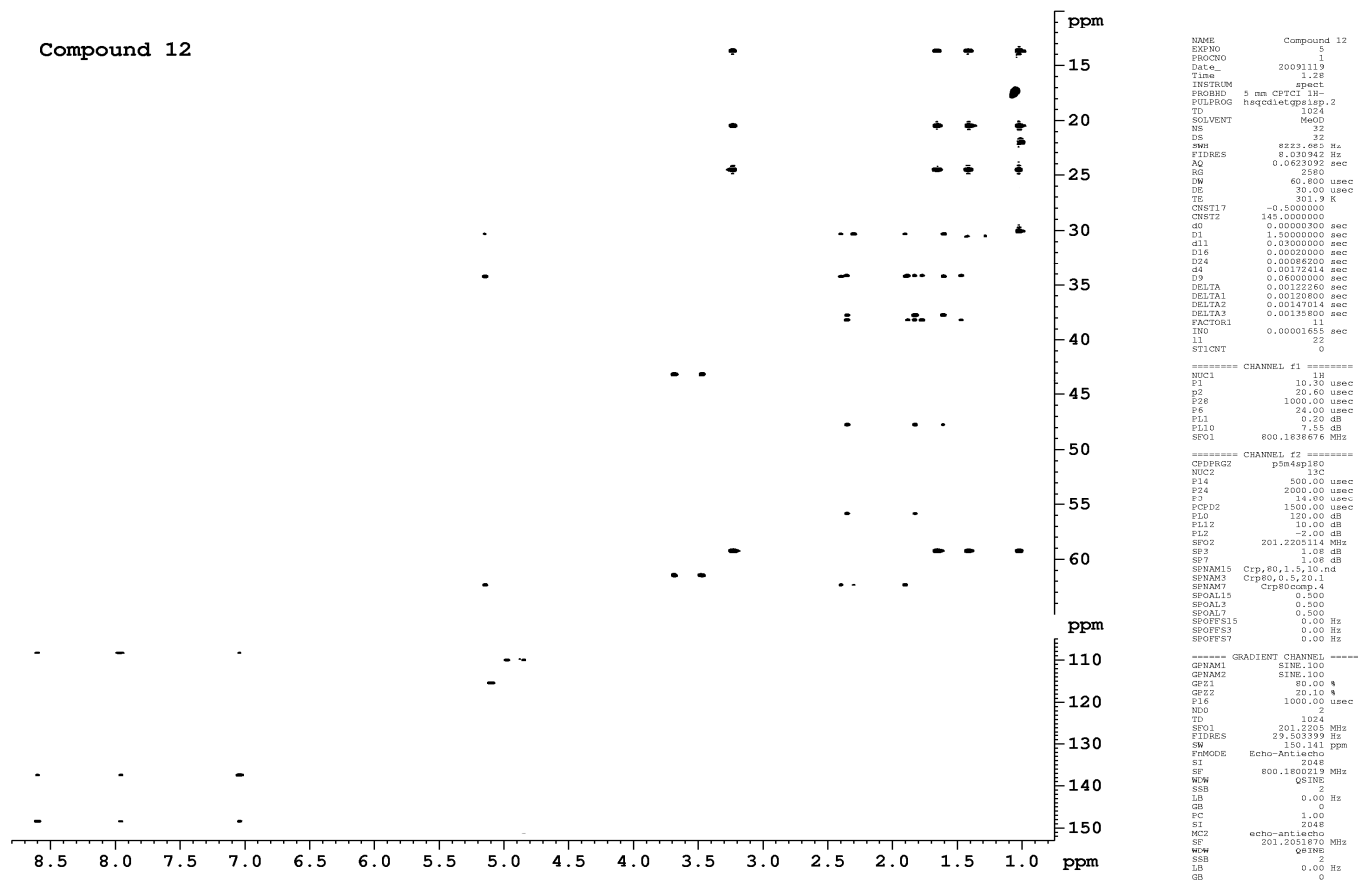
2D TOCSY spectrum of **12** in CD₃OD at 25° C.

Compound 12

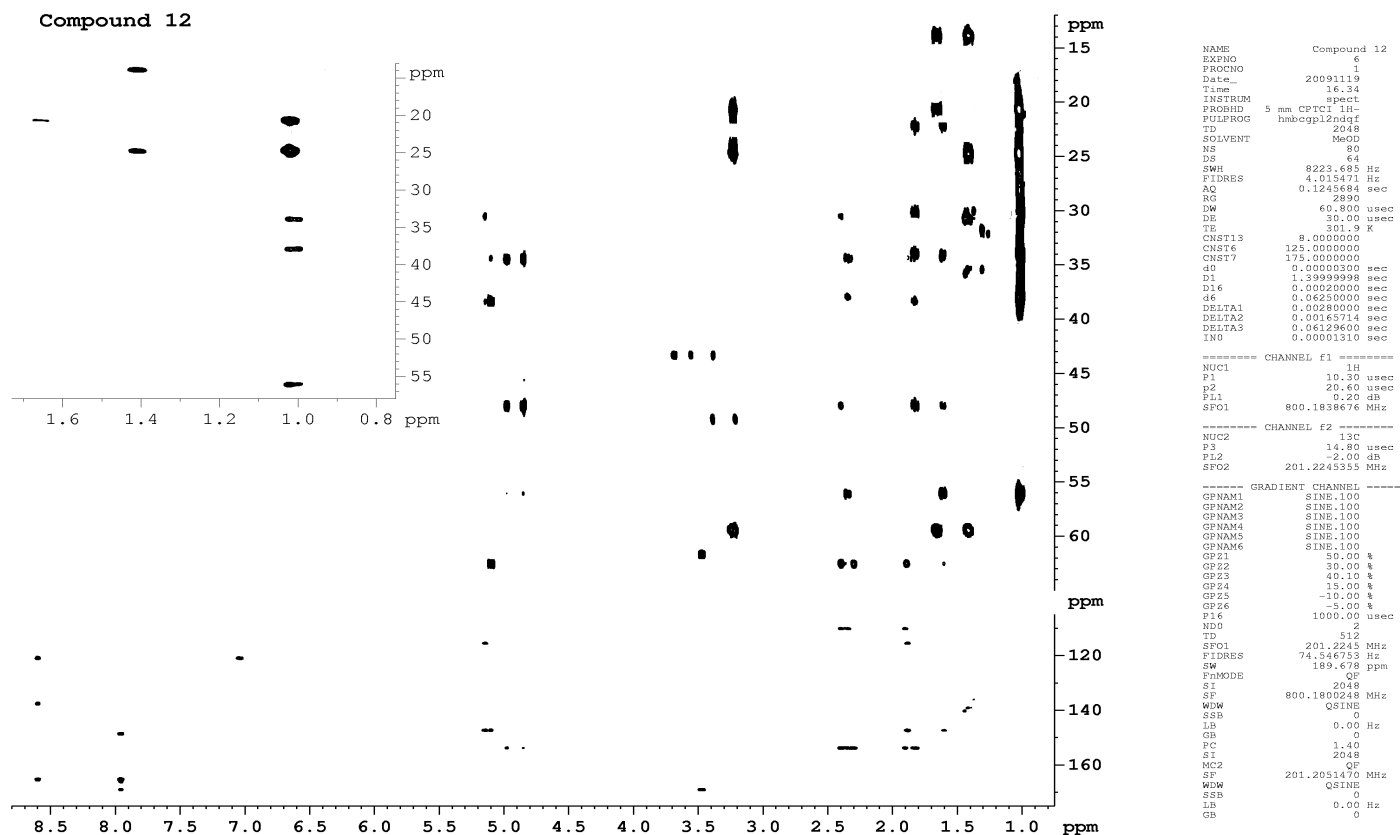


2D ^{13}C -HSQC spectrum of **12** in CD_3OD at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively. The black cross-peaks correspond to CH_3 and CH groups, the red cross-peaks correspond to CH_2 groups.

Compound 12

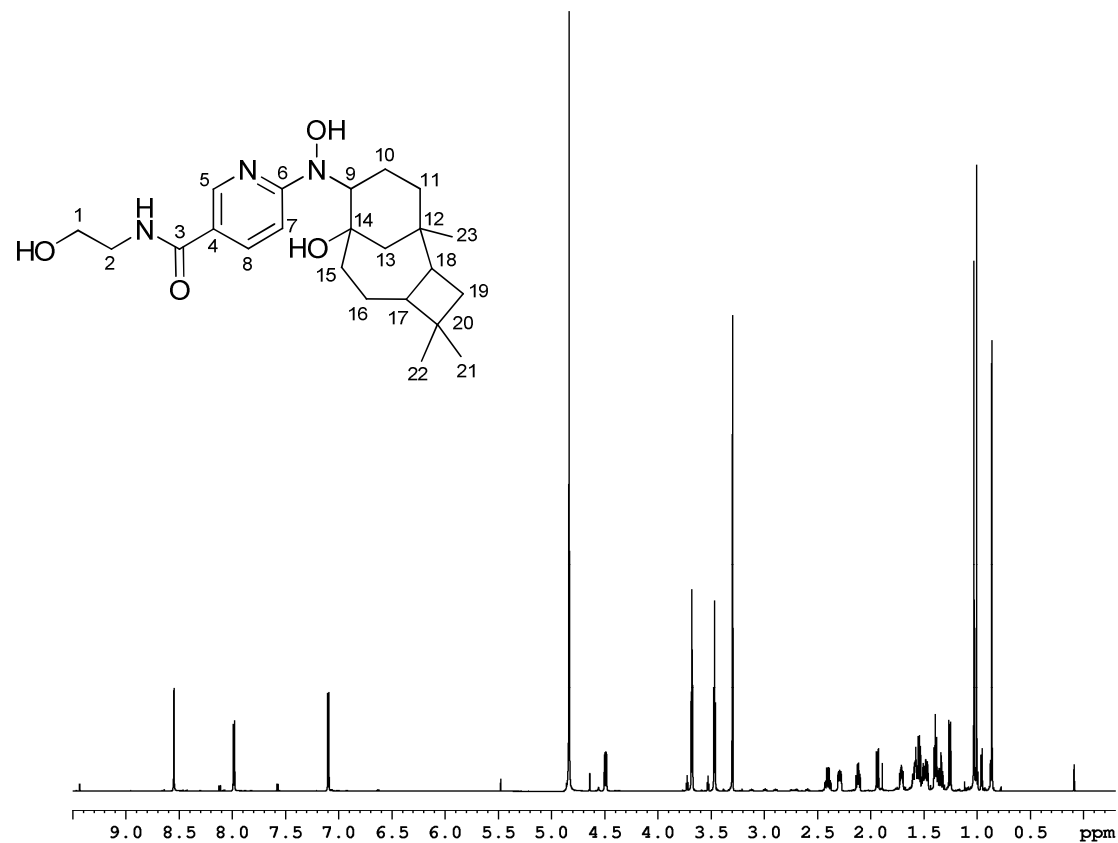


2D ^{13}C -HSQC-TOCSY spectrum of **12** in CD_3OD at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively.



2D ^{13}C -HMBC spectrum of **12** in CD_3OD at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively. The insert in the upper left corner represents an expansion of the high field region plotted with reduced cross-peaks intensities for a better clarity.

Compound 13



```

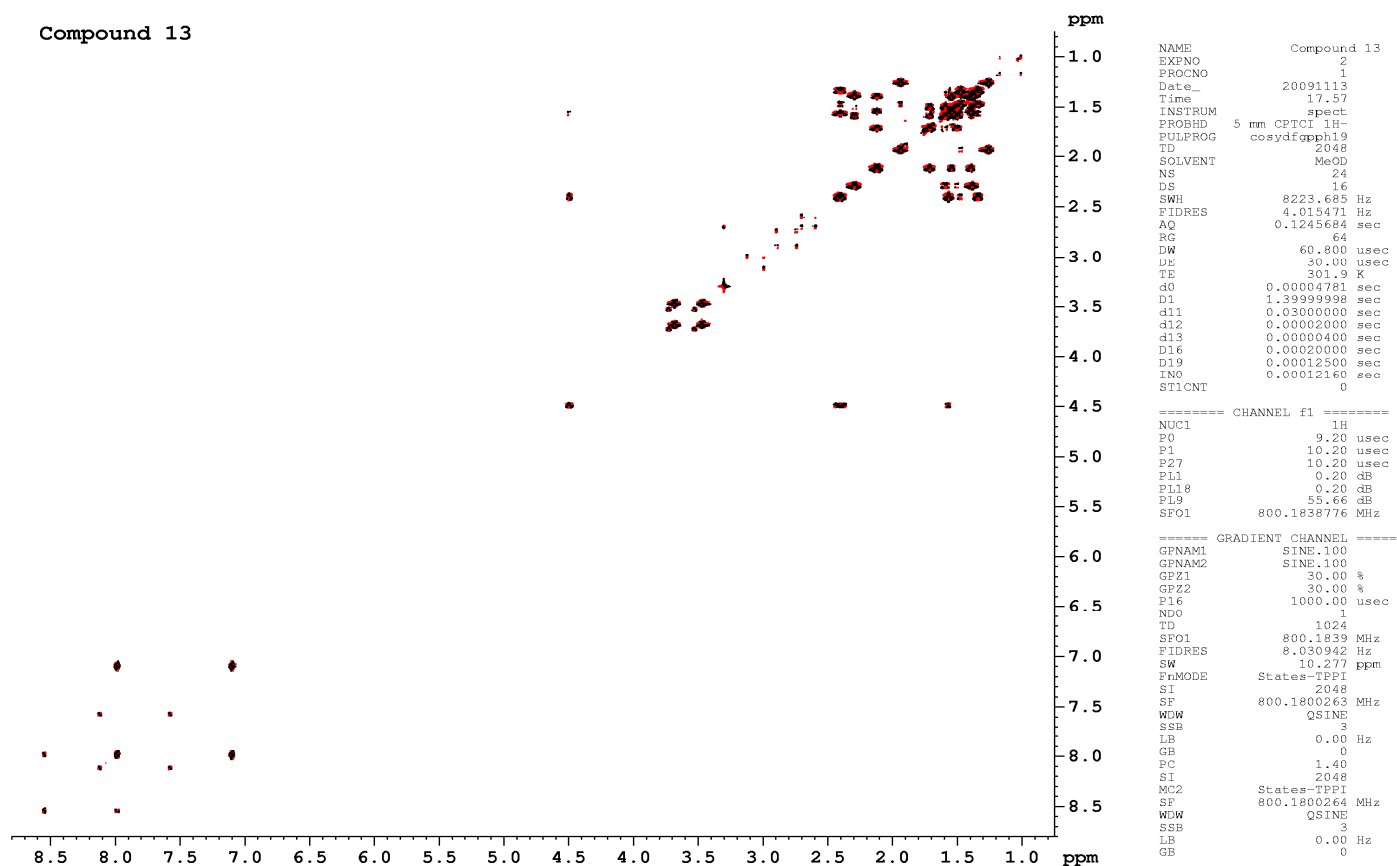
NAME          Compound 13
EXPNO         1
PROCNO        1
Date_         20091113
Time          17.27
INSTRUM       spect
PROBHD        5 mm CPTCI 1H-
PULPROG       zg
TD            65536
SOLVENT       MeOD
NS            64
DS            0
SWH           8223.685 Hz
FIDRES        0.125483 Hz
AQ            3.9846387 sec
RG            32
DW            60.800 usec
DE            30.00 usec
TE            301.9 K
D1            0.10000000 sec
TD0           1
    
```

```

===== CHANNEL f1 =====
NUC1          1H
P1            6.00 usec
PL1           0.20 dB
SFO1          800.1837640 MHz
SI            131072
SF            800.1800262 MHz
WDW           no
SSB           0
LB            0.00 Hz
GB            0
PC            1.00
    
```

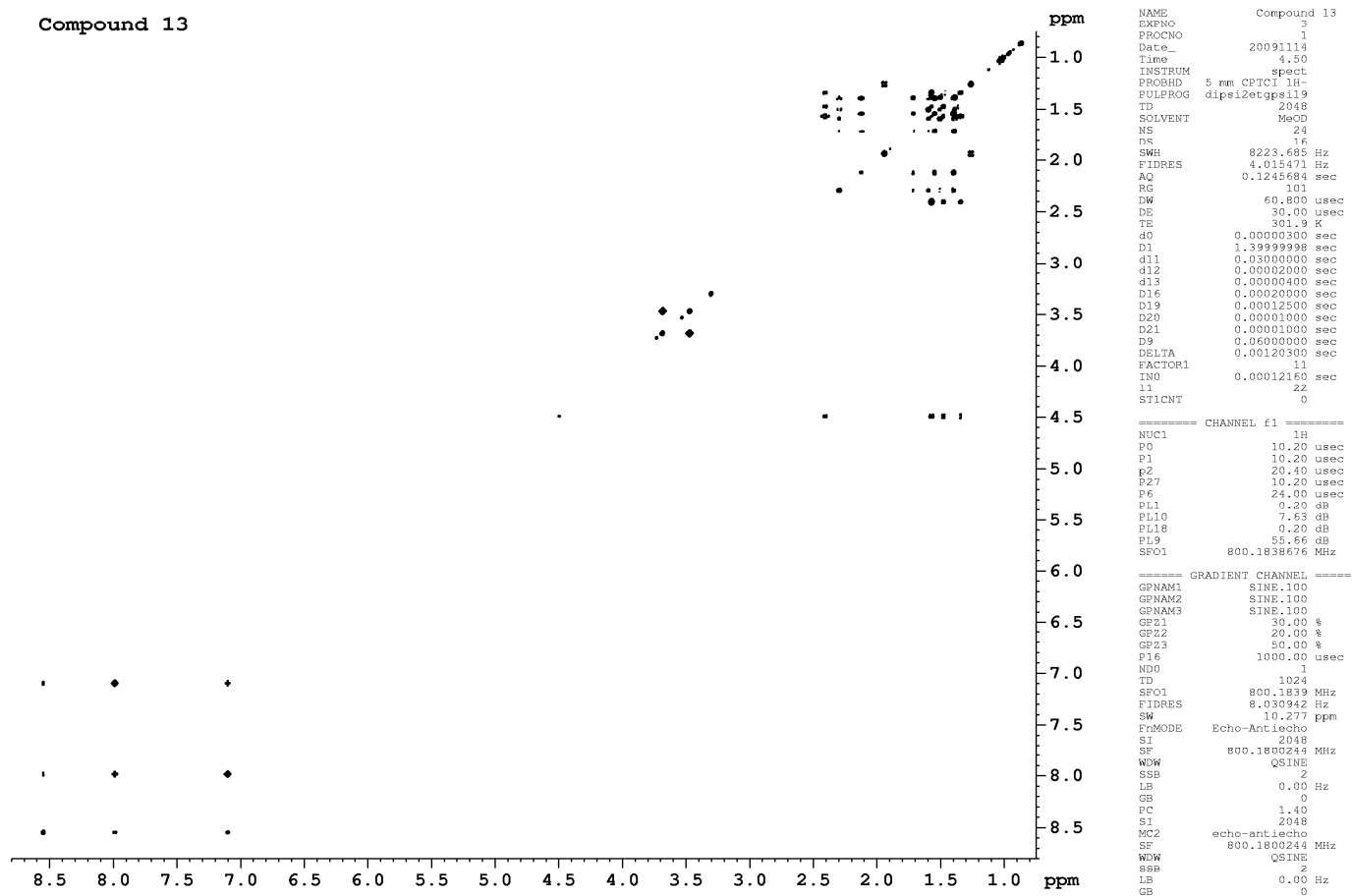
¹H spectrum of **13** in CD₃OD at 25° C.

Compound 13



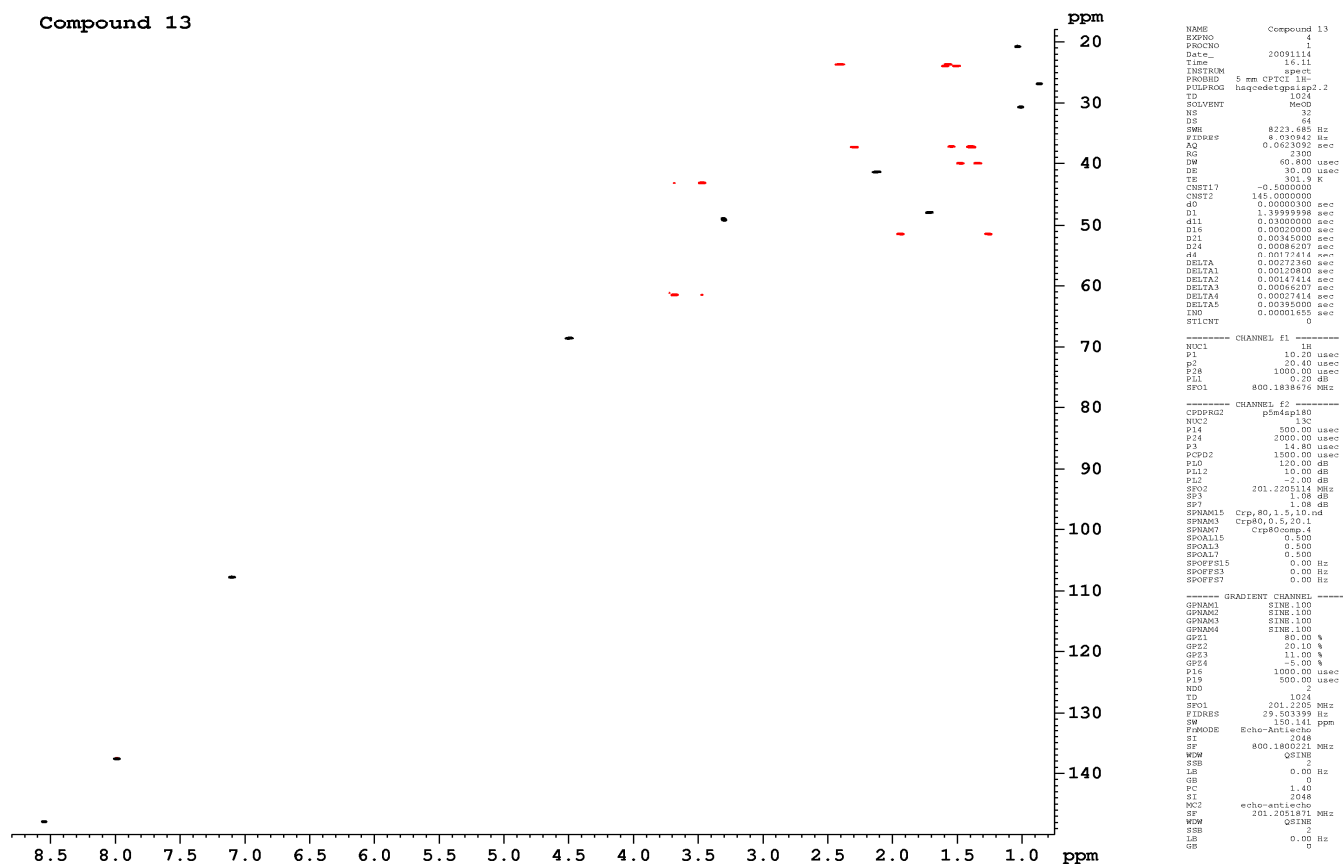
2D DQF-COSY spectrum of **13** in CD₃OD at 25° C. The black and red contours represent positive and negative peaks, respectively.

Compound 13



2D TOCSY spectrum of **13** in CD₃OD at 25° C.

Compound 13



```

NAME          Compound 13
EXPNO         4
PROCNO        1
Date_         20091114
Time          15.11
INSTRUM       spect
PROBHD        5 mm CPTCI 1h-
PULPROG       hsqcetdetspp2.2
TD            1024
SOLVENT       MeCD
NS            32
DS            64
SWS           8223.685 Hz
FIDRES        8.039842 Hz
AQ            0.0623993 sec
RG            7800
DM            60.800 usec
DE            30.00 usec
TE            301.2 K
CNS117        -0.5000000
CNS122        145.0000000
d0            0.0000000 sec
D1            1.39999996 sec
d11           0.0300000 sec
D16           0.0002000 sec
D21           0.0034000 sec
D24           0.0008620 sec
d4            0.0017614 sec
DELTA         0.0027286 sec
DELTA1        0.0012000 sec
DELTA2        0.00147414 sec
DELTA3        0.0006000 sec
DELTA4        0.00027414 sec
DELTA5        0.00395000 sec
IND           0.00001655 sec
STLCNT        0

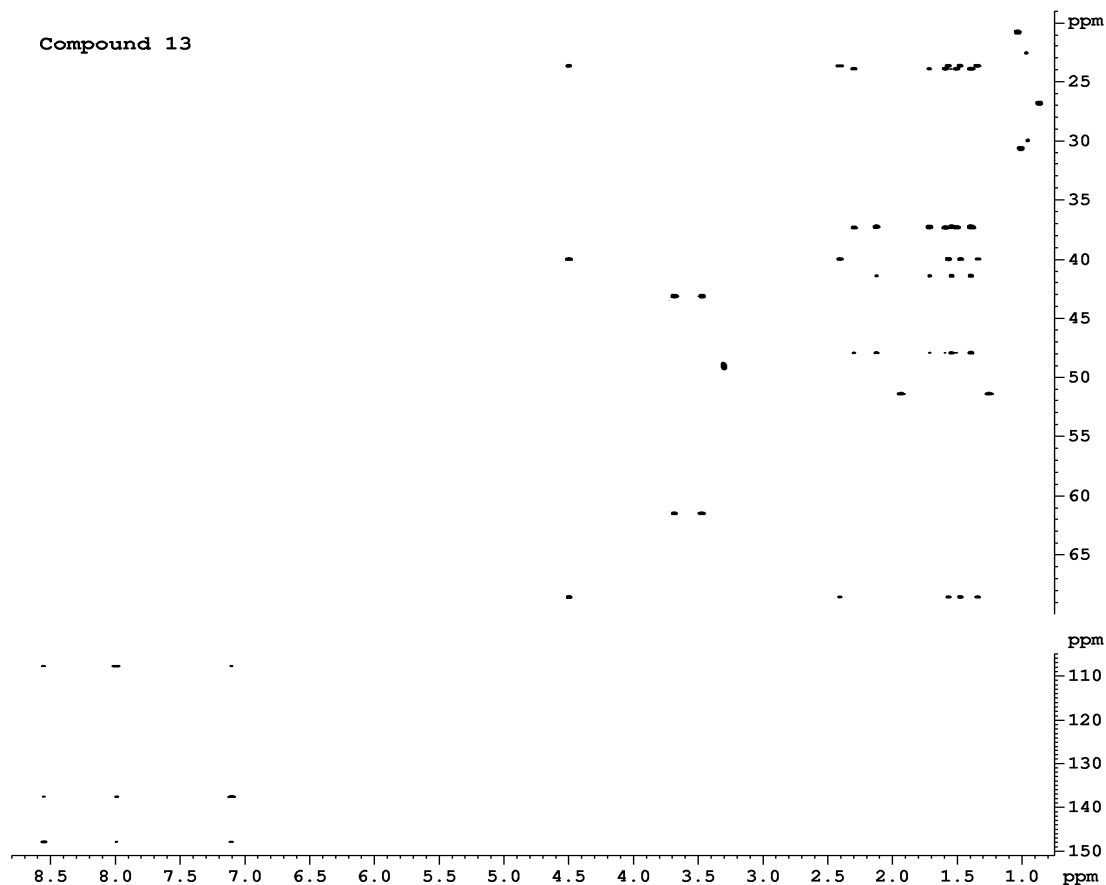
----- CHANNEL f1 -----
NUC1          1H
P1            10.00 usec
P2            20.40 usec
P2A           1000.00 usec
P21           0.25 dB
SFO1          800.1803479 MHz

----- CHANNEL f2 -----
CPDPRG2      pnm4e180
NUC2          13C
P14           500.00 usec
P24           2000.00 usec
P3            14.80 usec
PCPD2         1500.00 usec
P10           120.00 dB
P12           10.00 dB
P13           -2.00 dB
SFO2          201.2205114 MHz
SF3           1.00 dB
SF7           1.00 dB
SFRAMA15     Ccp,80,1.5,10.0d
SFRAMA3      Ccp80,0.5,20.1
SFRAMA7      Ccp80cm8,4
SFOAL15      0.500
SFOAL3       0.500
SFOAL7       0.500
SPOFF15      0.00 Hz
SPOFF3       0.00 Hz
SPOFF7       0.00 Hz

----- GRADIENT CHANNEL -----
GPARAM1      SINE,100
GPARAM2      SINE,100
GPARAM3      SINE,100
GPARAM4      SINE,100
GP21         80.00 %
GP22         20.10 %
GP23         11.00 %
GP24         -5.00 %
P16          1000.00 usec
P19          500.00 usec
ND0          2
TD            1024
SFO1          201.2205 MHz
FIDRES        29.503399 Hz
SW            150.141 ppm
FMODE        Echo-AntiEcho
SI            1048
SF            800.1800221 MHz
WDM          QSINE
SDB          0.00 Hz
GB           0
PC            1.40
SI            2046
MC2          echo-antiEcho
SF            201.2201871 MHz
WDM          QSINE
SDB          0.00 Hz
GB           0
  
```

2D ^{13}C -HSQC spectrum of **13** in CD_3OD at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively. The black cross-peaks correspond to CH_3 and CH groups, the red cross-peaks correspond to CH_2 groups.

Compound 13



```

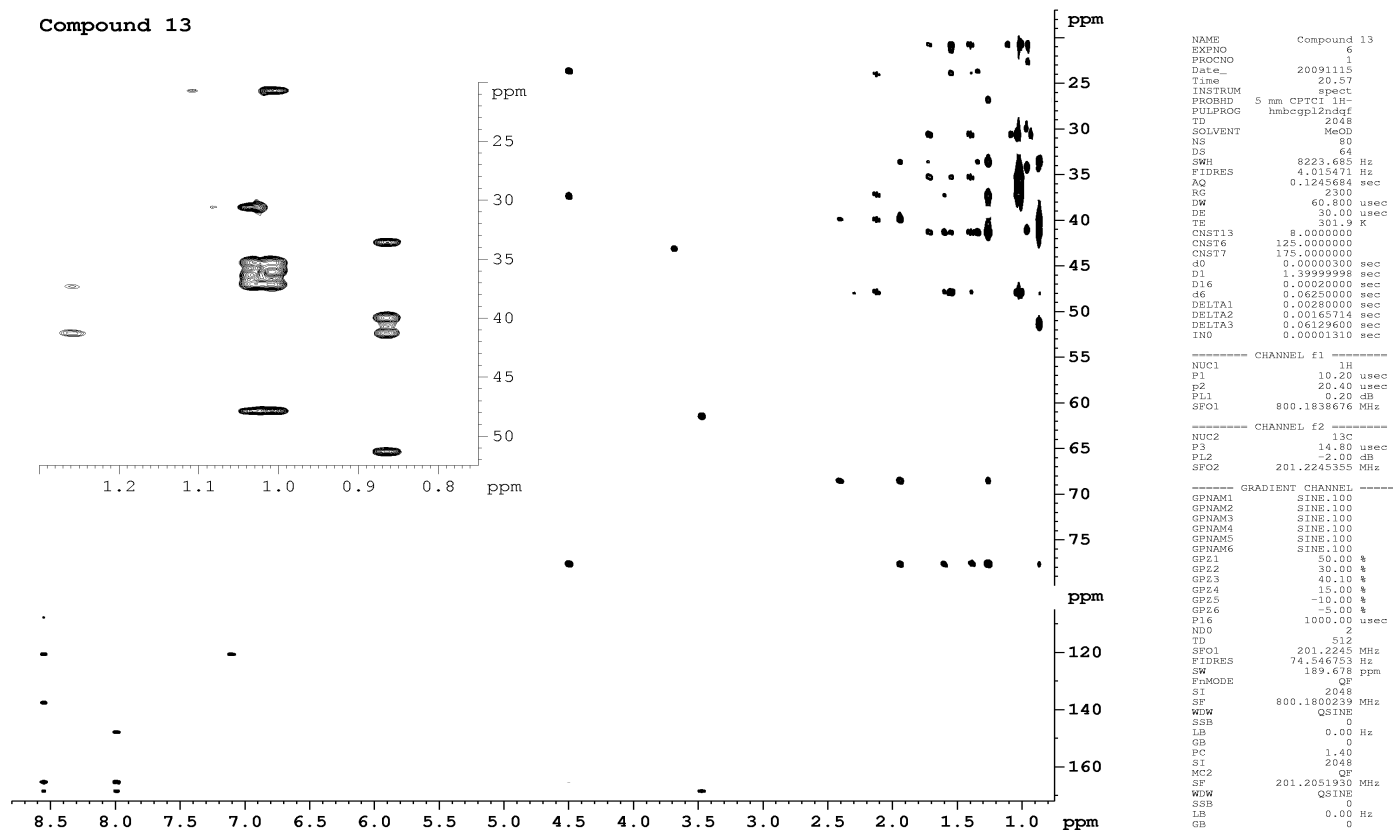
NAME          Compound 13
EXPNO         5
PROCNO        1
Date_         20091115
Time          5.50
INSTRUM       spect
PROBHD        5 mm CPTCI 1H-
PULPROG       hzgdcletqz4p.2
TD            1024
SOLVENT       MeOD
NS            32
DS            2
SHE          8223.685 Hz
FIDRES       0.030762 Hz
AQ           0.4983025 sec
RG           2300
DW           60.400 usec
DE           30.00 usec
TE           301.3 K
CNS117       -0.5000000
CNS12        149.0000000
d0           0.00000000 sec
d1           1.50000000 sec
d11          0.00000000 sec
d16          0.00020000 sec
d19         0.00086200 sec
d4           0.00172414 sec
d9           0.00000000 sec
DELTA        0.00122240 sec
DELTA1       0.00120800 sec
DELTA2       0.00147014 sec
DELTA3       0.00135800 sec
FACTOR1      11
INO          0.00001655 sec
l1           22
ST1CNT       0

===== CHANNEL f1 =====
NUC1          13
P1           10.20 usec
P2           20.40 usec
P28          1000.00 usec
P6           24.00 usec
PL1          0.20 dB
PL10         7.63 dB
SFO1         800.1338676 MHz

===== CHANNEL f2 =====
CPDPRG2      P5eap180
NUC2          13C
P14          500.00 usec
P24          2000.00 usec
P3           34.00 usec
PCPD2        1800.00 usec
P201         120.00 dB
PL12         10.00 dB
PL2          -2.00 dB
SFO2         201.2205114 MHz
SP3          1.08 dB
SP7          1.08 dB
SPNAM15      Crp, 80, 1.5, 10, nd
SPNAM3       Crp80, 0.3, 10, 1
SPNAM7       Crp80comp.4
SPNAM15      0.500
SPNAM17      0.500
SPNAM19      0.500
SPNAM17      0.500
SPOFFS15     0.00 Hz
SPOFFS3       0.00 Hz
SPOFFS7       0.00 Hz

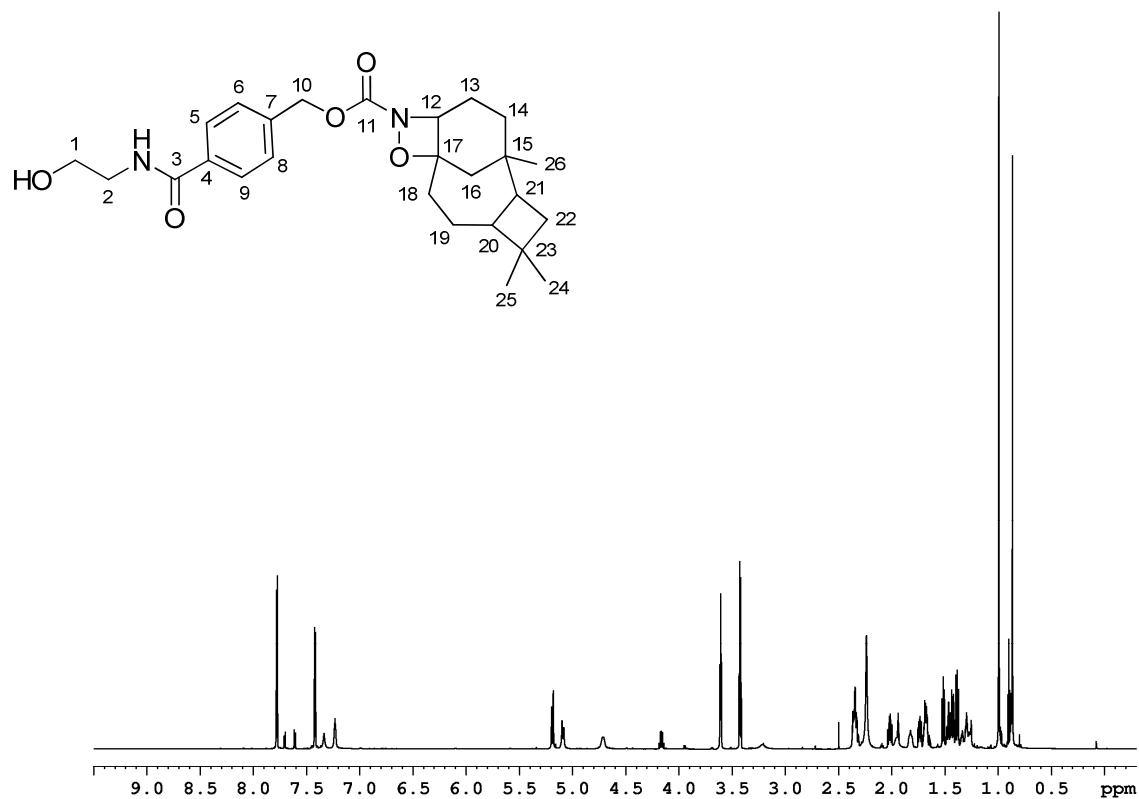
===== GRADIENT CHANNEL =====
GPNAM1       SINE.100
GPNAM2       SINE.100
GP21         80.00 %
GP22         20.10 %
P16          1000.00 usec
ND0          2
SFO1         201.2205 MHz
FIDRES       29.503399 Hz
SW           150.141 PPM
FMOD         Echo-Antiecho
SI           2048
SF           800.1800213 MHz
WDM          QSINE
SSB          0.00 Hz
GB           0
PC           1.00
SI           2048
MC2          echo-Antiecho
SF           201.2205170 MHz
WDM          QSINE
SSB          0.00 Hz
GB           0
  
```

2D ^{13}C -HSQC-TOCSY spectrum of **13** in CD_3OD at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively.



2D ^{13}C -HMBC spectrum of **13** in CD_3OD at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively. The insert in the upper left corner represents an expansion of the high field region plotted with reduced cross-peaks intensities for a better clarity.

Compound 16

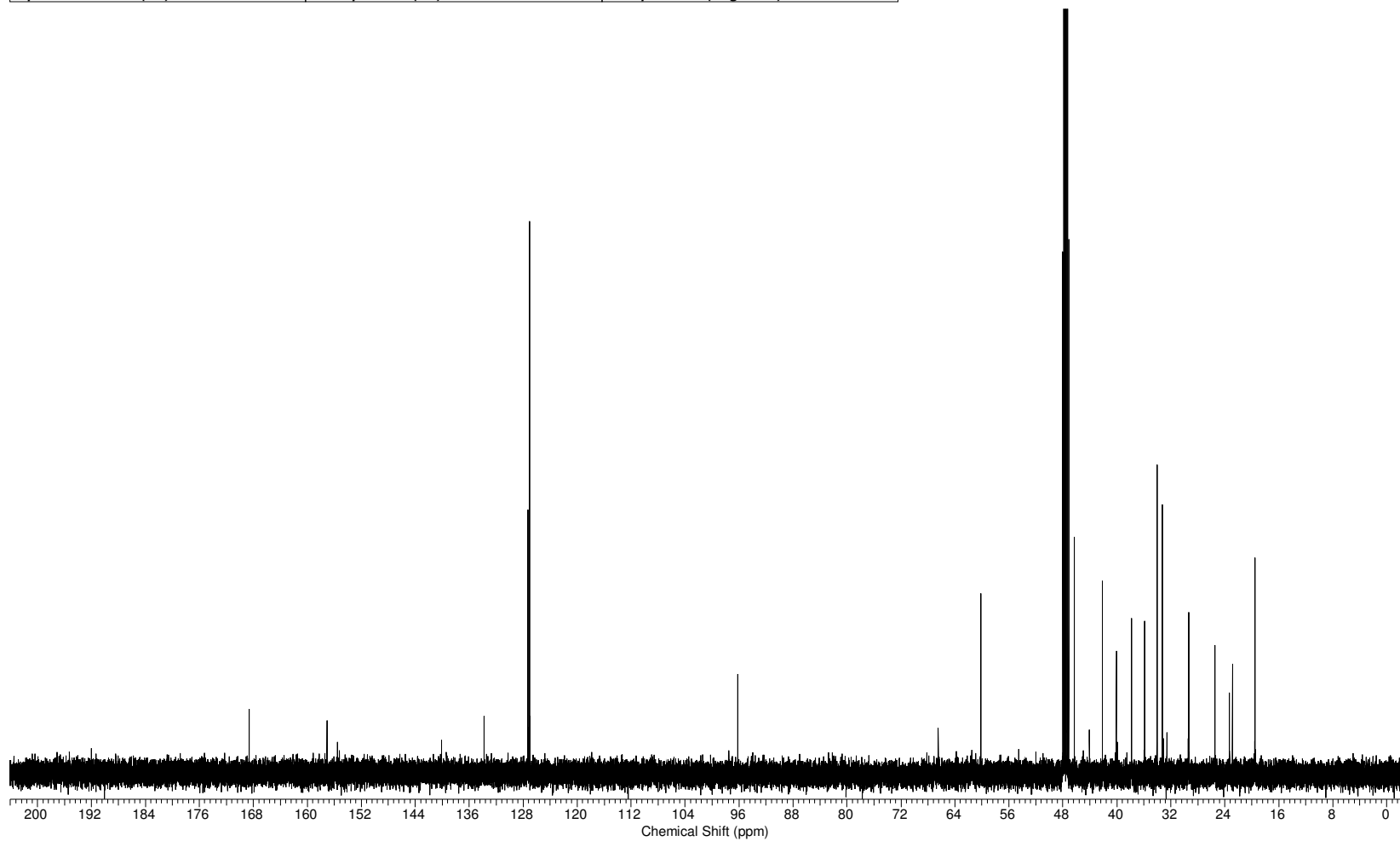


```
NAME          Compound 16
EXPNO          1
PROCNO         1
Date_         20091123
Time           9.15
INSTRUM       spect
PROBHD        5 mm CPTCI 1H-
PULPROG       zg
TD            65536
SOLVENT       CD3CN
NS            128
DS            4
SWH           8223.685 Hz
FIDRES        0.125483 Hz
AQ            3.9846387 sec
RG            32
DW            60.800 usec
DE            30.00 usec
TE            280.0 K
D1            0.10000000 sec
TD0           1
```

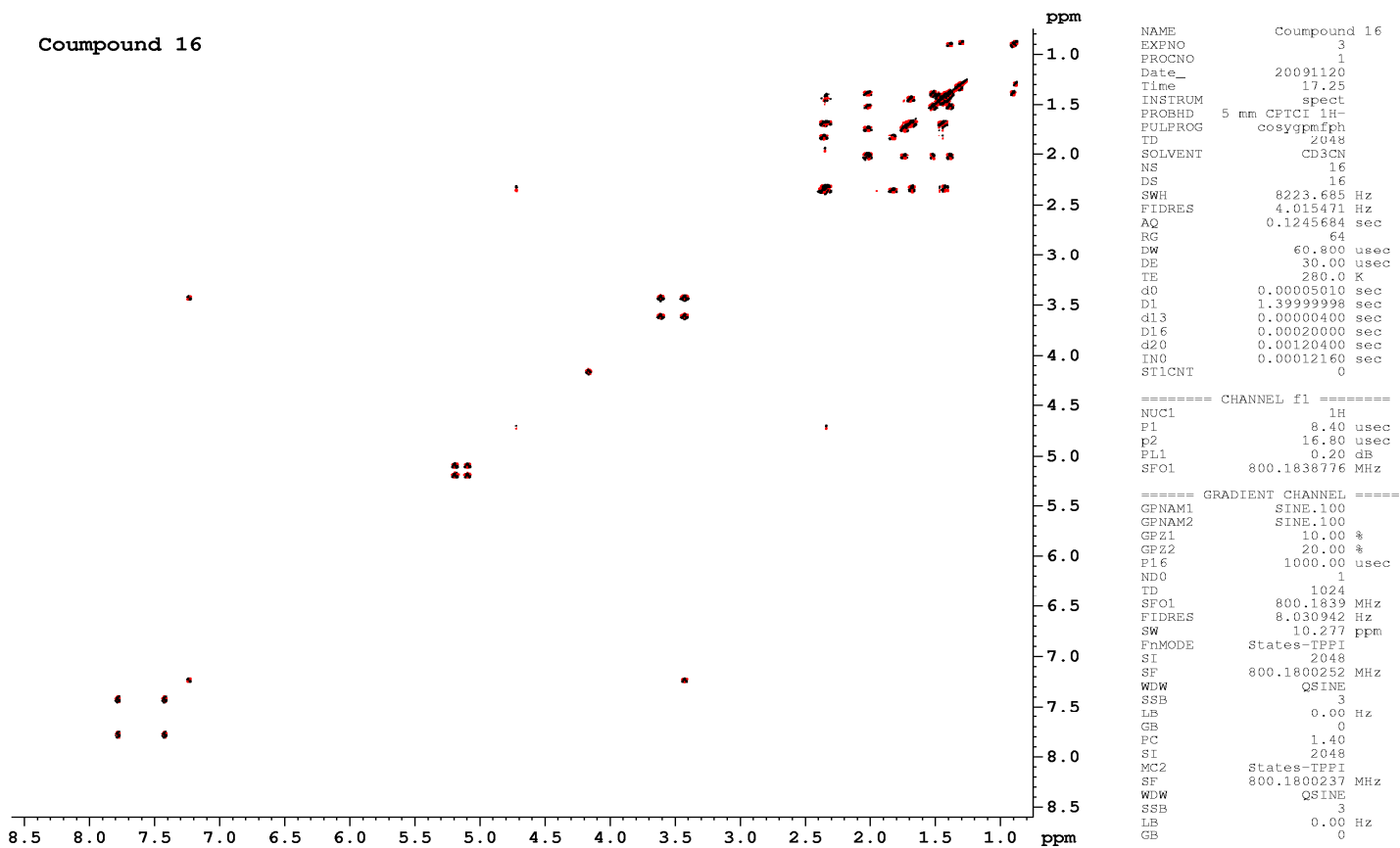
```
===== CHANNEL f1 =====
NUC1           1H
P1             8.40 usec
PL1            0.20 dB
SFO1          800.1837640 MHz
SI            131072
SF            800.1800262 MHz
WDW            no
SSB            0
LB             0.00 Hz
GB             0
PC             1.00
```

¹H spectrum of **16** in CD₃CN at 25° C.

Acquisition Time (sec)	1.7826	Comment	Compound 16	Date	May 27 2009	Date Stamp	May 27 2009
						Frequency (MHz)	150.85
Nucleus	¹³ C	Number of Transients	256	Original Points Count	65536	Points Count	65536
Pulse Sequence	s2pul	Receiver Gain	60.00	Solvent	METHANOL-d4		
Spectrum Offset (Hz)	15838.1133	Sweep Width (Hz)	36764.71	Temperature (degree C)	22.000		

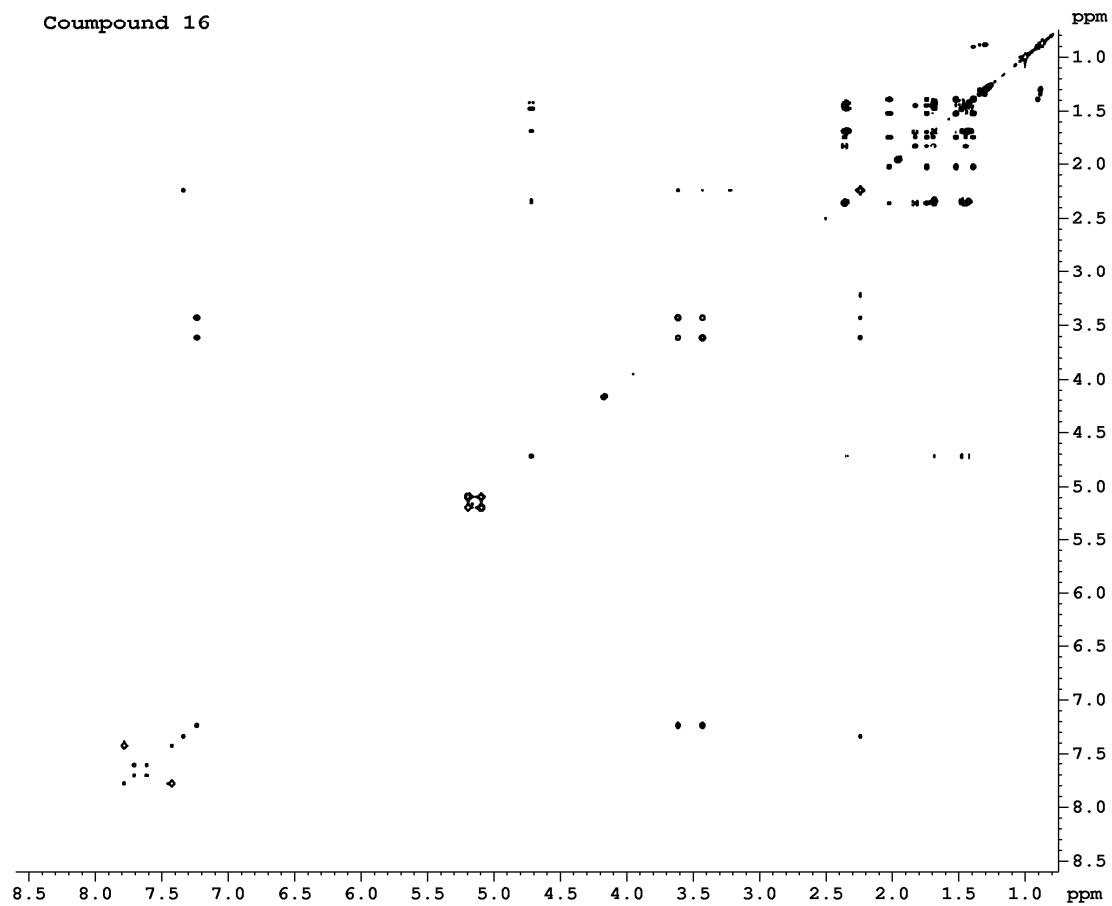


¹³C{¹H} spectrum of **16** in CD₃OD at 25° C.



2D DQF-COSY spectrum of **16** in CD₃CN at 25° C. The black and red contours represent positive and negative peaks, respectively.

Compound 16



```

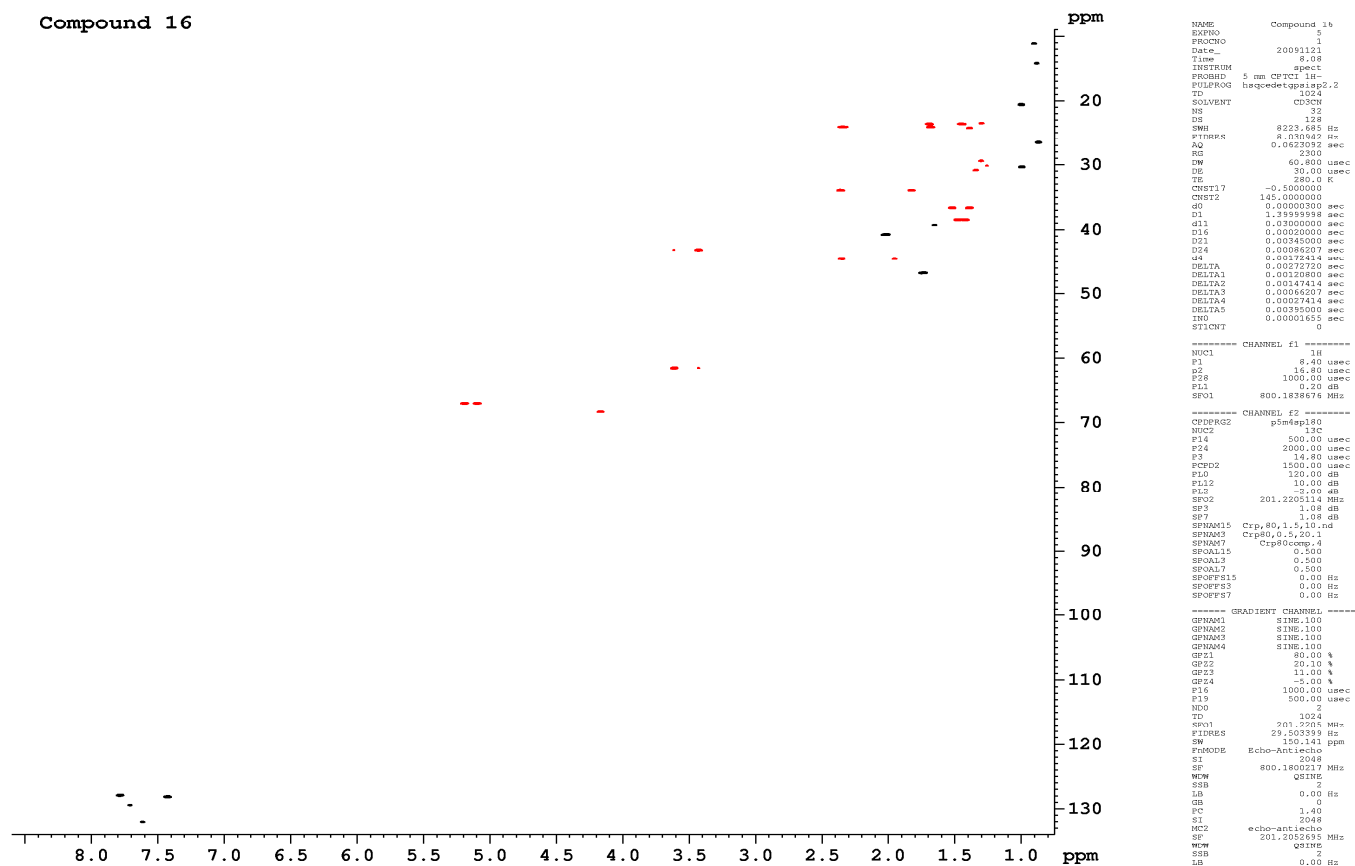
NAME          Compound 16
EXPNO         4
PROCNO        1
Date_         20091121
Time          0.32
INSTRUM       spect
PROBHD        5 mm CPTCI 1H-
PULPROG       dipssi2etgpsi
TD            2048
SOLVENT       CD3CN
NS            16
DS            16
SWH           8223.685 Hz
FIDRES        4.015471 Hz
AQ            0.1245684 sec
RG            64
DW            60.800 usec
DE            30.00 usec
TE            280.0 K
d0            0.00000300 sec
D1            1.39999998 sec
d11           0.03000000 sec
D16           0.00020000 sec
D20           0.00001000 sec
D21           0.00001000 sec
D9            0.06000000 sec
DELTA         0.00120300 sec
DELTA1        0.00120800 sec
FACTOR1       11
l1            22
INO           0.00012160 sec
STICNT        0

===== CHANNEL f1 =====
NUC1          1H
P1            8.40 usec
P2            16.80 usec
P6            24.00 usec
PL1           0.20 dB
PL10          9.32 dB
SFO1          800.1838676 MHz

===== GRADIENT CHANNEL =====
GPNAM1        SINE.100
GPNAM2        SINE.100
GPZ1          30.00 %
GPZ2          30.00 %
P16           1000.00 usec
ND0           1
TD            1024
SFO1          800.1839 MHz
FIDRES        8.030942 Hz
SW            10.277 ppm
FnMODE        Echo-Antiecho
SI            2048
SF            800.1800240 MHz
WDW           QSINE
SSB           2
LB            0.00 Hz
GB            0
PC            1.40
SI            2048
MC2           echo-antiecho
SF            800.1800239 MHz
WDW           QSINE
SSB           2
LB            0.00 Hz
GB            0
    
```

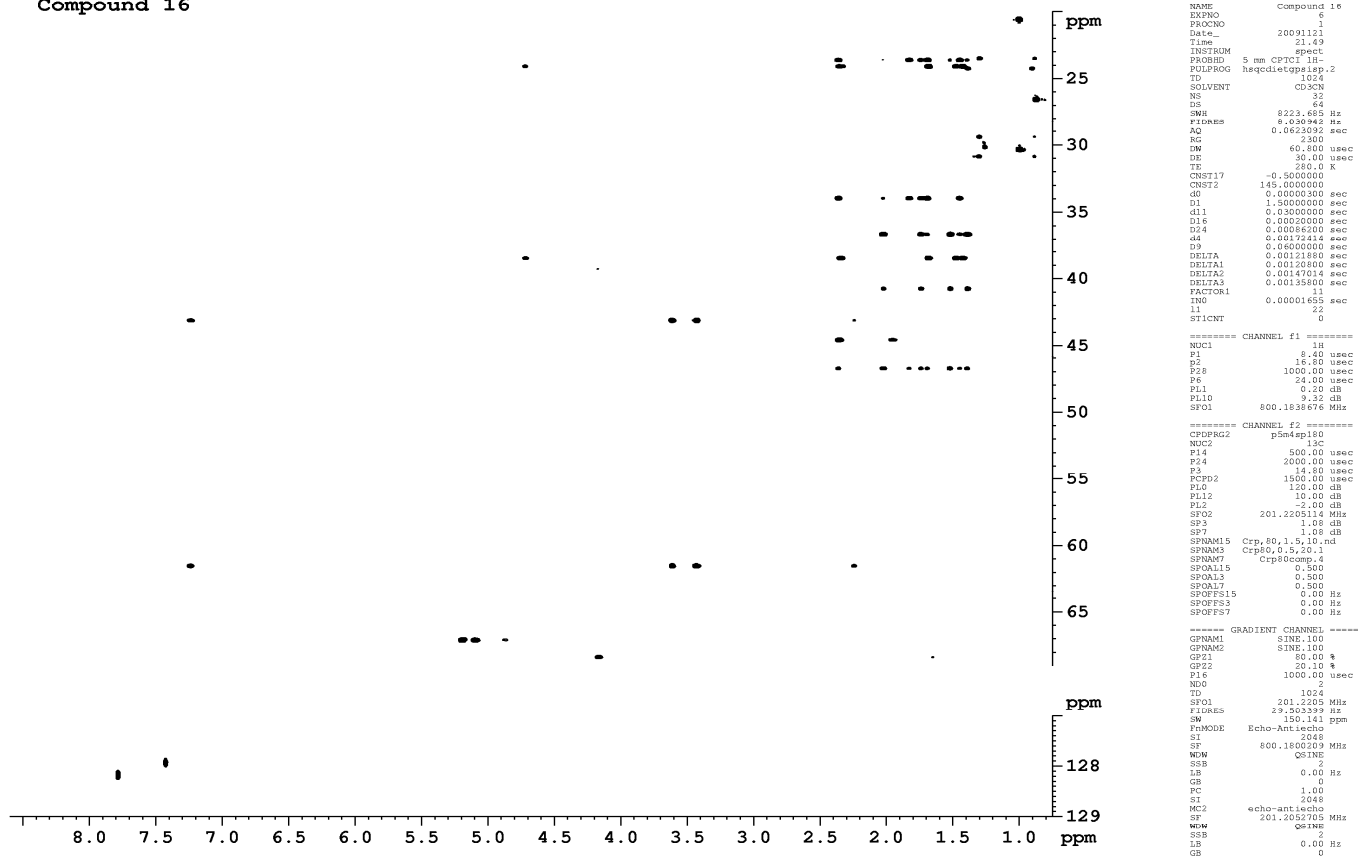
2D TOCSY spectrum of **16** in CD₃CN at 25° C.

Compound 16

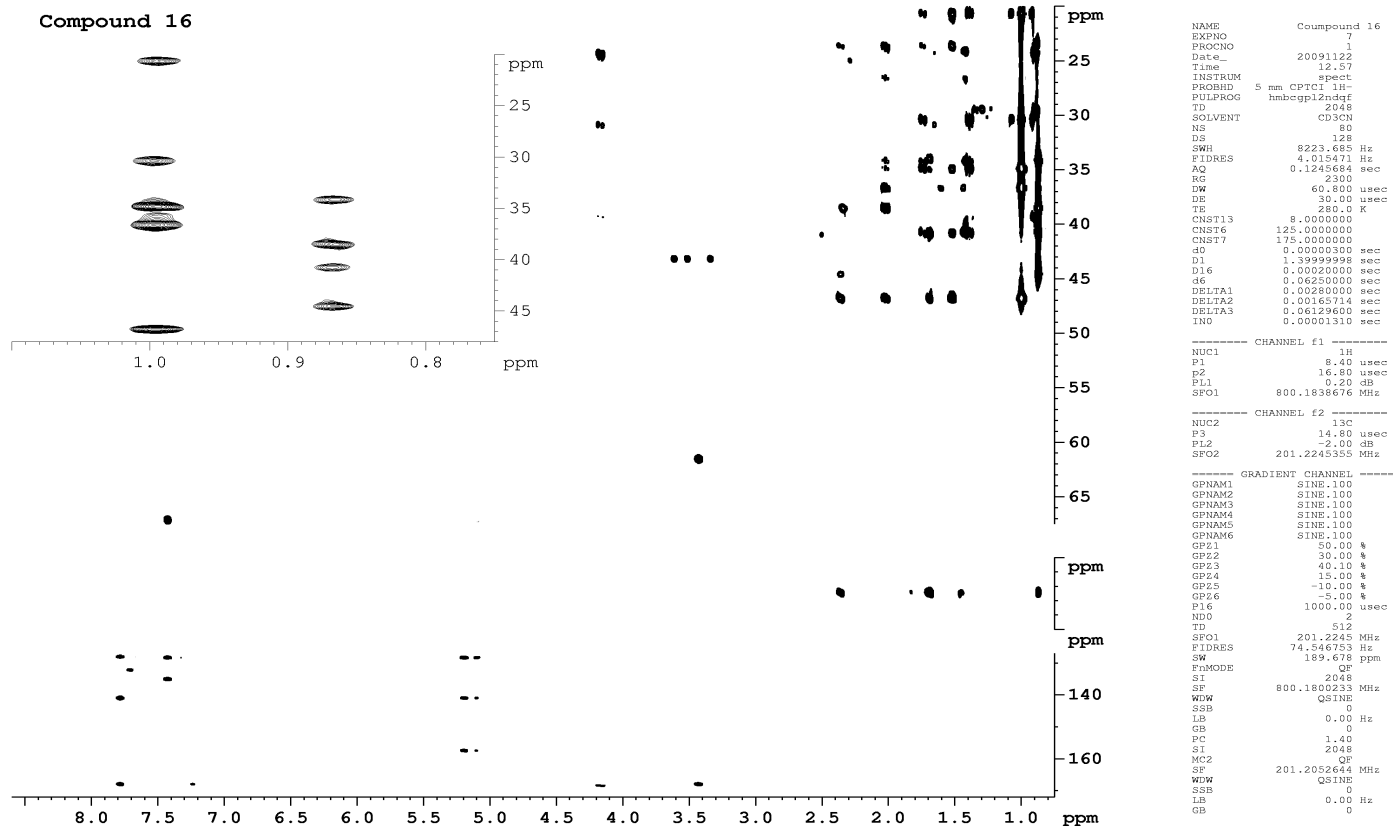


2D ^{13}C -HSQC spectrum of **16** in CD_3CN at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively. The black cross-peaks correspond to CH_3 and CH groups, the red cross-peaks correspond to CH_2 groups.

Compound 16

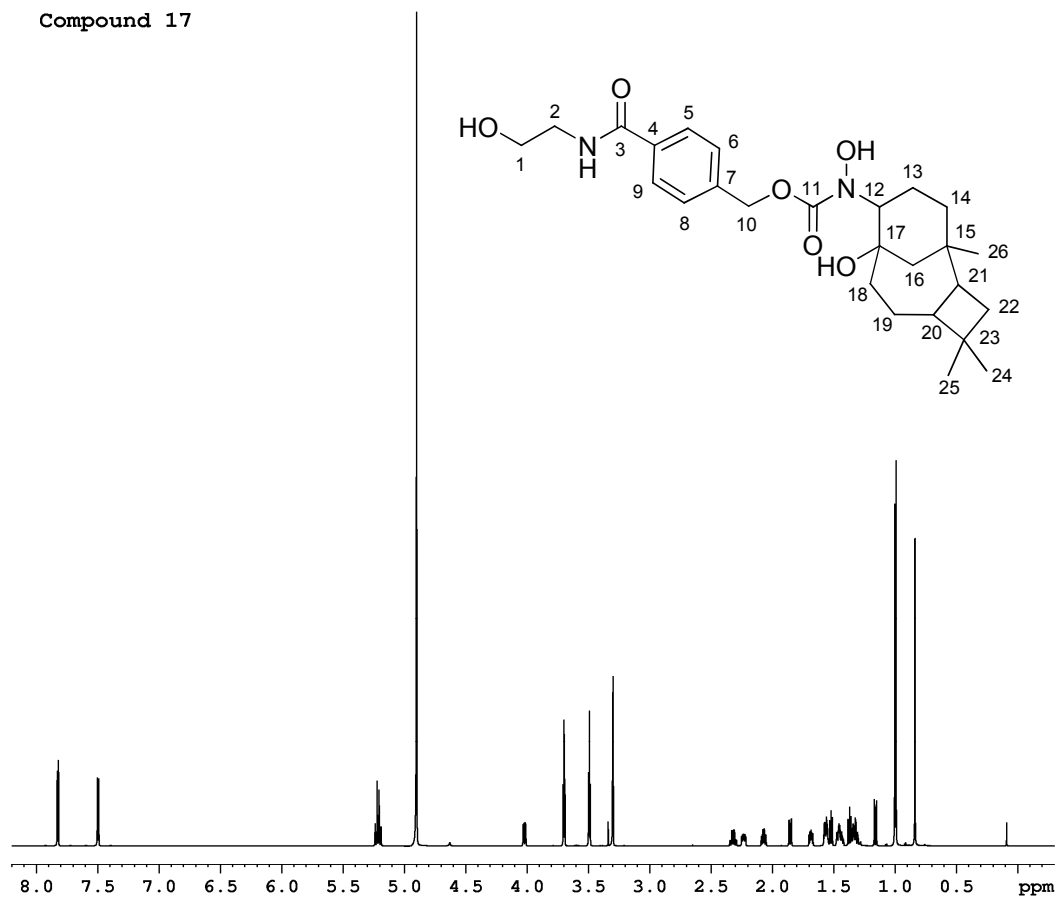


2D ^{13}C -HSQC-TOCSY spectrum of **16** in CD_3CN at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively.



2D ^{13}C -HMBC spectrum of **16** in CD_3CN at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively. The insert in the upper left corner represents an expansion of the high field region plotted with reduced cross-peaks intensities for a better clarity.

Compound 17



```

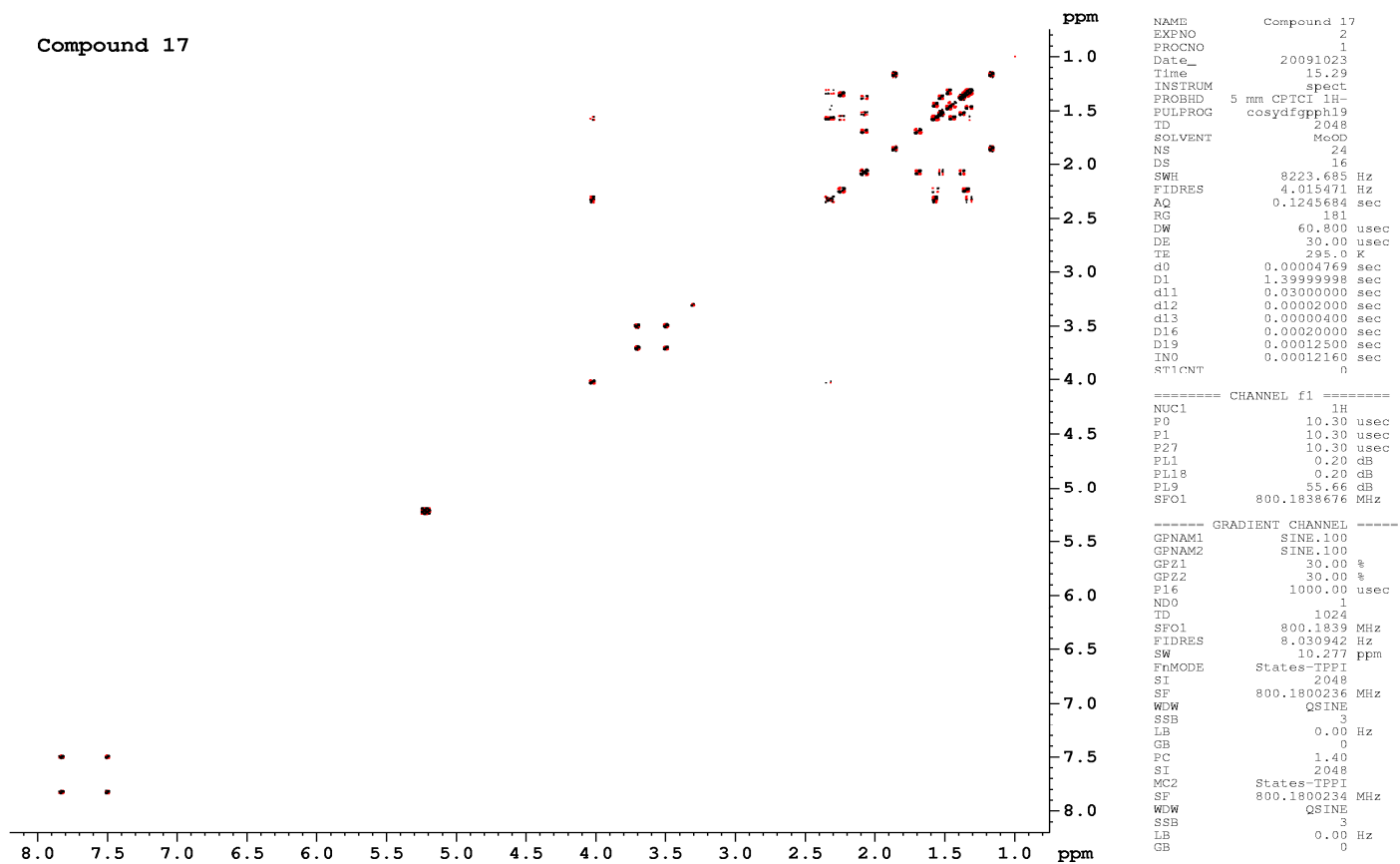
NAME          Coumpound 17
EXPNO         1
PROCNO        1
Date_         20091023
Time          15.28
INSTRUM       spect
PROBHD        5 mm CPTCI 1H-
PULPROG       zg
TD            65536
SOLVENT       MeOD
NS            128
DS            4
SWH           8223.685 Hz
FIDRES        0.125483 Hz
AQ            3.9846387 sec
RG            71.8
DW            60.800 usec
DE            30.00 usec
TE            295.0 K
D1            0.10000000 sec
TD0           1
    
```

```

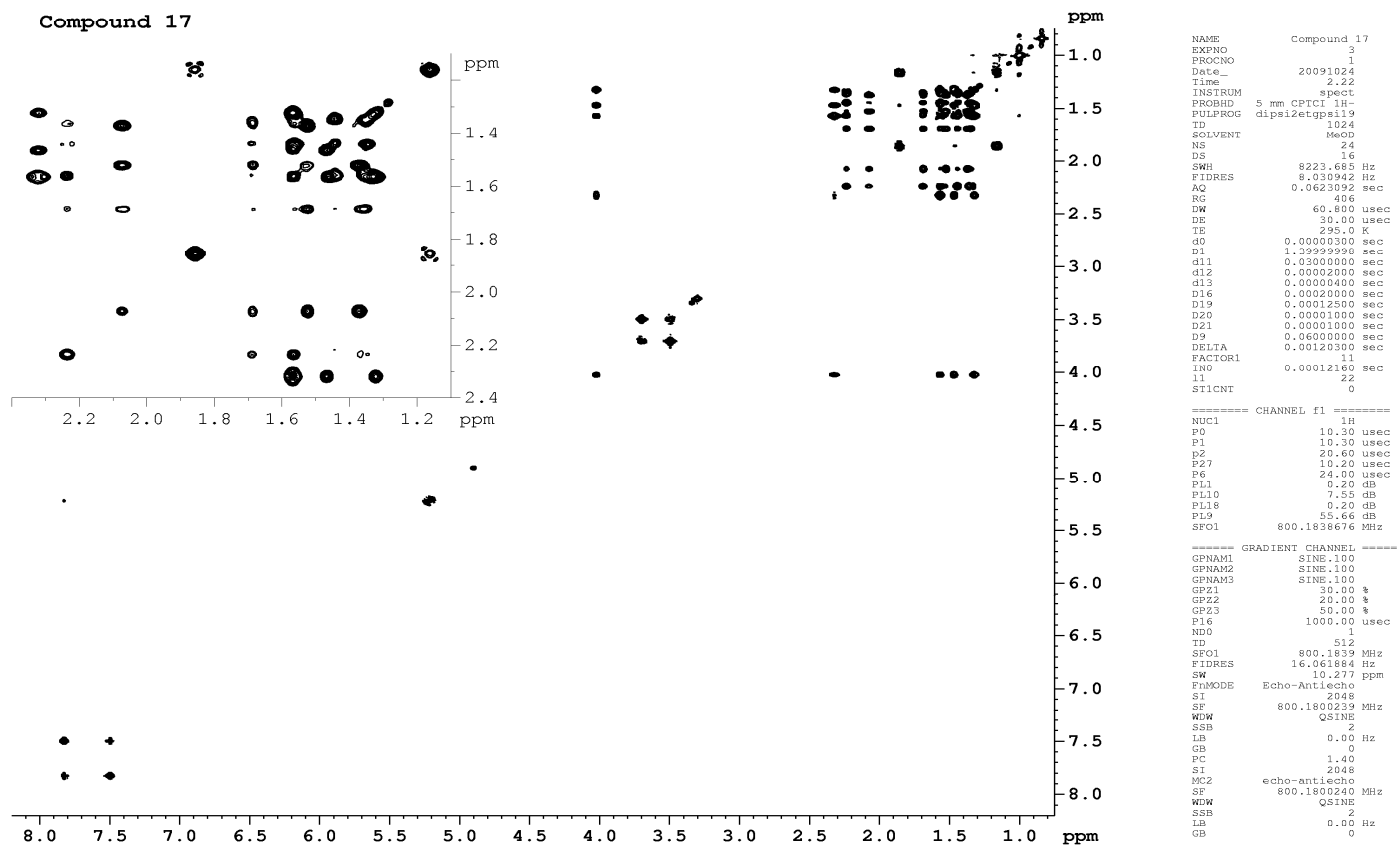
===== CHANNEL f1 =====
NUC1          1H
P1            10.30 usec
PL1           0.20 dB
SFO1          800.1837640 MHz
SI            131072
SF            800.1800265 MHz
WDW           no
SSB           0
LB            0.00 Hz
GB            0
PC            1.00
    
```

¹H spectrum of 17 in CD₃OD at 25° C.

Compound 17

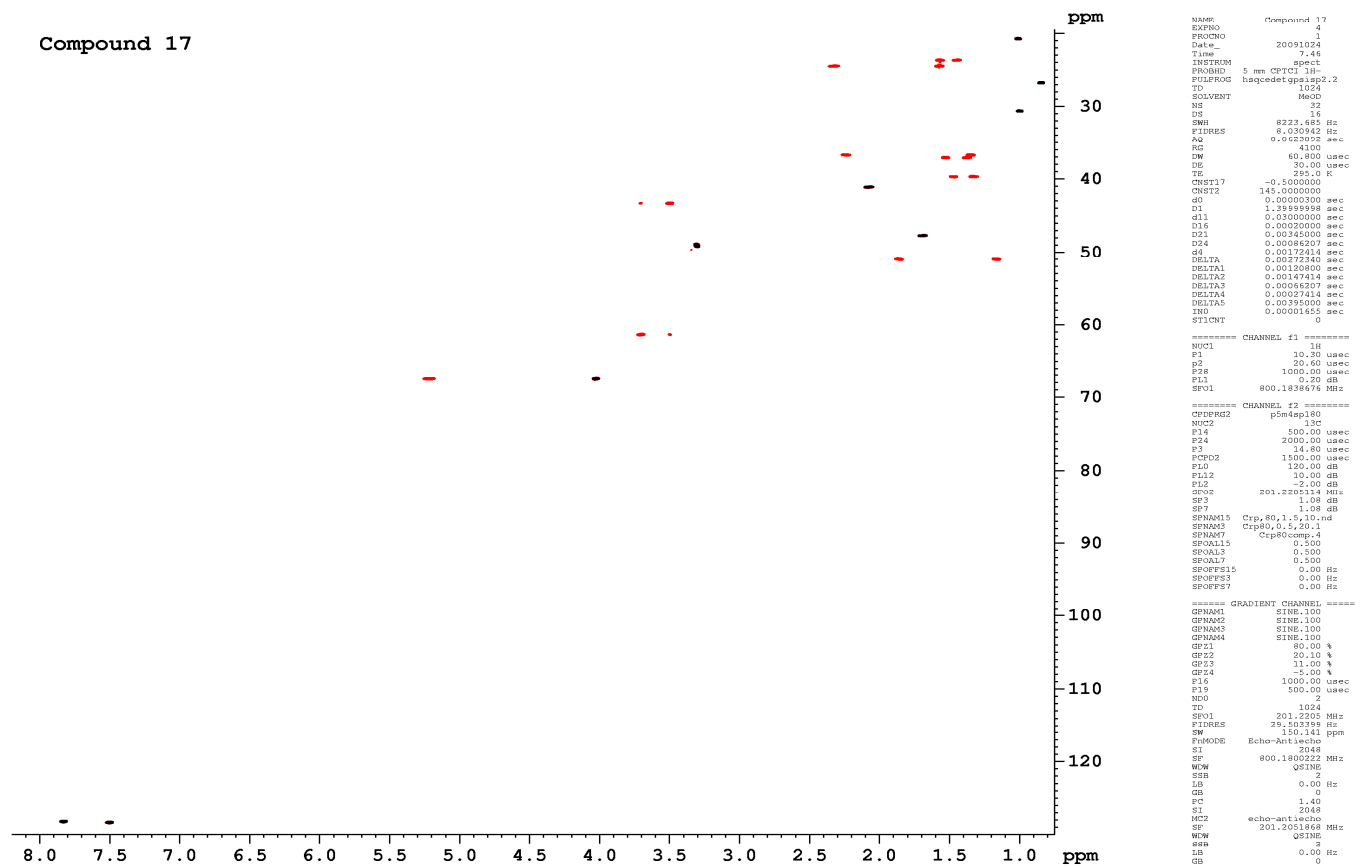


2D DQF-COSY spectrum of **17** in CD₃OD at 25° C. The black and red contours represent positive and negative peaks, respectively.



2D TOCSY spectrum of **17** in CD₃OD at 25^o C. The insert in the upper left corner represents an expansion of the high field region plotted with reduced cross-peaks intensities for a better clarity.

Compound 17



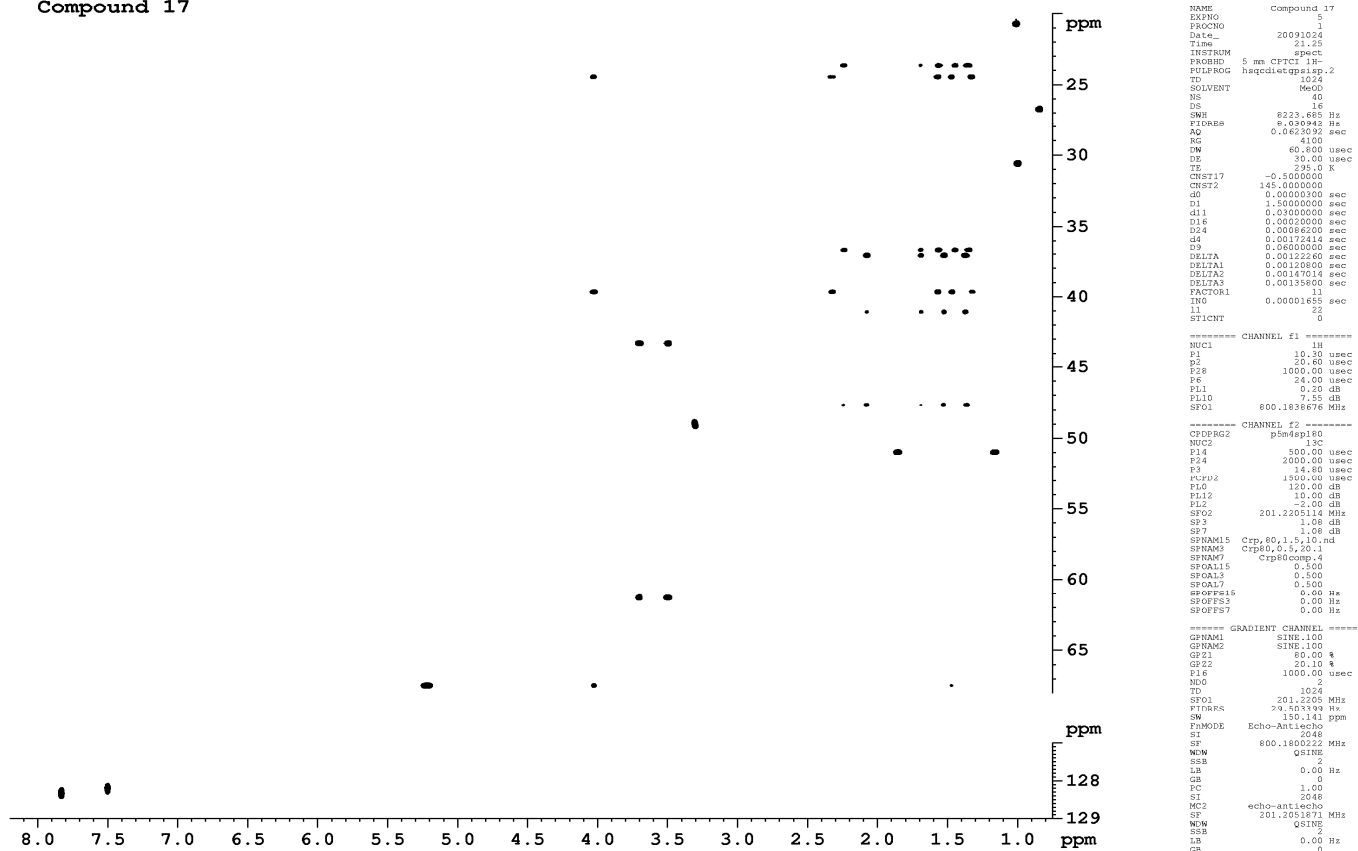
```

NAME Compound 17
EXPNO 4
PROCNO 1
DATE_ 20091024
TIME 7:46
INSTRUM spect
PROBHD 5 mm CPYCO 1H-
PULPROG zgpg30p2.2
TD 1024
SOLVENT MeOD
NS 32
DS 16
SWH 8223.685 Hz
FIDRES 0.020942 Hz
AQ 0.002023 sec
RG 4100
DW 60.800 usec
DE 30.00 usec
TE 300.0 K
CONST17 -0.5000000
CONST2 145.0000000
d0 0.0000000 sec
d1 1.3999999 sec
d11 0.0300000 sec
d16 0.0000000 sec
d21 0.0034500 sec
d24 0.0008207 sec
d3 0.0017214 sec
DELTA 0.0027340 sec
DELTA1 0.0010900 sec
DELTA2 0.0014714 sec
DELTA3 0.0008207 sec
DELTA4 0.0027414 sec
DELTA5 0.0000000 sec
IN0 0.0001655 sec
SFOFF 0
===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
P2 20.60 usec
P3 1000.00 usec
PL1 0.00 dB
PL2 0.00 dB
PL3 0.00 dB
PL4 0.00 dB
PL5 0.00 dB
PL6 0.00 dB
PL7 0.00 dB
PL8 0.00 dB
PL9 0.00 dB
PL10 0.00 dB
PL11 0.00 dB
PL12 0.00 dB
PL13 0.00 dB
PL14 0.00 dB
PL15 0.00 dB
PL16 0.00 dB
PL17 0.00 dB
PL18 0.00 dB
PL19 0.00 dB
PL20 0.00 dB
PL21 0.00 dB
PL22 0.00 dB
PL23 0.00 dB
PL24 0.00 dB
PL25 0.00 dB
PL26 0.00 dB
PL27 0.00 dB
PL28 0.00 dB
PL29 0.00 dB
PL30 0.00 dB
PL31 0.00 dB
PL32 0.00 dB
PL33 0.00 dB
PL34 0.00 dB
PL35 0.00 dB
PL36 0.00 dB
PL37 0.00 dB
PL38 0.00 dB
PL39 0.00 dB
PL40 0.00 dB
PL41 0.00 dB
PL42 0.00 dB
PL43 0.00 dB
PL44 0.00 dB
PL45 0.00 dB
PL46 0.00 dB
PL47 0.00 dB
PL48 0.00 dB
PL49 0.00 dB
PL50 0.00 dB
PL51 0.00 dB
PL52 0.00 dB
PL53 0.00 dB
PL54 0.00 dB
PL55 0.00 dB
PL56 0.00 dB
PL57 0.00 dB
PL58 0.00 dB
PL59 0.00 dB
PL60 0.00 dB
PL61 0.00 dB
PL62 0.00 dB
PL63 0.00 dB
PL64 0.00 dB
PL65 0.00 dB
PL66 0.00 dB
PL67 0.00 dB
PL68 0.00 dB
PL69 0.00 dB
PL70 0.00 dB
PL71 0.00 dB
PL72 0.00 dB
PL73 0.00 dB
PL74 0.00 dB
PL75 0.00 dB
PL76 0.00 dB
PL77 0.00 dB
PL78 0.00 dB
PL79 0.00 dB
PL80 0.00 dB
PL81 0.00 dB
PL82 0.00 dB
PL83 0.00 dB
PL84 0.00 dB
PL85 0.00 dB
PL86 0.00 dB
PL87 0.00 dB
PL88 0.00 dB
PL89 0.00 dB
PL90 0.00 dB
PL91 0.00 dB
PL92 0.00 dB
PL93 0.00 dB
PL94 0.00 dB
PL95 0.00 dB
PL96 0.00 dB
PL97 0.00 dB
PL98 0.00 dB
PL99 0.00 dB
PL100 0.00 dB
===== CHANNEL f2 =====
CPDPRG2 pSine40.80
NUC2 13C
P14 500.00 usec
P24 2000.00 usec
P3 141.80 usec
PCPD2 1500.00 usec
P10 150.00 dB
P12 10.00 dB
P13 -2.00 dB
P14 201.2205 MHz
P15 1.08 dB
P16 1.08 dB
SPAN15 Cfg, 80, 1, 6, 10, nd
SPAN25 Cfg, 0, 1, 2, 20, 1
SPAN7 Cfg, 80, 0, 4
SFOFF15 0.500
SFOFF25 0.500
SFOFF7 0.00 Hz
SFOFF15 0.00 Hz
SFOFF25 0.00 Hz
SFOFF7 0.00 Hz
===== GRADIENT CHANNEL =====
GPNAM1 SINE.100
GPNAM2 SINE.100
GPNAM3 SINE.100
GPNAM4 SINE.100
GP21 80.00 %
GP22 20.10 %
GP23 11.00 %
GP24 -2.00 %
P16 1000.00 usec
P15 500.00 usec
ND0 2
TD 1024
SFO1 201.2205 MHz
FIDRES 29.503399 Hz
SW 150.141 ppm
F2MODE Echo-AntiEcho
SI 2048
SF 800.1809222 MHz
WVW GSIW
SSB 2
LB 0.00 Hz
GB 0
FC 1.40
SI 2048
F2MODE Echo-AntiEcho
SF 201.2051868 MHz
WVW GSIW
SSB 2
LB 0.00 Hz
GB 0

```

2D ¹³C-HSQC spectrum of **17** in CD₃OD at 25° C. The horizontal and vertical axes display ¹H and ¹³C chemical shifts, respectively. The black cross-peaks correspond to CH₃ and CH groups, the red cross-peaks correspond to CH₂ groups.

Compound 17



```

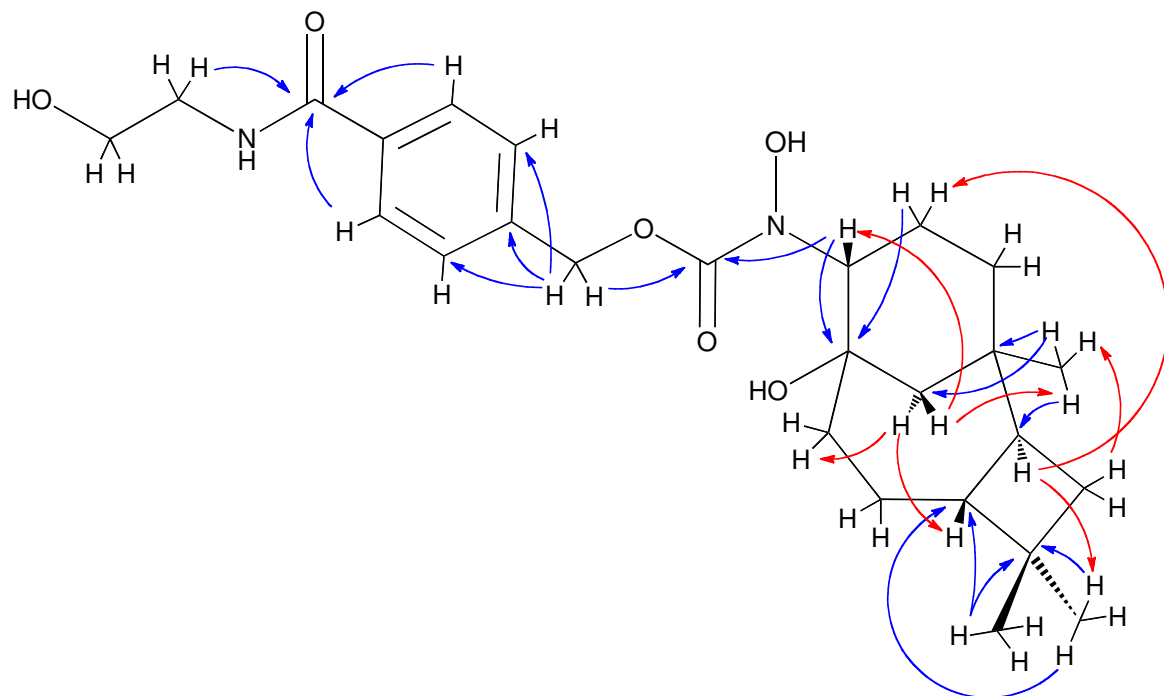
NAME          Compound_17
EXPNO         5
PROCNO        1
Date_         2001024
Time          21:25
INSTRUM       spect
PROBHD        5 mm CPTCI 1H-
PULPROG       hsqcletqp3sp.2
TD            1024
SOLVENT       MeOD
NS            40
DS            16
SWH           8222.465 Hz
FIDRES        0.000943 Hz
AQ            0.0623032 sec
RG            4100
DN            60.800 usec
DE            30.00 usec
TE            295.0 K
CNS17         -0.5000000
CNS22         145.0000000
d0            0.0000000 sec
D1            1.5000000 sec
d11           0.0300000 sec
D16           0.0000000 sec
D24           0.00086200 sec
d4            0.00172414 sec
D9            0.00000000 sec
DELTA2        0.00122800 sec
DELTA1        0.00120800 sec
DELTA2A       0.00147014 sec
DELTA2A3      0.00135800 sec
FACTOR1       11
IND           22
L1            0
STICNT        0

----- CHANNEL f1 -----
NUC1          1H
P1            10.10 usec
P2            20.60 usec
P28           1000.00 usec
P6            24.00 usec
PL1           0.20 dB
PL10          7.55 dB
SF01          800.1836676 MHz

----- CHANNEL f2 -----
CPOBPG2      p5m4sp160
NUC2          13C
P14           500.00 usec
P24           2000.00 usec
P9            14.80 usec
P17F2        1500.00 usec
PL12          120.00 dB
PL10          10.00 dB
PL2           -2.00 dB
SF02         201.2205114 MHz
SP3           1.08 dB
SP7           1.08 dB
SFXAM15      Crp, 60, 1.5, 10, nd
SFXAM3       Crp0, 0.5, 20, 1
SFXAM7       Crp0comp, 4
SFOAL15      0.500
SFOAL5        0.500
SFOAL7        0.500
SFOFFPS15    0.00 Hz
SFOFFPS3     0.00 Hz
SFOFFPS7     0.00 Hz

----- GRADIENT CHANNEL -----
GPNAM1       SINE, 100
GPNAM2       SINE, 100
GP21         80.00 %
GP22         20.10 %
FILE         1000.00 usec
TD           2
F0           1024
SF01         201.2205 MHz
FTRFPS       20.505599 Hz
SW           150.141 ppm
FMODE        Echo-Anti1echo
SI           2048
SF           800.1800222 MHz
NSW          GSINE
SSB          2
LB           0.00 Hz
GB           0
PC           1.00
SI           2048
MC2          echo-anti1echo
SF           201.2205171 MHz
NSW          GSINE
SSB          2
LB           0.00 Hz
GB           0
    
```

2D ^{13}C -HSQC-TOCSY spectrum of **17** in CD_3OD at 25°C . The horizontal and vertical axes display ^1H and ^{13}C chemical shifts, respectively.



Correlation diagram for compound **17**. The HMBC (in blue) and NOE (in red) correlations, together with proton-proton correlations in the DQF-COSY and TOCSY spectra and one bond proton-carbon correlations in the HSQC spectrum unambiguously establish the proposed structure of compound **17** and conformation of its β -caryophyllene unit (segment).

References and Notes

1. For a detailed account of this antibiotic susceptibility assay see: Afonin, S.; Glaser, R. W.; Berditchevskaja, M.; Wadhvani, P.; Gührs, K.-H.; Möllmann, U.; Perner, A.; Ulrich, A. S. "4-Fluoro-phenylglycine as a Label for ^{19}F -NMR Structure Analysis of Membrane Associated Peptides." *ChemBioChem* **2003**, 4, 1151-1163.

2. Murray, P. R.; Baron, E. J.; Tenover, F. C.; Tenover, F. C.; Tenover, F. C.; Yolken, R. H. *Manual of Clinical Microbiology*, 7th ed.; American Society for Microbiology: Washington, DC, **1999**.
3. The test organisms *Mycobacterium vaccae* IMET 10670 and *Mycobacterium smegmatis* mc²155 required longer incubation times for overnight LB cultures (24-48 hours) and were used in the agar diffusion assay directly from the LB culture broth without dilution and standardization. The test organism *Bacillus subtilis* ATCC 6633 was used in the agar diffusion assay directly from the LB culture broth without dilution and standardization.
4. Snapper, S. B.; Melton, R. E.; Mustafa, S.; Kieser, T.; Jacobs, W. R. "Isolation and Characterization of Efficient Plasmid Transformation Mutants of *Mycobacterium smegmatis*." *Mol. Microbiol.* **1990**, *4*, 1911-1919.
5. Zimmermann, W. "Penetration of β -Lactam Antibiotics into their Target Enzymes in *Pseudomonas aeruginosa*: Comparison of a Highly Sensitive Mutant with its Parent Strain." *Antimicrob. Agents Chemother.* **1980**, *18*, 94-100.