

Supplementary materials

Supplementary Table S1. Fatty acid composition (weight %) of yeast cells expressing genes when feeding with C16:1^{Δ7}.

Supplementary Table S2. Fatty acid composition (weight %) of yeast cells expressing genes when fed with C20:4^{Δ8,11,14,17} or C20:4^{Δ5,8,11,14}.

Supplementary Table S3. Fatty acid composition (weight %) of yeast cells expressing CeFAT-1.

Supplementary Fig. S1. Mass spectra of products produced by CeFAT-2. A-D are C16:2, C16:3, C18:2 and C18:3 respectively.

Supplementary Fig. S2. Mass spectra of minor FAME products or their DMOX adducts in yeast cells expressing CeFAT-2 when fed with C14:1^{Δ9}.



A. C14:2^{Δ9,12}; B. DMOX-C14:2^{Δ9,12}; C. DMOX-C16:1^{Δ11}; D. DMOX-C16:2^{Δ11,14}; E. DMOX-C18:1^{Δ11}; F. DMOX-C18:1^{Δ13}; G. DMOX-C18:2^{Δ11,14}.

Supplementary Fig. S3. Partial GC chromatogram of yeast feeding for CeFAT-2 with C15:0 and mass spectra of products.

A-C, FAMES of yeast S288C expressing pYES2 vector only, AtFAD2 and CeFAT-2, respectively, fed with C15:0; D, DMOX-C15:1^{Δ9}; E, DMOX-C17:1^{Δ9}; F, DMOX-C15:2^{Δ9,12}; G, DMOX-C17:2^{Δ9,12}. New desaturated products from AtFAD2 or CeFAT-2 are indicated by arrows.

Supplementary Fig. S4. Mass spectra of new product of the feeding substrate C18:3^{Δ6,9,12} by CeFAT-2.

A, Mass spectra of new peak C18:4 from CeFAT-2; B, Mass spectra of DMOX adduct of new product C18:4. The new desaturated products from AtFAD2 or CeFAT-2 are indicated by arrows.

Supplementary Fig. S5. Summary of CeFAT-2 substrates used by Δ12-desaturation and Δ15-desaturation activity. The symbol  indicates no desaturation activity on this fatty acid, while  indicates low or trace activity on this fatty acid substrate.

Supplementary Fig. S6. Partial amino acid sequence alignment around His boxes between Δ12-desaturases, Δ15-desaturases and dual functional desaturases. The conserved His residues in His boxes are in bold. The GenBank accession numbers for the sequences used are, *Arabidopsis thaliana* Δ12-desaturase (AtFAD2, P46313), *Mortierella alpine* Δ12-desaturase (Ma Δ12, Q9Y8H5), *Umbelopsis isabellina* Δ12-desaturase (Ui Δ12, P59668), *Claviceps purpurea* Δ12-desaturase (Cp Δ12, ABS18716), *A. thaliana* Δ15-desaturase (AtFAD3, P48623), *Caenorhabditis elegans* Δ15-desaturase (CeFAT-1, L41807), *C. elegans* Δ12-desaturase (CeFAT-2, AF240777), *C. purpurea* Δ12/Δ15 bifunctional desaturase (Cp Δ12/15, ABS18717), *Fusarium moniliforme* Δ12/Δ15 bifunctional desaturase (Fm Δ12/15, DQ271516), *Magnaporthe grisea* Δ12/Δ15 bifunctional desaturase (Mg Δ12/15, XP_362963), *Aspergillus nidulans* Δ12/Δ15 bifunctional desaturase (An Δ12/15, XP_664808), *Acanthamoeba castellanii* Δ12/Δ15 bifunctional desaturase (Ac Δ12/15, ABK15557), *Sorghum bicolor* Δ12/Δ15 bifunctional desaturase (Sb Δ12/15, ABN49521). These sequences are grouped as Δ12-desaturase (Δ12), Δ15-desaturase (Δ15) and bifunctional desaturase (bi). The two critical residues of *C. purpurea* Δ12/Δ15 bifunctional desaturase, V152 and A206,

which were shown to contribute to regiospecificity of the enzyme (Meesapyodsuk *et al.* 2007, J. Biol. Chem. 282:20191-20199) are in red. The residues in CeFAT-2 that may potentially be important to the Δ 15-desaturation in addition to Δ 12-desaturation activity are in blue.