

**Supplementary Figure 1. *papX* linker-insertional mutants do not have impaired growth rate.** Thirty 15-bp insertional mutants of *papX* in pDRM001, as well as WT *papX* in pDRM001, were electroporated into strain CFT073  $\Delta papX$ . Strains were diluted 1:100 in LB broth and OD<sub>600</sub> was automatically recorded every 15 min by a bioanalyzer. No significant difference in growth rate was detected among the constructs. Name of transforming plasmid is shown on right.

**Supplementary Figure 2. PapX is a dimer** — (A) Recombinant purified PapX-His<sub>6</sub> and molecular-weight standards were run through a Sephacryl S-200 size-exclusion column at 1 ml/min and OD<sub>280</sub> was measured in mAU over a 300 ml volume. Traces of each were superimposed by aligning the void-volumes. PapX-His<sub>6</sub> peak appears at approximately twice the predicted molecular weight of the monomer. (B) SDS-PAGE was performed on non-denatured recombinant PapX-His<sub>6</sub>. A major band appears at twice the molecular weight of the monomer (the minor band).

**Supplementary Figure 3. Non-radiolabeled fluorescent DNase protect assay** (A) 0.5  $\mu$ g of recombinant purified PapX-His<sub>6</sub> was incubated with a 6-carboxyfluorescein end-labeled fragment derived from the first 360 bases of the *flhD* promoter and exposed to low-levels of DNase I for 1-20 minutes. Sheared fragments were processed by electrophoresis as for DNA sequencing to produce chromatograms representing all sheared fragments. Fluorescence of the 6-carboxyfluorescein is shown on the y-axis. Position of the fragment is shown on the x-axis. (A) Representative chromatogram of a fragment exposed to DNase I for 1 minute in the absence of PapX protein (B) Representative chromatogram of a fragment exposed to DNase I for 1 minute in the presence of 0.5  $\mu$ g of PapX. (C) Representative chromatogram of a fragment exposed to DNase I for 20 minutes in the presence of 0.5  $\mu$ g of PapX. Ratios of peak heights remained the same between conditions.

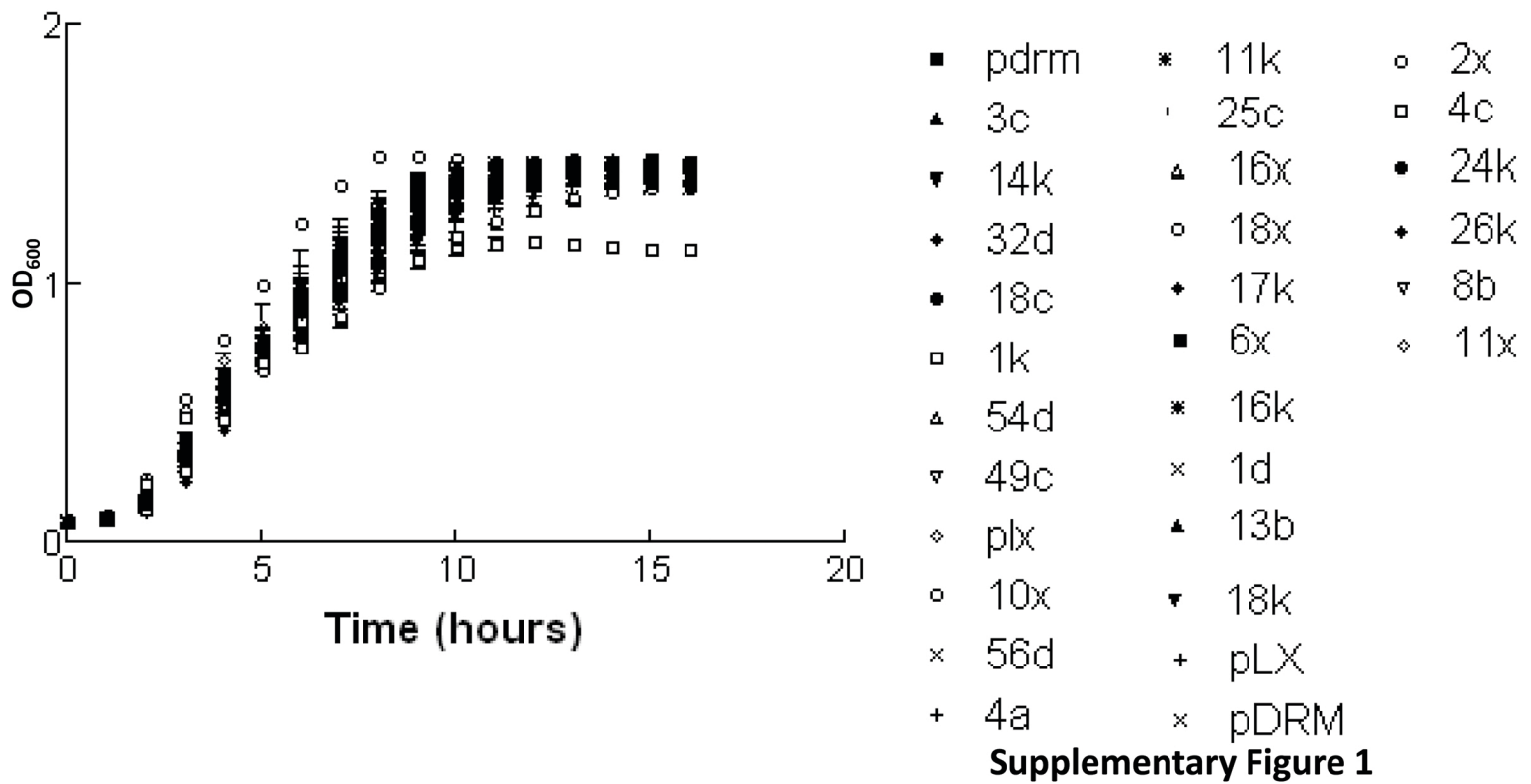
**Supplementary Table 1. Bacterial strains and plasmids used in this study.**

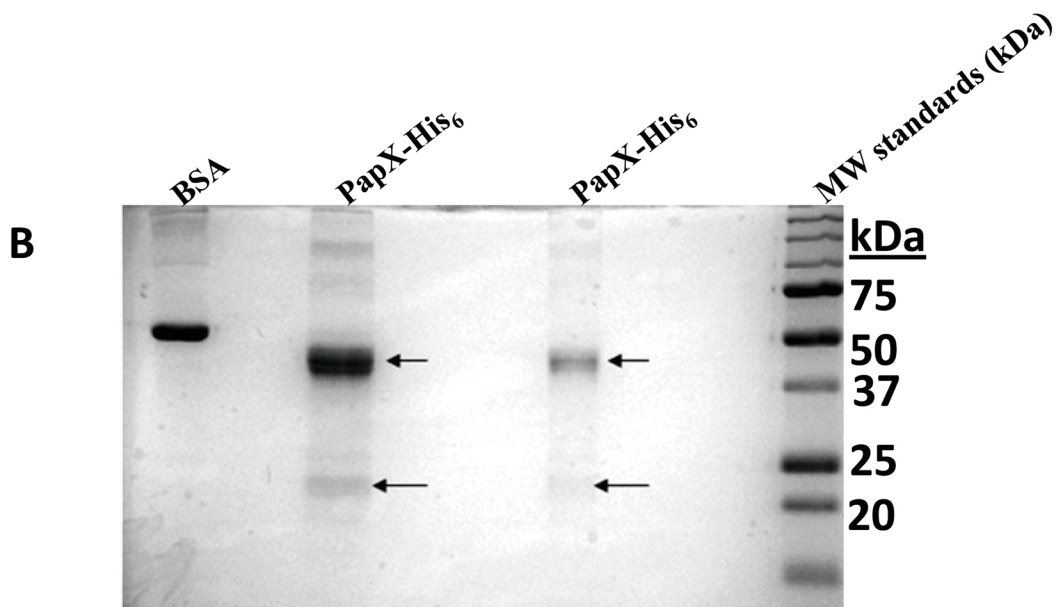
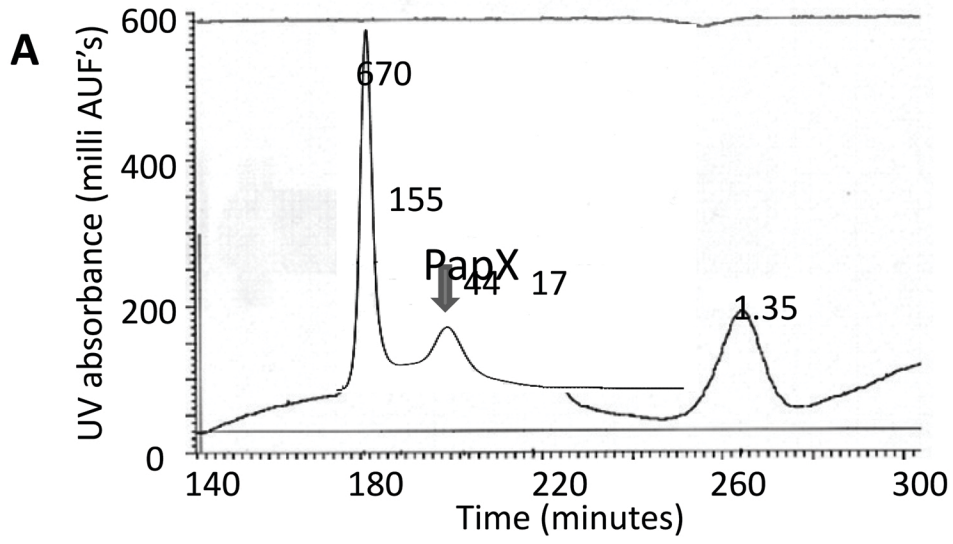
| Plasmid or strain name             | Function   |
|------------------------------------|--|
| pMCSG7                             | Recombinant PapX-His <sub>6</sub>                                      |
| pMCSG7_R127G                       | PapX-His <sub>6</sub> including an Arg <sup>127</sup> Gly point mutant |
| pRS551                             | Promoterless <i>lacZ</i> containing vector                             |
| pRS551 <sub>A</sub>                | <i>lacZ</i> fused to the first 130 bases of <i>flhD</i> promoter       |
| pRS551 <sub>B</sub>                | <i>lacZ</i> fused to the first 215 bases of <i>flhD</i> promoter       |
| pRS551 <sub>C</sub>                | <i>lacZ</i> fused to the first 360 bases of <i>flhD</i> promoter       |
| pRS551 <sub>D</sub>                | <i>lacZ</i> fused to the first 501 bases of <i>flhD</i> promoter       |
| pRS551 <sub>E</sub>                | <i>lacZ</i> fused to the first 597 bases of <i>flhD</i> promoter       |
| pRS551 <sub>F</sub>                | <i>lacZ</i> fused to the first 718 bases of <i>flhD</i> promoter       |
| pLX3607 or pVector                 | Empty IPTG-inducible vector  |
| pDRM001 or pPxWT                   | Wildtype <i>papX</i> made in pLX3607                                   |
| pDRM002 or pR127G                  | <i>papX</i> _Arg <sup>127</sup> Gly constructed in pDRM001             |
| pDRM003 or pK54N                   | <i>papX</i> _Lys <sup>54</sup> Asn constructed in pDRM001              |
| pDRM004 or pR127K                  | <i>papX</i> _Arg <sup>127</sup> Lys constructed in pDRM001             |
| pDRM005 or pT21A                   | <i>papX</i> _Thr <sup>21</sup> Ala constructed in pDRM001              |
| pDRM006 or pR51A                   | <i>papX</i> _Arg <sup>51</sup> Ala constructed in pDRM001              |
| pDRM008 or pR55G                   | <i>papX</i> _Arg <sup>55</sup> Gly constructed in pDRM001              |
| pDRM009 or pH71A                   | <i>papX</i> _His <sup>71</sup> Ala constructed in pDRM001              |
| pDRM010 or pR101A                  | <i>papX</i> _Arg <sup>101</sup> Ala constructed in pDRM001             |
| pDRM011 or pR106A                  | <i>papX</i> _Arg <sup>106</sup> Ala constructed in pDRM001             |
| pDRM012 or pK113N                  | <i>papX</i> _Lys <sup>113</sup> Asn constructed in pDRM001             |
| pDRM013 or pK118A                  | <i>papX</i> _Lys <sup>118</sup> Ala constructed in pDRM001             |
| pDRM014 or pR119A                  | <i>papX</i> _Arg <sup>119</sup> Ala constructed in pDRM001             |
| pDRM015 or pR123A                  | <i>papX</i> _Arg <sup>123</sup> Ala constructed in pDRM001             |
| pDRM016 or pR126A                  | <i>papX</i> _Arg <sup>126</sup> Ala constructed in pDRM001             |
| pDRM017 or pK129A                  | <i>papX</i> _Lys <sup>129</sup> Ala constructed in pDRM001             |
| pDRM018 or pK130A                  | <i>papX</i> _Lys <sup>130</sup> Ala constructed in pDRM001             |
| pKD3                               | λ red encoding chloramphenicol resistance cassette                     |
| pKD4                               | λ red encoding kanamycin resistance cassette                           |
| pCP20                              | λ red encoding recombinase to remove cassettes                         |
| pKD46                              | λ red encoding recombinase to insert cassettes                         |
| CFT073                             | Prototypical strain of pyelonephritogenic UPEC                         |
| CFT073 Δ <i>papX</i>               | Deletion of <i>papX</i> in strain CFT073                               |
| CFT073 Δ <i>papX</i> Δ <i>lacZ</i> | Deletion of <i>papX</i> and <i>lacZ</i> in strain CFT073               |
| CFT073 Δ <i>lacZ</i>               | Deletion of <i>lacZ</i> in strain CFT073                               |
| K12 MG1655                         | Fecal/commensal strain K12 MG1655                                      |

**Supplementary Table 2. Primers used in this study.**

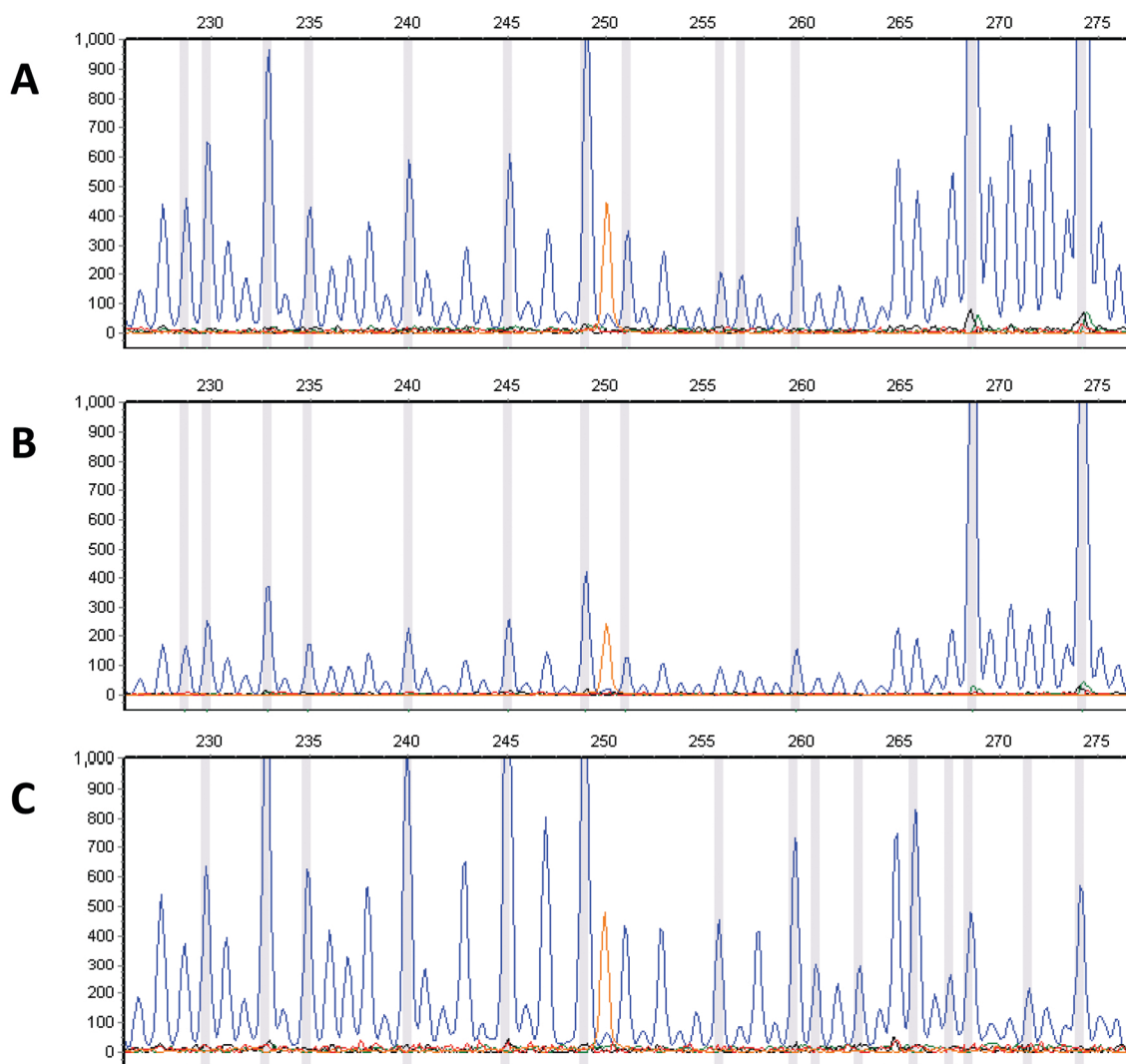
| Primer                 | Sequence (5'→3')   | Function             |
|------------------------|--|----------------------|
| <i>papX</i> H1P1       | ...TGCCTGAAGCTATCCGGCATACTCAGGCATTTCCAGCCTTTGTGTAGGCTGGAGCTGCTTC | λ red mutants        |
| <i>papX</i> H2P2       | ..AGAAATCAGGCAGGAATAAAATTTAGTGGAAAGTCGATGTATGGGAATTAGCCATGGTCC   | λ red mutants        |
| <i>lacZ</i> H1P1       | .....TTACGCGAAATACGGGCGAGACATAGCCTGCCCGTTATTA                    | λ red mutants        |
| <i>lacZ</i> H2P2       | .....ATGACTATGATTACGGATTCTCTGGCCGTCGTATTACAAC                    | λ red mutants        |
| <i>gapA</i> 29bp fwd   | .....ACTCCACTCAGGCGGTTTCGACGGTACCG                               | Gel shift            |
| <i>gapA</i> 29bp rev   | .....CGGTACCGTCGAAACGGCCGTGAGTGGAGT                              | Gel shift            |
| <i>papX</i> 29bp fwd   | .....TTACGGTGAGTTATTTAACTGTGCGCAAC                               | Gel shift            |
| <i>papX</i> 29bp rev   | .....GTTGCGCACAGTAAAAAATACTCACCCTAA                              | Gel shift            |
| EMSA154_F              | .....TCGCTACACTGGCAAGCAA   | Gel shift            |
| EMSA154_R              | .....GTTGTGCGGTAAGTGTTTGT  | Gel shift            |
| <i>papX</i> RTfwd      | .....CGCATATCCCTTTCCGGGAGCATT                                    | qPCR                 |
| <i>papX</i> RTrev      | .....TCCGCTGCGAATAATGTGGTGAGA                                    | qPCR                 |
| <i>gap</i> ARTfwd      | .....CGTTAAAGGCGCTAACTTCG  | qPCR                 |
| <i>gap</i> ARTrev      | .....ACGGTGGTCATCAGACCTTC  | qPCR                 |
| <i>flh</i> DRTfwd      | .....TCCGCTATGTTTGGTCTCGGCATA                                    | qPCR                 |
| <i>flh</i> DRTrev      | .....ACCAGTTGATGGTTCTCGCCAGC                                     | qPCR                 |
| <i>flh</i> CRTfwd      | .....AAACTGGCTTGTGAATGGCGCTG                                     | qPCR                 |
| <i>flh</i> CRTrev      | .....TCAACAAACCCACCAATGTCCAG                                     | qPCR                 |
| T <sup>21</sup> Afwd   | ...GTAATATGGAGAATAAATGAAATGAATAACGCAGACACATAGAAAAATAATCAGACAC    | Site-directed mutant |
| T <sup>21</sup> Arev   | ...CATTATACCTCTTATTACTTTACTTATTGGCTCTGTGTAATCTTTTTATTAGTCTGTG    | Site-directed mutant |
| R <sup>51</sup> Afwd   | .....GGAGCATTGTGTGATGCAACTCTGTATTGCCACAAATAAAGAATGCAG            | Site-directed mutant |
| R <sup>51</sup> Arev   | .....CTGCATTCTTTATTTGTGGCAATCAGAGTTGCATCAACAAATGTCC              | Site-directed mutant |
| K <sup>54</sup> Nfwd   | .....AGATATATTATCTCGCATTCTGTTATTTGTGCGAATACAGAGTTGCA             | Site-directed mutant |
| K <sup>54</sup> Nrev   | .....TGCAACTCTGTATTGCACAAATAACAGAATGCAGGATAATATATCT              | Site-directed mutant |
| R <sup>55</sup> Gfwd   | .....AGATATATTATCTCGCATTCTTTATTTGTGCGAATACAGAGTTGC               | Site-directed mutant |
| R <sup>55</sup> Grev   | .....GCAACTCTGTATTGCACAAATAAAGGAATGCAGGATAATATATCT               | Site-directed mutant |
| H <sup>71</sup> Afwd   | .....CTGAATTTCTGGGGTGTATGGAATAAATGCTCAGTATATATGGTTCT             | Site-directed mutant |
| H <sup>71</sup> Arev   | .....AGAACCATATATCTGAGGCATTTATCCATACACCCAGAAATTCAG               | Site-directed mutant |
| R <sup>104</sup> Afwd  | .....TAAGCCAGAACTTCAGTTTACCGCACTAATATTACCCGCATTACAG              | Site-directed mutant |
| R <sup>104</sup> Arev  | .....CTGTAATGCGGGTAATATTAGTTGCGGTAACCTGAAGTTCTGGCTTA             | Site-directed mutant |
| R <sup>106</sup> Afwd  | .....TCAGTTTACCAGAACTAATAATTACCAGCATTACAGATTTTGTAGAAAAAGCC       | Site-directed mutant |
| R <sup>106</sup> Arev  | .....GGCTTTTCTAAAAAATCTGTAATGGCGGTAATATTAGTTCTGGTAAACTGA         | Site-directed mutant |
| K <sup>113</sup> Nfwd  | .....TAATATTACCCGCATTACAGATTTTGTAGAAAAAGCCGGATATGTAAAAAG         | Site-directed mutant |
| K <sup>113</sup> Nrev  | .....CTTTTACATATCCGGCTTTCTAAAAAATCTGTAATGCGGGTAATATTA            | Site-directed mutant |
| K <sup>118</sup> Afwd  | .....TTACAGATTTTGTAGAAAAAGCCGGATATGTAGCAAGGATGGATAGCAGGG         | Site-directed mutant |
| K <sup>118</sup> Arev  | .....CCCTGCTATCCATCTTGTCTACATATCCGGCTTTTCTAAAAAATCTGTAA          | Site-directed mutant |
| R <sup>119</sup> Afwd  | .....AGATTTTGTAGAAAAAGCCGGATATGTAAAAAGGATGGATAGCAGGGAGG          | Site-directed mutant |
| R <sup>119</sup> Arev  | .....CCTCCCTGCTATCCATCGCTTTTACATATCCGGCTTTTCTAAAAAATCT           | Site-directed mutant |
| R <sup>123</sup> Afwd  | .....GTAAAAAGGATGGATAGCGGGGAGGATCGCCCTGTCTAA                     | Site-directed mutant |
| R <sup>123</sup> Arev  | .....TTAGCACGGCGATCTCCGGCTATCCATCTCTTTTAC                        | Site-directed mutant |
| R <sup>126</sup> Afwd  | .....AAAAAGGATGGATAGCAGGGAGGATGCCCTGTCTAAAAA                     | Site-directed mutant |
| R <sup>126</sup> Arev  | .....TTTTTGTAGCAGGGATCTCCCTGCTATCCATCTTTTT                       | Site-directed mutant |
| R <sup>127</sup> Gfwd  | .....TAGCAGGGAGGATCGCGGTGCTAAAAAATCAGT                           | Site-directed mutant |
| R <sup>127</sup> Grev  | .....ATCGTCCCTCCTAGCGCCACGATTTTGTAGTCA                           | Site-directed mutant |
| R <sup>127</sup> Kfwd  | .....GATGGATAGCAGGGAGGATCGCAAGGTAATAAATCAGTCTGCAT                | Site-directed mutant |
| R <sup>127</sup> Krev  | .....ATGTCAGACTGATTTTGTAGCCTTGCATCTCCCTGCTATCCATC                | Site-directed mutant |
| K <sup>129</sup> Afwd  | .....CAGGGAGGATCGCCGTGCTGCAAAAAATCAGTCTGCATCTGT                  | Site-directed mutant |
| K <sup>129</sup> Arev  | .....CAGATGTCACTGATTTTGTAGCACGGCGATCTCCCTG                       | Site-directed mutant |
| K <sup>130</sup> Afwd  | .....GGGAGGATCGCCGTGCTAAAAAGCAATCAGTCTGCATCTGAA                  | Site-directed mutant |
| K <sup>130</sup> Arev  | .....TTCAGATGTCACTGATTTGCTTAGCAGCGGATCTCC                        | Site-directed mutant |
| <i>papX</i> MltplxRev  | .....TCCGCTGCGAATAATGTGGTGAGA                                    | Multiplex PCR        |
| <i>X</i> MltPlexfwd    | .....GCGCGTTGTACACAGACAGTGTG                                     | Multiplex PCR        |
| <i>focX</i> MltPlexfwd | .....CAGGCGTGTGGAAGCTGATG  | Multiplex PCR        |
| <i>papX</i> MltPlexfwd | .....CCTGACCATCGGCAGTCGCCTC                                      | Multiplex PCR        |
| DNase <i>flh</i> Drev  | .....TCAGCAACTCGGAGGTATGC  | DNase protect assay  |
| DNase <i>flh</i> DAfwd | .....ACCAAAAAGGTGGTCTGCT   | DNase protect assay  |
| DNase <i>flh</i> DBfwd | .....AAAATCGCAGCCCCCTCCG   | DNase protect assay  |
| DNase <i>flh</i> DCfwd | .....TTTGCTTGCAGTGTAGCGA   | DNase protect assay  |
| DNase <i>flh</i> DDfwd | .....GTTGTGCGGTAAGTGTGTTGT                                       | DNase protect assay  |
| DNase <i>gap</i> Afwd  | .....ACGTGACTGATTCTA   | DNase protect assay  |
| DNase <i>gap</i> Arev  | .....CAGTATTTGTTAGTG   | DNase protect assay  |
| <i>flh</i> DEHrev      | .....TCAGCAACTCGGAGGTATGC  | <i>lacZ</i> -fusions |
| <i>flh</i> DAfwd       | .....ACCAAAAAGGTGGTCTGCT   | <i>lacZ</i> -fusions |
| <i>flh</i> DBfwd       | .....AAAATCGCAGCCCCCTCCG   | <i>lacZ</i> -fusions |
| <i>flh</i> DCfwd       | .....TTTTGCTTGCAGTGTAGCGA  | <i>lacZ</i> -fusions |
| <i>flh</i> DDfwd       | .....GTTGTGCGGTAAGTGTGTTGT                                       | <i>lacZ</i> -fusions |
| <i>flh</i> DEFwd       | .....ATGTAAGTATTTCCATATT   | <i>lacZ</i> -fusions |
| <i>flh</i> DFfwd       | .....TGGAGAACGACCAATCCC  | <i>lacZ</i> -fusions |

Growth kinetics of CFT073  $\Delta papX$  linker-insertional transformants





Supplementary  
Figure 2



**Supplementary Figure 3**