

Supplementary Table

Table S1. Comparison of the *TmLamCD* models in different space groups. RMSD (Å) values on the upper-right were calculated by PDBeFold.

	1	2	3	4	5	6	7	8	9	10	11
1		0.43	0.60	0.35	0.42	0.46	0.48	0.63	0.43	0.39	0.36
2			0.74	0.44	0.24	0.39	0.58	0.77	0.29	0.21	0.45
3				0.68	0.73	0.76	0.75	0.35	0.74	0.72	0.71
4					0.41	0.42	0.36	0.65	0.38	0.41	0.23
5						0.38	0.55	0.76	0.27	0.19	0.41
6							0.45	0.77	0.30	0.37	0.40
7								0.68	0.47	0.54	0.37
8									0.75	0.75	0.66
9										0.27	0.35
10											0.41
11											

1, chain A, open form, in $P2_12_12_1$; 2, chain A, closed form, in $P2_12_12_1$; 3, chain B in $P2_12_12_1$; 4, chain C in $P2_12_12_1$; 5, chain D in $P2_12_12_1$; 6, chain A in $C2$; 7, chain B in $C2$; 8, chain A in $P4_3$; 9, chain B in $P4_3$; 10, chain C in $P4_3$; 11, chain D in $P4_3$

Supplementary Figure Legends

Fig. S1. The topology of the (A) *TmLamCD* and (B) *TmCel12A*. The secondary structure is generated by the ESPript software. The η denotes 3_{10} helix.

Fig. S2. Stereoview of a double occupancy of the flexible GASIG loop was observed in some *TmLamCD* chains. (A) The difference Fourier map ($mF_o - DF_c$) of the GASIG loop is shown in gray mesh contoured at 1.5σ . (B) The flexible GASIG fragment in the *TmLamCD* coordinate. The closed configuration is shown in hot pink, and the open form is in cyan.

Fig. S3. Stereoview of superimposed C α traces of catalytic domains of laminarinase structures. Chain A was selected for each structure to compare the diversity. 2HYK is shown in blue, 2VY0 in green, 3ILN in orange, 3DGT in cyan, *TmLamCD* with the open form loop in magenta and closed form loop in red. Variable regions of the secondary structures are labeled according to the annotation in Fig.1B.

Fig. S4. Ligplot of gluconolactone complexed in laminarinase showing H-bonds and hydrophobic interactions around the active site. Lgc represents gluconolactone, and 16A means cetyltrimethylammonium (ctab) molecule. A, B, C in the parenthesis means the chain code in the asymmetric unit. (D) The ligplot shows the amino acid residues around the inhibitor with hydrophobic contacts.

Fig. S5. Electrostatic potential surface diagram of *TmLam* catalytic domain shows the charge character of the catalytic groove. (A) A modeled long chain β -1,3-glucan moves outwards the active site of *TmLamCD*. (B) Modeled laminarihexaose penetrates the entranceway of the *TmLamCD*. Negative potentials are colored in red, positive in blue, and neutral in white. Ligands are shown in green sticks.

Fig. S6. Calcium coordinates. (A) Stereo image of the surrounding Lam residues coordinating with the calcium ion (chocolate). Calcium coordinates are shown in green. (B) Ligplot to show each distance of coordinates in green.

Fig. S7. Structural comparison of the catalytic grooves among glucanases or the carbohydrate binding module. (A) A laminarihexaose molecule was bound on the carbohydrate-binding module of *T. maritima* laminarinase (PDB: 1GUI) (27). (B) A cellotetraose locates in the cellulase12A from *T. maritima* (PDB of *TmCel12A*: 3AMM) (35). (C) An exo- β -1,3-glucanase from *Candida albicans* in complex with laminaritriose (PDB: 3N9K) (D) A *p*-nitrophenyl- β -D-glucopyranoside molecule located in the β -glucosidase E193D mutant protein from *Neotermes koshunensis* (PDB of *NkBgl-E193D-pNPG*: 3AI0) (37). Figures on the left represent the surface view, and the detailed interactive residues are shown on the right ones. All ligands are shown in thick stick. Protein residues interacting with ligands by electrostatic forces are shown in yellow sticks, whereas hydrophobic interacting residues are in magenta. Potential interactive residues are in cyan. Hydrogen bonds are indicated by dashed lines.

Fig. S8. Comparison of *TmLamCD* and *TmCel12A* structures. (A) Overlay of chain A of both *TmLamCD* (in cyan) and *TmCel12A* structures (in gray) by superposing residues 123-140 (in yellow) of *TmLamCD* with residues 126-143 of *TmCel12A*. (B) and (C) represent the surface view of the substrate-binding cleft of *TmLamCD* and *TmCel12A*, respectively. The gluconolactone, laminaritriose, and cellotetraose molecules are shown in cyan, blue and purple sticks.

Fig. S9. Thin-layer chromatography of the digested products of laminarin by *TmLamCD*. Glucose (G1, lane1), maltose (G2, lane 2), laminaritriose (G3, lane 3) and laminaritetraose (G4, lane 4) were used as standards. Five μ g *TmLamCD* protein were used to digest 2.4% laminarin in 25 mM sodium phosphate buffer (pH 7.0) at 45°C for 30 sec (lane 5), 1 min (lane 6), 5 min (lane 7) and 18 hours (lane 8). Lane 9 and lane 10 represent controls as 5% laminarin without *TmLamCD* and 5 mg

TmLamCD without laminarin, respectively, in 25 mM sodium phosphate buffer (pH 7.0) at 45°C for 18 hours. Solutions of lanes 1-4, 8, and 9 were frozen before TLC analysis.

Fig. S10. A modeled long chain laminarin at the reducing end of carbohydrate in the *TmLamCD* catalytic cavity. Subsites of glucosyl residues in the active site are numbered in black. Amino acid residues around the enclosure at the catalytic cavity of *TmLamCD* are labeled in gray.

Fig. S11. Superimposed C α traces of catalytic domains of laminarinase structures. 2HYK is shown in blue, 2VY0 in green, 3ILN in orange, open form of *TmLamCD* in magenta, and the closed form GASIG loop of *TmLamCD* in red. Side chains of Trp residues corresponding to Trp232 in *TmLamCD* are shown in sticks and numbered according to their deposited PDB. (A) Stereoview of the superimposed structures shown in cartoon loop. (B) Surface view of each structure showing their catalytic cavity.

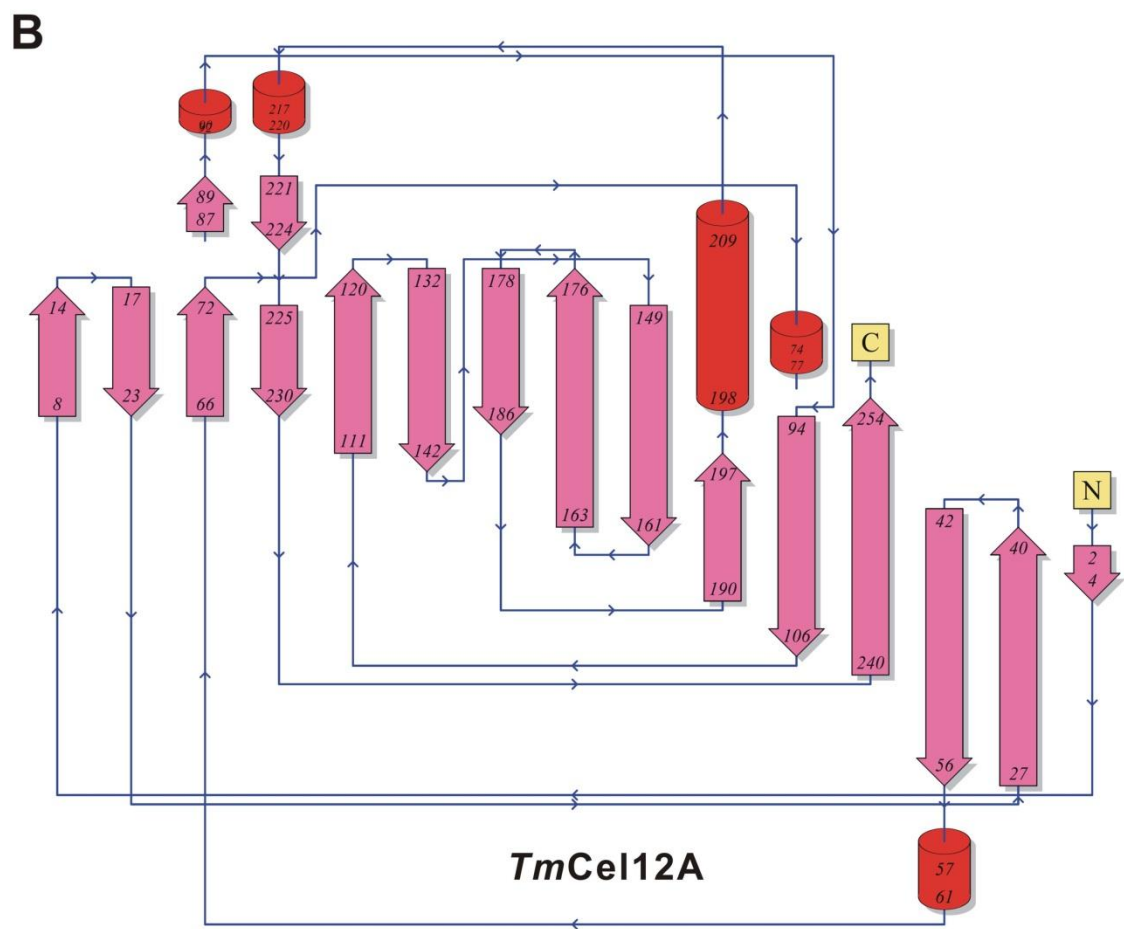
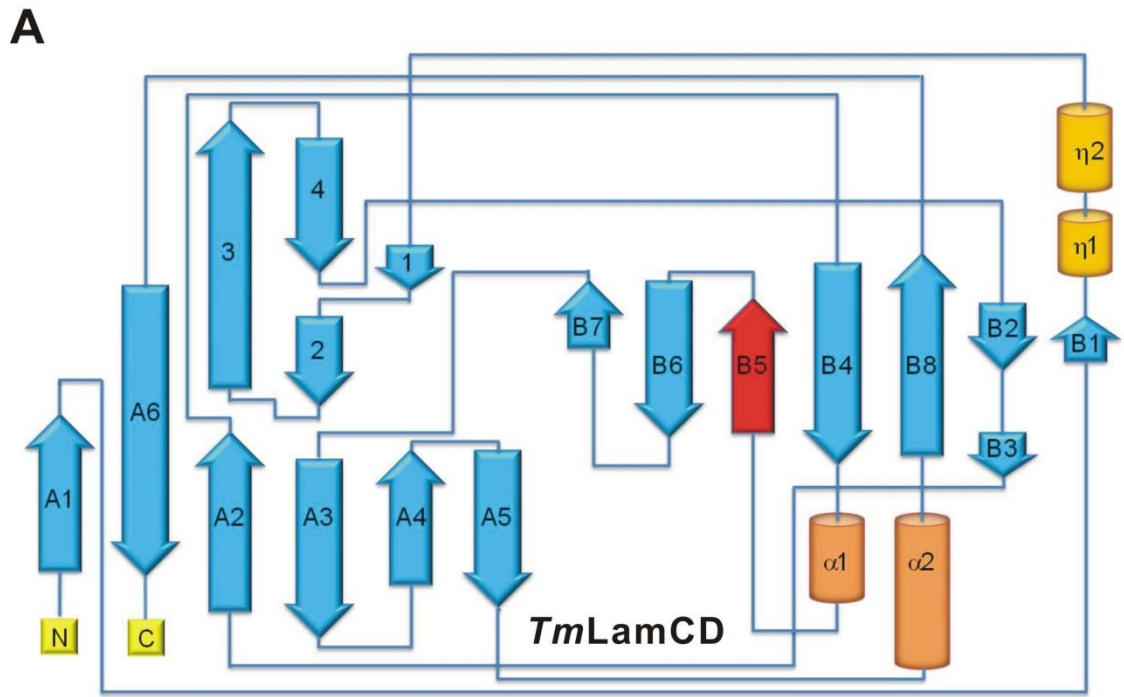


Figure S1

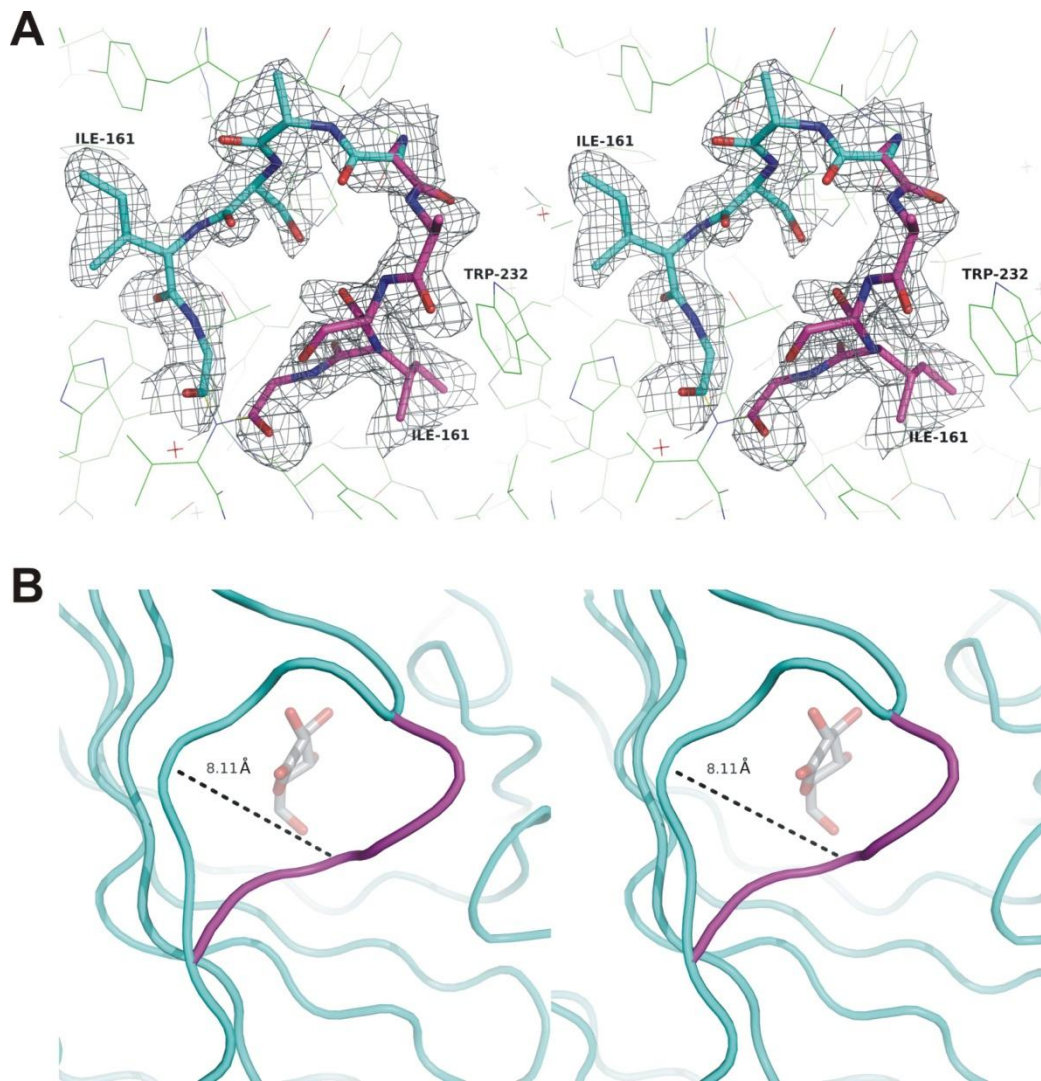


Figure S2

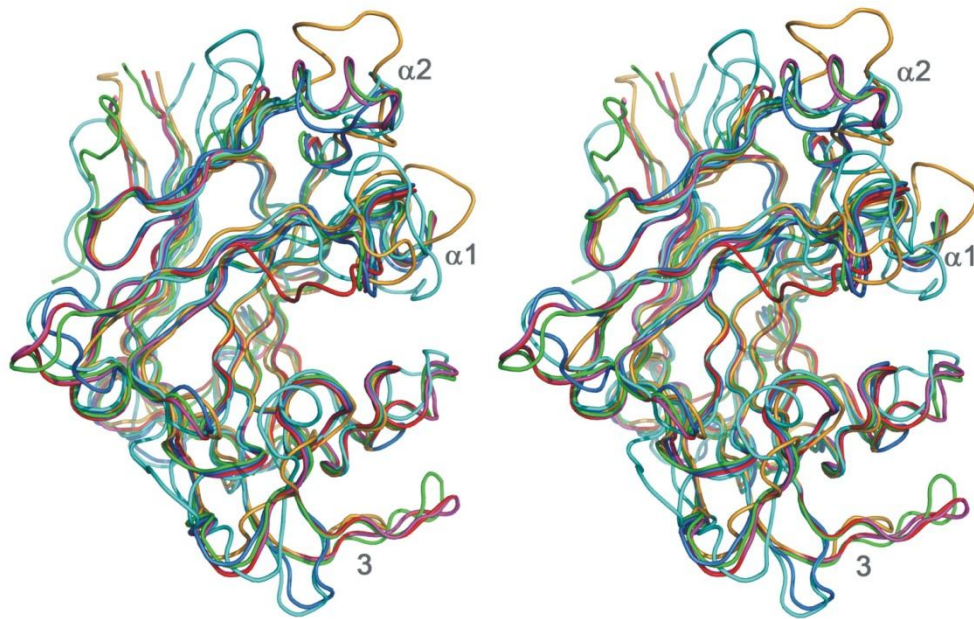


Figure S3

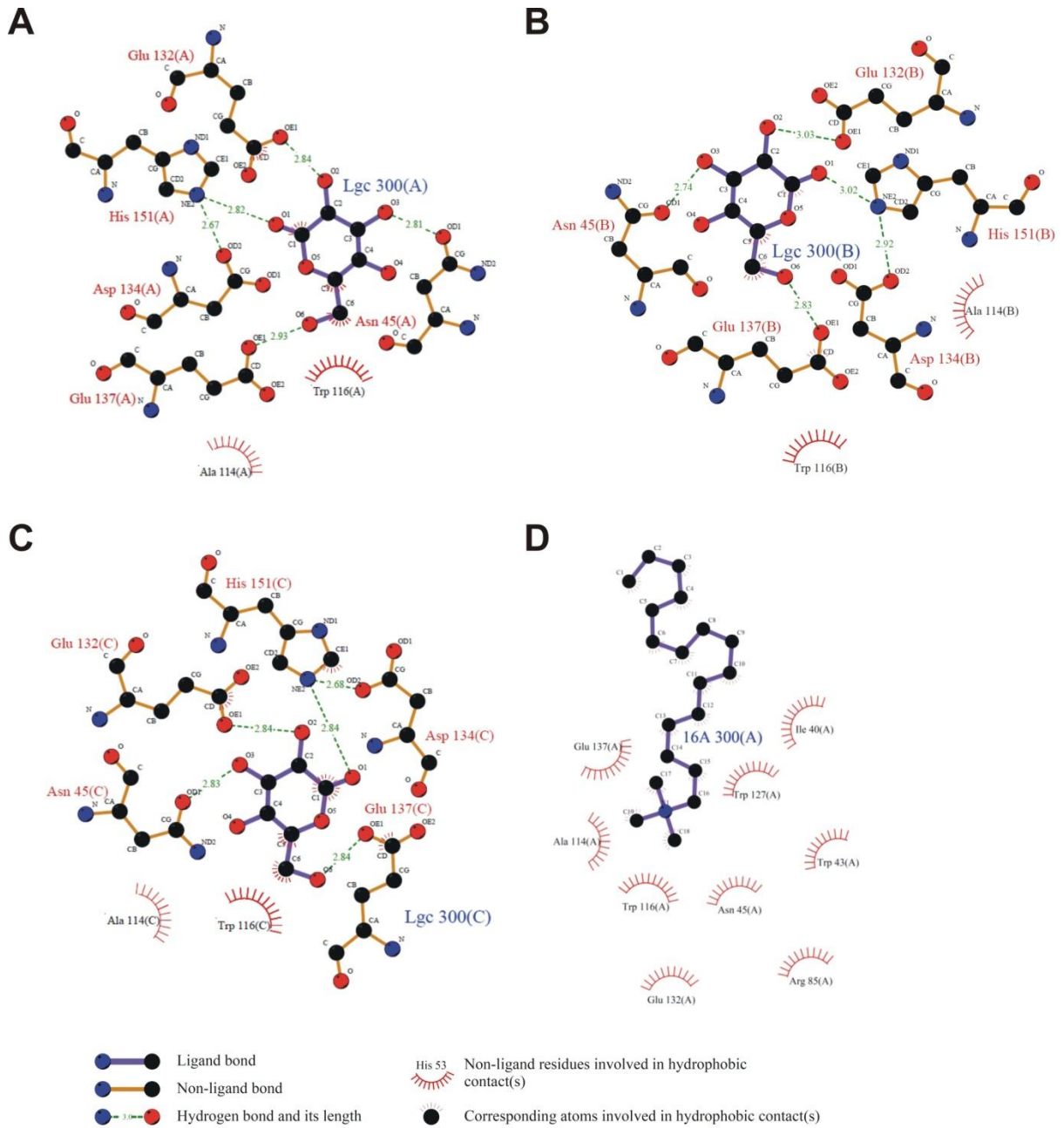
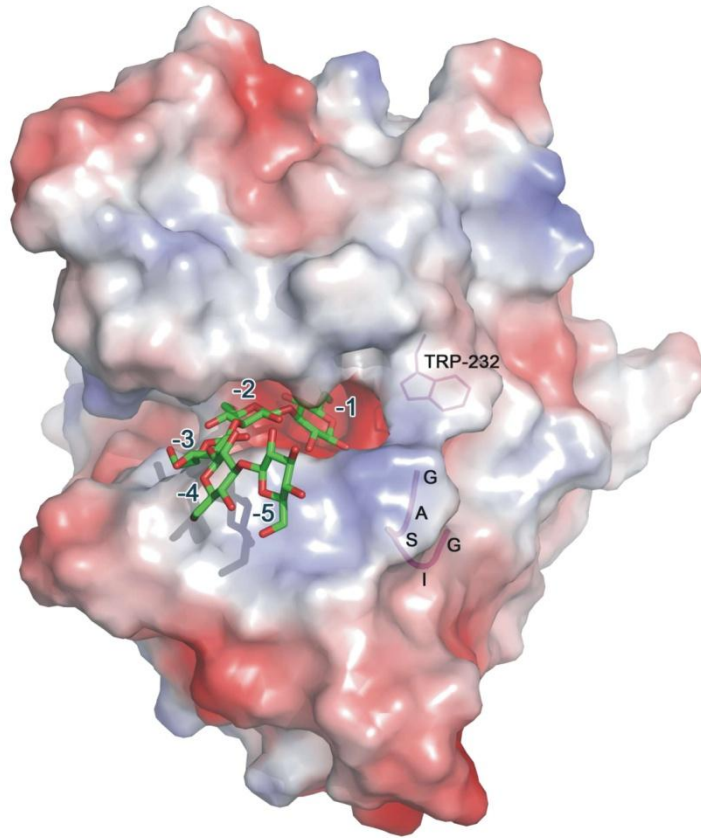


Figure S4

A



B

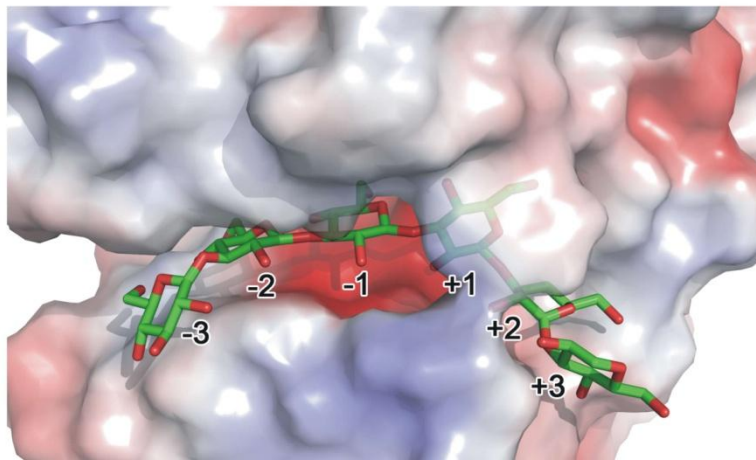


Figure S5

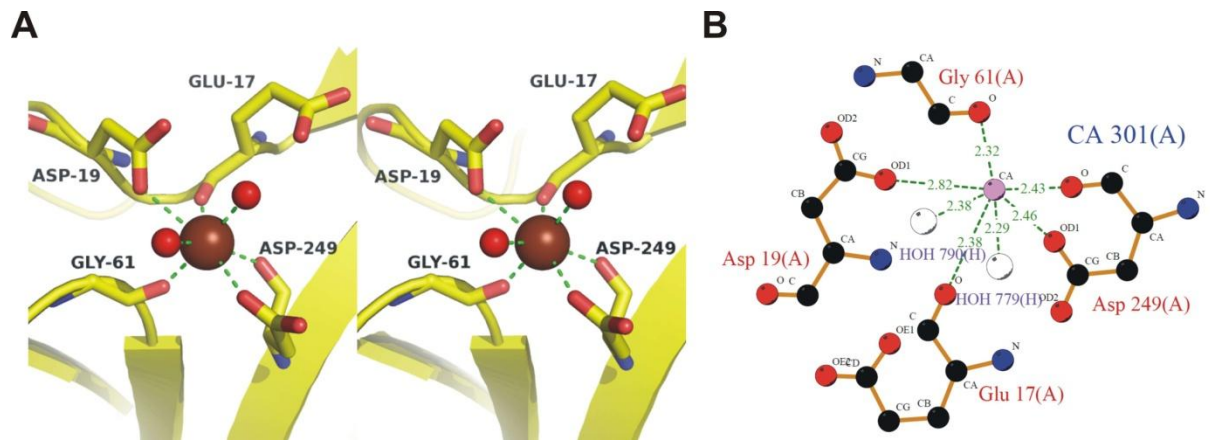


Figure S6

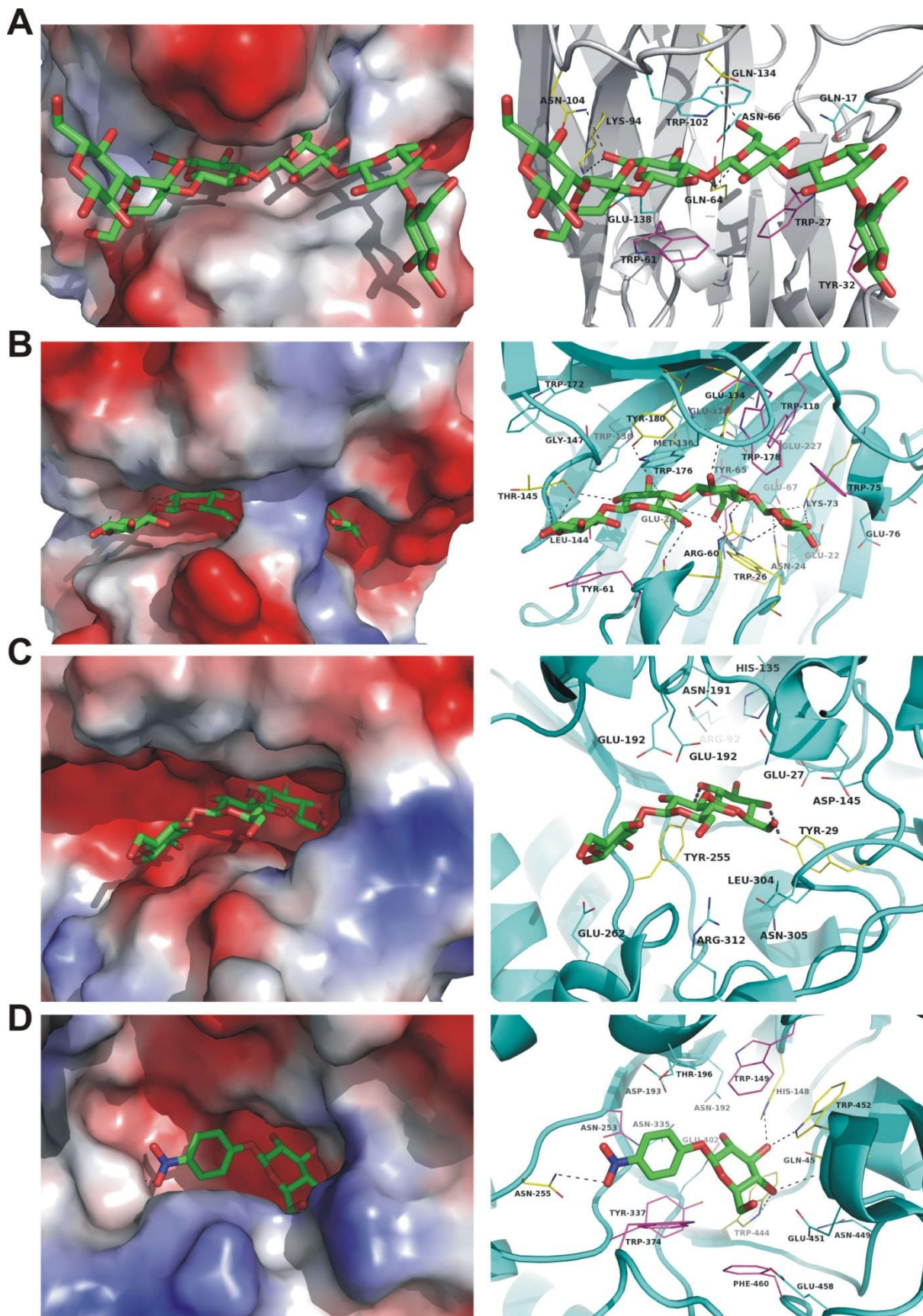


Figure S7

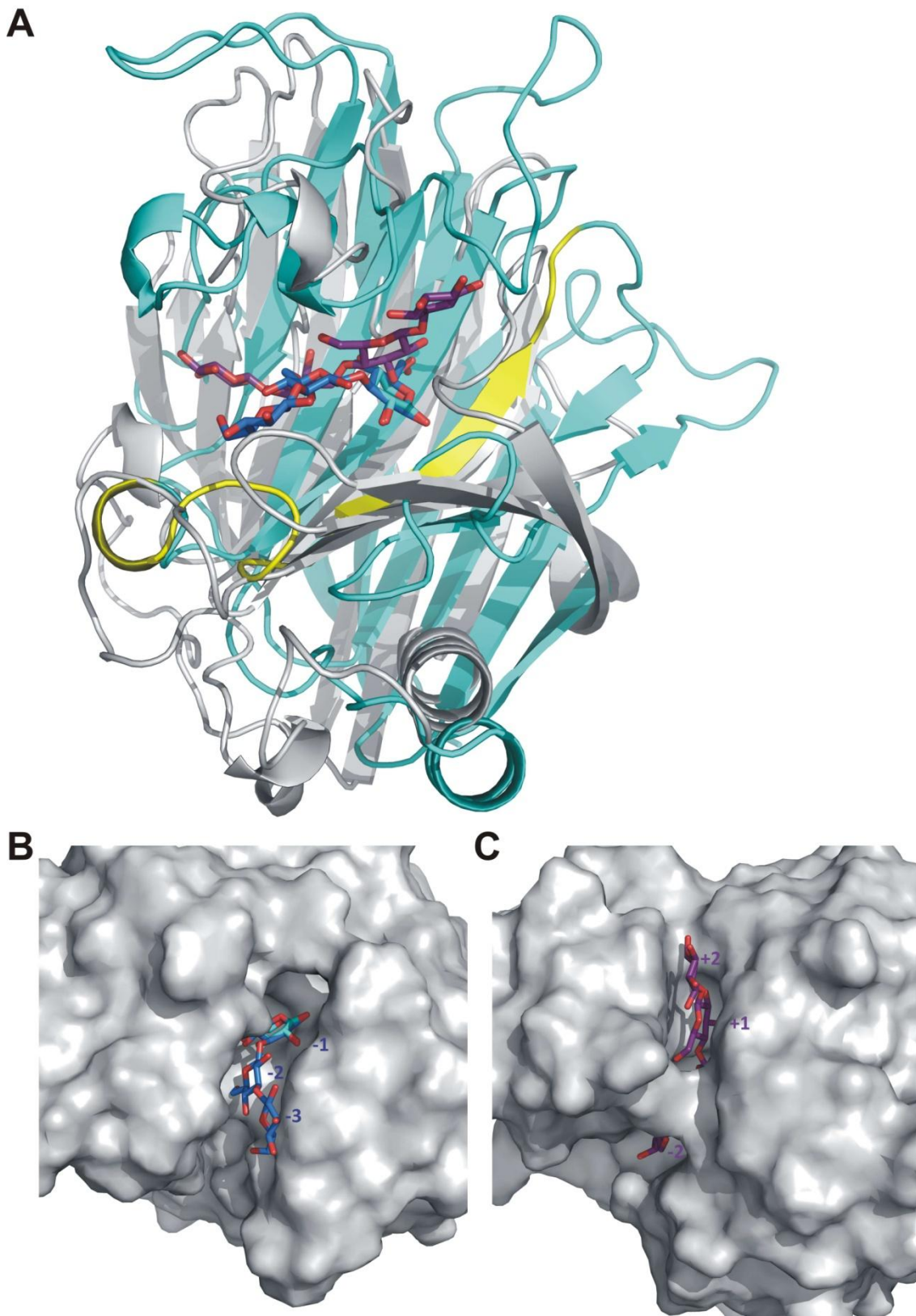


Figure S8

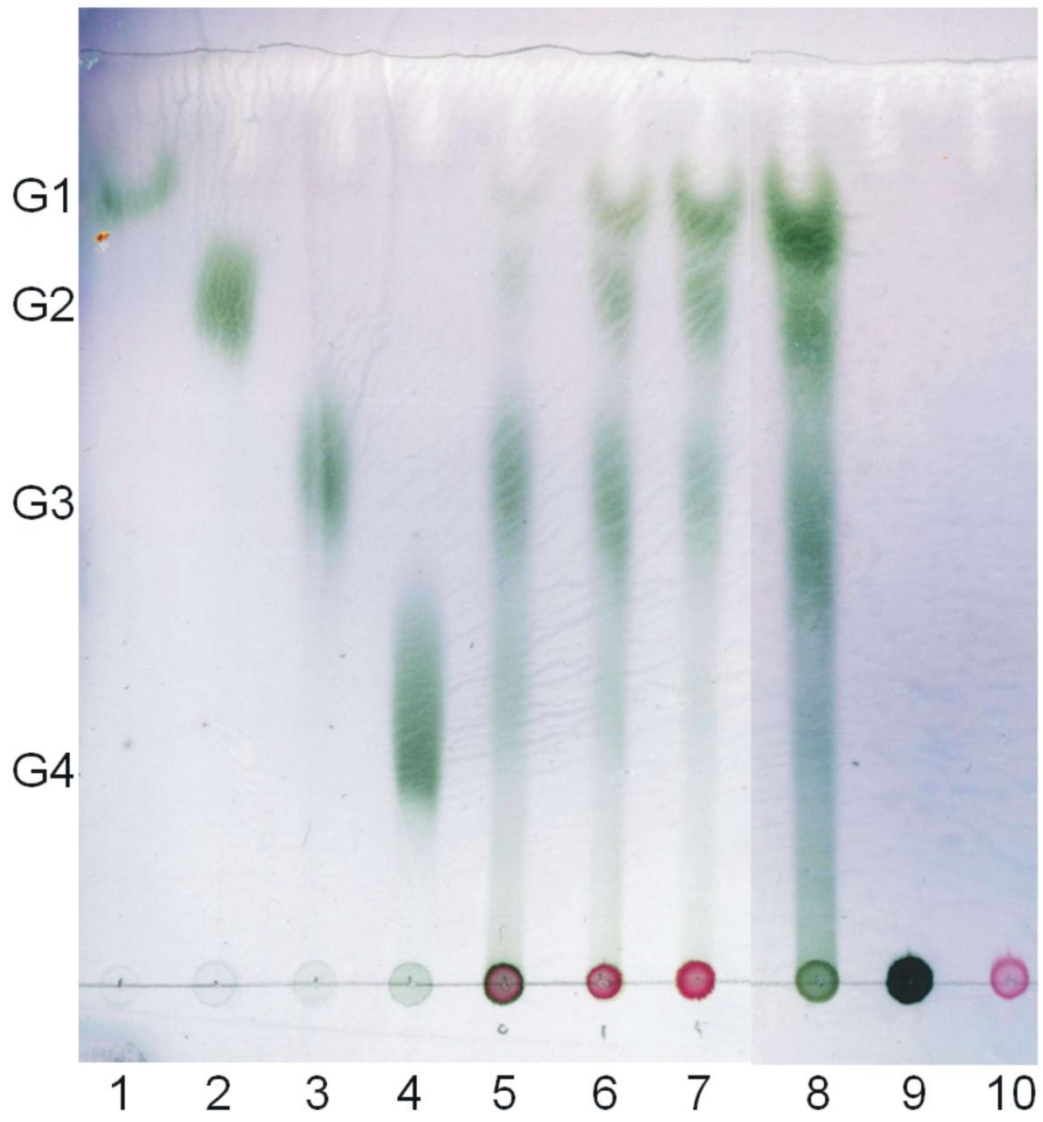


Figure S9

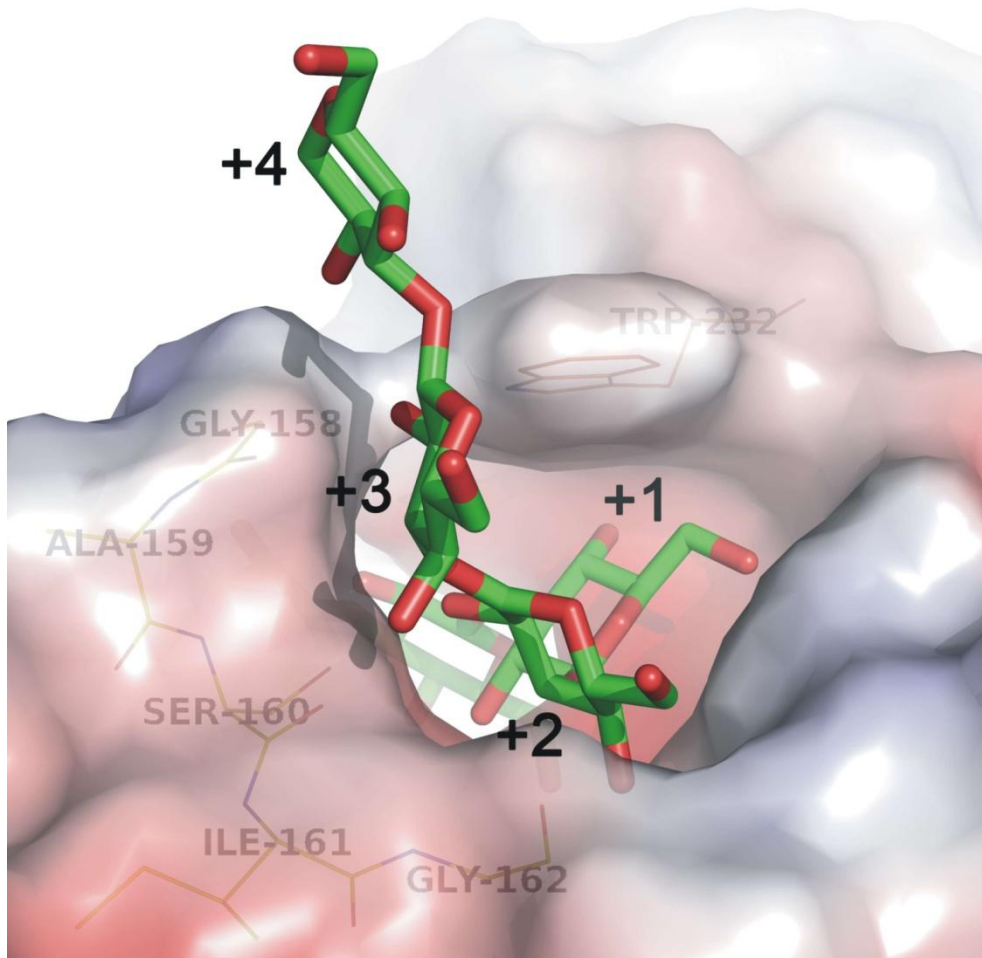


Figure S10

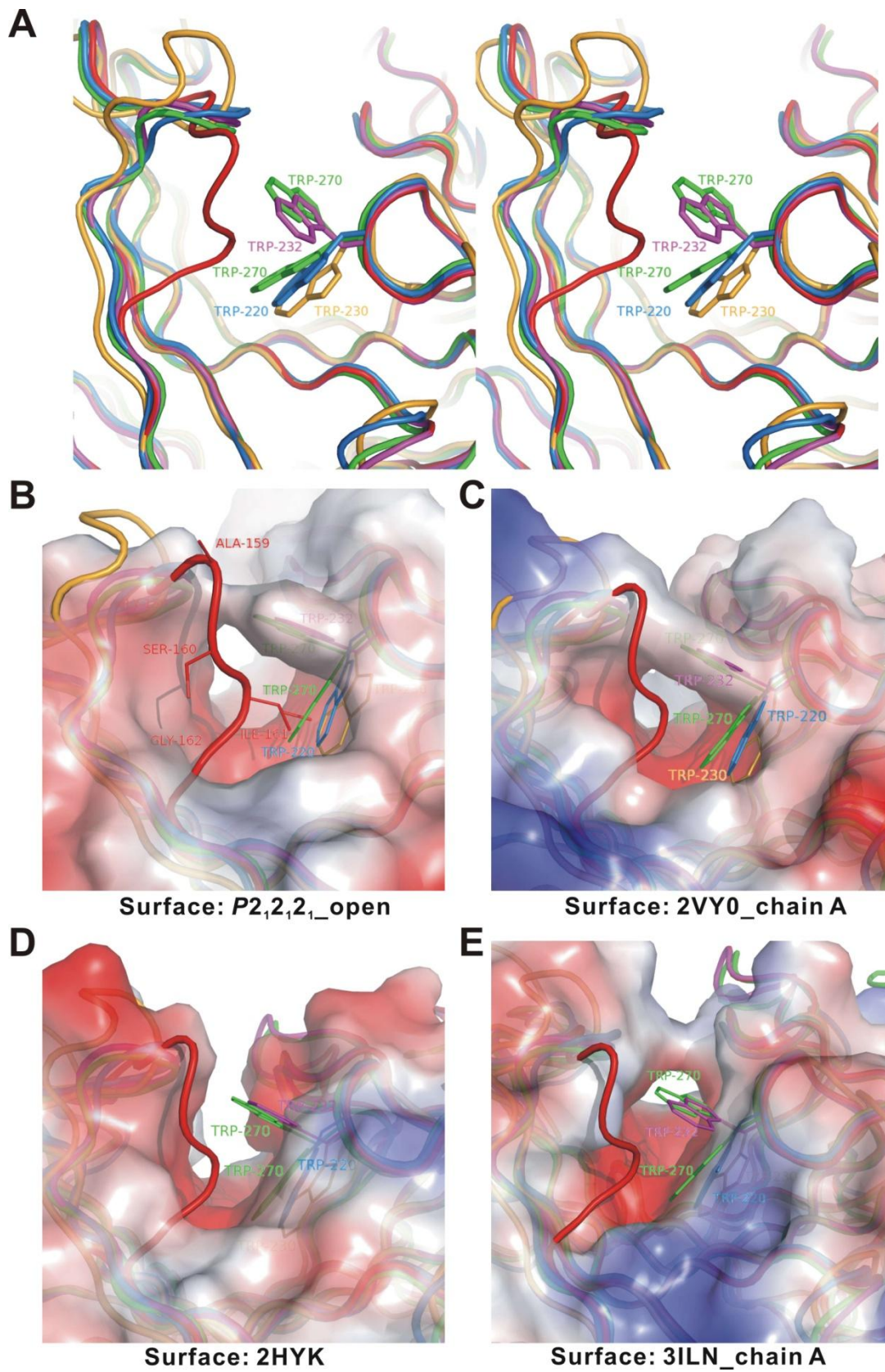


Figure S11