

Supporting Information

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SI Methods

Mice. To generate the *Let-7* transgenic mice, a genomic region on chromosome 13 comprising the genes for *Let-7a*, *Let-7d*, and *Let-7f* was amplified from C57BL/6 DNA using the following primers: forward, 5'-TTTGTCTCATGAACATCTTTTCC-3'; reverse, 5'-CCTGCTTCCCTCATTTGGTA-3'. The fragment was then subcloned into the pDONR221 gateway vector (Invitrogen) to create pENTR221-*Let-7a,d,f*. Concomitantly, attB sites for Gateway cloning were introduced into the Sall site of the CAG-Z-EGFP 1.0 vector (Invitrogen). Then the *Let-7a,d,f* DNA fragment from pENTR221-*Let-7a,d,f* was transferred by LR-Clonase (Invitrogen) recombination reaction into the pDEST-CAG-Z-EGFP vector to create pEXP-CAG-Z-EGFP-*Let-7a,d,f*. pEXP-CAG-Z-EGFP-*Let-7a,d,f* was linearized with Spe I and Swa I and used to generate transgenic mice by pronuclear injection. Founder mice were screened by lacZ staining of toe tips and PCR for the transgene construct. Transgene-positive offspring, *Let-7* Tg OFF mice, were then crossed to appropriate *Cre* lines to activate the transgene construct globally or in specific tissues. C57BL/6 mice, *CAG-Cre*, *MCK-Cre*, *Albumin-Cre*, *Nestin-Cre*, and *Fabp4-Cre* transgenic mice were purchased from Jackson Laboratory. Mice were housed in a SPF facility. The high-fat diet (HFD), containing 60 kcal% fat, was obtained from Research Diets (D12492).

Northern Blot Analysis. RNA was isolated using TRIzol (Invitrogen). For Northern blot analysis, 5 μ g of total RNA was denatured for 5 min at 70 °C in a buffer containing 50% formamide

and 10 mM EDTA. After electrophoresis on a 17% polyacrylamide gel, RNA was transferred onto a Hybond N membrane (Amersham) in 0.5 \times Tris/Borate/EDTA buffer at 80 V for 1 h. Hybridization was then done at 39 °C with ³²P-labeled Star-Fire oligonucleotide probes (Integrated DNA Technologies) directed against individual *Let-7* family members.

Histological and Immunohistological Examinations. For immunohistology, tissues were fixed with 4% paraformaldehyde (PFA) in PBS, embedded in paraffin wax, cut into 5- μ m-thick sections, and then deparaffinized. The primary antibodies were anti-insulin (Dako) and anti-glucagon (Millipore) in 1:300 dilutions. Nuclei were counterstained with DAPI. For neutral lipid staining, liver sections were fixed overnight in 4% PFA, cryoprotected with increasing sucrose concentrations (10–18%), cryoembedded, stained with Oil Red O, and counterstained with hematoxylin.

Glucose and Insulin Measurements. Blood glucose was measured with an Accu-Check Compact Plus Blood Glucose Meter (Roche) using whole blood. For glucose tolerance tests (GTTs), mice were fasted for 7 h before receiving i.p. glucose injection (1.5 mg/g body weight for transgenic mice and 2.0 mg/g lean mass for antimicro-treated mice). Blood glucose levels were measured before injection and at 15, 30, 60, and 90 min after glucose stimulation. For insulin tolerance tests (ITTs), insulin (1.0 U/kg body weight) was injected i.p. in fed mice. Blood glucose levels were measured before injection and at 15, 30, 45, 60, and 90 min after insulin injection.

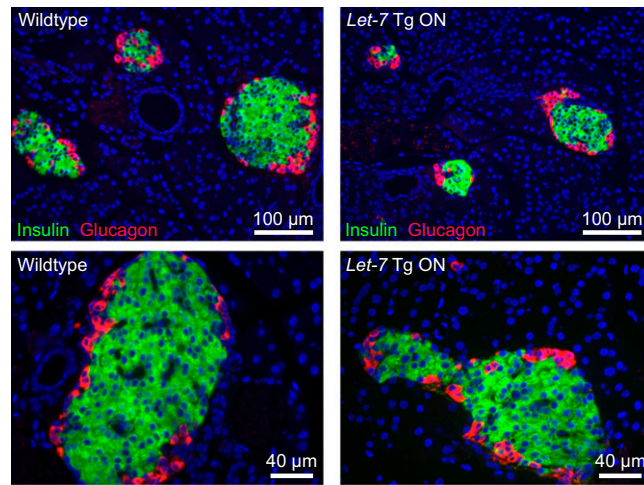


Fig. S4. *Let-7* overexpression does not change pancreatic islet structure. PFA-fixed sections of pancreata from 6-wk-old WT and *Let-7* Tg ON mice were deparaffinized and then used for immunofluorescence staining for insulin (green) and glucagon (red). Nuclei were counterstained with DAPI (blue).

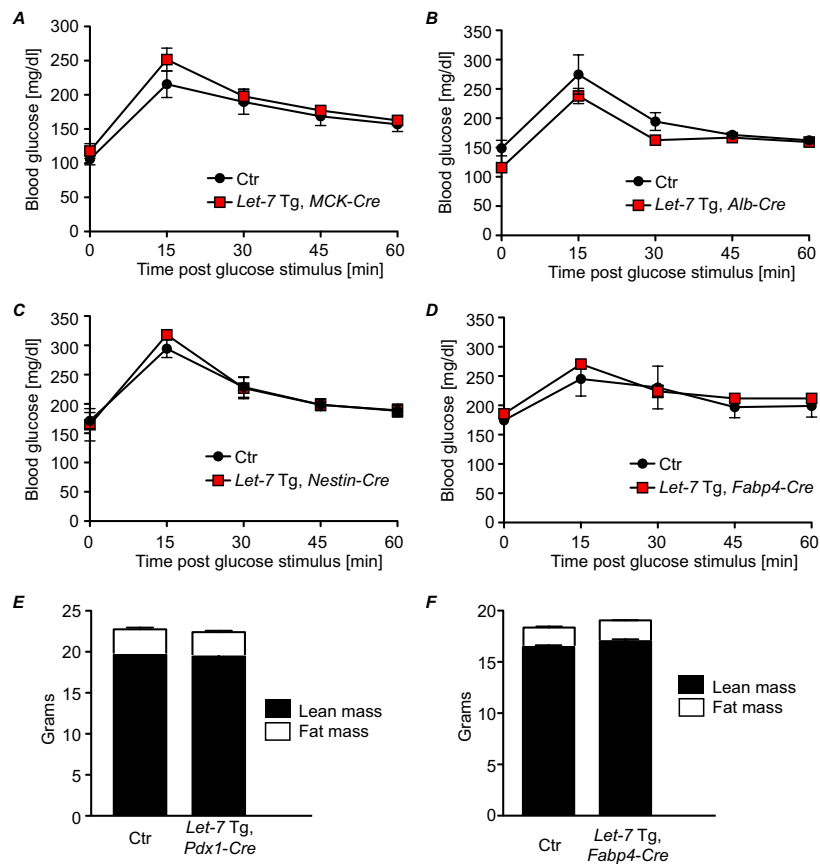


Fig. S5. Overexpression of *Let-7* in CNS, fat, muscle, or liver does not result in impaired glucose tolerance under baseline conditions. (A–F) To enrich *Let-7* expression in specific tissues, *Let-7* Tg OFF mice were crossed with transgenic mice expressing *MCK-Cre* (muscle), *Albumin-Cre* (liver), *Nestin-Cre* (CNS), and *Fabp4-Cre* (fat). To determine glucose tolerance, 6-wk-old double-transgenic mice were fasted for 7 h, after which glucose levels were measured before and at 15, 30, 45, and 60 min after glucose stimulation (1.5 mg/g body weight i.p.). WT, *Let-7* Tg OFF, or *Cre*⁺ single transgenic littermates served as controls (Ctr). $n = 3–5$ (E and F) Body composition was assessed at age 6 wk in control (Ctr) vs. *Let-7*, *Pdx1-Cre* and in Ctr vs. *Let-7*, *Fabp4-Cre* double-transgenic mice using an EchoMRI-100 body composition analyzer (Echo Medical Systems).

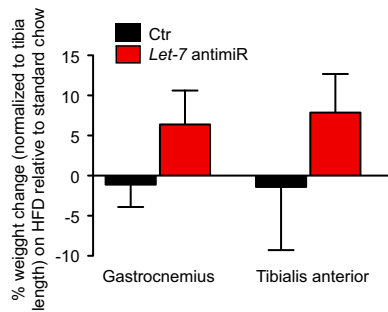


Fig. S6. *Let-7* anti-miR treatment enhances muscle growth. Male C57BL/6 mice were fed for 10 wk with HFD and either treated once a week with *Let-7* anti-miR (20 mg/kg body weight) or saline control. At the end of the study, gastrocnemius and tibialis anterior muscles were harvested and muscle weights were compared with mice on a standard diet. $n = 8$.

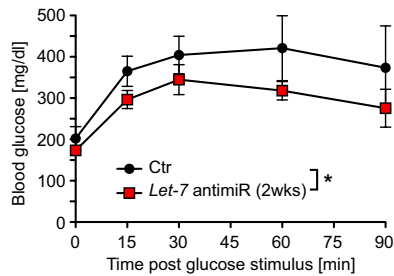


Fig. S7. The 2-wk treatment with *Let-7* anti-miR is sufficient to improve impaired glucose tolerance. A GTT was performed at 2 wk after onset of *Let-7* anti-miR administration in the treatment study. $*P < 0.05$ for Ctr vs. *Let-7* anti-miR-treated mice by ANOVA.