

## Supplementary Discussion

Extrasynaptic GABA<sub>A</sub> receptors are morphologically distinct from synaptic GABA<sub>A</sub> receptors as, in the main, they contain the  $\delta$  subunit. This conveys both an increased sensitivity to low concentrations of GABA and reduced desensitization properties compared with the  $\gamma 2$  subunit containing GABA<sub>A</sub> receptors located post-synaptically (Belelli *et al.* 2009). This morphological distinction is important as it confers on the extrasynaptic GABA<sub>A</sub> receptors compared with the synaptic GABA<sub>A</sub> receptors a relative insensitivity to benzodiazepines (in line with the lack of effect of lorazepam on 1ms SICI (Ziemann *et al.* 1996)) but an increased sensitivity to neurosteroids (in line with the cyclical changes in GABA observed by MRS ((Harada *et al.* 2010; Epperson *et al.* 2005)). The presence of an extrasynaptic GABAergic “tone” in humans has been confirmed in both pyramidal cells and interneurons within temporal neocortical brain slice preparations from epilepsy patients (Scimemi *et al.* 2006).

Our data do not provide a clear explanation as to *how* increased activation of the extrasynaptic GABA<sub>A</sub> receptors confers greater inhibition measured by 1ms SICI but we can speculate on possible mechanisms. Tonic GABA currents reduce spontaneous firing rates in interneurons (Semyanov *et al.* 2003), via an increase in chloride ion (Cl<sup>-</sup>) conductance and therefore, as the equilibrium potential of Cl<sup>-</sup> is lower than the resting membrane potential, hyperpolarize the membrane. However, hyperpolarization of the membrane leads to a *decrease* in the duration of the relative refractory period, (Kiernan & Bostock, 2000), meaning cells should be more responsive to the test stimulus, demonstrated as *reduced*

inhibition. The GABA-induced hyperpolarization of the membrane therefore cannot directly explain the relationship described here, where higher GABA levels are associated with greater inhibition at 1ms ISI, presumably reflecting a longer relative refractory period. However, the relationship may be explained by a difference in intracellular chloride ion concentration ( $[Cl^-]_i$ ) between the cell body and the axons. TMS primarily acts on axons (Di Lazzaro *et al.* 2004).  $[Cl^-]_i$  is higher in neuronal axons than in the neuronal cell bodies, and critically, is found at a concentration high enough to generate a potential greater than the reversal potential of  $Cl^-$  (Khirug *et al.* 2008; Stell, 2011). This means that, at the axon, an influx of  $Cl^-$  will cause depolarization rather than hyperpolarization of the cell (Khirug *et al.* 2008; Rojas *et al.* 2011), which will lengthen the relative refractory period (Kiernan & Bostock, 2000), thus explaining the relationship demonstrated in our data.

## Figure Legends

### Supplementary Figure 1

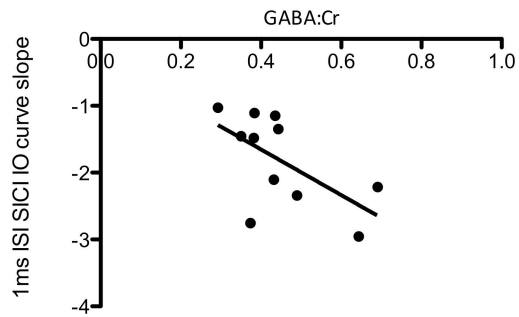
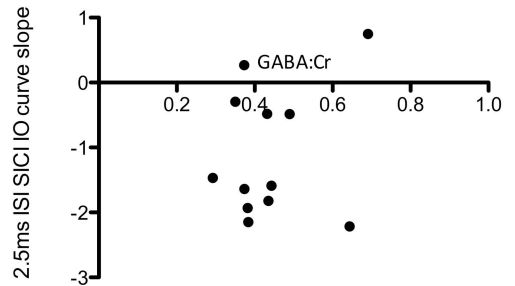
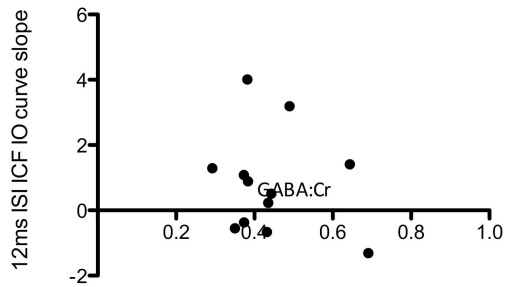
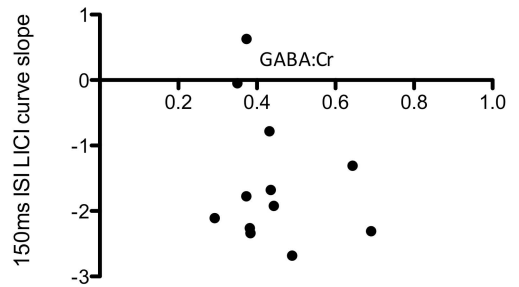
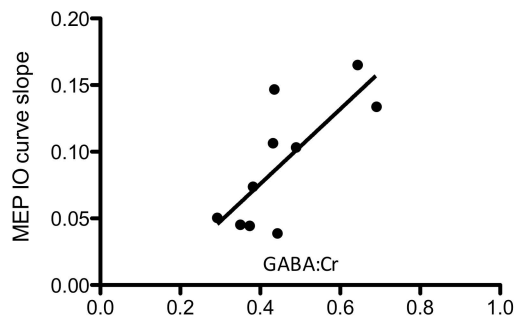
Summary figure of all correlations performed between MRS-GABA and TMS measures. A significant relationship was observed between MRS-GABA and the 1ms SICI measure (A) and between MRS-GABA and MEP IO curve (E), although this was in a counter-intuitive direction, with higher GABA levels being correlated with steeper IO curves. The remaining relationships were non-significant.

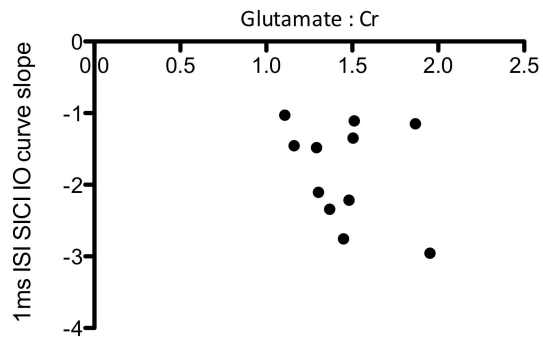
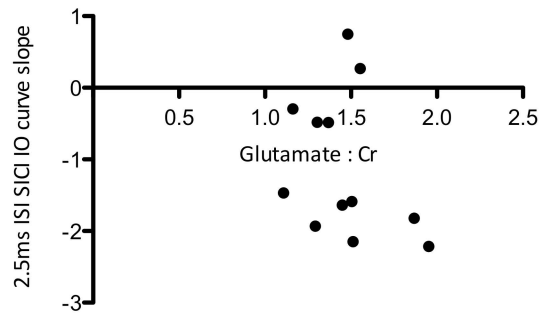
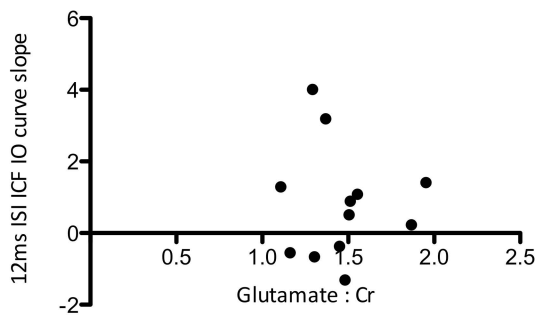
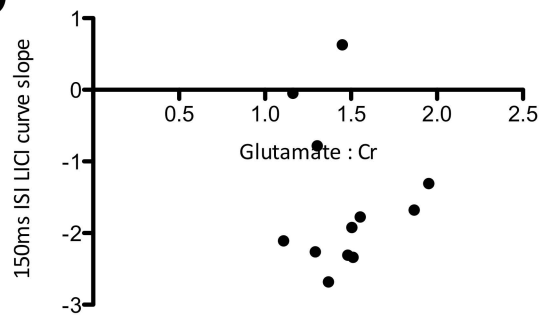
## **Supplementary Figure 2**

Summary figure of all correlations performed between MRS-Glutamate and TMS measures. A significant relationship was observed between MRS-Glutamate and MEP IO curve (E). The remaining relationships were non-significant.

## **Supplementary Figure 3**

Summary figure of all correlations performed between the TMS inhibitory measures. All relationships were non-significant.

**A****B****C****D****E**

**A****B****C****D****E**