

Crystal structures of the extracellular domain of LRP6 and its complex with DKK1

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Supplementary Information

This PDF file includes:
Supplementary Figures 1 to 6

LRP5_1 SPPLLFFANRRDVRLVDAGG-VKLESTIVVSGLEAAVDFQFSKGAVYWTDVSEEAIKQT 90
 LRP6_1 APLLLYANRRLDRLVDAATN-GKENATIVVGGLEDAAAVDFVFSHGLIYWSDVSEEAIKRT 78
 LRP5_3 EAFLVFTSRAAIHRISLET-NNNDVAIPLTGVEASALDFDVSNNHIYWTDVSLKTISRA 702
 LRP6_3 EAFLLFSSRRADIRRISLET-NNNNVAIPLGVKEASALDFDVTDNRIYWTDISLKTISRA 689
 LRP5_2 EEVLLLARRTDLRRISLDTPDFTDIVLQVDDIRHAIAIDYDPLEGYVYWTDEVRAIRRA 400
 LRP6_2 TELLLARRTDLRRISLDTPDFTDIVLQLEDIRHAIAIDYDPVEGYIYWTDEVRAIRRS 387
 LRP5_4 TTFLLFSQKSAISRMIPDDQHSPDILPLHGLRNKAIDYDPLDKFIYWVDGRQN-IKRA 1003
 LRP6_4 TTFLLFSQKSAINRMVIDEQQSPDIILPIHSRNVRAIDYDPLDKQLYWIDSQRQMIRKA 992
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 LRP5_1 YLNQTGAAVQNVVISGLVS---PDGLACDWVGKKLYWTDSETNRIEVANLNGTSRKVLF 146
 LRP6_1 EFNKT-ESVQNVVVSGLLS---PDGLACDWLGKLYWTDSETNRIEVSNLGDSLRKVLF 133
 LRP5_3 FMNGS--SVEHVVVEFGLDY---PEGMAVDWMGKNLWADTGTNRIEVARLDGQFRQVLV 756
 LRP6_3 FMNGS--ALEHVVVEFGLDY---PEGMAVDWLGNLWADTGTNRIEVSKLDGQHRQVLV 743
 LRP5_2 YLDGS--GAQTLVNTIEIND---PDGIAVDWVARNLYWTDGTDRIEVTRLNGTSRKILV 454
 LRP6_2 FIDGS--GSQFVVTAQIAH---PDGIAVDWVARNLYWTDGTDRIEVTRLNGTMRKILI 441
 LRP5_4 KDDGTQPF-VLTSLSQGQNPDRQPHDLSIDIYSRTLFWTCEATNTINVHRLSGEAMGVVL 1062
 LRP6_4 QEDGSQGFTVVVSSVPSQNLEIQPYDLSIDIYSRYIYWTCEATNVI_NVTRLDGRSGVVL 1052
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 LRP5_1 WQDLDQPRALDPAHGYMYWTDWGET-PRIERAGMDGSTRKIIIVDSDIYWPNGLTIDLE 205
 LRP6_1 WQELDQPRALDPPSSGFMYWTDWGEV-PKIERAGMDGSSRFIIINSEIYWPNGLTLDYE 192
 LRP5_3 WRDLDNPRSLALDPTKGYYWTEWGGK-PRIVRAFMGTCMTLVD-KVGRANDLTIDYA 814
 LRP6_3 WKDLDSPRALALDPAEGFMYWTEGGK-PKIDRAAMDGSERTTLVP-NVGRANGLTIDYA 801
 LRP5_2 SEDLDEPRAIALHPVMGLMYWTDWGEN-PKIECANLDGQERRVLVNASLGWPNGLALDLQ 513
 LRP6_2 SEDLEEPRAIVLDPMVGYMYWTDWGEI-PKIERAALDGSRVVLVNTSLGWPNGLALDYD 500
 LRP5_4 RGDRDKPRAIVVNAERGYLYFTNMQDRAAKIERAALDGTEREVLFTGLIRPVALVVDNT 1122
 LRP6_4 KGEQDRPRAIVVNPEKGYMFNLQERSPKIERAALDGTEREVLFFSGLSKPIALALDSR 1112
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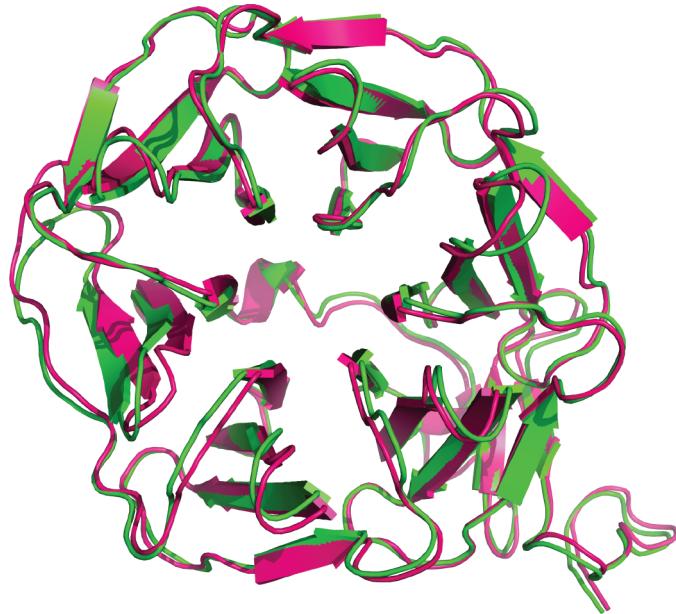
 LRP5_1 EQKLYWADAKLSFIHRANLDGSFRQKVVEGSLTHPFALTLSGDTLYWTDWQTRSIHACNK 265
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 LRP5_3 DQRLYWTDLDTNLIESSNMLGLNR-EVIADDLPHPFGLTQYQDYIYWTDWNLHSIERADK 873
 LRP6_3 KRRLYWTDLDTNLIESSNMLGLNR-EVIADDLPHPFGLTQYQDYIYWTDWNLHSIERADK 860
 LRP5_2 EGKLYWGDAKTDKIEVINVDGKRRRLLEDKLPHIFGFTLLGDFIYWTDWQRRSIERVHK 573
 LRP6_2 EGKLYWGDAKTDKIEVMNTDGTGRRVLVEDKIPHIFGFTLLGDFIYWTDWQRRSIERVHK 560
 LRP5_4 LGKLFWVDADLKRIESCDLSGANRNLTEDANIVQPLGLTILGKHLWIDRQQQMIEKIDM 1182
 LRP6_4 LGKLFWADSDLRRIESSDLSGANRIVLEDSNILQPVGLTVFENWLYWIDKQQQMIEKIDM 1172
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 LRP5_1 RTGGKRKEILSALYSPMDIQVLSQERQPF-FHTRCEEDNGGCSHLCLLSPSEPFYTCACP 324
 LRP6_1 YTGEGLREIHSDIFSPMDIHAFSQQRQP_N-ATNPCGIDNGGCSHLCLMSPVKPFYQCACP 311
 LRP5_3 TSGRNRTLIQGHLDVFMDILVFHSSRQD--GLNDCMHNNGQCGQLCLAIP--GGHRCGCA 929
 LRP6_3 TSGQ_NRTIIQGHLDVFMDILVFHSSRQD--GWNECASSNGHCSHLCLAVPV-GGFVCGCP 917
 LRP5_2 VK-ASRDVIIDQLPDLMGLKAVNFKV--GTNPACDRNGGCSHLCKFTP--ATRCGCP 628
 LRP6_2 RS-AEREVIIDQLPDLMGLKATNVHVRV--GSNPCEENGGSCHLCLYRPQ--GLRCACP 615
 LRP5_4 TTGDKRTRIQGRVAHLTGIHAVEEVSLLEFSAHPCARDNGGCSHICIAKGD-GTPRCSCP 1241
 LRP6_4 TGREGRTKVQARIAQLSDIHAVKELNLQEYRQHPCAQDNGGCSHICLVKGD-GTTRCSCP 1231
 : : * * * . : * : * . * .

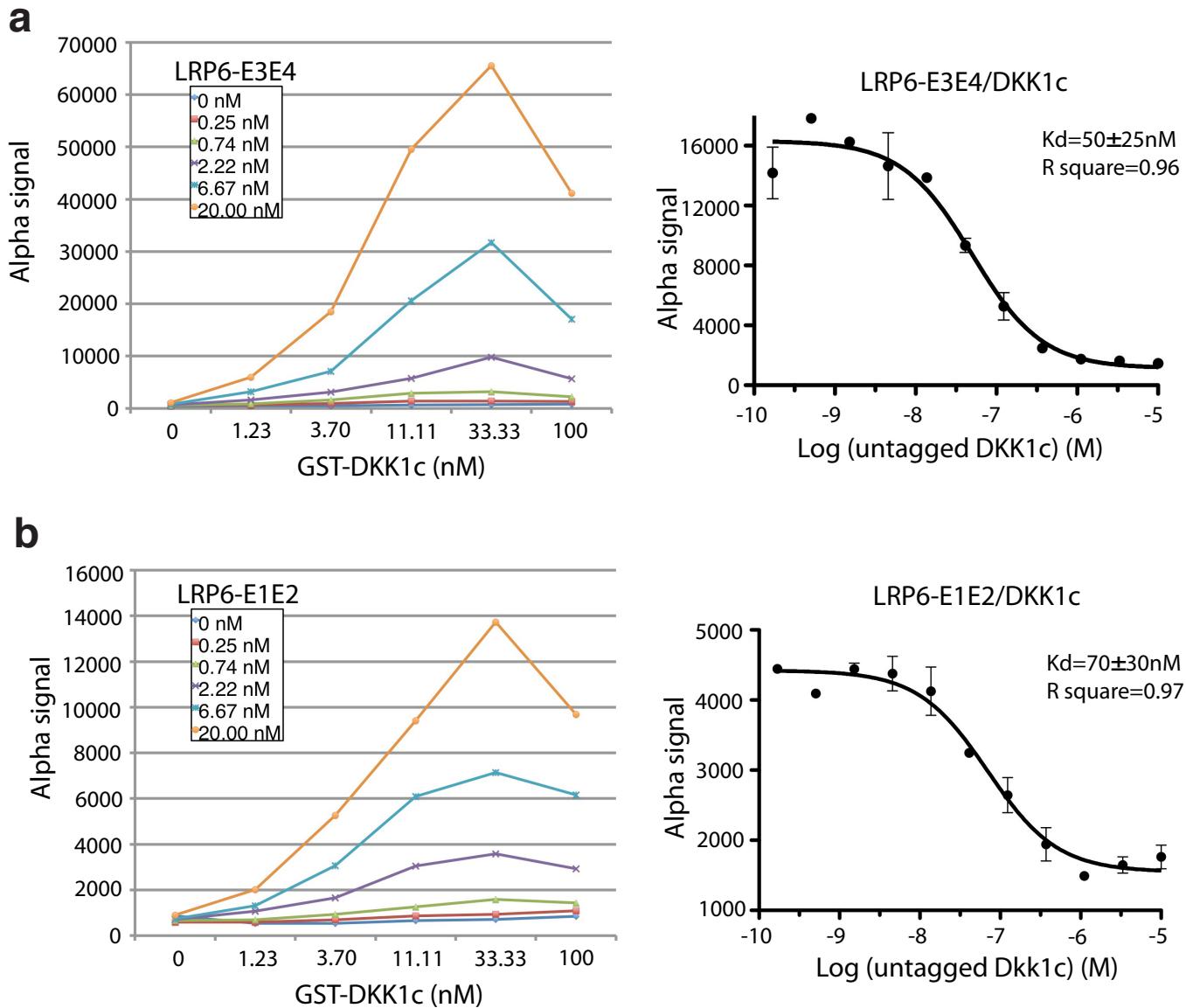
 LRP5_1 TGVQLQDNGRTCKAGA 340
 LRP6_1 TGVKLLENGKTKDGA 327
 LRP5_3 SHYTLDPSSRNCSPP- 944
 LRP6_3 AHYSLNAD_NRTCSAP- 932
 LRP5_2 IGLELLSDMKTCIVP- 646
 LRP6_2 IGFELISDMKTCIVP- 630
 LRP5_4 VHLVLLQNLITCGEPP 1257
 LRP6_4 MHLVLLQDELSCGEPP 1247
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Supplementary Figure 1. Sequence alignment of LRP5/6 propellers. LRP6 residues on interfaces A and B are highlighted by green and cyan background, respectively. Residues used by both interfaces are highlighted with red background. Seven observed N-glycosylation sites (three in LRP6-E1E2 and four in E3E4) are in purple font and underlined, whereas corresponding disease associated missense mutation sites in LRP5/6 are in bold red font.

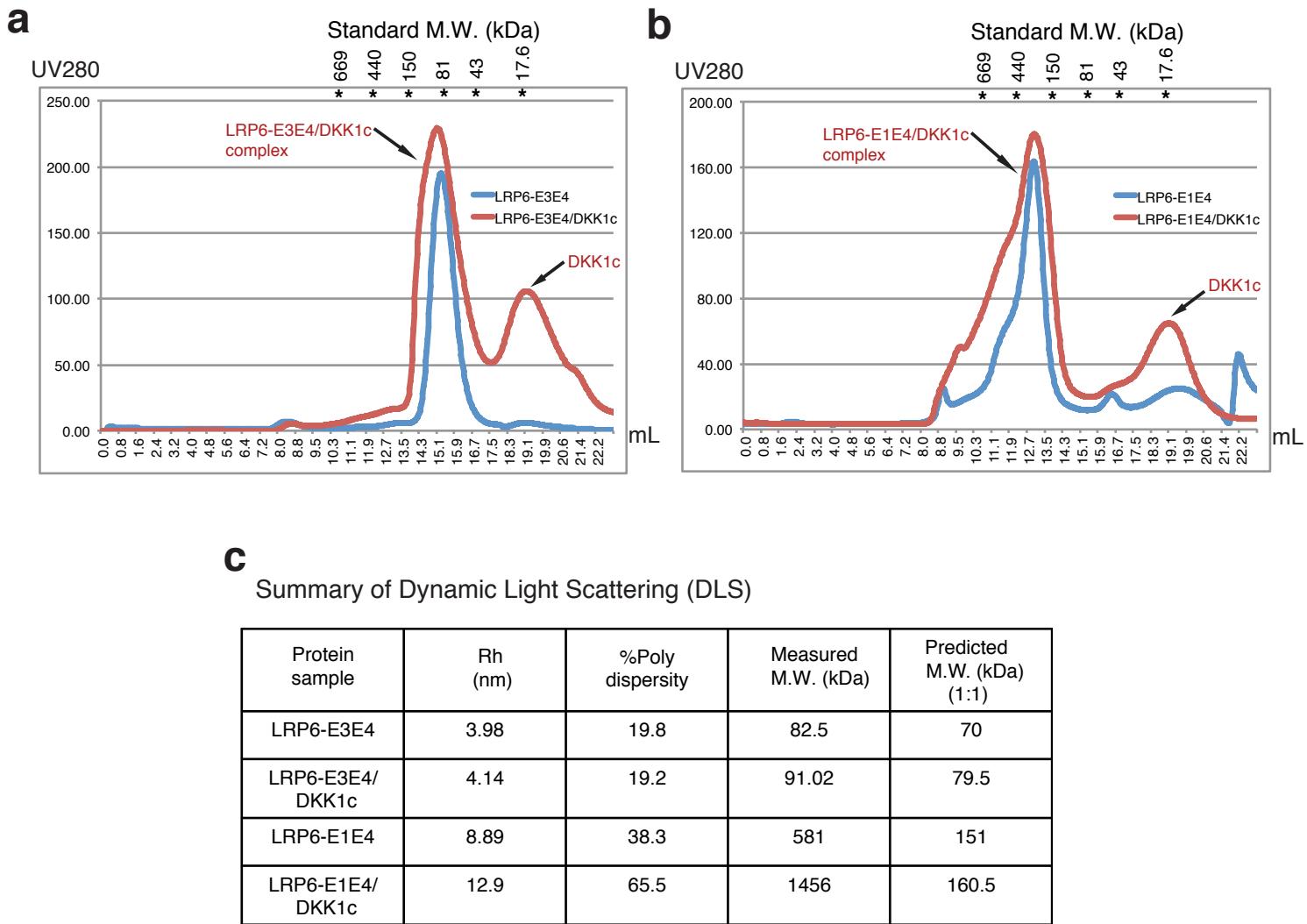
LRP6-E3 Apo vs. Dkk1c-bound LRP6-E3



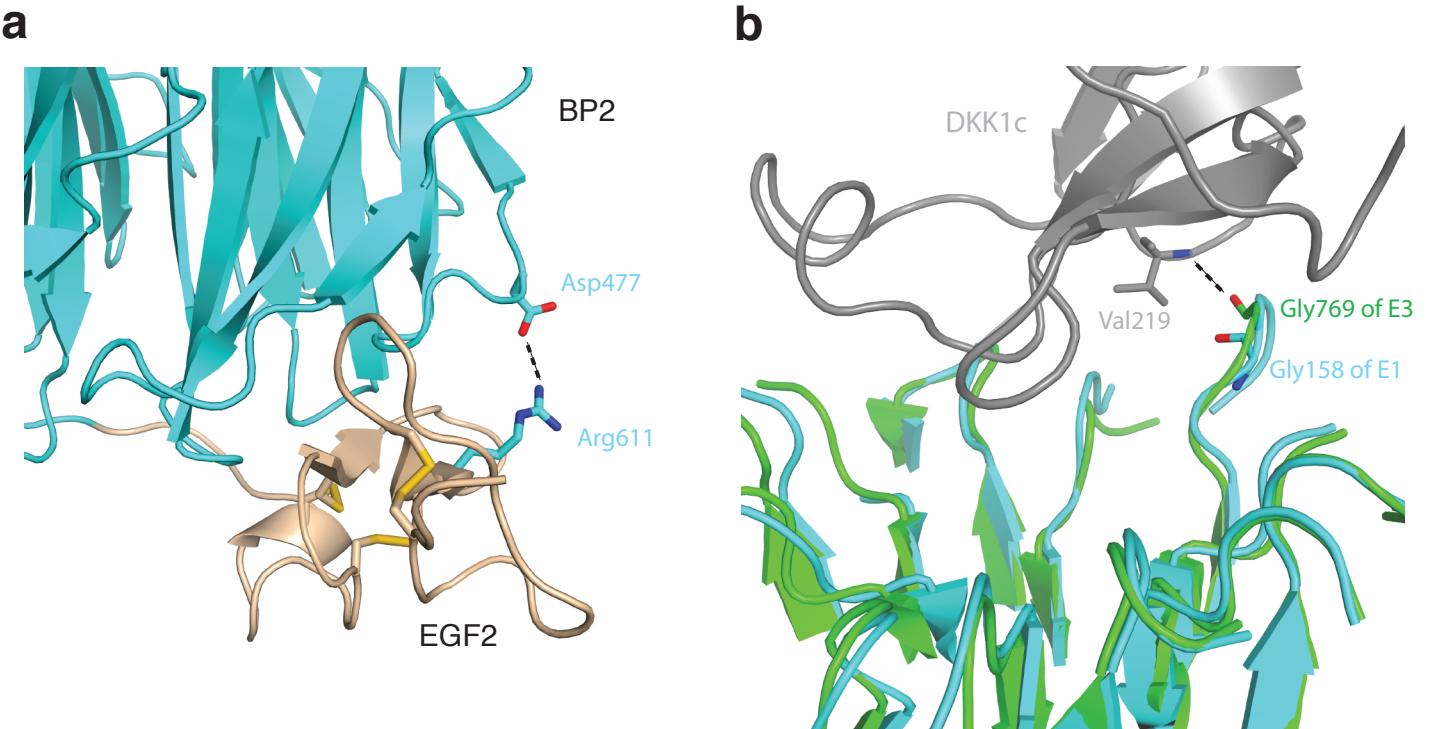
Supplementary Figure 2. Structural comparison of LRP6-E3 in bound and unbound states. There is no significant conformational change on the top surface of LRP6-E3 propeller.



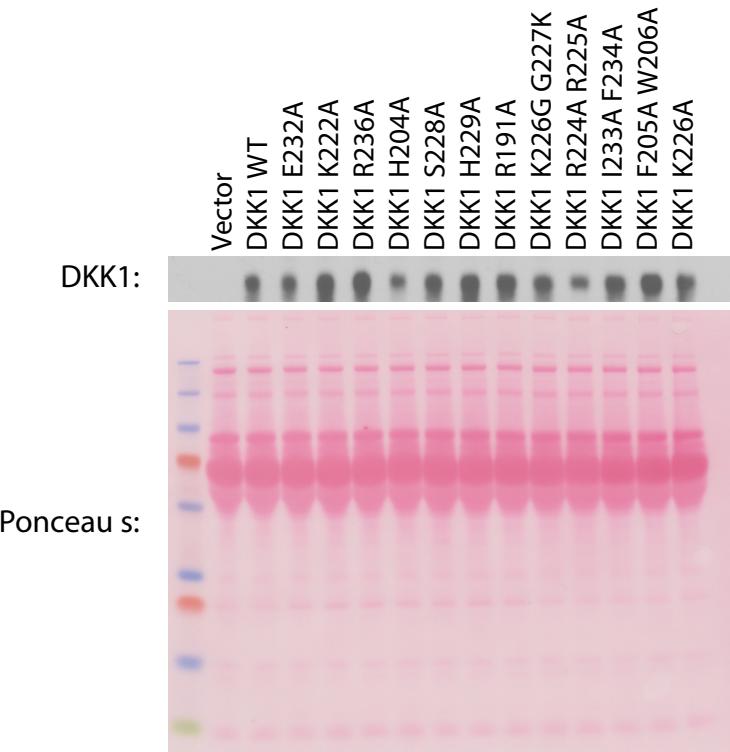
Supplementary Figure 3. Interactions between DKK1c and LRP6-E1E2/E3E4. Binding affinities between DKK1c and two LRP6 rigid blocks (E3E4 and E1E2) were measured using Alpha Technology binding assay (Perkin Elmer, MA). Streptavidin Donor beads and Glutathione Acceptor beads were used in this assay. The experimental procedure is based on the “Determining Kd with an Alpha assay” protocol from PerkinElmer. (a) (Left): Saturation curves of the interaction between biotinylated LRP6-E3E4 and GST-tagged DKK1c. (Right): Competition binding assay to determine the binding affinity Kd. (b) (Left): Saturation curves of the interaction between biotinylated LRP6-E1E2 and GST-tagged DKK1c. (Right): Competition binding assay to determine the binding affinity Kd. The error bars for each data point indicate standard deviations, whereas the “ \pm ” signs associated with Kd values indicate that Kd ranges as determined by the software GRAPHPAD PRISM.



Supplementary Figure 4. Size Exclusion Chromatography (SEC) and Dynamic Light Scattering (DLS) analysis of LRP6-E3E4 and LRP6-E1E4 with/without DKK1c. (a) SEC of LRP6-E3E4 and its complex with DKK1c. Individually purified proteins were mixed with excess of DKK1c and incubated for 1 hour at 4°C prior to SEC. Standard molecular weight was labeled with “*” in corresponding positions. (b) SEC of LRP6-E1E4 and its complex with DKK1c. LRP6-E1E4 by itself tends to form oligomers, while DKK1c binding promotes this oligomerization to higher orders with a peak much broader than that of LRP6-E1E4 alone. (c) DLS analysis of the above samples. The derived molecular weight of LRP6-E3E4 and LRP6-E3E4/DKK1c from DLS is close to the calculated molecular weight and indicates 1:1 binding at the 1 mg/ml. The 12.5 kDa difference of LRP6-E3E4 can be accounted for by glycosylation. The deduced molecular weight of LRP6-E1E4 is much larger than its calculated molecular weight, suggesting that LRP6-E1E4 by itself tends to form oligomers. The observation of a much larger size of the LRP6-E1E4/DKK1c complex is consistent with the broadened SEC peak, suggesting DKK1c induced LRP6-E1E4 aggregation.



Supplementary Figure 5. Structural basis of pathological LRP5/6 mutations. (a) LRP6 R611C mutation. **(b)** LRP5 G171V mutation corresponding to LRP6 G158V.



Supplementary Figure 6. Expression of FLAG-tagged DKK1 and DKK1 mutants in conditioned media. 10 µl of conditioned media containing the indicated FLAG-tagged wildtype (WT) or mutant DKK1 proteins was separated by SDS-PAGE and transferred to nitrocellulose. The nitrocellulose was stained with Ponceau S to show equal loading followed by western blot analysis using an anti-FLAG primary antibody. It should be noted that some of the DKK1 mutants were expressed in higher levels than WT. However all of these expressed in higher levels were found to be inactive – even with higher protein levels. Therefore all our conclusions about mutations that can abolish DKK1 activity hold true.