

Supporting Information

Li et al. 10.1073/pnas.1118560109

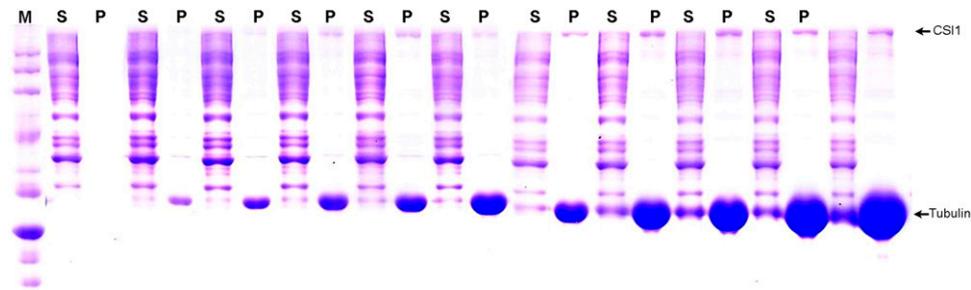


Fig. S1. Microtubule-binding assay of CESA interactive protein 1 (CSI1). For measurement of binding affinity, increasing amounts of taxol-stabilized tubulin were incubated with $0.7 \mu\text{M}$ purified CSI1 protein for 30 min at room temperature. After centrifugation, $5 \mu\text{L}$ of each pellet was resolved by SDS/PAGE and visualized by Coomassie blue staining. A representative gel from three technical replicates used for dissociation constant calculation is shown. The positions of CSI1 and tubulin are indicated by arrows. S, supernatant fraction; P, pellet fraction.

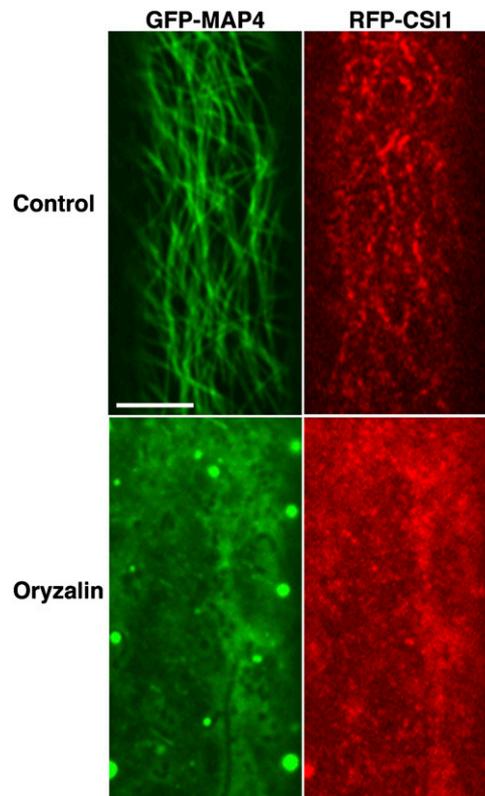


Fig. S2. CSI1 localization is microtubule dependent. Epidermal cells in 3-d-old dark-grown hypocotyls coexpressing GFP-MAP4-MBD and RFP-CSI1 were incubated in Murashige and Skoog liquid solution containing diluted methanol control or $20 \mu\text{M}$ oryzalin for 7 h. A single optical section of GFP-MAP4-MBD or RFP-CSI1 was acquired. Representative images from 22 cells in nine seedlings upon oryzalin treatment are shown. (Scale bar, $5 \mu\text{m}$.)

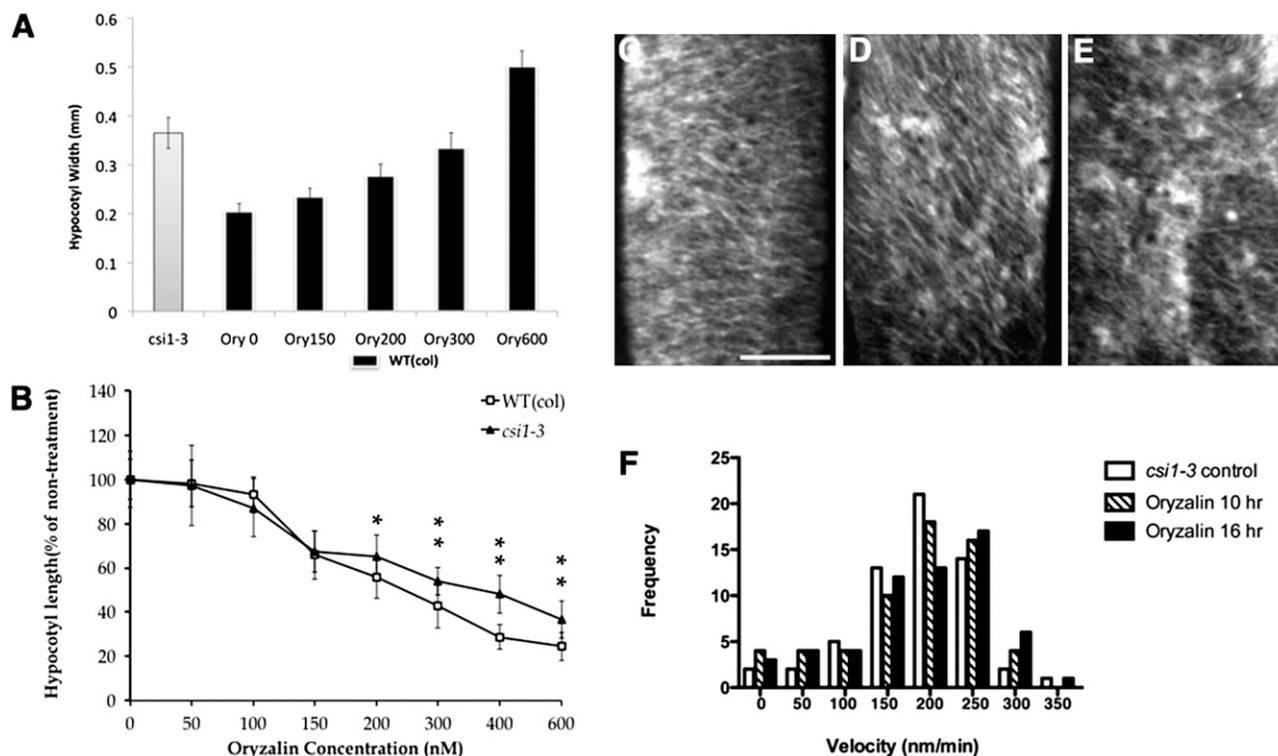


Fig. 55. Quantification of the effect of oryzalin on growth morphology and CESA complexes dynamics in *csi1*. (A) Quantification of hypocotyl width in wild type treated with various concentrations of oryzalin and *csi1-3*. (B) Dose–response curve of the effect of oryzalin on hypocotyl length. Seedlings were grown on agar for 4 d on Murashige and Skoog plates supplemented with indicated concentrations of oryzalin. $**P < 0.01$; $*P < 0.05$. Error bars indicate SD. (C–E) Time average of 61 frames (duration 300 s, 5-s interval) showing YFP-CESA6 in *csi1-3* with mock treatment (C), with oryzalin treatment for 10 h (D), and with oryzalin treatment for 16 h (E). (Scale bar, 5 μm .) (F) Histogram of YFP-CESA6 particle velocities in mock treatment or oryzalin treatment for indicated time. The average velocity of GFP-CESA6 particles in *csi1-3* ($n = 15$ cells, 213 ± 70 nm/min) was no different from those treated with 20 μM oryzalin for 10 h ($n = 12$ cells, 207 ± 87 nm/min) or those treated with 20 μM oryzalin for 16 h ($n = 13$ cells, 217 ± 89 nm/min).

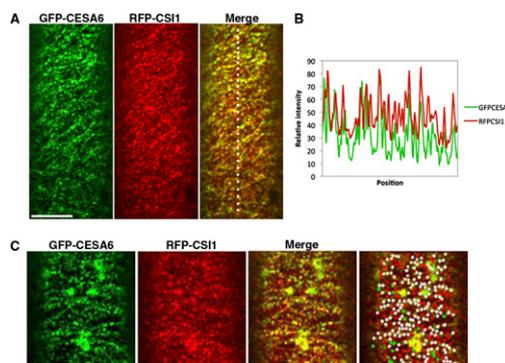
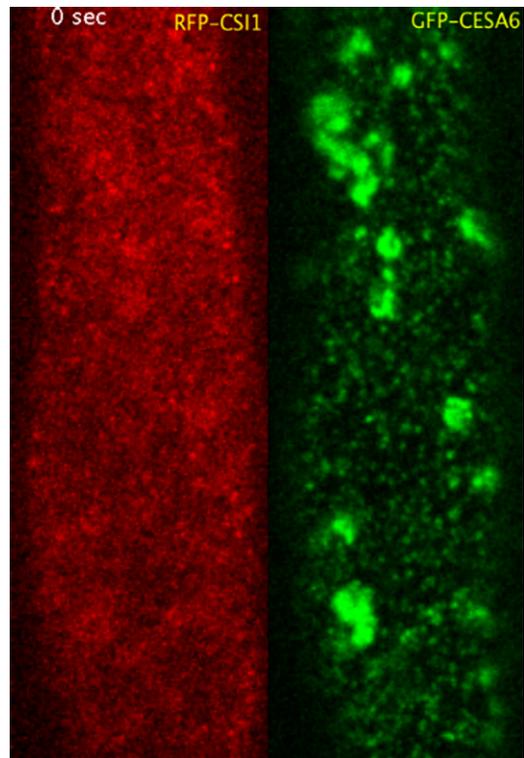
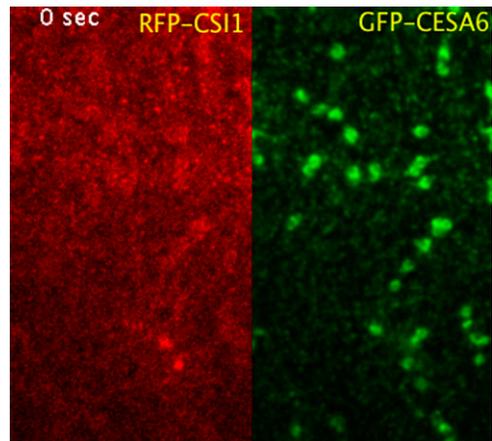


Fig. 56. Colocalization of cellulose synthase (CESA) complexes and CSI1. (A) Two-channel imaging of epidermal cells in 3-d-old dark-grown hypocotyls expressing markers for CESA complexes (GFP-CESA6) and CSI1 (RFP-CSI1). A single optical section was acquired. (Scale bar, 5 μm .) (B) Plot of a line scan showing a strong correlation between GFP-CESA6 and RFP-CSI1; line is indicated in merge image (A). (C) Quantification of colocalization of CESA complexes and CSI1. White dots represent colocalized GFP-CESA6 and RFP-CSI1. RFP-CSI1 and GFP-CESA6 that did not colocalize are green and red, respectively. Analysis was performed in five cells from five seedlings (Table 1). (Scale bar, 10 μm .)



Movie S2. Two-channel imaging of epidermal cells in 3-d-old dark-grown hypocotyls coexpressing RFP-CSI1 and GFP-CESA6. Two-day-old dark-grown hypocotyls were incubated in Murashige and Skoog liquid solution containing 20 μ M oryzalin for 10 h before imaging. Time series are 5 min long at 5-s intervals. Frame rate = 7 fps.

[Movie S2](#)



Movie S3. Two-channel imaging of epidermal cells in 3-d-old dark-grown hypocotyls coexpressing RFP-CSI1 and GFP-CESA6. Two-day-old dark-grown hypocotyls were incubated in Murashige and Skoog liquid solution containing 20 μ M oryzalin for 16 h before imaging. Time series are 5 min long at 5-s intervals. Frame rate = 7 fps.

[Movie S3](#)

