

Supporting Information

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SI Materials and Methods

Cloning and Purification of Rpn6. Rpn6 from *Drosophila melanogaster* was cloned as N-terminal 6xHis-tag fusion protein into a modified pTipRC1 plasmid with a tobacco etch virus (TEV) protease cleavage site in between 6xHis-tag and the *rpn6* gene. Protein expression was carried out in *Rhodococcus erythropolis* (L-88) (1), where the resulting strain was grown at 30 °C in 5 L of LB medium. Protein production was induced with 0.5 µg mL⁻¹ thioestrepton for another 24 to 48 h. Cells were sedimented at 4,000 g and washed with sterile water. The pellet (20 g) was resuspended in 50 mL lysis buffer (50 mM sodium phosphate pH 8.0, 10 mM imidazole, 300 mM NaCl). Complete protease inhibitor cocktail (Roche Biotech) and 1 mg mL⁻¹ lysozyme were added, and the mixture incubated for 2 h on ice. Subsequently, 100 ppm Benzonase was added, followed by ultrasonication on ice. Cell debris was removed by ultracentrifugation at 28,000 × g. His-tagged Rpn6 was purified by affinity chromatography using Ni-NTA beads (GE Healthcare) according to the supplier recommendations. Fractions containing Rpn6 were pooled, augmented with TEV protease for removal of the 6xHis-tag, and incubated for 12 h at 4 °C in a dialysis chamber equilibrating against 25 mM Tris HCl pH 7.5. TEV protease was removed by MonoQ anion exchange chromatography, using a linear salt gradient to 1 M NaCl in 25 mM Tris HCl pH 7.5. Size-exclusion chromatography (SEC), using Superose-12 (GE Healthcare) equilibrated with 20 mM Hepes NaOH pH 7.5, 300 mM NaCl, and 1 mM DTT was used as final purification step.

Limited proteolysis. Full-length Rpn6 at 0.5 mg mL⁻¹ was subjected to limited proteolysis, using increasing concentrations of Proteinase-K (0.025–0.1 mg mL⁻¹). After 30 min incubation at 20 °C, samples were analyzed by SDS-PAGE followed by in-gel digestion and peptide mass spectrometry. Samples with prominent digestion products were subjected to liquid chromatography–mass spectrometry analysis to measure apparent molecular masses to be able to determine actual proteolytic sites.

Crystallization. Crystals of Rpn6(30–422) were grown by the sitting drop vapor diffusion method at 4 °C and 18 °C, mixing equal volumes of Rpn6(30–422) (25 mg mL⁻¹ in 20 mM Hepes NaOH pH 7.5, 300 mM NaCl, and 1 mM DTT) with a precipitant containing 100 mM Tris HCl pH 7.5 200 mM Li₂SO₄, 12% PEG-3350. For cryoprotection, crystals were transferred stepwise into 100 mM Tris HCl pH 7.5, 300 mM Li₂SO₄, 15% PEG-3350 and 20% glycerol before being flash-frozen in liquid nitrogen.

Structure Determination. Diffraction data were collected at the European Synchrotron Radiation Facility (ESRF) in Grenoble, France. The data were processed with XDS (2) and transferred into the CCP4 format using Pointless (3), Scala (4), and Truncate (5). The structure was solved using crystals soaked with 0.5 mM GdCl₃. Four Gd sites were found in a MAD dataset using SHLXD (6) as implemented in HKL2MAP (7). This solution was further refined with Sharp (8). Density modification was subsequently carried out using Resolve (9). A preliminary model was manually built in the resulting map using Coot (10). For final model building and refinement, nearly isomorphous native data

were used. Iterative cycles of manual model building and refinement with Refmac (11), as implemented in the CCP4 interface (12), were carried out. The final model contains Rpn6 residues 38 to 390, two sulphate, two glycerol, and 49 water molecules. Nonglycine residues facing solvent channels without discernable side-chain density were modeled as alanines.

Site-Directed Mutagenesis. Site-directed mutations in Rpn6 were introduced with the QuikChange site-directed mutagenesis kit (Stratagene) using pTipRC-Rpn6 as the template.

Coprecipitation Assay. Different subunits from the *D. melanogaster* regulatory particle were used for the coprecipitation assay. Binary interactions of these subunits were studied by mixing Rpn6 with other subunits having a 6xHis-tag. Proteins were mixed at 1:1 molar ratio and incubated with Ni-NTA Superflow beads at 20 °C for 45 min. The mixtures were applied to spin column bodies (Qiagen), washed with the washing buffer, and the retained proteins were eluted with the elution buffer. The fractions were analyzed by SDS-PAGE.

EM Density Fitting. The exact position and orientation of Rpn6 in the electron density map of the 26S proteasome was determined by an exhaustive six-dimensional search procedure. The atomic coordinates of Rpn6 were converted into a gray-scale volume by assigning the sums of atomic numbers for all atoms contained in the corresponding voxels. This Rpn6 density volume was low-pass filtered to a resolution of 6.4 Å and used as a template for a cross-correlation based search by screening three translational and three rotational parameters. The rotational search was performed with an angular increment of 2° using MOLMATCH (13). The position of the maximal normalized cross-correlation coefficient ($CCC_{max} = 0.48$) and the corresponding rotational parameters were determined, and the original atomic coordinates of Rpn6 were transformed accordingly (see also Fig. S5). In addition, we computed a Z-score for the orientation-specificity for each of the determined positions as described previously (14).

Bioinformatics Methods. A Dali search (15) of the Protein Data Bank (PDB) using the solenoid part of Rpn6 revealed several prokaryotic proteins with structurally similar repeats but disparate biological functions: DrR162B (PDB ID code 3GW4), PlcR [PDB ID code 2QFC (16)], and MalT [PDB ID code 1HZ4 (17)].

HHpred (18) was used for identification of structural templates and their alignment to the target sequence of *Schizosaccharomyces pombe* Rpn6. Our *D. melanogaster* Rpn6 crystal structure covered residues 38 to 387, and TOM70 (PDB ID code 2GW1) was used as an additional template for residues 4 to 100. The C-terminal residues 388–421 were not modeled. Comparative models were built using MODELLER (19) and further refined in the context of the EM map using MDFF (20).

Structural figures were prepared using PyMOL (<http://www.pymol.org>) and Chimera (<http://www.cgl.ucsf.edu/chimera/>). Alignment figures were created with ESPript (21).

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1      10      20      30      40
D.melanogaster . . . . .MAGATLFERAQALSSVNREEQDSSLLNKLVRDQEG. . . . .AEENDE
H.sapiens . . . . .MAAAAVVEFQRAQSLSTDRASIDILHSIVKRDIO. . . . .ENDE
O.dioica . . . . .MAGAACVAQPT. . . . .NNDLQQLYHDLMSAKE. . . . .RSDQ
C.albicans SIHHLQPLQNMSSGQLLEEARAKAVTNKQYDEAEIKYKQIINPSESSTSTTTTTTTTFTA
S.cerevisiae . . . . .MSLPGS.KLEEARRLVNEKQYNEAEQVYLSLID. . . . .KDSSQSSAAAGASVDD
Y.lipolytica . . . . .MTLEETLASAKTAGDKGNLVEAEKLYREILD. . . . .QKAGTNE
C.posadasii . . . . .MAPNSETAALLKEATALS.KTDPSSKAEITFCQVLSLG. . . . .TGSFE
P.brasiliensis . . . . .MAG.PDDASLLAEAKNLV.KTDPKAESEYQKILSKG. . . . .VGSFE
T.stipitatus . . . . .MAST.SASQRTBEAKSLA.SKDPKASEQIYRDRVSSG. . . . .VGKTE
N.crassa . . . . .MAQG.EASERVREAQKVV.STDPRQAEQIYKDIISKPP. . . . .SVTSD
T.melanosporum . . . . .MGRLEEAGEIQ.KRDPARAEAMYKEMISKP. . . . .PGMND
S.pombe . . . . .MSSKSSLELANNVKSNDIEKAIKYKELVNLKG. . . . .VSKDE
D.discoideum . . . . .MNNWKEQLEEIGNCO. . . . .DSNKATQDYNKILLTIQ. . . . .E
A.thaliana . . . . .MVSYRAT.TETISLALAEANSSEAITLYQVILE. . . . .DPSSSP
P.patens . . . . .MLREVIY. . . . .HPASTA
C.rheinhardtii . . . . .MSAELETRLKSATELGATDVPGAAAKLKLGLVLE. . . . .ESSNDA
N.gruberi MTDNTTSDQQTFYTTLFEKATKEEEKQNTAEAIKIFTQVINGE. . . . .MSKNE
T.brucei . . . . .MEKDLDLWDTAEDFIADGRRTEARGVLEDIVSTD. . . . .VTADDA
C.elegans . . . . .MSATPVTLKAVQSEVSAQT. . . . .AKSSE
P.tetraurelia . . . . .MATLIEKFOALLQEVNARLDTNNLEAALLKLLTFDSK. . . . .SE
T.vaginalis .MDNEIPFVERLGINVDPKDFNVPLTQRAALLEDLIDEVLOGD. . . . .AHIE
consensus>50 . . . . .v. . . . .e

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a1 a2 n1 a3
50 60 70 80 90 100
D.melanogaster RIRIKRQGLLQGEIYKQEGKAKEADLLKVTTRPFLSSISKAKAAKLVRSLVDMFLDMD
H.sapiens EAVVVKQSIIELELGSLLAKGQAAELGGLIKYVTRPFLNSISKAKAARLVRSLLDLFLDME
O.dioica ETRKNEAKIIELELGEWLQKGNAKELEGLVKNVTRPFLGSSKAKAARLVRLVDFLDM
C.albicans KQLQQQESAIIELELGIYETINEPTKSDLIADSRSLGNFAKSKTAKIVKTLIEDFDKILIS
S.cerevisiae KRRENEQESLIELELQLYVTMGAKDKREFIPHSTIEMMGEFAKSKTKVVKLTLIEKFEOVP
Y.lipolytica KSIQIQESALINLGTLYASNNKQDADLIHTSLTVMGFAKSKTAKIIRNLIIDLFKAVP
C.posadasii AASRDYEAALVGLGELYRDKRPKELAEILRTSRSSFSFAKAKSAKLVRLDLPFAAIP
P.brasiliensis AASRDYEAALVGLGELYRDKRPKELAEILRTSRSSFSFAKAKSAKLVRLDLPFAAIP
T.stipitatus SASRDYEAALVGLGELYRDKRKAHEADLIKSRDFAFSFAKAKTAKLVRLDLDLSEIP
N.crassa AAIREYEAALVGLGELYRDKNSQGLVLDVLTQSRVTLSSFAKAKTAKLVRLDLDLFEAIP
T.melanosporum AELEREYEAALVGLGELYRDARRTGEAEELIKASRSMSSFAKAKTAKIVRLDLDLFTTIP
S.pombe KVANECQALTNISDLVYRNRHNDLAQLVQSPMLMANSKAKSAKIVRLDLDLDFSGK
D.discoideum SFDIIEKSALELRLAKLFEVKGKGDQLETLIRSVRPFDFKSKPKTKIVRNLIDLDFSPF
A.thaliana EALIKKQATNLCDAITEKRGEDLRKLTKEPFSFLKAKTAKIVRNLIDLDAVAFIP
P.patens EVLVVEQATLALDLSQEKKAELRGLITDRLRPYFNLFKAKTAKIVRNLIDLDAVAFIP
C.rheinhardtii EAVRIKQATISQLCELYIKANAQAADLITSLRGFNFNLFKAKTAKIVRNLIDSLIAKVP
N.gruberi VIKKKESAIYSLGKIYATKGAQAAMELNKSRPFFQDLKAKTAKIVRNLIDLVEVTKK
T.brucei IGLPAKRAIYRAELISVFKQTDMLVQLLSAIRSFFALLPKAKATRMVVRKMFIDLNLNSG
C.elegans AEVKRCEDLILSVSRQLAKFKDITGTRTLVESIRSFYDLVGCARASKLIRDLIVEHALTID
P.tetraurelia EHLKHKESAYNKLSSLYCKNKPQLVFCIMKTHD.FSGFNQTRAAKIMRQIDVQSOVLE
T.vaginalis ERVGFQVNHLELYVOANSEKFAILLDLAEYFTKLPKARTAKLVRIIIOALRKVP
consensus>50 . . . . .E. . . . .l. . . . .y. . . . .l. dl. . . . .r. . . . .kak.ak.vr.li#. . . . .

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a4 a5 a6
110 120 130 140 150
D.melanogaster A. . . . .GTEIYVCKDCLEWAKQEKRTFQRQSLAHLIALLYFDTALVTEATLGAQ
H.sapiens A. . . . .GQVEVLCLECIEMAKSEKRTFRQALEARLVSLYFDTKRYQEAHLGSGQ
O.dioica S. . . . .AGVEVDLCKECIQWAMEEKRTYRQALEARLIMALLYFDTNNYKELCTLGGQR
C.albicans AKD. . . . .EKSIDIQIDVTKSSMQWAIIESKLSFRQSLQLRLSDLLYQKHHKHAALINR
S.cerevisiae D. . . . .SDDQIFVCEKSEIEFAKREKRVFKHSLSLKATLTIYQKQVYKDSALIND
Y.lipolytica S. . . . .DCIDLOIAITOKCIEWAVSEKRNFRQSLQTRLVSLFLEKTYVYDALIND
C.posadasii N. . . . .TIDIQISVIKSCIEWAVSERRSFRQNLLETFLVTIYMOKQSYDDALTLINS
P.brasiliensis N. . . . .TIDIQIIVIKSCIEWAVSERRSFRQNLLETFLVTIYMOKQSYDDALTLINS
T.stipitatus N. . . . .TIDIQVAVIKSCIEWAVSERRSFRQNLLETFLVALIYMOKQSYDDALTLINS
N.crassa D. . . . .SDDIQISVTKSCIEWATSERRSFRQNLLETFLVALIYMOKQSYDDALTLING
T.melanosporum D. . . . .TIDIQISVTKSCIEWAIQERRSFRQNLLETFLVGLYQKHSYTEALSLINS
S.pombe K. . . . .SFLQIEVANDCIKWAIKEKRTFRQALETRLISLYDNSSYTDAALNLINT
D.discoideum D. . . . .NLTLLIEVYKNIQWKDTRNRYRQRLTEKLFITLMEFAKDYANALSGLIT
A.thaliana G. . . . .TDLQITLCKEMVEWTRAEKRTFRQRVEARLAALLMENKEYVEALALLST
P.patens G. . . . .TDLQISLQREMEVWTRAEKRTFRQRVEARLAALLMENKEYSEALSLINT
C.rheinhardtii G. . . . .SLOVDVCKGQVEWATEKRTFRQRVIELRLASLYMQRDRYPALALITR
N.gruberi G. . . . .SIAIQIEICRESIEWATEKRTFRQRVIESFLANLFLIKFEPESLEITR
T.brucei A. . . . .SRQMEVCRDMIAWARQEKRTFRQRLQHRLAEVQFARNERQEAALTLQA
C.elegans QGVGFALDHGKKEKIDLLNCGIWAATSNKREFRSLQARLIRLYNDIRDTNNAQKLAQD
P.tetraurelia G. . . . .TLEQVEHCQFLTDCAKDKNFKHWQIRLALTIYNEKFSQSGEITDK
T.vaginalis G. . . . .TLEIQADLCKQKWITWAKDQERTFRQRIETEISELLEQHKWNEAELIQR
consensus>50 . . . . .qi. . . . .iewa. . . . .e.r.flrq.le.rL.l.q. . . . .y.eal.l.l.

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Table S1. Data collection and refinement statistics

Dataset	MAD			Native
	ESRF, ID23-1 <i>peak</i>	<i>inflection</i>	<i>remote</i>	
Beamline	ESRF, ID23-1 <i>peak</i>	<i>inflection</i>	<i>remote</i>	ESRF, ID29
Wavelength, Å	1.71024	1.71072	1.03320	1.00686
Space group	$P6_1$			$P6_1$
Cell dimensions, a, b, c; Å	161.25, 161.25, 42.08;			161.30, 161.30, 42.10;
$\alpha, \beta, \gamma, ^\circ$	90, 90, 120			90, 90, 120
Resolution limits, Å*	40.42–3.4 (3.58–3.4)	40.47–3.4 (3.58–3.4)	38.89–3.0 (3.16–3.0)	52.78–2.5(2.65–2.5)
R_{merge}^*	0.063 (0.335)	0.054 (0.352)	0.048 (0.489)	0.046 (0.303)
I/σ^*	20.1 (5.2)	13.5 (2.9)	14.5 (1.9)	14.4 (2.6)
Multiplicity [†]	7.3 (7.6)	3.6 (3.7)	3.6 (3.7)	3.3 (3.3)
Completeness, %*	97.9 (98.5)	97.4 (97.7)	97.9 (96.6)	99.3 (99.0)
Phasing				
Sites	4 Gd			—
Phasing power ano	2.172	1.405	0.447	
Phasing power iso	1.158	1.113	—	
Mean FoM			0.277	—
Refinement				
Resolution range	—	—	20–3.0	20–2.5
Reflections (test set)	—	—	11819 (638)	20825 (1146)
R_{work}	—	—	0.205	0.216
R_{free}	—	—	0.251	0.265
No. of atoms	—	—	2731	2788
rmsd bonds, Å	—	—	0.012	0.012
rmsd angles, °	—	—	1.384	1.294
Ramachandran plot [†]				
% most favored region	—	—	88.7	91.8
% additionally allowed	—	—	11.0	8.2

*Values in parenthesis for outer shell.

[†]As defined in Procheck (1).1 Laskowski RA, MacArthur MW, Moss DS, Thornton JM (1993) PROCHECK: A program to check the stereochemical quality of protein structures. *J Appl Crystallogr* 26:283–291.