

Supplementary Figure 1. Expression of Ova in the epidermis after induction with doxycycline. Ova-specific RT-PCR of epidermal cell suspensions isolated from K5/TGO mice after treatment with topical doxycycline ("Cutaneous Dox") or maintenance on doxycycline chow ("Systemic Dox"). Data are normalized to background expression from epidermal cells isolated from wildtype Balb/c mice. sOVA group represents liver tissue harvested from mice constitutively expressing a soluble for of Ova (positive control). Results are representative of 2 replicate experiments.



Supplementary Figure 2. Expression of Ova in the periphery and continued generation of Ova-specific T cells in K5/TGO/DO11 mice maintained on

doxycycline. (a) Flow cytometry of CFSE-labeled DO11 cells adoptively transferred into K5/TGO/DO11 mice that were either left untreated (No dox) or maintained on doxycycline for 97 days. The 2 right panels (Dox (97 days)) are results from two separate mice, both of whom had completely resolved skin disease at the time of adoptive transfer. (b) Flow cytometry of thymocytes and SDLN cells isolated from K5/TGO/DO11 mice maintained on doxycycline for 97 days or left untreated. Cells are pre-gated on CD4 single-positive cells. (c) Numbers of SDLN CD4⁺KJ⁺ cell in SDLNs from mice maintained on doxycycline for 64 days or left untreated. Results are representative of 3 replicate experiments with 2-4 mice/group.



Supplementary Figure 3. Depletion of Treg cells prior to antigen induction results in non-resolving lethal skin inflammation. Survival of K5/TGO/DO11 mice treated i.p. with 2 doses of PC61 monoclonal antibody or isotype control antibody at 10 and 3 days prior to beginning doxycycline. Results are representative of 3 replicate experiments with 3-4 mice/group.



Supplementary Figure 4. **Anti-CD25 antibody treatment depletes nTreg cells**. Flow cytometry of SDLN cells isolated from TGO/DO11 at 7 days and 11 days after i.p. injection of a single dose of anti-CD25 antibody (PC61; 0.5mg/mouse). Cells are gated on CD4⁺KJ⁺ cells. Results are representative of 3 replicate experiments.



Supplementary Figure 5. Treg cells from K5/TGO/DO11 mice that have resolved skin inflammation express higher levels of Foxp3 and CD25. Flow cytometry of SDLN cells isolated from K5/TGO/DO11 mice left untreated (unshaded) or treated with doxycycline for 63 days (shaded). Mice treated with doxycycline for 63 days had completely resolved skin inflammation. Cells are gated on CD4⁺KJ⁺ cells. Results are representative of 3 replicate experiments.



Supplementary Figure 6. Antigen expression is effectively 'turned off' by 30 days after discontinuing doxycycline. (a) LN cells from thy1.1+ DO11.10 mice were labeled with CFSE and adoptively transferred into K5/TGO/DO11 mice maintained on doxycycline for greater than 30 days (positive control), mice that had been off of doxycycline chow for 20 or 29 days, and TGO/DO11 mice (negative control). CFSE dilution and CD44 expression was assayed by flow cytometry 3 days after adoptive transfer. Endogenous DO11 cells from the same K5/TGO/DO11 mice were assayed for CD69 expression by flow cytometry. Adoptively transferred DO11 cells are gated on Thy1.1+CD4+KJ+ cells and endogenous DO11 cells are gated on Thy1.1-CD4+KJ+ cells. (b) Ova-specific RT-PCR of skin harvested from K5/TGO/DO11 mice maintained on doxycycline for greater than 30 days ("On dox", positive control) and mice that had been on doxycycline for >30 days and off for 32 days, and TGO/DO11. Three individual mice are shown. CT values for Ova mRNA were normalized to β-actin controls from each sample. Data shown are β -actin-normalized values expressed as fold increases over mean single transgene (TGO/DO11) negative controls. Results are from 2-3 mice/group.



Supplementary Figure 7. DO11 T cells return to baseline numbers after cessation of doxycycline. Absolute numbers of CD4+KJ+ cells isolated from SDLNs of TGO/DO11 mice (Control) or K5/TGO/DO11 mice that had been on doxycycline for 30 days and subsequently taken off doxycycline for 24 or 30 days (">23 days"). Results shown are means of 4 TGO/DO11 mice and 4 K5/TGO/DO11 mice (2 K5/TGO/DO11 mice had been off doxycycline for 24 days and 2 K5/TGO/DO11 mice had been off doxycycline for 32 days). Error bars represent standard deviation of samples within each group.



Supplementary Figure 8. Antigen-experienced DO11 T cells induce skin disease when adoptively transferred into antigen-expressing hosts. To test whether effector T cells are rendered functionally unresponsive after exposure to tissue antigen, antigenexperienced DO11 cells were isolated from K5/TGO/DO11 mice that were maintained on doxycycline for >100 days (and had resolved disease) or antigen-experienced DO11 cells from K5/TGO/DO11 mice that were on doxycycline for >50 days and off doxycycline for >50 days. Sorted cells were adoptively transferred into Ova-expressing K5/TGO/TCR α^{-1} hosts. K5/TGO/TCR α^{-1} mice were utilized as hosts because these mice do not have endogenous Treg cells, allowing for specific examination of cell-intrinsic anergy of adoptively transferred effector T cells. (a) Gating strategy for cells sorted from antigen-experienced mice. (b) Clinical disease upon adoptive transfer of antigenexperienced DO11 effector cells into antigen-expressing hosts. Approximately 6.5x10⁵ CD4+KJ+CD44+CD25- SDLN cells from K5/TGO/DO11 mice that were maintained on doxycycline for >100 days (On Dox) or from K5/TGO/DO11 mice that were on doxycycline for >50 days and off doxycycline for >50 days (Off Dox) were adoptively transferred into K5/TGO/TCRα^{-/-} hosts. As a positive control, 6.5x10⁵ CD4+KJ+CD25-SDLN cells from antigen-naïve K5/TGO/DO11 mice (Naïve) were transferred into K5/TGO/TCR α^{-1} hosts, and as negative control 'On Dox' cells were transferred into TGO/TCR $\alpha^{-/-}$ hosts (Neg Con). Recipient mice were started on doxycycline 1 day prior to adoptive transfer. (c) Intracellular cytokine staining from SDLNs harvested at 14 days after adoptive transfer. Two replicate mice from each group are shown. Gated on Live CD4+KJ+Foxp3- cells. Results are from 3 mice/group. Error bars represent standard deviation of samples within each group.