SUPPLEMENTARY INFORMATION

Figure 1S

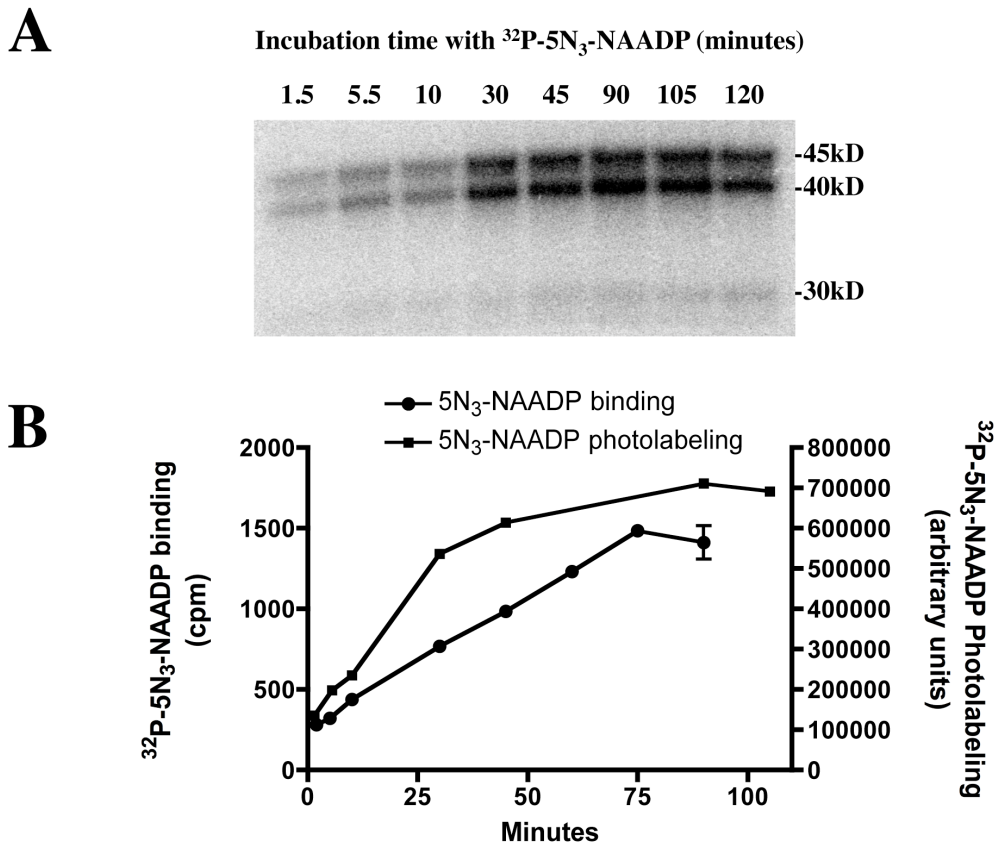

Figure 1S: Time course of ^{32}P-5N$_3$-NAADP photolabeling and binding. Sea urchin egg homogenates were incubated with ^{32}P-5N$_3$-NAADP for the indicated times at 4°C and then either exposed to UV photolysis for 2 minutes or subjected to a conventional binding assay using vacuum filtration. Panel A shows the results of ^{32}P-5N$_3$-NAADP photolabeling. Panel B shows the quantitative results of the photolabeling experiment shown in Panel B (squares --intensity of the 45, 40 and 30kDa bands combined) compared to the results obtained with a conventional binding assay (circles).

Figure 2S

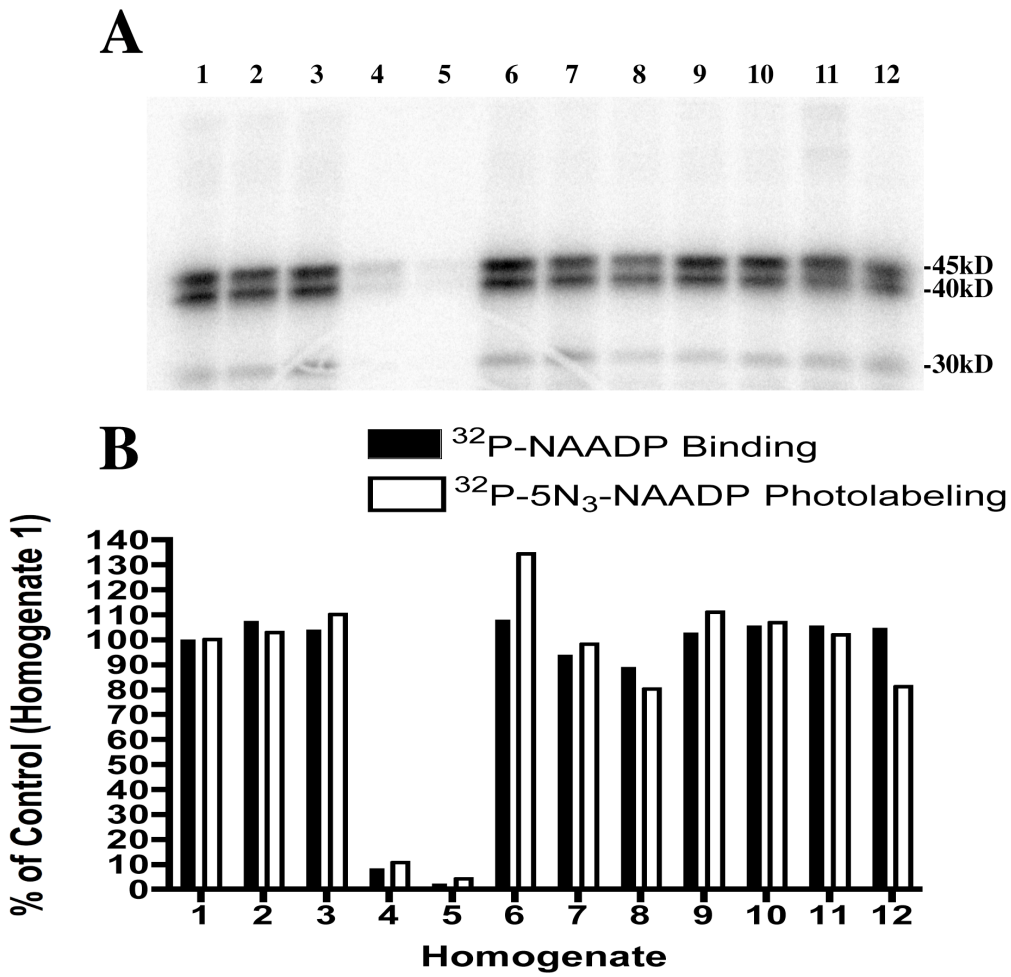

Figure 2S. ^{32}P-5N$_3$-NAADP photolabeling correlates with ^{32}P-NAADP binding. Panel A shows ^{32}P-5N$_3$-NAADP photolabeling of twelve distinct *S. purpuratus* egg homogenate preparations. Panel B shows the comparison of the densitometric analyses of the photolabeling shown in Panel A (white bars) 45, 40 and 30kDa bands combined) with ^{32}P-NAADP binding in these preparations (black bars). The data for photolabeling and binding is normalized to the results of homogenate #1, which was used for most of the studies described here. Note that photolabeling and binding correlate. Homogenates that have low ^{32}P-NAADP binding activity do not photo label efficiently (i.e. homogenates #4 and 5). This correlation between binding and photolabeling applies to the 32 preparations we have examined. This correlation also extends to the ability of NAADP to induce calcium release.