Purification of HIV RNA from serum using a polymer capture matrix in a microfluidic device

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7.1 PCR

The PCR mix for amplification of the ssDNA oligonucleotide contained 26 μ l of H₂O, 1 μ l of recovered sample, 4.1 μ l of 10x PCR buffer, 4.1 μ l of 25 mM MgCl₂, 3.25 μ l of dNTP mix (contains 10 mM of each nucleotide), 1.2 μ l of each primer (10 μ M stock solutions) and 0.6 μ l of AmpliTaq Gold (PCR reagents from Applied Biosystems, Foster City, CA, primers from Operon Biotechnologies). Thermal cycling conditions were: 10 minutes at 95 °C, followed by 36 cycles of 95 °C for 30 seconds, 52 °C for 30 seconds and 62 °C for 1 minute. The RT-PCR mixture for amplification of recovered RNA contained 12.5 μ l of each primer (10 μ M stock solution), and 5 μ l of recovered sample. Thermal cycling conditions for RT-PCR were: 30 minutes at 55 °C, 5 minutes at 94 °C, 42 cycles of 94 °C for 15 seconds, 55 °C for 30 seconds, 68 °C for 30 seconds, and a final extension at 68 °C for 5 minutes. Primers for PCR and RT-PCR are:

PCR

5' - TGC TTG ATG TCC CCC CAC T 5' - TTG TAA AAC GAC GGC CAG T

RT-PCR

5' - ACT GGG GGG ACA TCA AGC AGC CAT GCA AAT 5' - TGC TAT GTC ACT TCC CCT TGG TTC TCT

The ssDNA oligonucleotide target used in this study was:

5'- TTGTAAAACG ACGGCCAGTC ACACACACA ACACACACA ACACACACA ACAGTGGGGG GACATCAAGCA

7.2 Pfold Analysis of the HIV-B gag region

Pfold (http://www.daimi.au.dk/~compbio/pfold/) was used to analyze the HIV-B gag region for potential secondary structure. The red, italicized sequences are regions of secondary structure predicted by Pfold. The blue, underlined sequences are the SK462 and SK431 primer regions. The bold brackets indicate the capture sequences used in this study. The capture sequences were shorter than the primers to lower the dehybridization temperature necessary for the recovery step. Pfold predicts a six base region of the 22 bases targeted by the SK462 capture sequence to be hybridized to another region of the gag sequence.

CAAATGGTAC	ATCAGGCCAT	ATCACCTAGA	ACTTTAAATG	<i>CATGGGT</i> AA
AAGTAGTAGA	AGAGAA <i>GGCT</i>	TTC AGCC CAG	AAGTAA <i>TACC</i>	<i>CATGT</i> TTTCA
GCATTATCAG	AAGGAGCCA <i>C</i>	<i>CCCAC</i> AAGAT	TTAAACACCA	TGCTAAACAC
AGTGGGGGGA	CATCAAGCAG	CC]ATGCAAAT	GTTAAAAGAG	ACCATCAATG
S	SK462			
AGGAAGCTGC	AGAATGGGAT	AGAGTACATC	CAGTGCATGC	AG <i>GGCCT</i> ATT
GCACCAGGCC	AGATG AGAGA	ACCAAGGGGA	AGTGACAT]AG	CAGGAACTAC
		SK431		
TAGTACCCTT	CAGGAACAAA	TAGGA <i>TGGAT</i>	GACAAATA AT	CCACCTATCC
<i>CAG</i> TAGGAGA	AATTTATAAA	AGATGGATAA	TC <i>CTGGGAT</i> T	ΑΑΑΤΑΑΑΤΑ
GTAAGAATGT	ATAGCCCTAC	CAGCATTCTG	GACATAAGAC	AAGGACCAAA
GGAACCC				