

## **Reynolds et al. Supplementary Information**

### **Supplementary Figures**

#### **Supplementary Figure 1. ChIP for NuRD component proteins.**

ChIP-qPCR for putative NuRD targets for proteins other than Mi2 $\beta$  which are unique to NuRD (Mbd3, MTA2) or which are not uniquely found in NuRD (HDAC1). qPCR is shown relative to level in IgG control either at regions close to TSS for individual genes, or across a single target locus, *Htra1*. Distance from TSS is shown in base pairs.

**Supplementary Figure 2. Distribution of histone modifications.** (A) At the whole gene level. Distance is shown in base pairs for upstream, 3kb Metagene, and downstream regions. (B) At transcription termination sites. Profiles, normalized to input, are of up-regulated, bivalent genes for  $\alpha$ H3K27me3 and  $\alpha$ H3K27ac ChIP samples (solid line) relative to that for all RefSeq genes in each sample (dotted line).

**Supplementary Figure 3. Distribution of histone modifications for genes bound by Mi2 $\beta$ .** Profiles, normalized to input for genes bound by Mi2 $\beta$  at transcription termination site (TSS) for  $\alpha$ H3K27me3 and  $\alpha$ H3K27ac ChIP samples. Profiles shown are for genes bound by Mi2 $\beta$  (solid line) relative to that for all Ref-Seq genes in each sample (dotted line).

**Supplementary Figure 4. NuRD and PRC2 complexes do not co-immunoprecipitate in ES cells.** (A) Immunoprecipitation of Mi2 $\beta$  or IgG control probed for presence of Jarid2, MTA2 and Mi2 $\beta$ . (B) Avi-tagged Mbd3 pulled down using streptavidin beads,

probed for presence of PRC2 component Ezh2 or known NuRD interactor Sall4. Pull-downs were performed in the presence or absence of nucleases as indicated. (C) Immunoprecipitation of either MTA2 or Jarid2, probed for the presence of Suz12 or MTA2. All precipitations were carried out using nuclear extracts from wild type ES cells.

**Supplementary Figure 5. Effect of TSA treatment on transcription over time.**

Quantitative RT-PCR from cells grown in TSA for varying times, or from *Mbd3*<sup>-/-</sup> cells shown relative to wild type expression levels in the absence of TSA. Error bars indicate sem.

**Supplementary Figure 6. Distribution of histone modifications for genes at which PRC2 occupancy is dependent on NuRD.** Profiles, normalized to input at transcription start site (TSS) for  $\alpha$ H3K27me3 and  $\alpha$ H3K27ac ChIP samples at those genes bound by Suz12 in wt cells but not in *Mbd3*<sup>-/-</sup> and which are also bound by Mi2 $\beta$  (solid line) relative to that for all Ref-Seq genes in each sample (dotted line).

**Supplementary Table 1. Results of microarray analysis comparing *Mbd3*<sup>-/-</sup> to wild type cell lines.**

**Supplementary Table 2. Summary of genomic regions bound by Mi2 $\beta$  in wild type ES cells.**

**Supplementary Table 3. Summary of ChIP-Seq data for H3K27 modifications in wild type and *Mbd3*<sup>-/-</sup> ES cells.**

**Supplementary Table 4. Comparison of gene expression patterns in *Mbd3*-null and PRC-null ES cells.**

**Supplementary Table S5. Summary of genomic regions bound by Suz12 in wild type and *Mbd3*<sup>-/-</sup> ES cells.**

**Supplementary Table 6a. Primers For Expression Analysis**

Gene	Forward	Reverse
<i>β Actin</i>	GTGGGCCGCTCTAGACACCA	CGGTTGGCCTTAGGGTTCAGGGGGG
<i>Htra1</i>	ACTTCGGA ACTCCGATATGG	CGTGGGACTCTGTCAAGAAC
<i>Klf2</i>	CTAAAGGCGCATCTGCGTA	TAGTGGCGGGTAAGCTCGT
<i>Klf4</i>	CGGGAAGGGAGAAGACACT	GAGTTCCTCACGCCAACG
<i>Klf5</i>	CCGGAGACGATCTGAAACAC	CAGATACTTCTCCATTTACATCTTG
<i>Lefty2</i>	GCAGGTCCAGGTACATCTCC	ACACGCTGGACCTCAAGGAC
<i>Mcm6</i>	GAGAAACACGCTGGTTGTGA	AAGGTCTTCAAGGCTCGACA
<i>Pp1A</i>	CACGGGGGCCTGTCTCCAGA	GTCAGGCACGTCTGTGGGCC
<i>Ppp2r2c</i>	TTCCCGCTGGAAGATAACC	CGCGGAAAATTAACCACAGC
<i>Smad7</i>	TCTCCCCCTCCTCCTTACTC	TCCAGAAGAAGTTGGGAATC
<i>Sohlh2</i>	TTCTGATTTGTCCTGGCAGC	TATTCCATGACTGCTGCAGG
<i>Sox9</i>	CCACGGAACAGACTCACATCTCTC	CTGCTCAGTTCACCGATGTCCACG
<i>T</i>	TGCTTCCCTGAGACCCAGTT	GATCACTTCTTTCCTTTGCATCAAG
<i>Tbx3</i>	GAACCTACCTGTTCCCGGAAA	CCATTGCCAGTGTCTCGAAAAC

**Supplementary Table 6b. Primers For Chromatin IP Analysis**

Gene	Forward	Reverse	Distance of mid-point
------	---------	---------	-----------------------

			from TSS (bp)
<i>β Actin</i>	GCCTAGTAACCGAGACAT TGA	AGAAAGCGAGATTGAGG AAG	-3250
<i>Cdx2</i>	AAGCCTGCCTTTCTGGACT T	TACGAGCTTCCTCCTTCCA A	-280
<i>Htral1</i>	AGCAGCACCCCTTGATCCTA A	GGGATGCCAGACAGAAA GAA	-2040
<i>Htral1</i>	TGGCGAGGTGCATGGGGA ACT	AGTCCCAGCGCTCGGGCA AA	-696
<i>Htral1</i>	TCCAGCGCTCGGGCAA TC	TCTGCCTCCGGGGTGACA GT	-646
<i>Htral1</i>	ACTTGAACTAGGTCTGG GC	GGCACTTAACAGAGGGAA AC	-236
<i>Htral1</i>	GGTTTCCCTCTGTTAAGTG C	GGCTCAGTTTCTCATTCTA GG	-109
<i>Htral1</i>	GTCACCGCCGCTAGGCCA ATG	GGGTCTTGGGGACAGCGG GT	65
<i>Htral1</i>	GCGGCTCCTTCCTTGGCGT T	TGGGATCGCAGTGCTCGG GA	250
<i>Htral1</i>	CTCGCCAACTCAGCCCGA CC	TGCTTCTCTGCACCTCCGC A	708
<i>Htral1</i>	GGGTGTTCTAGGCATCCA GT	GATGCTTGGCGCAGAAAT AG	795
<i>Htral1</i>	GAGGGGCCCACTGGGGAT GAA	TGTCGCCACCACCACGTC CA	1356
<i>Htral1</i>	GGCCCCAGTGGCCCCCTAA GT	AGTCAGGGCGAAGCCAGG CT	2733
<i>Htral1</i>	TCTGCTAACAGTGCCAATG C	ACAGGTTACGATCCACA CA	3643
<i>Klf2</i>	GGGGCTTGAGGCTGGGGA GA	GGCGACGGCGTCAACAAA CC	-265
<i>Klf4</i>	CTCACCCCCACCCTACG	ATTATCCGCGTGACTCAT CC	-570
<i>Klf5</i>	GTCGGAGGCGGGACCTCG TG	GTCGGAGGCGGGACCTCG TG	-66
<i>Lefty2</i>	GAACACACATCAGGTGGT GG	CAGGGCAGACTTCTTTGA GG	-4500
<i>Mcm6</i> (Peng et al, 2009)	CTTATCGGAGGCACCTATA GTGAT	CTAACTCTCCTAGCCTCCT GACAC	993
<i>Ppp2r2c</i>	CCGCGATATCCTCGCGCTC C	AGCGGTCACCTCCGGAAC CA	-186
<i>Smad7</i>	GGTGGCAGTAACTGGGAG G	CGTCTAGACACCCTGTCTG CT	-397

<i>Sohlh2</i>	CCATTGGTTCTCAAGTCAG C	GGCGGTTTCTTTAATTCAG GAT	-328
<i>Sox9</i> 1	TATTAGAGACCCTGAGCT GGAAGT	CTGGACTGAAACTGGTAA AGTTGT	24
<i>T</i>	GGGAGTGGGGACTGCCCG AA	GTGCCAGGGAATGACGGG CC	-3265
<i>T</i>	GCGGAGGCTCAGGCACGA AG	CTGGGCACTGCTGGCTGC TT	-2117
<i>T</i>	GGTCGGGGTTGGGCGCAA ATG	AGCCAACCTCTGGGGTGG GATG	-1399
<i>T</i>	CAGTCCATGGGGCGAGGG GA	GGCGTCTCCCGGGTCTCC TT	-629
<i>T</i>	CTGCGCCCGACGCTTTCCT TA	CTCCCGCAAGGCGCGACA AG	-234
<i>T</i>	TCTCGGTGCTCCTTTGGCG AAT	CGCTGAGCAGGTGGTCCA CTC	64
<i>T</i>	CCTACCTGCCGTTCTTGGT CACA	GTGGGCCTGGAGGAGAGC GA	278
<i>T</i>	CACCGAACGCGAACTGCG AG	ACAAAGTCGGCCGGTGG GAA	312
<i>T</i>	CTTTGTTTCTTCCCGCTGA G	GCAAACCTGGTCATTCCA GT	474
<i>T</i>	CTGGCTCTGGCCATAGGTG AGC	GTGCAGAGTAGCCAGTGT CCCCT	1311
<i>T</i>	GCCCGCAACGCATGATCA C	GGGACTCCCTTCCGAAAC CTCAC	1796
<i>T</i>	ACGGGAAAGTCCTGGCAA AGGCT	GCCCCAGGGGACTGATGA CACA	2113
<i>Tbx3</i>	AGGCACCCAGAGATAAGT GTGATG	GAGCTGCCGCTCTGGGCT TG	-817

## Supplementary Table 7

### Antibodies

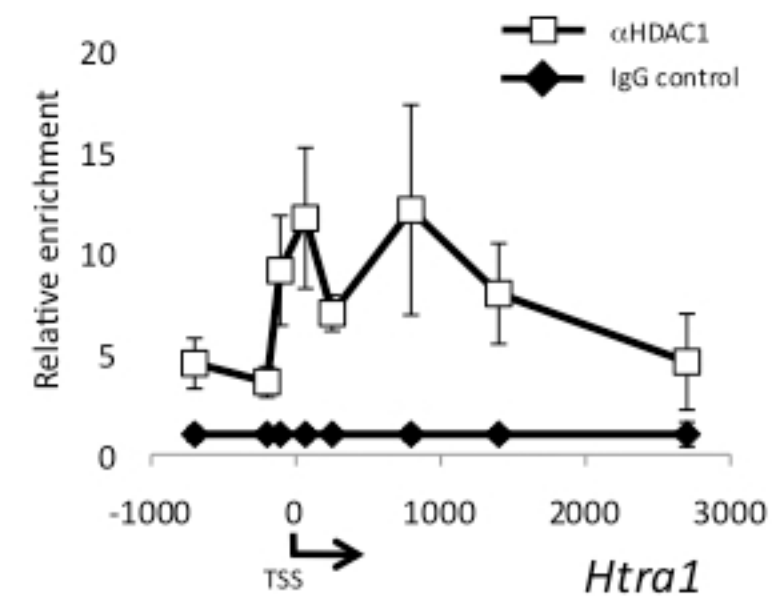
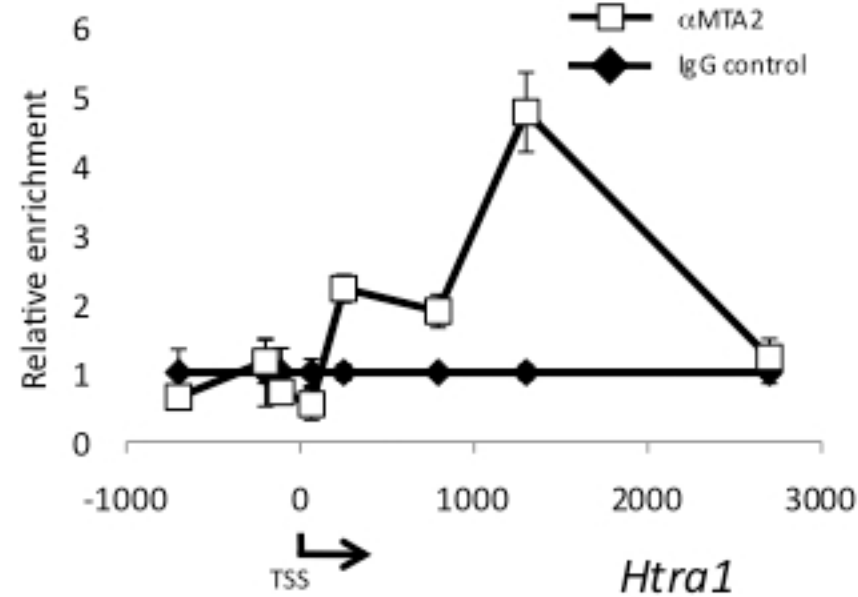
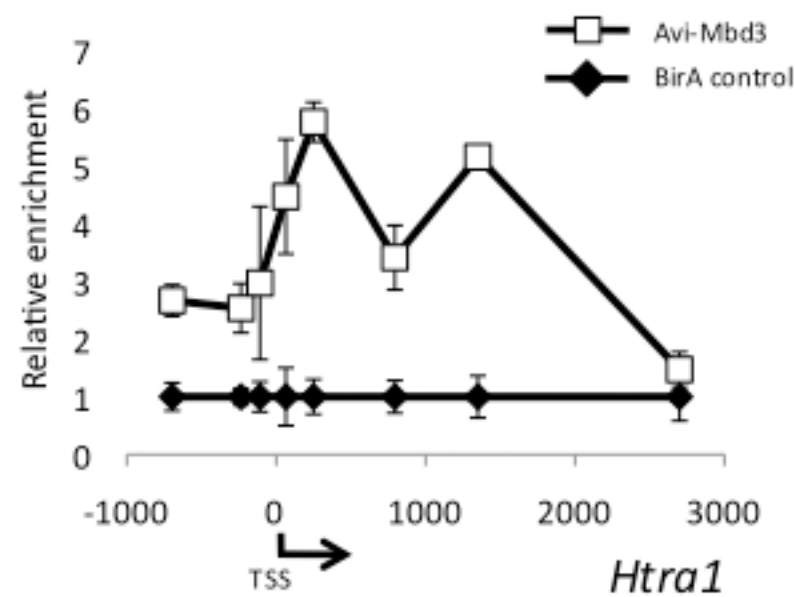
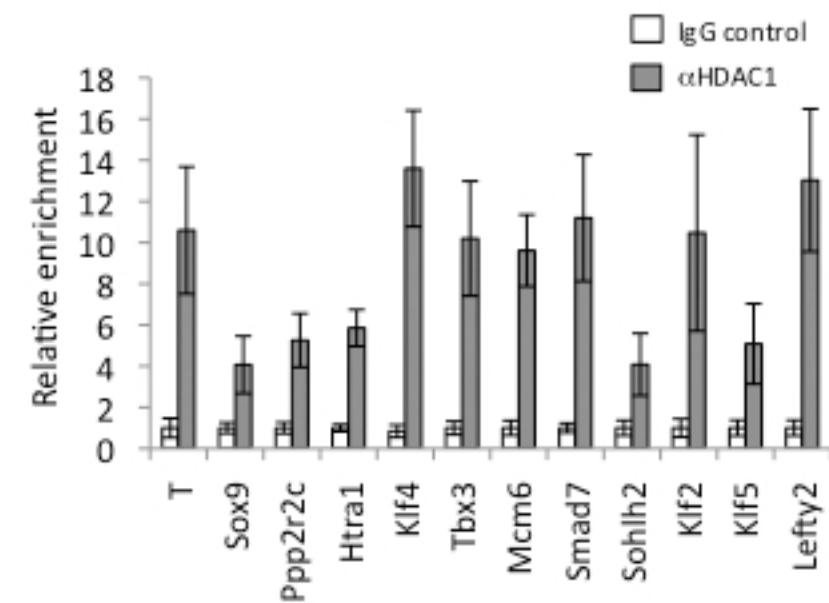
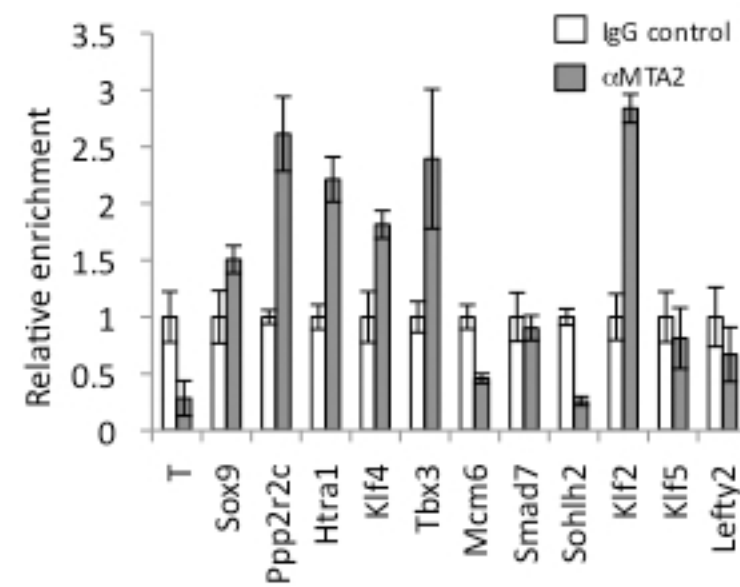
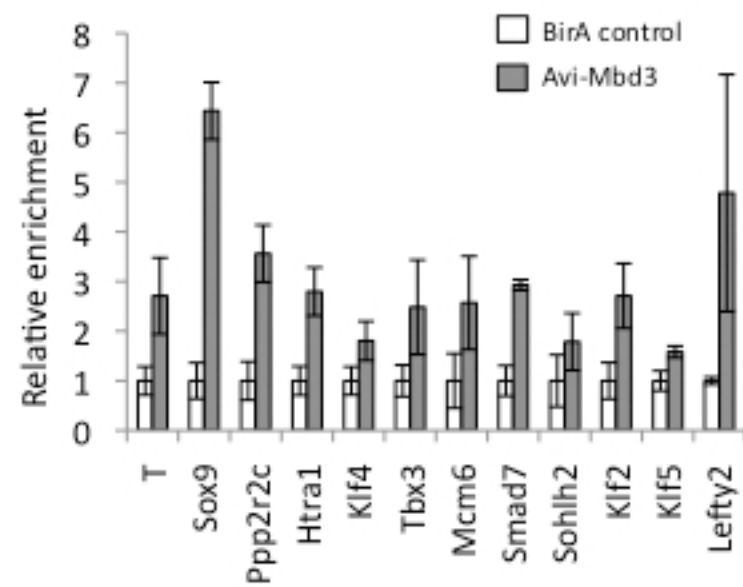
Antibody	Source	Reference
$\alpha$ Mi2 $\beta$	Abcam	ab70469
$\alpha$ MTA2	Santa Cruz	sc-9447
$\alpha$ Mbd3	Santa Cruz	sc-9402
$\alpha$ Suz12	Cell Signalling	3737s
$\alpha$ Ezh2	Active Motif	39104
$\alpha$ Eed	Millipore	09-774

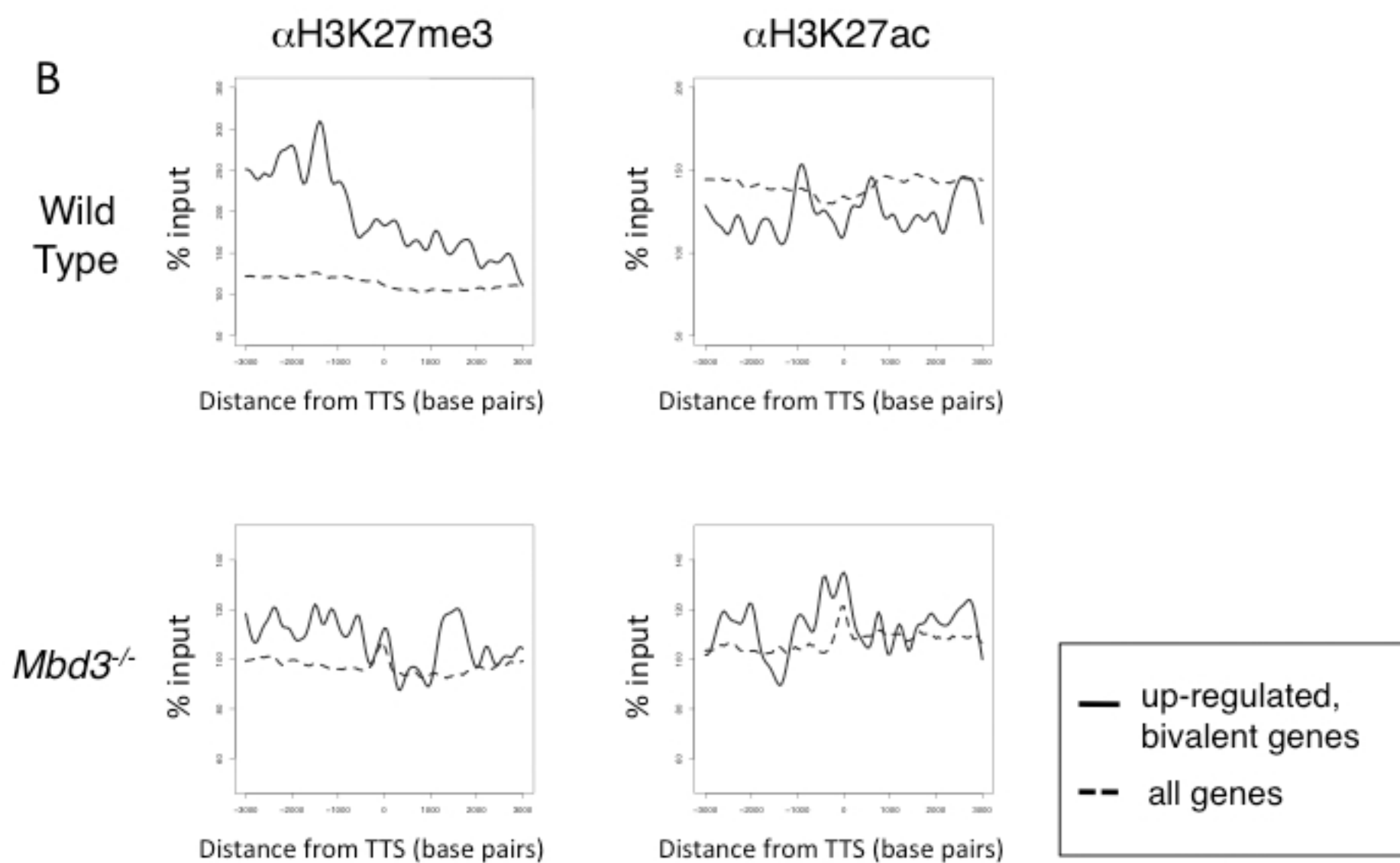
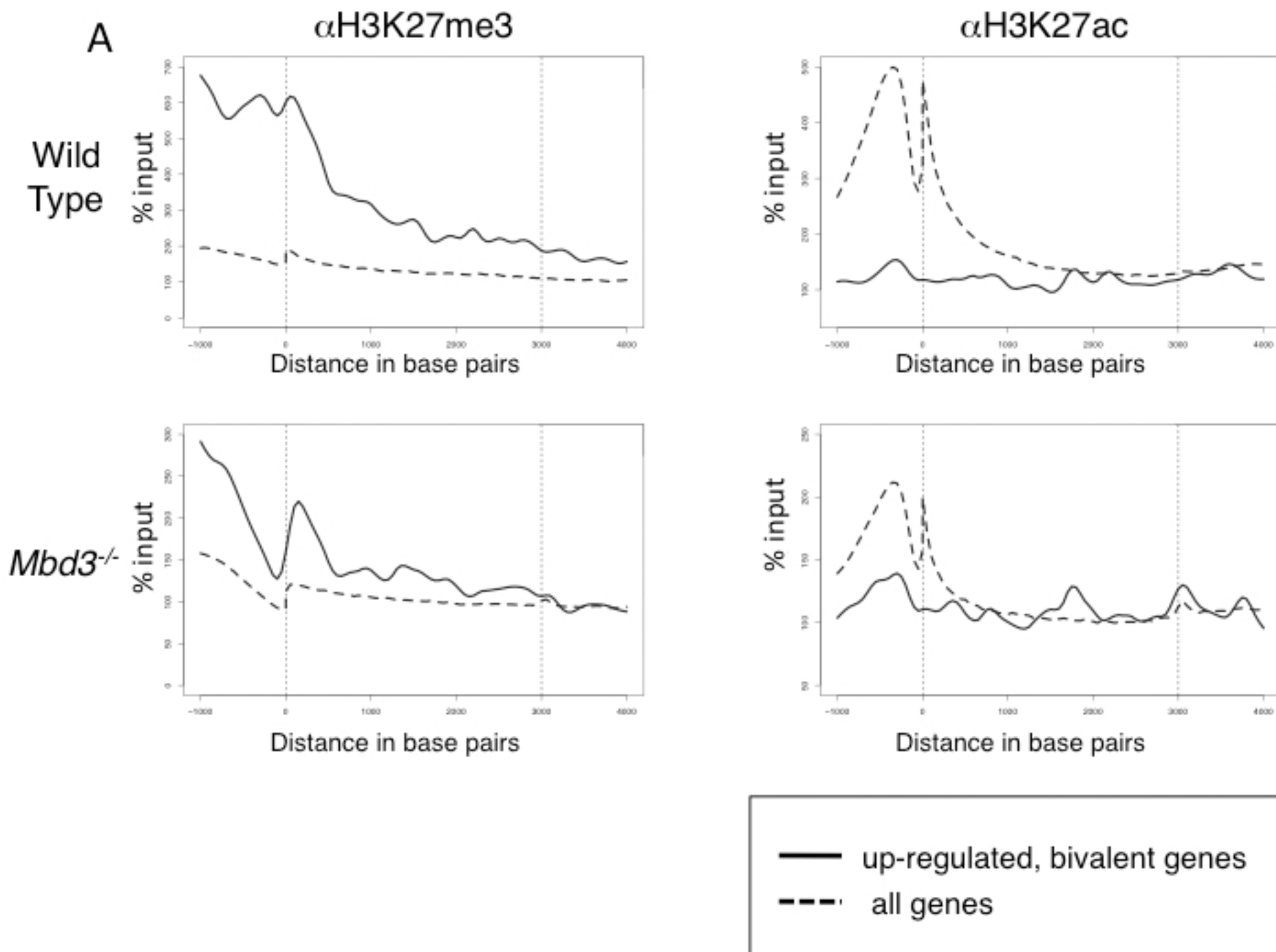
$\alpha$ Jarid2	Novus Biologicals	NB100-2214
$\alpha$ H3K4me3	Millipore	04-745
$\alpha$ H3K9ac	Millipore	06-942
$\alpha$ H3K9me3	Abcam	ab8898
$\alpha$ H3K14ac	L. O'Neill, University of Birmingham, UK.	
$\alpha$ H3K36ac	Millipore	07-540
$\alpha$ H4K16ac	L. O'Neill, University of Birmingham, UK.	
$\alpha$ H4K5ac	L. O'Neill, University of Birmingham, UK.	
$\alpha$ H3K27ac	Abcam	ab4729-25
$\alpha$ H3K27me3	Millipore	07-449
$\alpha$ H3	Abcam	ab1791
$\alpha$ tubulin	Santa Cruz	sc-5286
$\alpha$ ER $\alpha$	Santa Cruz	Sc-543
Rabbit IgG from serum	Sigma	I8140
Mouse IgG from serum	Sigma	I8765

### Supplementary References

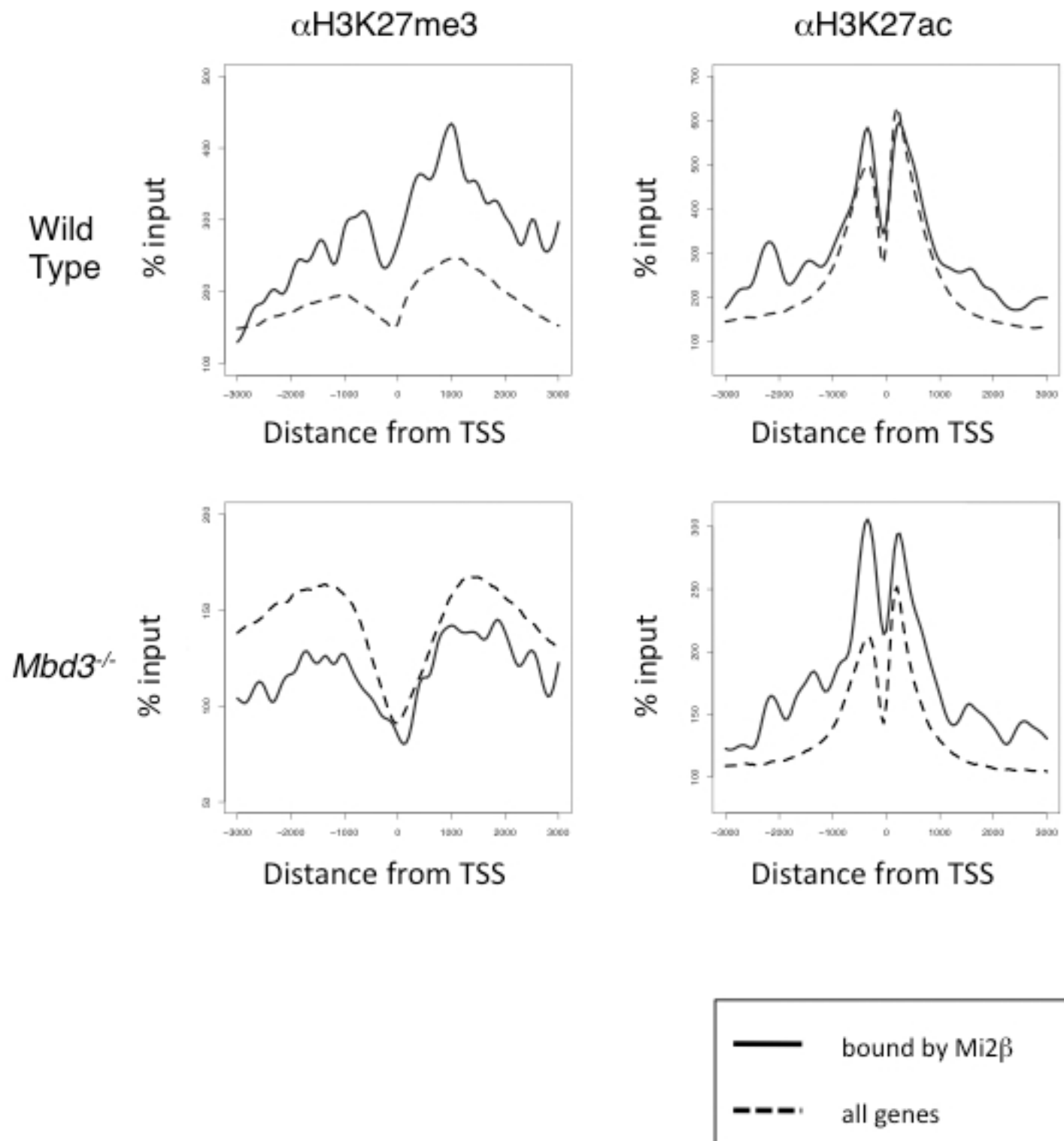
Leeb M, Pasini D, Novatchkova M, Jaritz M, Helin K, Wutz A (2010) Polycomb complexes act redundantly to repress genomic repeats and genes. *Genes & development* **24**: 265-276

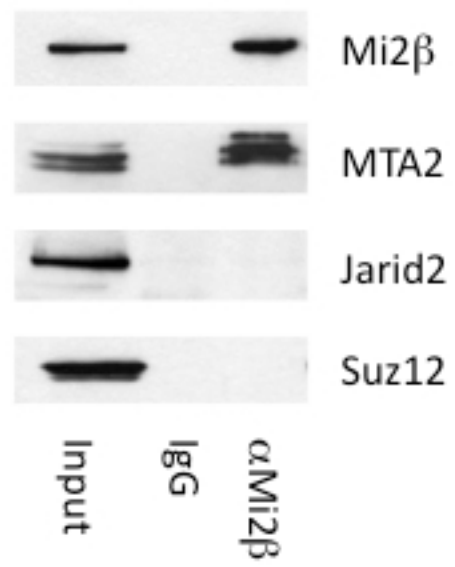
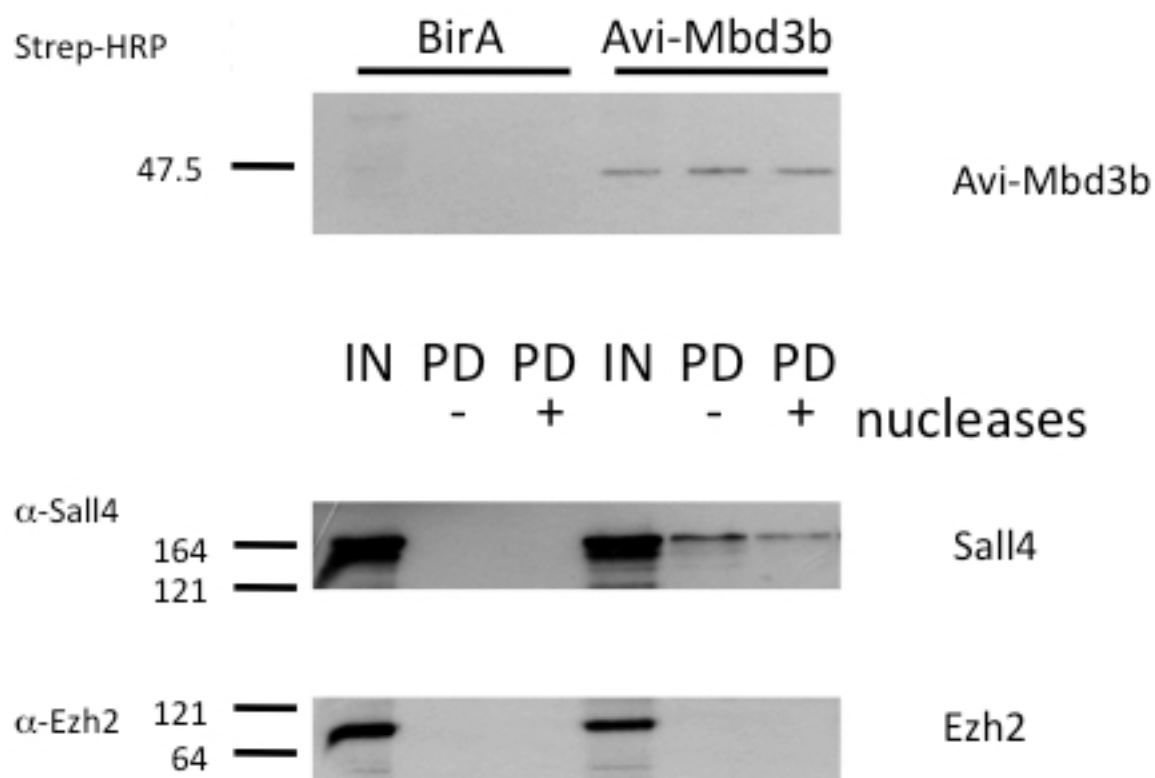
Peng JC, Valouev A, Swigut T, Zhang J, Zhao Y, Sidow A, Wysocka J (2009) Jarid2/Jumonji coordinates control of PRC2 enzymatic activity and target gene occupancy in pluripotent cells. *Cell* **139**: 1290-1302









**A****B****C**