The nucleotide sequence of Euglena cytoplasmic phenylalanine transfer RNA. Evidence for possible classification of Euglena among the animal rather than the plant kingdom

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ABSTRACT

INTRODUCTION

The origin of mitochondria and chloroplasts is the subject of much controversy. Perhaps the most widely accepted theory to explain their origin is the endosymbiotic hypothesis (1-3). However, an alternative hypothesis suggests that these organelles may have arisen internally from different genomes within an ancestral eukaryotic cell (4,5).

Previously we have reported the nucleotide sequence of the chloroplastic tRNA^{Phe} from <u>Euglena</u> (6-8). A comparison of the sequence of this tRNA with those of other tRNA^{Phe}s (three from prokaryotes and three from eukaryotic cytoplasms) showed that both in the nature of its modified nucleosides and in its sequence chloroplastic tRNA^{Phe} appears to be more homologous with prokaryotic tRNA^{Phes} (although all tRNA^{Phe} sequences show great homology) (9). These observations indicate that perhaps <u>Euglena</u> chloroplasts and extant prokaryotes shared a common ancestor. Such interpretations might be enhanced by comparing the nucleotide sequences of chloroplastic and cytoplasmic tRNA^{Phe}s from the same organism. In this paper we therefore present the nucleotide sequence of <u>Euglena</u> cytoplasmic tRNA^{Phe} and discuss its structure in terms of the origin of <u>Euglena</u> chloroplasts and the taxonomy of Euglena itself.

MATERIALS AND METHODS

Cytoplasmic tRNA^{Phe} from <u>Euglena</u> gracilis was purified as described previously (8). The nucleotide sequence of this tRNA was established using the scheme of <u>in vitro</u> labelling described previously (10-12).

RESULTS

Sequence Analysis. Euglena cytoplasmic tRNA was digested with Tl RNase or pancreatic RNase. The fragments produced were labelled at their 5'-ends. separated by two-dimensional high voltage electrophoresis and analyzed for their 5'-end and sequence. Derivation of the total sequence of this tRNA required, in addition, sequence information for the longer oligonucleotide fragments of the tRNA (13). These were obtained by partial digestion of the tRNA with pancreatic RNase, by specific chemical cleavage of the tRNA at the phosphodiester bond adjacent to m^7G and by partial nuclease Pl digestion on a tRNA sample labelled with ³²P at the 3'-end. The results of these analyses are summarized in Figure 1. The cloverleaf form (14) of Euglena cvtoplasmic tRNA^{Phe} is shown in Figure 2b. Except for a m⁵C, all the other modified nucleosides known to be present in the tRNA have been located in this sequence. The nucleosides at positions 13 and 14 are Cm and A respectively. These are different from those previously published for the same tRNA (7) which were C and $m^{6}A$ at the corresponding positions. This discrepancy is due to a misinterpretation of the two-dimensional homochromato-

² ₽GCCGACUUAĞCUC**LAGDDGGGAGAGCG**♥♥AGACLUGLAAYAYCUAAAGGUCCCUGGT♥CGĞUCCCGGGAG♥CGGCACCAoH



Figure 1. Sequence of Euglena cytoplasmic tRNA^{Phe} in linear form (top) and large fragments necessary for the establishment of the sequence. Fragments (a) through (d) are large oligonucleotides produced by partial pancreatic RNase digestion. The sequence shown in (e) was obtained by a partial nuclease Pl digestion of the 3'-end labelled tRNA. The sequence of fragment (f) was established by 3' half of the tRNA resulting from m^7G cleavage. Oligonucleotides (g) through (j) are those produced in complete Tl RNase digest, and (k) from complete pancreatic RNase digest.



Cloverleaf models of Euglena cytoplasmic tRNA^{Phe} (b) Euglena chloroplastic tRNA^{Phe} (a) and beef liver tRNA^{Phe} (c). Hatched areas indicate sequence differences when <u>Euglena</u> cytoplasmic tRNA^{Phe} is compared with the corresponding chloroplastic tRNA^{Phe} (a and b) and beef liver tRNA^{Phe} (b and c). Figure 2.

graphic patterns for the corresponding fragments during our earlier work.

For the purposes of comparison, the cloverleaf structures of <u>Euglena</u> chloroplastic tRNA^{Phe} (Fig. 2a) and that of mammalian cytoplasmic tRNA^{Phe} (Fig. 2c) are also shown in the same figure. Like 17 other tRNA^{Phe}s thus far sequenced, <u>Euglena</u> cytoplasmic tRNA^{Phe} also has a chain length of 76 nucleotides. Based on the modified nucleosides present in this tRNA and their location within the tRNA and its overall sequence homologies with other tRNA^{Phe}s (9) <u>Euglena</u> cytoplasmic tRNA^{Phe} is a typical eukaryotic tRNA. Thus it contains m¹A, m⁵C, m²G, m²G, and Y, modified nucleosides which are characteristic of eukaryotic tRNA^{Phe}s.

DISCUSSION

Besides the finding that the general features of Euglena cytoplasmic tRNA^{Phe} are more similar to those of eukaryotic cytoplasmic rather than prokaryotic tRNA^{Phe}s, the most surprising result of this work is that the sequence of Euglena cytoplasmic tRNA^{Phe} is virtually identical to that of mammalian cytoplasmic tRNAPhe (15-17) (Fig. 2b and c) and differs from it in only 4 out of 76 nucleotides. This extent of sequence homology (94.7%) is significantly higher than the homology of Euglena cytoplasmic tRNA to the corresponding tRNA from wheat germ or pea seedling (78.9%), baker's yeast (80.3%) and Schizosaccharomyces pombe cytoplasm (67.1%) (18). This result raises an interesting question about the taxonomy of Euglena, which because of their "plant-like" or "animal-like" characteristics has long been an enigma to biologists. The evidence presented here indicates that Euglena would more appropriately be classified as an animal rather than as a plant. It would clearly be of interest to investigate this further and examine whether this sequence homology between Euglena and mammalian cytoplasmic tRNA^{Phe} extends to other tRNAs as well.

The sequence of <u>Euglena</u> cytoplasmic tRNA^{Phe} is quite different from that of the corresponding chloroplastic tRNA. As shown in Figures 2a and 2b, these two tRNAs differ in 27 out of 76 nucleotides. This finding is in accord with the hypothesis that the chloroplastic genome and nuclear genome may have evolved from different ancestors (1-3). A similar difference between bean chloroplastic tRNA^{Phe} and bean leaf cytoplasmic tRNA^{Phe} has also been described (19).

In contrast to the substantial sequence differences between the chloroplastic and the corresponding cytoplasmic $tRNA^{Phe}s$, the two chloroplastic $tRNA^{Phe}s$ that have been sequenced have virtually identical sequences

| Sourse | B.sub- | E. | A. ni- | Bean | Eugl. | Yeast | Yeast | S. | Wheat | Mamm. | Eugl. |
|--|--------|--------------|----------------------|------------------------------|--------------------------------------|--|--|--|--|--|--|
| of tRNA | tilis | coli | dulans | Chl. | Chl. | mito. | cyto. | pombe | germ | | cyto |
| Mycoplasma B. subtilis E. coli A. nidulans Bean Chl. Eugl. Chl. Yeast mito. Yeast cyto. S. pombe Wheat germ Mammal | 82.9 | 72.4 75.8 | 78.9 81.6 84.2 | 69.7 76.3 78.9 86.8 | 68.4 71.8 70.0 82.9 93.4 | 57.8 56.6 63.1 55.3 60.5 63.1 | 63.2 60.5 63.2 59.2 59.2 61.8 61.8 | 55.3 64.5 56.6 59.2 59.2 61.8 56.9 64.5 | 64.5 65.8 64.6 69.7 67.1 69.7 61.8 84.2 64.5 | 61.8 63.2 71.0 69.7 67.1 67.1 60.5 77.6 72.4 81.6 | 61.8 64.5 68.4 67.1 64.5 59.2 80.3 67.1 78.9 94.7 |

TABLE I. Sequence Homology between Various Phenylalanine tRNA (%)^a

^{a.} Comparison of the number of identical parental nucleotides between two different tRNA^{Phes.}

(Table I), 93.4% homology (19). Furthermore, the two chloroplastic tRNAs are generally more homologous to prokaryotic tRNA^{Phe}s (68.4-86.8%) than to eukaryotic tRNA^{Phe} (59.2-69.7%) and within the prokaryotes more to blue green algae tRNA^{Phe} (Chang <u>et al</u>., unpublished results) than to the corresponding tRNA from mycoplasma, <u>B. subtilis or E. coli</u>. These results are in accord with the hypothesis that chloroplasts evolved from a common prokaryotic endosymbiont, which is more likely similar to current day blue green algae. Furthermore, analysis of oligonucleotides present in Tl RNase digests of red algae chloroplastic 16s rRNA (20) and a more recent paper on sequence of 3'-terminal of maize chloroplastic 16s rRNA (21) also provide independent evidence for the prokaryotic origin of chloroplasts.

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