

# Synthesis and Anticancer Activity of Sclerophytin-Inspired Isobenzofurans

*T. David Bateman, Aarti L. Joshi, Kwangyul Moon, Elena N. Galitovskaya, Meenakshi Upreti, Timothy C. Chambers, and Matthias C. McIntosh*

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**Materials and Instrumentation.** All commercially available compounds were purchased from Aldrich Chemical Co., Alfa Aesar Organics, or TCI and used as received unless otherwise noted. Ether, THF, CH<sub>3</sub>CN, and CH<sub>2</sub>Cl<sub>2</sub> were dried over alumina using the Solv-Tec<sup>®</sup> ST-002 solvent purification system. Microwave reactions were carried out using a CEM Discover<sup>®</sup> series reactor. Bruker 300 and 400 MHz spectrometers and a JEOL 270 MHz spectrometer were employed for NMR data collection.

## Experimental Procedures

**Anti-alcohol 3a.** To a solution of diisopropylamine (49.9 mmol, 1.5 eq) in dry THF (200 mL) was added n-BuLi (3.20 g, 49.9 mmol, 1.5 eq, 2.81 M in hexane) dropwise at -78 °C. After 15 min, (*S*)-(+)-carvone (5.00 g, 33.3 mmol, 1 eq) was added dropwise, followed after 30 min by slow addition of 2-bromobenzaldehyde (7.4 g, 39.9 mmol, 1.2 eq) over 45 min. The mixture was allowed to stir at -78 °C until no starting material was observed by TLC (~ 6 h). Glacial HOAc (3.00 g, 49.9 mmol, 1.5 eq) was then added to the reaction mixture at -78 °C. After ~5 min the reaction mixture was warmed to rt and diluted with ether (100 mL), washed with water (150 mL) and extracted with ether (3 x 100 mL). The combined organic extracts were then washed with brine, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The crude product was purified via flash chromatography over silica gel (10:90 EtOAc/hexanes) to deliver the desired *anti*-alcohol **3a** as a colorless oil in 77 % yield (8.56 g, 28.6 mmol). <sup>1</sup>H-NMR (270 MHz, CDCl<sub>3</sub>) δ 7.74–7.62 (d, *J*=7.55 Hz, 1H), 7.60–7.47 (d, *J*=7.99 Hz, 1H), 7.36–7.34 (t, *J*=7.55 Hz, 1H), 7.14–7.11 (t, 1H), 6.70 (s, 1H), 5.22–5.18 (t, *J*=6.15 Hz, 1H), 4.85 (s, 1H), 4.80 (s, 1H), 3.20–3.18 (d, *J*=5.93 Hz, 1H), 3.00–2.98 (m, 1H), 2.64–2.68 (m, 2H), 1.77 (s, 3H), 1.67 (s, 3H). <sup>13</sup>C-NMR (67 MHz, CDCl<sub>3</sub>) δ 200.7, 145.1, 143.7, 141.3, 135.3, 132.7, 129.3, 129.1, 127.6, 113.1, 72.5, 55.1, 44.3, 29.4, 20.9, 15.8. IR (film) 3435, 1659 cm<sup>-1</sup>. HRMS calculated for C<sub>17</sub>H<sub>19</sub>BrO<sub>2</sub>Na 357.0466, found 357.0466.

**Anti-alcohol 3b.** Prepared as for **3a**. The desired alcohol **3b** was obtained as a colorless oil in 58% yield (3.89 g, 11.6 mmol) from 3.00 g (20.0 mmol) of (*S*)-(+)-carvone. <sup>1</sup>H-NMR (270 MHz, CDCl<sub>3</sub>) δ 7.51 (s, 1H), 7.35–7.31 (m, 1H), 7.23–7.20 (t, *J*=7.79 Hz, 1H), 6.71–6.68 (m, 1H), 4.91–4.89 (d, *J*=6.38 Hz, 2H), 4.81–4.76 (dd, *J*=8.60, 3.50 Hz, 1H), 3.22 (d, *J*=8.86 Hz, 1H), 2.94–2.85 (m, 2H), 2.40–2.43 (m, 2H), 1.74 (s, 3H), 1.53 (s, 3H). <sup>13</sup>C-NMR (67 MHz, CDCl<sub>3</sub>) δ 202.50, 145.91, 144.64, 143.92, 135.95, 130.90, 130.50, 129.83, 129.61, 126.20, 122.04, 114.66, 77.34, 77.02, 76.71, 73.94, 53.62, 44.35, 31.26, 19.05, 15.63. Anal. (C<sub>17</sub>H<sub>19</sub>BrO<sub>2</sub>) C 60.94, H 5.66.

**Anti-alcohol 3c.** Prepared as for **3a**. The desired alcohol **3c** was obtained as a colorless oil in 75% yield (8.17 g, 25.1 mmol) from 5.00 g (33.2 mmol) of (*S*)-(+)-carvone. <sup>1</sup>H-NMR (270 MHz, CDCl<sub>3</sub>) δ 7.66–7.63 (d, 1H), 7.38–7.36 (d, 1H), 7.28–7.25 (t, 1H), 6.71–6.69 (m, 1H), 5.24–5.21 (dd, *J*=7.32 Hz, 4.55 Hz, 1H), 4.89–4.87 (m, 2H), 3.28–3.25 (d, *J*=7.32 Hz, 1H), 3.05–3.00 (dd, *J*=9.20, 4.55 Hz, 2H), 2.87–2.79 (m, 1H), 2.53–2.52 (m, 1H), 1.74 (s, 1H), 1.39 (s, 3H). <sup>13</sup>C-NMR (67 MHz, CDCl<sub>3</sub>) δ 202.50, 145.91, 144.64, 143.92, 135.95, 130.90, 130.50, 129.83, 129.61, 126.20, 122.04, 114.66, 77.34, 77.02, 76.71, 73.94, 53.62, 44.35, 31.26, 19.05, 15.63. Anal. (C<sub>17</sub>H<sub>18</sub>Cl<sub>2</sub>O<sub>2</sub>) C 62.65, H 5.64.

**Anti-alcohol 3d.** Prepared as for **3a**. The desired alcohol **3d** was obtained as a white solid in 66% yield (7.13 g, 21.9 mmol) from 5.00 g (33.2 mmol) of (*S*)-(+)-carvone; mp 110–112 °C; <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 7.65 (d, 1H), 7.31–7.24 (m, 2H), 6.74–6.65 (m, 1H), 5.15–5.08 (dd, *J*=7.32, 4.75 Hz, 1H), 4.88 (s, 1H), 3.26 (d, *J*=7.32 Hz, 1H), 3.01–2.95 (dd, *J*=4.75, 9.3 Hz, 1H), 2.89–2.79 (m, 1H), 2.49 (m, 2H), 1.74 (s, 6H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 200.65, 145.04, 144.81, 138.07, 135.55, 133.51, 133.15, 130.29, 128.92, 126.66, 113.62, 77.46, 77.04, 76.61, 69.46, 52.83, 43.19, 30.52, 19.12, 16.07.

**Anti-alcohol 3e.** Prepared as above for **3a**. The desired alcohol **3e** was obtained in 65% yield as a solid (2.22 g, 7.25 mmol) from 1.96 g (13.1 mmol) of (*S*)-(+)-carvone; mp 123-124 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.27 (d, J = 7.6 Hz, 1H), 8.13 (d, J = 7.5 Hz, 0H), 7.94 – 7.78 (m, 3H), 7.62 (d, J = 7.2 Hz, 1H), 7.60 – 7.41 (m, 5H), 6.72 (s, 1H), 5.58 (t, J = 6.1 Hz, 1H), 5.16 (s, 1H), 4.82 (d, J = 18.7 Hz, 2H), 3.38 (d, J = 6.5 Hz, 1H), 3.30 – 3.20 (m, 1H), 2.72 – 2.61 (m, 1H), 2.61 – 2.32 (m, 2H), 1.84 (d, J = 1.4 Hz, 3H), 1.61 (s, 3H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 201.86, 145.62, 143.77, 137.80, 135.55, 131.34, 128.85, 128.75, 126.52, 126.05, 125.59, 125.50, 123.84, 113.38, 71.11, 63.87, 44.66, 29.39, 21.09, 16.16. Anal. (C<sub>21</sub>H<sub>22</sub>O<sub>2</sub>) C 82.23, H, 7.36.

**Anti-alcohol 3f.** Prepared as above for **3a**. The desired alcohol **3f** was obtained in 28 % yield as an oil (1.92 g, 7.5 mmol) from 4.00 g (26.6 mmol) of (*S*)-(+)-carvone. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.50 (d, J = 4.8 Hz, 1H), 7.68 (td, J = 1.7, 7.7 Hz, 1H), 7.51 (d, J = 7.6 Hz, 1H), 7.13 (dd, J = 4.9, 7.4 Hz, 1H), 6.74 – 6.65 (m, 1H), 4.94 (dd, J = 10.4, 11.8 Hz, 2H), 4.84 (d, J = 7.9 Hz, 1H), 3.90 (d, J = 9.1 Hz, 1H), 3.50 (dd, J = 2.1, 12.4 Hz, 1H), 3.13 (ddd, J = 5.1, 10.5, 12.4 Hz, 1H), 2.65 – 2.47 (m, 1H), 2.39 (dt, J = 5.2, 9.5 Hz, 1H), 1.85 (s, 3H), 1.68 (s, 3H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 200.91, 163.01, 148.18, 145.60, 144.46, 136.58, 135.88, 121.65, 119.69, 114.48, 72.07, 54.52, 46.27, 31.26, 19.12, 15.84. Anal. (C<sub>16</sub>H<sub>19</sub>NO<sub>2</sub>) C 74.45, H, 7.50.

**Anti-alcohol 3g.** Prepared as for **3a**. The desired alcohol **3g** was obtained as an oil in 47% yield (1.15 g, 4.7 mmol) from 1.5 g (10.0 mmol) of (*S*)-(+)-carvone. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 7.30 (dd, J = 0.8, 1.8 Hz, 1H), 6.78 (ddd, J = 1.3, 2.4, 6.1 Hz, 1H), 6.29 (dd, J = 1.8, 3.2 Hz, 1H), 6.16 (d, J = 3.3 Hz, 1H), 5.20 (d, J = 10.9 Hz, 1H), 4.91 – 4.83 (m, 1H), 4.75 (dd, J = 4.5, 11.0 Hz, 2H), 2.95 (dd, J = 4.4, 12.8 Hz, 1H), 2.64 – 2.50 (m, 1H), 2.50 – 2.34 (m, 1H), 2.33 – 2.19 (m, 1H), 1.81 (dt, J = 1.3, 2.4 Hz, 3H), 1.74 (s, 4H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 202.69, 154.94, 154.81, 144.03, 141.68, 135.78, 114.37, 110.23, 108.17, 68.24, 52.10, 45.29, 31.17, 18.74, 15.74. Anal. (C<sub>15</sub>H<sub>18</sub>O<sub>3</sub>) C 73.39, H, 7.43.

**Anti-alcohol 3h.** Prepared as for **3a**. The desired alcohol **3h** was obtained in 50% yield as an oil (1.79 g, 6.52 mmol) from 1.96 g (13.1 mmol) of (*S*)-(+)-carvone. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 7.35 (td, J = 1.7, 7.6 Hz, 1H), 7.30-7.14 (m, 1H), 7.07 (dt, J = 3.8, 7.5 Hz, 1H), 6.96 (ddd, J = 1.2, 8.2, 10.6 Hz, 1H), 6.72 (ddd, J = 1.4, 3.6, 4.9 Hz, 1H), 5.39 (dd, J = 5.1, 8.1 Hz, 1H), 4.71 (dd, J = 12.7, 14.1 Hz, 2H), 4.12 (d, J = 8.2 Hz, 1H), 3.03 (dd, J = 5.1, 9.8 Hz, 1H), 2.84 – 2.68 (m, 1H), 2.51 – 2.25 (m, 2H), 1.77 (s, 3H), 1.58 (s, 3H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 202.01, 161.92, 158.66, 145.11, 135.90, 129.28, 129.16, 129.11, 128.99, 124.07, 115.49, 115.19, 113.26, 67.87, 53.89, 43.54, 30.52, 19.83, 16.04. Anal. (C<sub>17</sub>H<sub>19</sub>FO<sub>2</sub>) C 74.70, H 7.03.

**Glycolate 4a.** To a stirring solution of alcohol **3a** (8.65 g, 25.80 mmol) in anhydrous DMF (75mL) was added Ag<sub>2</sub>O (9.00 g, 38.7 mmol, 1.5 eq), followed by dropwise addition of ethyl bromoacetate (6.46 g, 38.7 mmol, 1.5 eq) at rt. After stirring ~10 min, 2,6-lutidine (4.15 g, 38.7 mmol) was added via syringe pump (~2 mL/h) and stirring was continued for 24 h at rt. The crude mixture was then filtered through a short silica gel column eluted with diethyl ether (100 mL). The filtrate was washed with 3N HCl (100 mL) and extracted with hexane (3x60 mL). The combined extracts were washed sequentially with saturated aqueous NaHCO<sub>3</sub> and brine, then dried over MgSO<sub>4</sub>, and concentrated in vacuo. The crude product was purified via flash chromatography over silica gel (20:80 EtOAc/hexanes) to deliver glycolate **4a** as a yellow oil in 77 % yield (8.47 g, 20.1 mmol). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.57 (dd, J = 1.7, 7.8 Hz, 1H),

7.48 (dd, J = 1.1, 8.0 Hz, 1H), 7.36 (t, J = 7.5 Hz, 1H), 7.23 – 7.10 (m, 1H), 6.69 (s, 1H), 5.45 (d, J = 7.0 Hz, 1H), 4.71 (dd, J = 8.1, 9.3 Hz, 3H), 4.33 – 4.14 (m, 3H), 4.02 (d, J = 16.4 Hz, 1H), 3.84 (d, J = 16.4 Hz, 1H), 3.33 (s, 1H), 3.08 – 2.89 (m, 2H), 2.45 (d, J = 19.3 Hz, 1H), 1.75 (d, J = 1.4 Hz, 3H), 1.57 (s, 3H), 1.37 – 1.21 (m, 3H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) 198.3, 169.9, 141.7, 129.8, 129.0, 128.8, 128.1, 127.6, 112.2, 80.7, 77.6, 77.1, 76.6, 65.6, 64.9, 60.7, 55.4, 43.5, 28.4, 21.4, 16.3, 14.2. IR (film) 1750, 1671 cm<sup>-1</sup>. HRMS Calcd for C<sub>21</sub>H<sub>25</sub>BrNaO<sub>4</sub>: 443.0834, found 443.0830.

**Glycolate 4b.** Prepared as for **4a**. Glycolate **4b** was obtained as a colorless oil in 77% yield (1.56 g, 3.70 mmol) from 1.61 g (4.80 mmol) of alcohol **3b**. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 7.44 – 7.34 (m, 2H), 7.20 (q, J = 7.7 Hz, 2H), 6.69 (s, 1H), 4.89 (d, J = 7.0 Hz, 1H), 4.74 (s, 1H), 4.67 (s, 1H), 4.26 – 4.13 (m, 2H), 4.08 (d, J = 16.6 Hz, 1H), 3.84 (d, J = 16.6 Hz, 1H), 3.25 (s, 1H), 3.04 – 2.90 (m, 1H), 2.80 (dd, J = 4.0, 7.0 Hz, 1H), 2.44 (d, J = 19.6 Hz, 1H), 1.71 (s, 4H), 1.58 (d, J = 14.2 Hz, 6H), 1.27 (t, J = 7.1 Hz, 6H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) 197.8, 170.1, 146.1, 143.4, 138.0, 135.5, 132.6, 129.6, 129.4, 127.6, 112.3, 79.4, 72.6, 68.0, 66.2, 61.0, 55.7, 41.5, 28.2, 21.2, 16.3, 14.3.

**Glycolate 4c.** Prepared as for **4a**. Glycolate **4c** was obtained as a colorless oil in 83% yield (7.41 g, 18.0 mmol) from 7.06 g of alcohol **3c** (21.7 mmol). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.55 (dd, J = 1.5, 7.9 Hz, 1H), 7.45 (dd, J = 1.6, 7.9 Hz, 1H), 7.30 (dd, J = 4.7, 12.7 Hz, 2H), 6.67 (s, 1H), 5.22 (d, J = 5.7 Hz, 1H), 4.83 (dd, J = 7.9, 9.2 Hz, 2H), 4.28 – 4.10 (m, 2H), 4.03 (d, J = 16.3 Hz, 1H), 3.73 (d, J = 16.3 Hz, 1H), 2.99 (t, J = 5.6 Hz, 1H), 2.89 – 2.69 (m, 2H), 2.43 (d, J = 18.5 Hz, 1H), 1.82 (s, 3H), 1.73 (s, 3H), 1.27 (t, J = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 197.71, 169.62, 145.56, 142.22, 139.41, 135.71, 132.88, 131.37, 129.87, 127.68, 127.47, 112.89, 79.45, 77.35, 77.03, 76.71, 66.40, 60.83, 54.26, 44.36, 28.82, 20.74, 16.19, 14.18.

**Glycolate 4d.** Prepared as for **4a**. Glycolate **4d** was obtained as a colorless oil in 80% yield (9.60 g, 23.3 mmol) from 9.48 g (29.2 mmol) of alcohol **3d**. <sup>1</sup>H-NMR (270 MHz, CDCl<sub>3</sub>) δ 7.53–7.49 (d, 1H), 7.44–7.40 (d, 1H), 7.30–7.26 (m, 1H), 6.64 (s, 1H), 5.20–5.18 (d, 1H), 4.81–4.76 (d, 2H) 4.20–4.11 (m, 2H), 4.03–3.97 (d, J=16.03 Hz, 1H), 3.72–3.66 (d, J=16.03 Hz, 1H), 3.31–3.25 (m, 1H), 2.94–2.90 (m, 2H), 2.44–2.34 (m, 1H), 1.72, (s, 3H), 1.48 (s, 3H), 1.31–1.26 (t, J=7.07 Hz, 3H). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) δ 198.12, 170.14, 145.91, 143.38, 141.50, 135.24, 131.18, 130.27, 129.96, 125.78, 122.56, 112.21, 80.74, 77.33, 77.01, 76.70, 66.00, 60.85, 57.54, 41.25, 29.71, 28.11, 21.23, 16.14, 14.16.

**Glycolate 4e.** Prepared as for **4a**. Glycolate **4e** was obtained as an oil in 25% yield (8.17 g, 25.13 mmol) from *anti* alcohol **1g** (3.27 mmol). <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 8.09 (d, J = 7.6 Hz, 1H), 7.91 – 7.83 (m, 1H), 7.79 (d, J = 8.2 Hz, 1H), 7.60 (d, J = 7.0 Hz, 1H), 7.56 – 7.41 (m, 3H), 6.75 (s, 1H), 5.86 (d, J = 5.1 Hz, 1H), 4.59 (s, 2H), 4.13 (d, J = 16.4 Hz, 1H), 3.92 (d, J = 10.3 Hz, 1H), 3.70 (s, 3H), 3.28 (dd, J = 4.4, 9.8 Hz, 1H), 3.09 – 2.90 (m, 2H), 2.47 – 2.31 (m, 1H), 1.72 (s, 3H), 1.27 (s, 3H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 198.92, 170.89, 146.63, 144.20, 135.41, 134.25, 133.96, 131.11, 129.22, 128.72, 126.54, 125.73, 125.26, 125.22, 122.98, 112.11, 79.47, 66.40, 55.66, 51.85, 41.25, 29.00, 20.97, 16.37. HRMS Calcd for C<sub>24</sub>H<sub>26</sub>O<sub>4</sub>Na: 401.1729, found: 401.1719.

**Glycolate 4f.** Prepared as for **4a**. Glycolate **4f** was obtained as an oil in 86% yield (1.43 g, 4.34 mmol) from 1.3g (5.05 mmol) of alcohol **3f**. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 8.50 (d, J = 4.8, 1H), 7.70 (dt, J = 7.0, 20.6 Hz, 2H), 7.23 – 7.10 (m, 1H), 6.63 (s, 1H), 4.83 (s, 1H), 4.75 (d, J = 5.2 Hz, 1H), 4.20 (d, J = 16.2 Hz, 1H), 3.93 (d, J = 16.2 Hz, 1H), 3.69 (s, 3H), 3.17 (dd, J = 5.2, 8.0 Hz, 1H), 2.89 – 2.70 (m, 1H), 2.50 (m, 2H), 1.80 – 1.71 (m, 6H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 197.99, 170.30, 160.59, 148.74, 145.81, 142.32, 136.52, 135.60, 122.39, 121.32, 113.28, 83.42, 67.59, 55.15, 51.80, 44.20, 29.28, 20.03, 16.15. Anal. (C<sub>19</sub>H<sub>23</sub>NO<sub>4</sub>) C 68.99, H 7.09.

**Glycolate 4g.** Prepared as for **4a**. Glycolate **4g** was obtained as an oil in 48% yield (679 mg, 2.13 mmol) from 1.1 g (4.46 mmol) of alcohol **3g**. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 7.40 (dd, J = 0.7 Hz, 1.7, 1H), 6.68 – 6.59 (m, 1H), 6.33 – 6.20 (m, 2H), 4.88 (d, J = 7.6 Hz, 1H), 4.74 (d, J = 1.2 Hz, 1H), 4.66 (s, 1H), 4.12 (d, J = 16.8 Hz, 1H), 3.96 – 3.87 (m, 1H), 3.73 – 3.66 (m, 3H), 3.21 (dd, J = 4.4, 9.4 Hz, 1H), 3.09 (dd, J = 4.4, 7.5 Hz, 1H), 2.98 – 2.81 (m, 1H), 2.42 (ddd, J = 2.5, 5.6, 19.6 Hz, 1H), 1.68 (s, 7H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 198.03, 171.00, 151.32, 145.97, 143.28, 143.06, 112.17, 110.34, 110.18, 74.74, 65.50, 54.79, 51.92, 41.73, 28.02, 21.52, 16.28. Anal. (C<sub>18</sub>H<sub>22</sub>O<sub>5</sub>) C 67.71, H 6.96.

**Glycolate 4h.** Prepared as for **4a**. Glycolate **4h** was obtained as an oil in 19 % yield (0.25 g, 0.71 mmol) from 1.05 g of alcohol **3h** (3.8 mmol). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.44 (td, J = 1.8, 7.5 Hz, 1H), 7.31 – 7.21 (m, 2H), 7.15 (t, J = 7.0 Hz, 1H), 6.97 (dd, J = 8.4, 10.1 Hz, 1H), 6.66 (s, 1H), 5.26 (d, J = 7.6 Hz, 1H), 4.68 (d, J = 11.6 Hz, 2H), 4.08 (d, J = 16.5 Hz, 1H), 3.87 (d, J = 16.5 Hz, 1H), 3.73 (s, 3H), 3.31 (s, 1H), 3.06 – 2.80 (m, 2H), 2.43 (d, J = 19.6 Hz, 1H), 1.72 (s, 3H), 1.59 (s, 3H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 198.18, 170.72, 162.47, 159.21, 146.07, 143.16, 135.39, 129.84, 128.75, 126.14, 125.97, 124.48, 124.43, 115.46, 115.18, 112.25, 74.74, 66.05, 56.64, 51.90, 41.50, 27.91, 21.48, 16.24. Anal. (C<sub>20</sub>H<sub>23</sub>FO<sub>4</sub>) C 68.98, H 6.61.

**Isobenzofuran 5a.** To a solution of glycolate **4a** (4.13 g, 9.80 mmol) in dry THF (100 mL) was added KHMDS (2.35 g, 11.8 mmol, 1.1 eq, 0.5 M soln. in toluene) quickly at -78 °C, followed immediately by rapid addition of 1.2 eq HOAc (0.71 g, 11.8 mmol). The reaction mixture was allowed to warm to rt, then water (100 mL) was added. The organic layer was extracted with ether (3 x 60 mL). The combined organic extracts were washed with brine and dried over MgSO<sub>4</sub>, filtered and concentrated in vacuo. The crude product was purified via flash chromatography over silica gel (10:90 EtOAc/hexanes) to yield cycloaldol product **5a** as a colorless oil in 80% yield (3.30 g, 7.84 mmol). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.28 (dd, J = 1.6, 7.9 Hz, 1H), 7.52 (d, J = 8.0, 1H), 7.43 (t, J = 7.6, 1H), 7.22 – 7.14 (m, 1H), 5.78 (s, 1H), 5.24 (d, J = 8.8 Hz, 1H), 4.84 (d, J = 3.3 Hz, 2H), 4.46 (s, 1H), 4.37 – 4.22 (m, 2H), 3.08 (s, 1H), 2.76 (dd, J = 5.3, 8.8 Hz, 1H), 2.54 (d, J = 15.8 Hz, 1H), 2.31 (d, J = 4.6 Hz, 2H), 1.86 (s, 3H), 1.58 (s, 3H), 1.35 (t, J = 7.1 Hz, 3H). <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>) 171.0, 147.0, 139.4, 132.9, 132.4, 130.4, 129.7, 128.3, 125.6, 123.9, 112.2, 82.6, 81.9, 80.7, 61.3, 55.5, 39.1, 27.7, 21.4, 17.7, 14.2. IR (film) 3536, 1743 cm<sup>-1</sup>. HRMS Calcd for C<sub>21</sub>H<sub>25</sub>BrO<sub>4</sub>Na<sup>+</sup>: 443.0834, found 443.0834.

**Isobenzofuran 5b.** Prepared as for **5a**. Isobenzofuran **5b** was obtained as a white solid in 73% yield (0.56 g, 1.34 mmol) from 0.77g (1.83 mmol) of glycolate **4b**. mp 131-136 °C; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 7.51 (s, 1H), 7.34 (d, J = 7.2 Hz, 1H), 7.20 (d, J = 7.1 Hz, 1H), 7.08

(d,  $J = 7.7$  Hz, 1H), 5.70 (s, 1H), 5.60 (s, 1H), 4.64 (s, 1H), 4.30 (d,  $J = 4.0$  Hz, 2H), 4.20 (d,  $J = 15.9$  Hz, 2H), 3.12 (s, 1H), 2.74 – 2.58 (m, 1H), 2.46 – 2.29 (m, 1H), 2.00-1.77 (m, 6H), 1.61 (s, 1H), 1.42 – 1.26 (m, 7H).  $^{13}\text{C}$ -NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  171.41, 145.68, 141.05, 133.43, 130.42, 130.26, 128.94, 125.89, 125.72, 121.90, 111.83, 83.50, 83.46, 82.88, 77.35, 77.03, 76.71, 61.53, 53.84, 40.35, 31.67, 19.40, 18.00, 14.15. Anal. ( $\text{C}_{21}\text{H}_{25}\text{BrO}_4$ ) C 60.13, H 6.04.

**Isobenzofuran 5c.** Prepared as for **5a**. Isobenzofuran **5c** was obtained as a pale yellow oil in 77% yield (2.61 g, 6.34 mmol) from 3.39 g (8.24 mmol) of glycolate **4c**.  $^1\text{H}$  NMR (270 MHz,  $\text{CDCl}_3$ )  $\delta$  8.22 (dd,  $J = 1.7, 7.8$  Hz, 1H), 7.38 (dd,  $J = 1.7, 7.9$  Hz, 1H), 7.27 (t,  $J = 7.8$  Hz, 1H), 5.70 (s, 1H), 5.30 (d,  $J = 8.2$  Hz, 1H), 4.80 (s, 3H), 4.42 (s, 1H), 4.25 (qd,  $J = 2.7, 7.1$  Hz, 2H), 3.06 (s, 1H), 2.74 – 2.59 (m, 1H), 2.27 (s, 3H), 1.79 (s, 3H), 1.53 (s, 3H), 1.29 (t,  $J = 7.1$  Hz, 3H).  $^{13}\text{C}$ -NMR (67 MHz,  $\text{CDCl}_3$ ) 171.1, 146.7, 140.8, 132.9, 132.6, 131.5, 130.1, 128.4, 128.1, 125.7, 112.5, 81.9, 80.9, 80.7, 61.4, 55.4, 39.7, 27.7, 21.3, 17.7, 14.2.

**Isobenzofuran 5d.** Prepared as for **5a**. Isobenzofuran **5d** was obtained as a pale yellow oil in 74% yield (3.50 g, 8.51 mmol) from 4.73 g (11.5 mmol) of glycolate **4d**.  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.27 (d,  $J = 9.1$ , 1H), 7.41 – 7.34 (m, 2H), 5.77 (s, 1H), 5.26 (d,  $J = 8.3$  Hz, 1H), 4.85 (s, 2H), 4.46 (s, 1H), 4.31 (ddd,  $J = 5.6, 8.3, 16.5$  Hz, 3H), 3.16 (s, 1H), 2.69 (dd,  $J = 5.9, 8.3$  Hz, 1H), 2.36 (m, 4H), 1.85 (d,  $J = 1.5$  Hz, 4H), 1.58 (d,  $J = 6.7$  Hz, 9H), 1.42 – 1.21 (m, 9H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  171.10, 146.70, 136.85, 134.34, 132.78, 131.21, 128.77, 128.06, 126.96, 125.72, 112.41, 81.72, 80.77, 79.71, 77.33, 77.21, 77.01, 76.70, 61.46, 59.84, 55.17, 39.68, 27.73, 21.23, 17.68, 14.13.

**Isobenzofuran 5e.** Prepared as for **5a**. Isobenzofuran **5e** was obtained in 59 % yield (0.22 g, 0.59 mmol) as an oil from 0.41 g (1.08 mmol) of glycolate **4e**.  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.34 (d,  $J = 8.0$  Hz, 1H), 8.16 (d,  $J = 7.2$  Hz, 1H), 7.86 (t,  $J = 9.1$  Hz, 2H), 7.52 (dt,  $J = 6.9, 14.5$  Hz, 3H), 5.76 (s, 1H), 5.62 (d,  $J = 7.2$  Hz, 1H), 4.91 (s, 1H), 4.83 (s, 1H), 4.59 (s, 1H), 3.85 (s, 3H), 3.26 (s, 1H), 3.03 (t,  $J = 7.2$  Hz, 1H), 2.39 (dd,  $J = 6.5, 12.7$  Hz, 1H), 2.29 (s, 2H), 1.87 (s, 3H), 1.55 (s, 3H).  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.29, 146.85, 135.72, 133.92, 133.46, 131.85, 128.99, 128.86, 126.11, 125.74, 125.63, 125.60, 125.52, 123.60, 113.03, 82.77, 81.51, 80.99, 53.68, 52.36, 42.04, 28.22, 20.68, 17.79. Anal. ( $\text{C}_{24}\text{H}_{26}\text{O}_4$ ) C 76.09, H 6.95.

**Isobenzofuran 5f.** Prepared as for **5a**. Isobenzofuran **5f** was obtained in 81 % yield (0.30 g, 0.90 mmol) as a solid from 0.36 g (1.10 mmol) of glycolate **4f**; mp 67-69 °C;  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.53 (d,  $J = 4.0$  Hz, 1H), 7.64 (td,  $J = 1.8, 7.7$  Hz, 1H), 7.19 (ddd,  $J = 3.6, 6.0, 7.8$  Hz, 2H), 5.58 (d,  $J = 1.5$  Hz, 1H), 4.88 (s, 1H), 4.82 (s, 1H), 4.55 (s, 1H), 3.69 (s, 3H), 2.55 – 2.32 (m, 2H), 2.07 (d,  $J = 3.1$ , 2H), 1.89 – 1.73 (m, 3H), 1.50 (s, 3H).  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  170.36, 159.63, 149.19, 146.18, 137.80, 133.65, 125.22, 123.42, 123.23, 113.50, 87.12, 84.83, 82.53, 54.58, 52.07, 46.56, 30.98, 19.36, 17.95. Anal. ( $\text{C}_{19}\text{H}_{23}\text{NO}_4$ ) C 68.98, H 6.86.

**Methyl ester 5a.i.** To a solution of ethyl ester **5a** (0.10 g, 0.24 mmol) in 15 mL of methanol was added 0.04 g (0.28 mmol, 1.2 eq) of  $\text{K}_2\text{CO}_3$  at room temperature. The reaction mixture was stirred at rt until no starting material was observed by TLC (ca. 20 min). The mixture was then diluted with  $\text{H}_2\text{O}$  (10 mL) and extracted with  $\text{CH}_2\text{Cl}_2$  (3 x 20 mL). The combined organic extracts were then washed with brine (50 mL), extracted with  $\text{CH}_2\text{Cl}_2$  (40 mL), dried over

MgSO<sub>4</sub>, and concentrated in vacuo. The crude product was filtered through a short silica gel column (EtOAc/Hexanes 1:1) to deliver methyl ester **5a.i** as a colorless oil in 97 % yield (94.8 mg, 23.3 mmol). <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 8.30 (dd, J = 1.7, 7.9 Hz, 1H), 7.54 (dd, J = 1.2, 8.0 Hz, 1H), 7.45 (t, J = 7.6 Hz, 1H), 7.24 – 7.15 (m, 1H), 5.79 (s, 1H), 5.25 (d, J = 9.2 Hz, 1H), 4.90 – 4.81 (m, 2H), 4.51 (s, 1H), 3.85 (s, 3H), 2.93 (s, 1H), 2.77 (dd, J = 4.9, 9.2 Hz, 1H), 2.66 – 2.51 (m, 1H), 2.32 (d, J = 15.0 Hz, 2H), 1.88 (d, J = 1.6 Hz, 3H), 1.58 (d, J = 5.8 Hz, 5H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 171.54, 147.20, 139.28, 132.82, 132.46, 130.27, 129.79, 128.39, 125.67, 123.84, 112.00, 82.51, 81.76, 80.50, 77.47, 77.05, 76.62, 73.47, 55.22, 52.22, 38.31, 27.49, 21.61, 17.67. Anal. (C<sub>20</sub>H<sub>23</sub>BrO<sub>4</sub>) C 59.16, H 5.80.

**Cyclopropylmethyl ester 5a.ii.** A mixture of ethyl ester **5a** (0.14 g, 0.33 mmol), cyclopropyl methanol (2.66 mL, 33.22 mmol, 100 eq), and Bu<sub>2</sub>SnO (0.06 g, 0.24 mmol, 0.75eq) was sealed in an 8 mL microwave reaction vessel. The mixture was irradiated for 30 min. at 150 °C and 300 W with continuous stirring. The mixture was then cooled to rt, diluted with ethyl acetate (10 mL), washed with sat. NaHCO<sub>3</sub> (30 mL) and extracted with EtOAc (3x20 mL). The combined organic extracts were then washed with brine, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The crude product was purified via flash chromatography over silica gel (10:90 EtOAc/hexanes) to deliver ester **5a.ii** as a colorless oil in 84% yield (0.125 g, 0.28 mmol). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.29 (dd, J = 1.7, 7.9 Hz, 1H), 7.53 (dd, J = 1.2, 8.0 Hz, 1H), 7.43 (t, J = 7.6 Hz, 1H), 7.24 – 7.11 (m, 1H), 5.78 (s, 1H), 5.26 (d, J = 8.6 Hz, 1H), 4.85 (s, 2H), 4.49 (s, 1H), 4.16 – 3.99 (m, 2H), 3.16 (s, 1H), 2.77 (dd, J = 5.7, 8.6 Hz, 1H), 2.53 (d, J = 15.6 Hz, 1H), 2.30 (d, J = 10.1 Hz, 2H), 2.19 (s, 1H), 1.88 (d, J = 1.4 Hz, 3H), 1.59 (d, J = 6.8 Hz, 3H), 1.30 – 1.13 (m, 2H), 0.62 (dt, J = 5.4, 5.9 Hz, 2H), 0.35 (q, J = 4.7 Hz, 2H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) 171.0, 146.8, 139.4, 132.9, 132.4, 130.4, 129.7, 128.3, 125.7, 112.2, 82.7, 81.9, 80.7, 70.2, 55.5, 39.5, 27.8, 21.27, 17.7, 9.6, 3.4.

**Cyclopentylmethyl ester 5a.iii.** Prepared as for **5a.ii**. Ester **5a.iii** was obtained in 91% yield as an oil (0.11 g, 0.24 mmol) from 0.11 g (0.26 mmol) of ester **5a**. <sup>1</sup>H-NMR (270 MHz, CDCl<sub>3</sub>) 8.29 (d, J = 6.2 Hz, 1H), 7.53 (d, J = 8.0 Hz, 1H), 7.44 (t, J = 7.5 Hz, 1H), 7.19 (dd, J = 4.5, 10.8 Hz, 1H) 5.79 (s, 1H), 5.25 (d, J = 9.0 Hz, 1H), 4.85 (s, 2H), 4.49 (s, 1H), 4.17 – 4.04 (m, 2H), 2.99 (s, 1H), 2.76 (dd, J = 5.0, 9.1 Hz, 1H), 2.64-2.52 (m, 1H), 2.32 (d, J = 7.3 Hz, 3H), 1.89-1.74 (m, 6H), 1.68-1.54 (m, 8H), 1.39-1.26 (m, 3H). <sup>13</sup>C-NMR (67 MHz, CDCl<sub>3</sub>) 171.1, 147.1, 139.5, 132.9, 132.4, 130.3, 129.7, 128.3, 125.5, 123.8, 112.0, 82.5, 81.9, 80.6, 69.3, 55.5, 38.5, 38.3, 29.3, 27.5, 25.4, 21.5, 17.6.

**Acid 5a.iv.** To a solution of 3:2:1 THF/H<sub>2</sub>O/MeOH and ester **5a** (0.18 g, 0.43 mmol) was added 0.02 g (0.85 mmol, 2 eq) of LiOH at rt. The reaction mixture was allowed to stir at rt until no starting material was observed by TLC (~1.5 h), then acidified to pH 2 with 3N HCl. The mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (30 mL), extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 x 30 mL), washed with brine (40 mL), dried over MgSO<sub>4</sub> and concentrated in vacuo to yield a yellow foam. The crude material was eluted through a plug of silica gel (1:1 EtOAc/hexanes) to deliver acid **5a.iv** as a white solid in 95% yield (0.16 g, 0.41 mmol); mp 52-56 °C. <sup>1</sup>H-NMR (270 MHz, CDCl<sub>3</sub>) δ 8.14- 8.09 (d, J=7.86 Hz, 1H), 7.59-7.54 (d, J=8.03 Hz, 1H), 7.47-7.40 (t, J=7.55 Hz, 1H), 7.25-7.18 (dd, J=4.47, 10.85 Hz, 1H), 5.81 (s, 1H), 5.28 (s, 1H), 5.25 (d, J=8.87 Hz, 1H), 4.85 (s, 2H), 4.51 (s, 1H), 2.79-2.72 (dd, J=5.18, 8.84 Hz, 1H), 2.60-2.47 ( m, 1H), 2.35-2.24 (m, 2H), 1.94 (s,

3H), 1.59 (s, 3H). <sup>13</sup>C-NMR (67 MHz, CDCl<sub>3</sub>) 146.6, 138.6, 132.9, 132.7, 130.0, 129., 128.4, 125.9, 124.1, 112.4, 82.7, 55.6, 27.6, 21.4, 17.7. IR (film) 3450, 1638 cm<sup>-1</sup>.

**Diene 6a.** Preparation was carried out as above for **5a** from glycolate **4a** (3.5 g, 8.31 mmol), but the reaction mixture was allowed to stir for 5 minutes before quenching with HOAc. Purification of the crude product via flash chromatography over silica gel (10:90 EtOAc/hexanes) delivered diene **6a** as a yellow oil in 15 % yield (0.42 g, 1.27 mmol) as well as isobenzofuran **5a** in 41% yield (1.43 g, 3.41 mmol). <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 7.64 (d, J = 6.5, 1H), 7.34 – 7.08 (m, 3H), 6.62 (s, 1H), 4.95 (s, 1H), 4.73 (s, 1H), 3.62 (s, 1H), 2.74 – 2.48 (m, 1H), 2.19 (s, 1H), 1.95 – 1.82 (m, 2H), 1.77 (s, 2H), 1.59 (s, 1H). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) δ 189.40, 145.37, 141.23, 139.35, 136.66, 136.31, 134.83, 132.82, 129.65, 129.62, 127.04, 125.00, 114.21, 77.35, 77.03, 76.71, 43.88, 29.72, 29.18, 21.78, 16.34.

**Diene 6e.** Prepared as for **6a**. Diene **6e** was obtained in 33% yield (0.10 g, 0.33 mmol) as a solid from 0.41 g (1.08 mmol) of glycolate **4e**; mp 113-115 °C; <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 8.34 (d, J = 8.0 Hz, 1H), 8.16 (d, J = 7.2 Hz, 1H), 7.86 (dd, J = 8.2, 10.1 Hz, 2H), 7.61 – 7.42 (m, 3H), 5.76 (s, 1H), 5.62 (d, J = 7.2 Hz, 1H), 4.87 (d, J = 26.1 Hz, 2H), 4.59 (s, 1H), 3.83 (s, 3H), 3.26 (s, 1H), 3.03 (t, J = 7.2 Hz, 1H), 2.39 (dd, J = 6.5, 12.7 Hz, 1H), 2.34 – 2.21 (m, 2H), 1.87 (s, 3H), 1.55 (s, 3H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 189.94, 146.02, 141.49, 140.18, 136.87, 133.88, 133.65, 133.24, 132.32, 128.97, 128.70, 126.58, 126.32, 126.02, 125.37, 124.92, 114.15, 44.28, 29.57, 22.06, 16.59. Anal: (C<sub>22</sub>H<sub>22</sub>O) C 87.78, H 7.26.

**Diene 6g.** Prepared as for **6a**. Diene **6g** was obtained in 41% yield (0.05 g, 0.19 mmol) as a solid from 0.15 g (0.46 mmol) of glycolate **4g**; mp 71-73 °C; <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 7.50 (d, J = 1.6 Hz, 1H), 7.42 (s, 1H), 6.70 – 6.61 (m, 1H), 6.58 (d, J = 3.4 Hz, 1H), 6.47 (dd, J = 1.8, 3.4 Hz, 1H), 4.87 – 4.74 (m, 1H), 4.65 (s, 1H), 4.37 (d, J = 6.1 Hz, 1H), 2.77 – 2.45 (m, 2H), 1.86 (s, 3H), 1.81 (s, 3H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 188.77, 152.31, 145.57, 144.27, 141.82, 136.68, 134.43, 122.87, 116.05, 112.65, 112.24, 43.29, 28.94, 22.11, 16.77. Anal. (C<sub>16</sub>H<sub>18</sub>O<sub>2</sub>) C 79.23, H 7.24.

**Diene 6h.** Prepared as for **10a**. Diene **6h** was obtained in 45% yield (0.07 g, 0.26 mmol) as an oil from 0.20 g (0.58 mmol) of glycolate **4h**. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.68 (s, 1H), 7.33 (dt, J = 4.5, 14.2 Hz, 2H), 7.12 (dd, J = 8.3, 16.2 Hz, 2H), 6.67 – 6.59 (m, 1H), 4.96 (s, 1H), 4.74 (s, 1H), 3.75 (s, 1H), 2.72 – 2.50 (m, 2H), 1.90 (s, 3H), 1.81 (s, 3H). <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 189.45, 162.81, 159.50, 145.36, 141.33, 140.24, 136.68, 130.42, 130.31, 129.93, 129.90, 128.54, 128.49, 124.03, 123.98, 123.86, 115.88, 115.59, 114.30, 44.28, 29.24, 21.92, 16.50. Anal. (C<sub>18</sub>H<sub>19</sub>FO) C 79.58, H 6.72.

**Enone 8a.** Isobenzofuran **5a** (0.37 g, 0.87 mmol) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (9.0 mL) was added to a mixture of finely ground pyridinium chlorochromate (2.00 g, 8.70 mmol) and silica gel (2.00 g) in dry CH<sub>2</sub>Cl<sub>2</sub> (20.0 mL) at rt. The reaction mixture was stirred at rt for 24 h. The crude solution was then filtered through a short silica gel column with ether (100 mL) and concentrated in vacuo. The dark brown residue was purified via flash chromatography over silica gel (15/85, EtOAc/hexanes) to deliver enone **6a** as a white solid in 55 % yield (0.20 g, 0.48 mmol) mp 126-127 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.08 – 7.98 (m, 1H), 7.49 (dd, J = 1.0, 8.1 Hz, 1H), 7.41

(t,  $J = 7.6$  Hz, 1H), 7.18 (td,  $J = 1.7, 8.0$  Hz, 1H), 5.28 (s, 1H), 5.21 (d,  $J = 10.0$  Hz, 1H), 4.74 (s, 1H), 4.47 (s, 1H), 4.40 – 4.25 (m, 2H), 3.26 (t,  $J = 10.3$  Hz, 1H), 2.93 – 2.70 (m, 1H), 2.54 – 2.30 (m, 2H), 2.01 – 1.87 (m, 3H), 1.58 (d,  $J = 8.2$  Hz, 1H), 1.36 (q,  $J = 7.1$  Hz, 3H), 1.27 (s, 3H).  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ ) 198.0, 171.0, 146.0, 142.0, 138.0, 132.9, 130.4, 130.0, 129.7, 128.3, 125.6, 114.2, 85.2, 78.5, 62.3, 51.5, 47.5, 42.0, 18.5, 14.7, 12.2. IR (film) 1743, 1674  $\text{cm}^{-1}$ . HRMS Calcd for  $\text{C}_{21}\text{H}_{24}\text{BrO}_4^+$ : 419.0858, found 419.0836. Anal. ( $\text{C}_{21}\text{H}_{23}\text{BrO}_4$ ) C 60.31, H 5.47.

**Enone 8b.** Prepared as for **8a**. Enone **8b** was obtained as a white foam in 54 % yield (0.07 g, 0.16 mmol) from 0.12g (0.29 mmol) of isobenzofuran **5b**.  $^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.42 (d,  $J = 8.8$  Hz, 1H), 7.18 (dd,  $J = 5.0, 10.5$  Hz, 2H), 6.98 (d,  $J = 7.8$  Hz, 1H), 5.61 (d,  $J = 8.9$  Hz, 1H), 5.56 (s, 1H), 4.92 (s, 1H), 4.77 (s, 1H), 4.30 (q,  $J = 7.1$  Hz, 2H), 3.70 (s, 1H), 2.50 (dd,  $J = 3.9, 16.3$  Hz, 1H), 2.23 (dd,  $J = 13.9, 16.3$  Hz, 1H), 1.98 (d,  $J = 2.0$  Hz, 3H), 1.93 – 1.80 (m, 1H), 1.61 (s, 1H), 1.53 (s, 3H), 1.35 (t,  $J = 7.1$  Hz, 3H).  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ ) 183.6, 156.0, 142.3, 132.6, 129.9, 127.9, 113.9, 85.0, 78.1, 66.6, 62.0, 58.0, 46.7, 42.4, 22.7, 18.02, 14.2, 11.4.

**Enone 8c.** Prepared as for **8a**. Enone **8c** was obtained as a white solid in 56% yield (0.22 g, 0.54 mmol) from 0.40 g (0.97 mmol) of isobenzofuran **5c**; mp 41-45  $^\circ\text{C}$ ;  $^1\text{H NMR}$  (270 MHz,  $\text{CDCl}_3$ )  $\delta$  7.93 (dd,  $J = 1.6, 7.8$ , 1H), 7.38 (dd,  $J = 1.7, 7.9$ , 1H), 7.25 (t,  $J = 7.9$ , 1H), 5.31 – 5.10 (m, 2H), 4.61 (s, 1H), 4.47 – 4.36 (m, 1H), 4.25 (qd,  $J = 1.3, 7.1$ , 2H), 3.14 (d,  $J = 10.4$ , 1H), 2.84 – 2.63 (m, 1H), 2.48 – 2.21 (m, 2H), 1.86 (dd,  $J = 0.9, 2.5$ , 3H), 1.30 (t,  $J = 7.1$ , 3H), 1.21 (s, 4H).  $^{13}\text{C-NMR}$  (67 MHz,  $\text{CDCl}_3$ ) 197.5, 169.8, 154.7, 142.1, 139.0, 132.5, 129.9, 127.4, 113.4, 82.6, 62.0, 50.4, 46.9, 42.3, 18.2, 14.1, 11.1.

**Enone 8d.** Prepared as for **8a**. Enone **8d** was obtained as a white solid in 67% yield (0.11 g, 0.27 mmol) from 0.17 g (0.41 mmol) of isobenzofuran **5d**; mp 96-100  $^\circ\text{C}$ ;  $^1\text{H NMR}$  (270 MHz,  $\text{CDCl}_3$ )  $\delta$  7.93 (dd,  $J = 1.6, 7.8$  Hz, 1H), 7.38 (dd,  $J = 1.7, 7.9$  Hz, 1H), 7.25 (t,  $J = 7.9$  Hz, 1H), 5.31 – 5.10 (m, 2H), 4.61 (s, 1H), 4.47 – 4.36 (m, 1H), 4.25 (qd,  $J = 1.3, 7.1$  Hz, 2H), 3.14 (d,  $J = 10.4$  Hz, 1H), 2.84 – 2.63 (m, 1H), 2.48 – 2.21 (m, 2H), 1.86 (dd,  $J = 0.9, 2.5$  Hz, 3H), 1.30 (t,  $J = 7.1$  Hz, 3H), 1.21 (s, 3H).  $^{13}\text{C-NMR}$  (67 MHz,  $\text{CDCl}_3$ ) 198.1, 169.8, 155.5, 142.4, 135.6, 134.7, 130.5, 130.4, 129.0, 127.8, 114.0, 82.0, 62.2, 50.2, 46.7, 42.3, 18.2, 14.3, 11.5.

**Tosylhydrazone 9a.** HOAc (0.01 g, 0.16 mmol) was added to a solution of enone **8a** (0.10 g, 0.24 mmol) and tosylhydrazide (0.06 g, 0.31 mmol) in  $\text{CH}_2\text{Cl}_2$  (2.20 mL). The reaction mixture stirred at rt for 24 h, then was washed with water (5.00 mL), dried over  $\text{MgSO}_4$ , filtered and concentrated in vacuo. The residue was purified via flash chromatography on silica gel (80:20, hexanes/EtOAc) to afford tosylhydrazone **9a** as white solid in 92 % yield (0.13 g, 0.22 mmol); mp 162-165  $^\circ\text{C}$ .  $^1\text{H-NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.02 – 7.79 (m, 3H), 7.46 (d,  $J = 9.5$  Hz, 2H), 7.36 (t,  $J = 8.7$  Hz, 3H), 7.14 (t,  $J = 7.7$  Hz, 1H), 5.28 (s, 1H), 5.07 (d,  $J = 9.8$  Hz, 1H), 4.70 (s, 1H), 4.48 (s, 1H), 4.25 (q,  $J = 7.1$  Hz, 2H), 2.99 (s, 1H), 2.58 – 2.34 (m, 6H), 2.10 – 1.85 (m, 4H), 1.34 (t,  $J = 7.1$  Hz, 3H), 1.22 (s, 3H).  $^{13}\text{C-NMR}$  (101 MHz,  $\text{CDCl}_3$ )  $\delta$  170.44, 153.31, 144.28, 143.92, 142.66, 138.39, 135.01, 132.51, 129.73, 129.69, 129.49, 128.25, 127.80, 127.78, 124.74, 114.17, 85.49, 78.23, 77.35, 77.04, 76.72, 61.70, 49.10, 44.80, 29.37, 21.66, 18.00, 14.18, 13.07. IR (film) 3210, 3070, 2980, 2919, 2256, 1739  $\text{cm}^{-1}$ . HRMS Calcd for  $\text{C}_{28}\text{H}_{31}\text{BrN}_2\text{O}_5\text{S}^+$ : 587.1215, found: 587.1201.

**Ester 10a.** Catecholborane (0.10 mL, 0.80 mmol) was added to a solution of tosylhydrazone **9a** (0.40 g, 0.70 mmol) in CHCl<sub>3</sub> (3.00 mL) at 0 °C. The reaction mixture stirred at 0 °C for 1 h, then NaOAc.3H<sub>2</sub>O (0.19 g, 1.30 mmol) was added in one portion. The reaction mixture was maintained for 1 h at 0 °C, diluted with CHCl<sub>3</sub> (1.80 mL), and heated under reflux for 12 h. The mixture was then cooled to rt and filtered through a pad of Celite. The filtrate was concentrated in vacuo and the residue was purified via flash chromatography over silica gel (90:10, hexanes/EtOAc) to afford ester **10a** as white solid in 68 % yield (0.17 g, 0.48 mmol); mp 37-40 °C; <sup>1</sup>H-NMR (270 MHz, CDCl<sub>3</sub>) δ 7.97 (d, J = 6.2 Hz, 1H), 7.49 (d, J = 8.0 Hz, 1H), 7.36 (t, J = 7.0 Hz, 1H), 7.12 (t, J = 7.6 Hz, 1H), 5.59 (s, 1H), 5.34 (d, J = 6.4 Hz, 1H), 4.79 (s, 2H), 4.38 (d, J = 6.4 Hz, 1H), 4.25 (dd, J = 4.9, 7.1 Hz, 2H), 3.01 (s, 1H), 2.37 (m, 3H), 2.21 – 2.04 (m, 1H), 1.73 (s, 3H), 1.60 (s, 4H), 1.31 (t, J = 7.1 Hz, 3H). <sup>13</sup>C-NMR (67 MHz, CDCl<sub>3</sub>) δ 173.01, 146.94, 140.85, 132.70, 130.21, 129.58, 129.37, 128.04, 123.25, 123.10, 112.16, 83.50, 81.30, 77.56, 77.09, 76.62, 61.33, 49.14, 47.31, 39.39, 28.04, 22.08, 21.00, 14.22.

## Biological Assays

**KB3 Assays.** All stock solutions of compounds were made at 10 mM in dimethyl sulfoxide (DMSO). DMSO and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) were from Sigma Chemical Co. (St. Louis, MO). Cell culture reagents were obtained from Life Technologies (Carlsbad, CA).

**Cell Culture.** The KB-3 human carcinoma cell line was maintained in monolayer culture at 37°C and 5% CO<sub>2</sub> in Dulbecco's Modified Eagle's Medium, supplemented with 10% fetal bovine serum, 2 mM L-glutamine, 50 units/mL penicillin, and 50 µg/mL streptomycin.

**MTT colorimetric assay.** Inhibition of cell proliferation was assessed by the MTT colorimetric assay. First described by Mosmann,<sup>1</sup> this assay is based on the ability of a mitochondrial dehydrogenase enzyme from viable cells to cleave the tetrazolium rings of the pale yellow MTT and form a dark blue formazan crystal product which is largely impermeable to cell membranes, thus resulting in its accumulation within healthy cells. Solubilization of the cells by the addition of solvent results in the liberation of the crystals which are solubilized. The level of the formazan product created is directly proportional to the number of living cells.

KB-3 cells (2000/well) were plated in 96-well dishes, and were treated after 24 h with different concentrations (0.1 nM -100 µM) of the compound being assayed. The final concentration of DMSO did not exceed 1%, and controls received vehicle alone. MTT assay was performed after 96 h as described.<sup>2,3</sup> Cells were incubated with 50 µg/well/0.2 mL MTT for 4 h at 37°C, the media was removed and the formazan crystals were dissolved in 150 µL of DMSO. The resulting color change was measured via spectrophotometry at a wavelength of 570 nm using an ELx800™ Absorbance Microplate Reader (Bio Tek Instruments, Inc., Winooski, VT). Values are the means of triplicate assays and are expressed as mean relative to untreated controls. IC<sub>50</sub>

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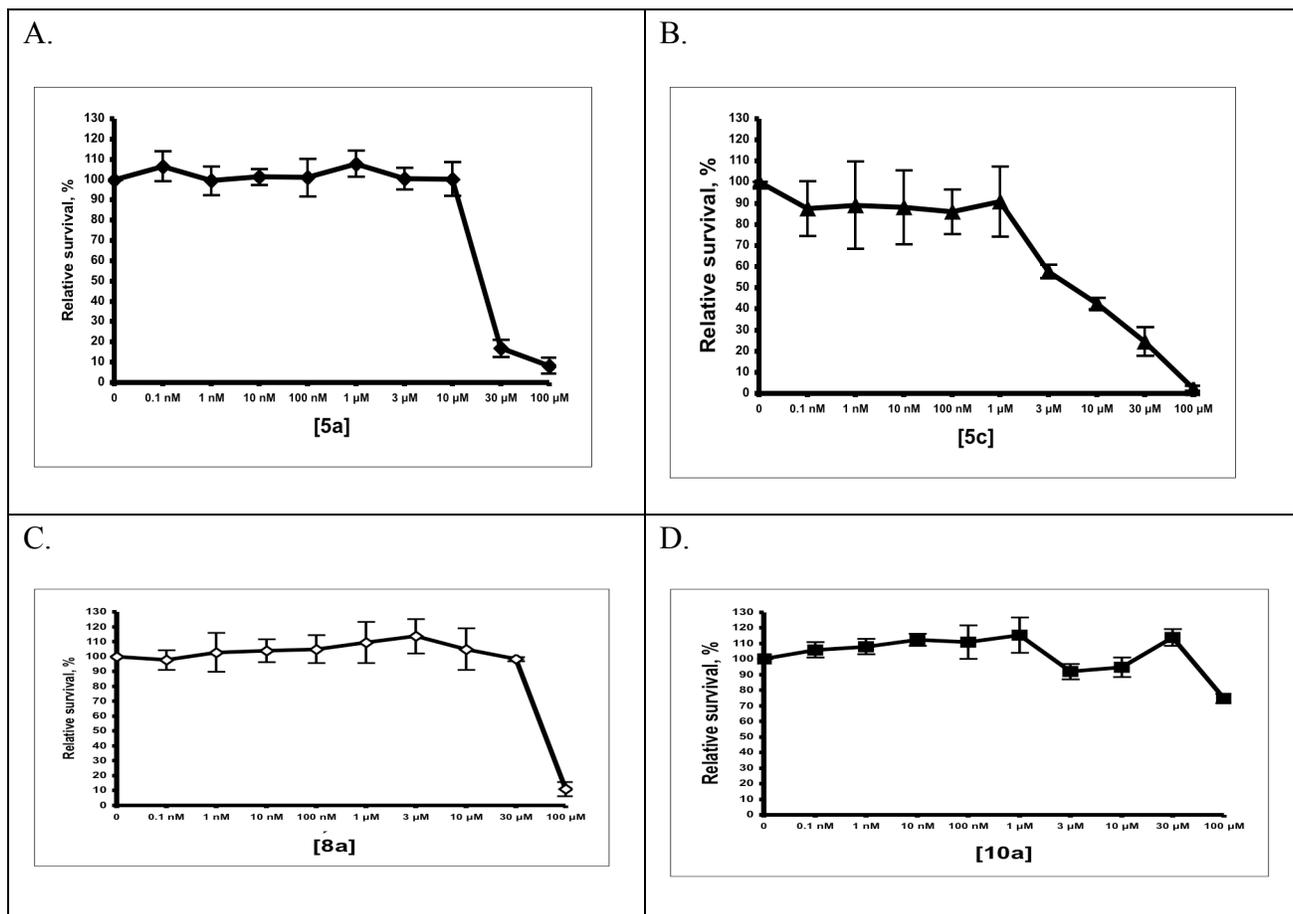
<sup>1</sup> Mosmann T. *J. Immunol. Methods* **1983**, *65*, 55-63.

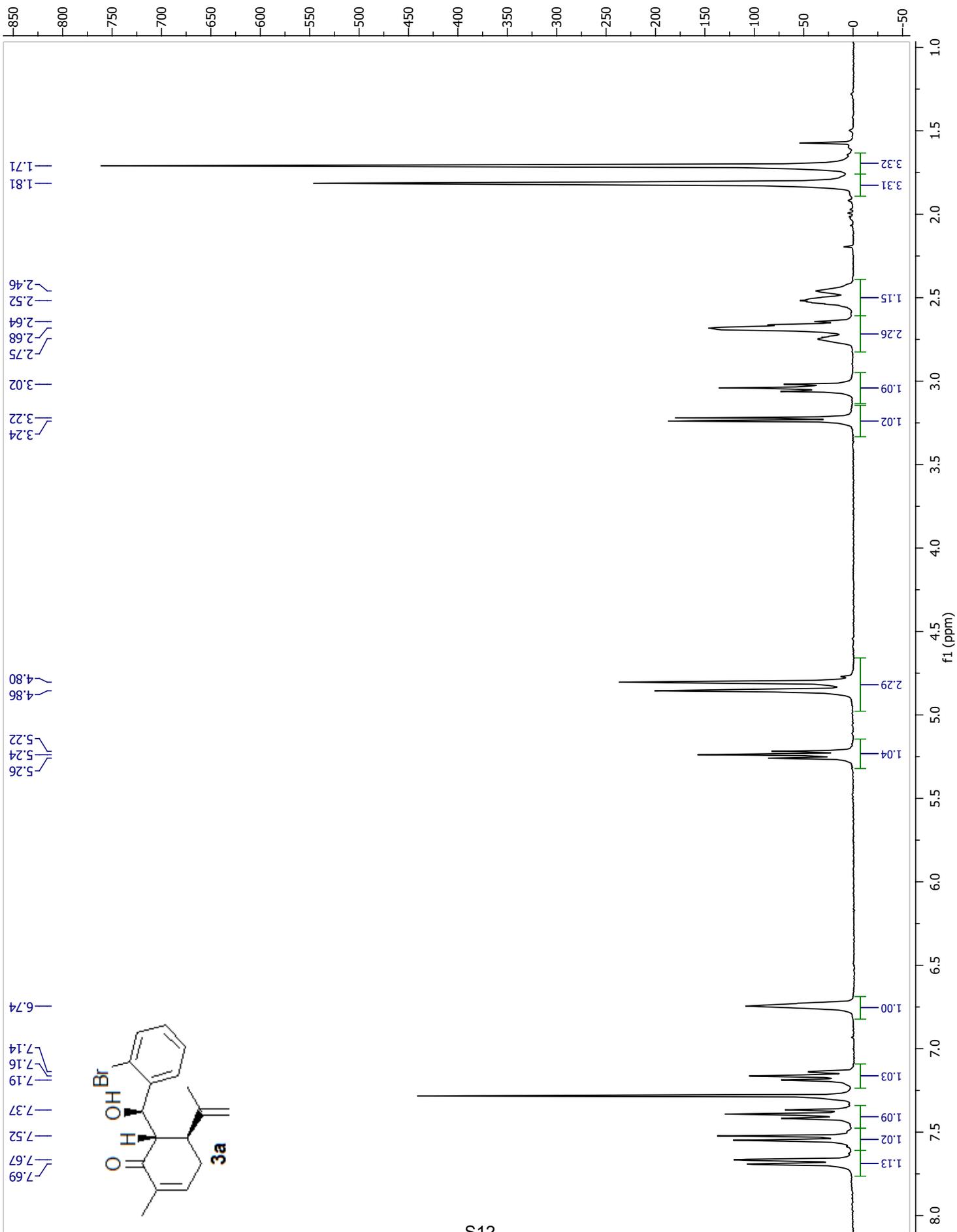
<sup>2</sup> Fan, M.; Du, L.; Stone, A. A.; Gilbert K. M.; Chambers, T. C. *Cancer Res.* **2000**, *60*, 6403-6407.

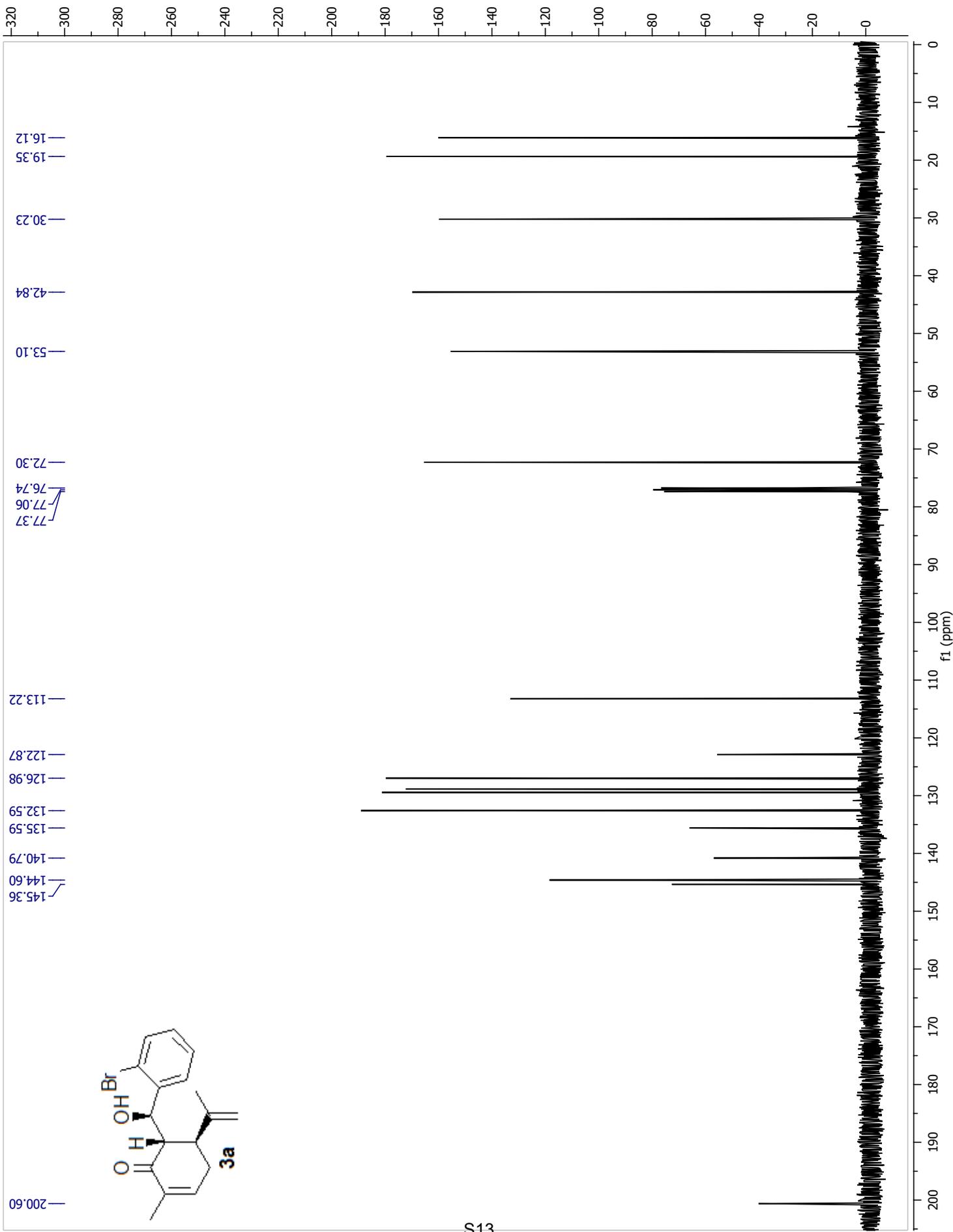
<sup>3</sup> Alley, M. C.; Scudiero, D. A.; Monks A.; Hursey, M. L.; Czerwinski, M. J.; Fine, D. L.; Abbott, B. J.; Mayo, J. G.; Shoemaker, R. H.; Boyd M. R. *Cancer Res.* **1988**, *48*, 589-601

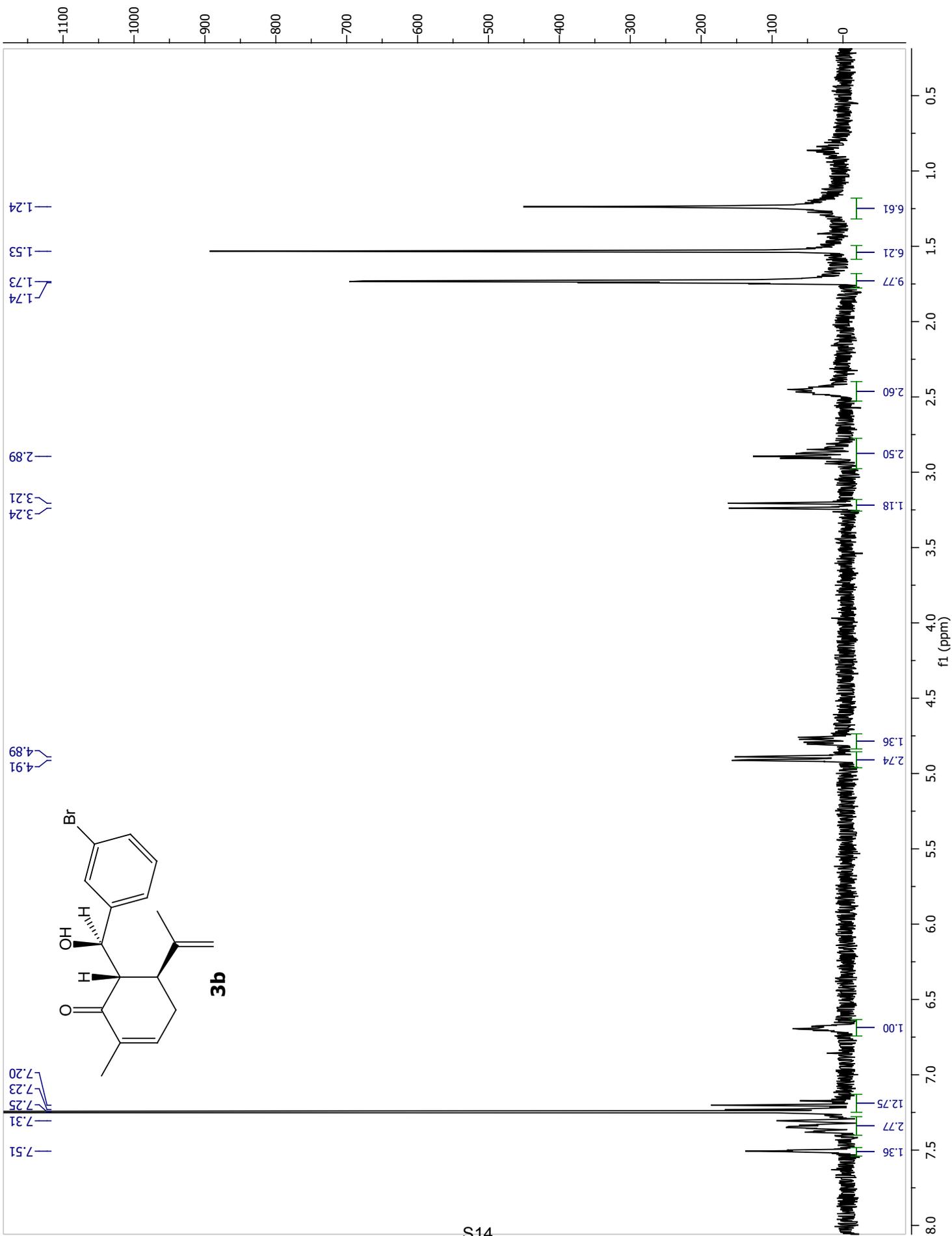
is the concentration that reduced survival to 50% of the control (no drug). Representative concentrations curves are shown below (Figure).

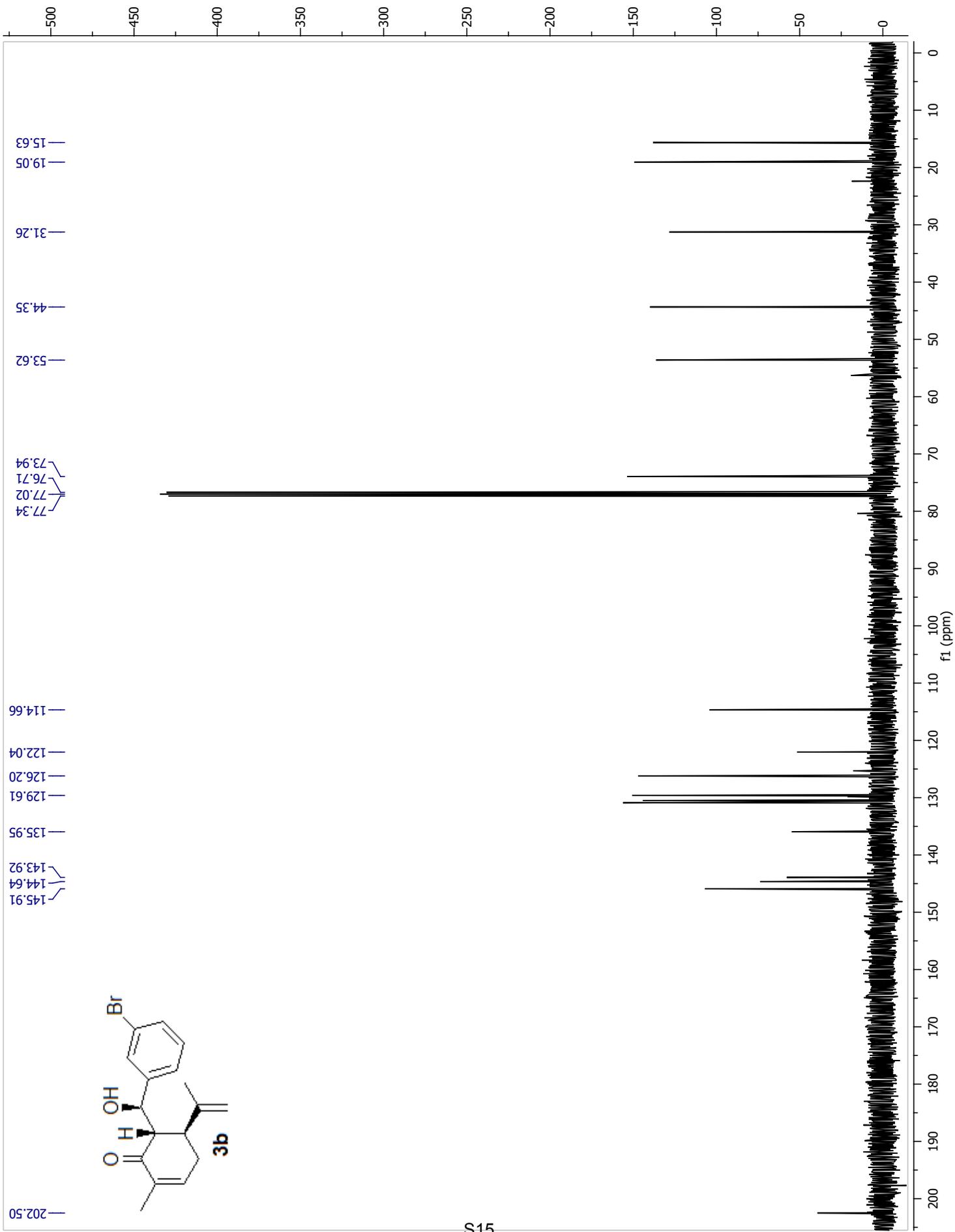
Figure. Concentration curves of Sclerologs **5a**, **5c**, **8a** and **10a**.

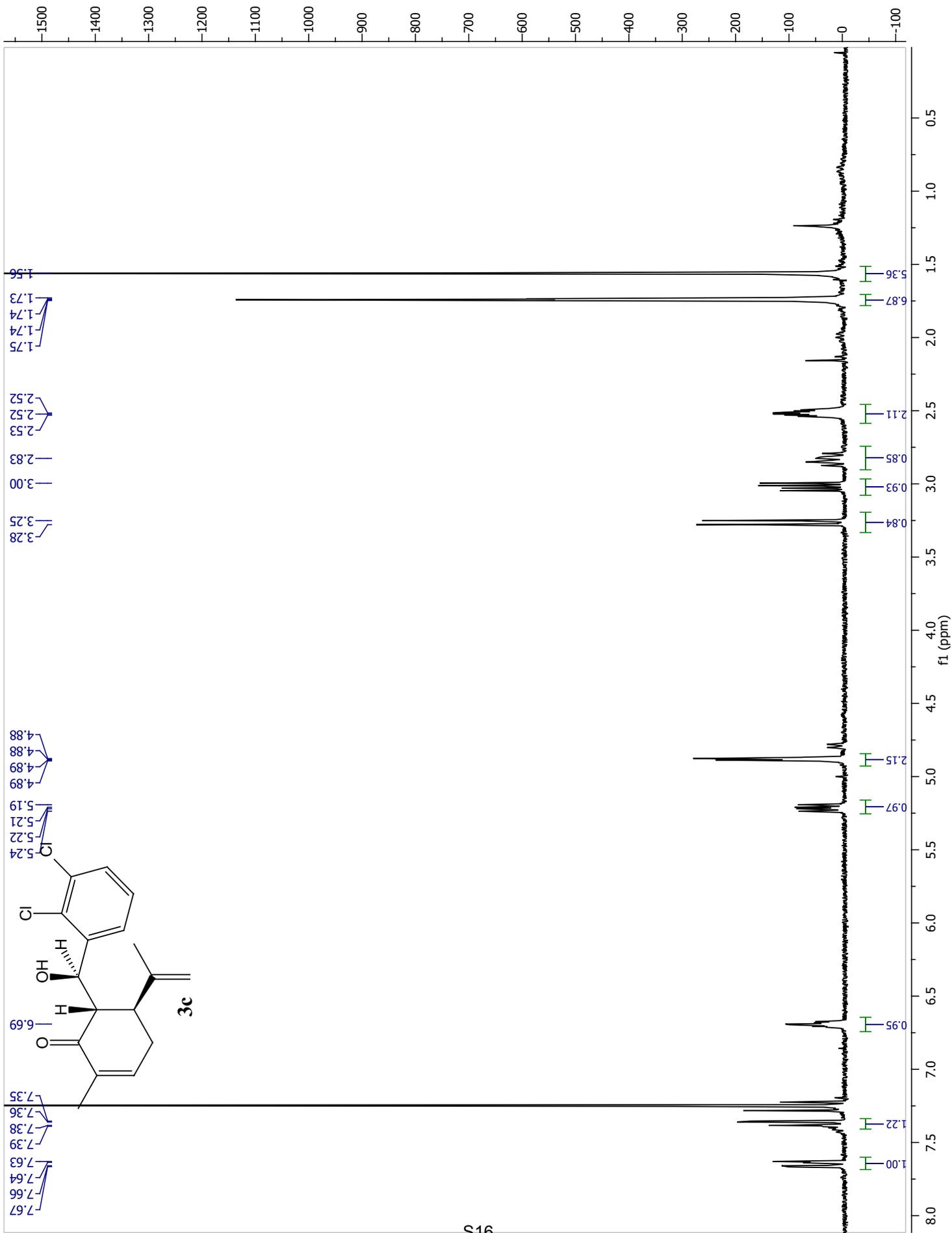


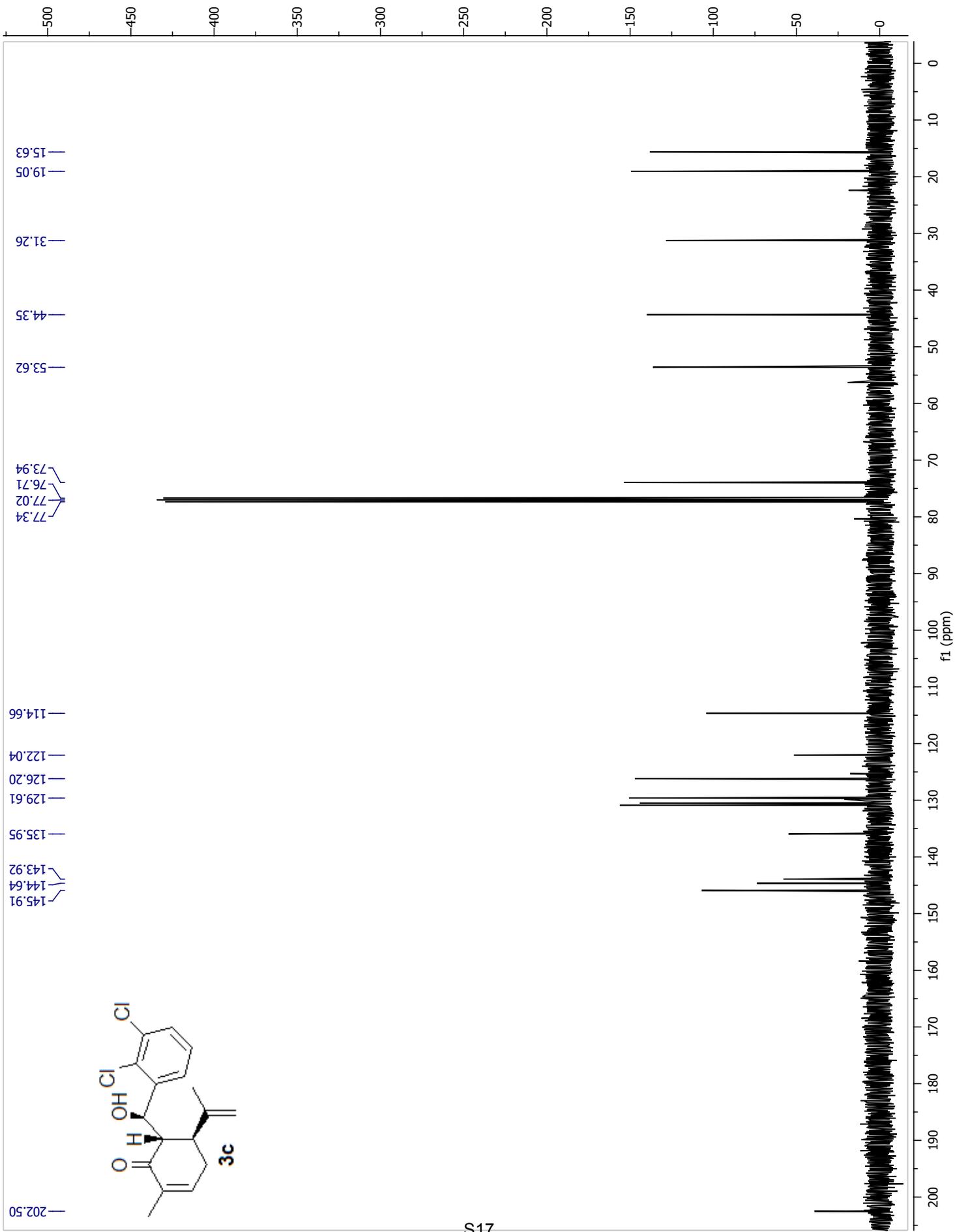


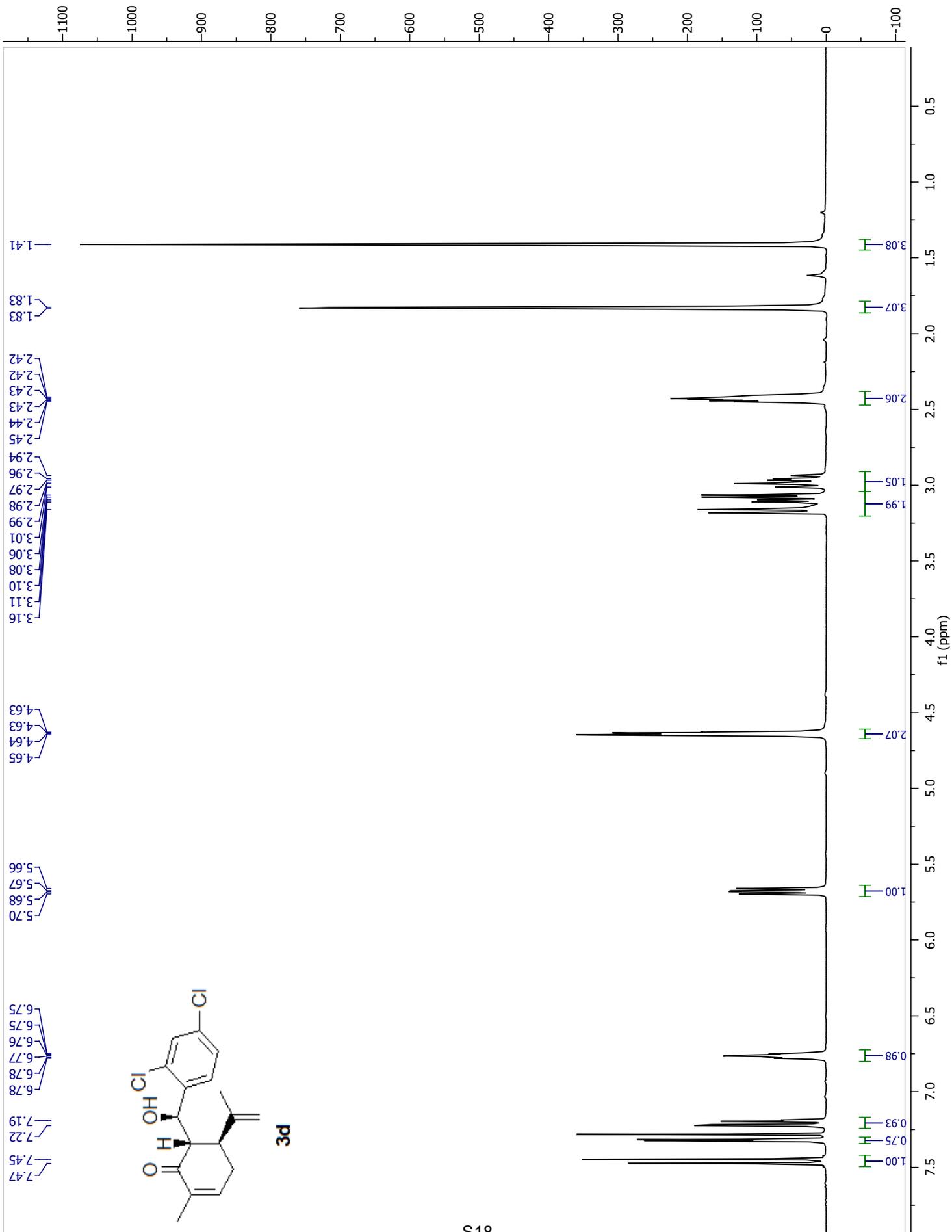


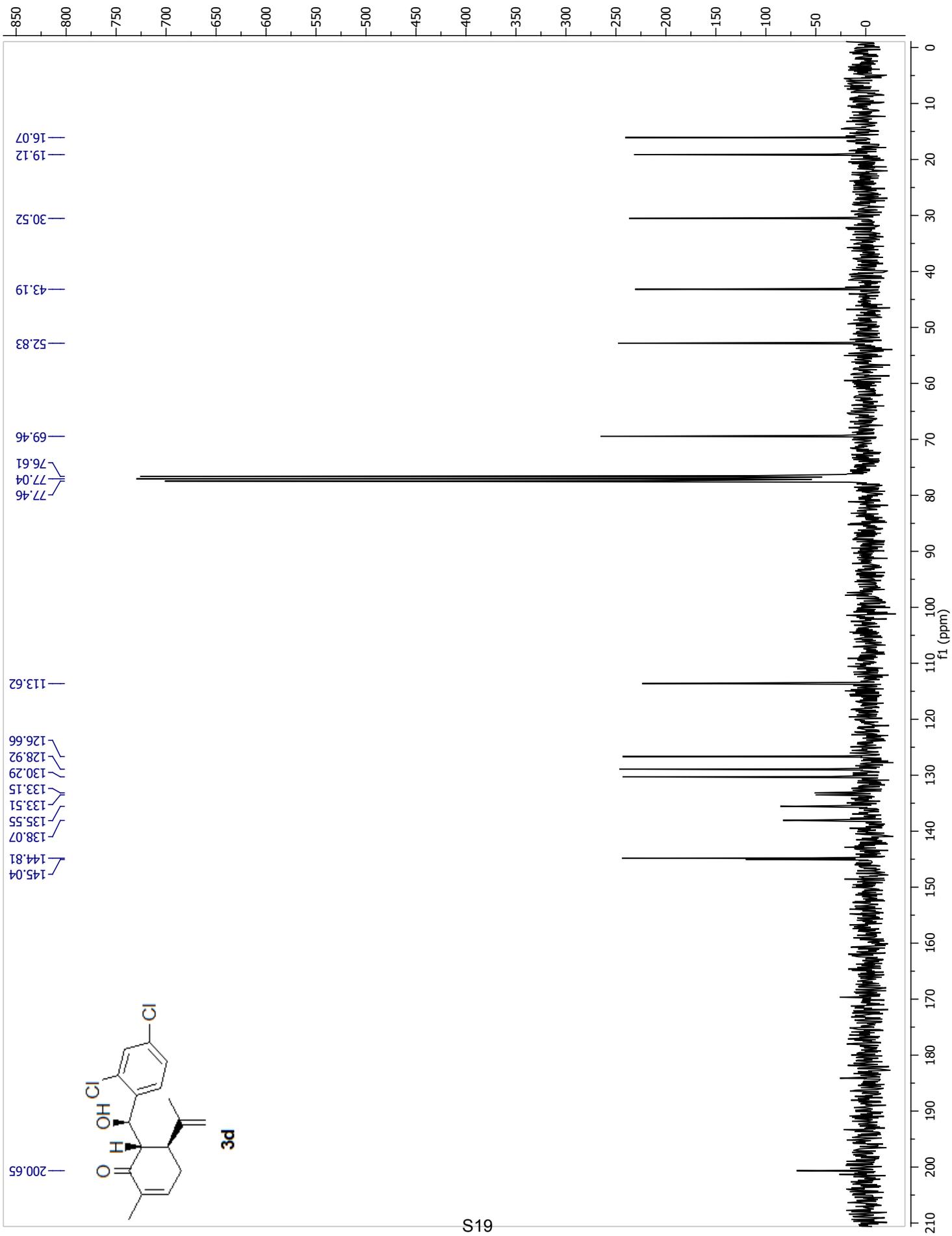












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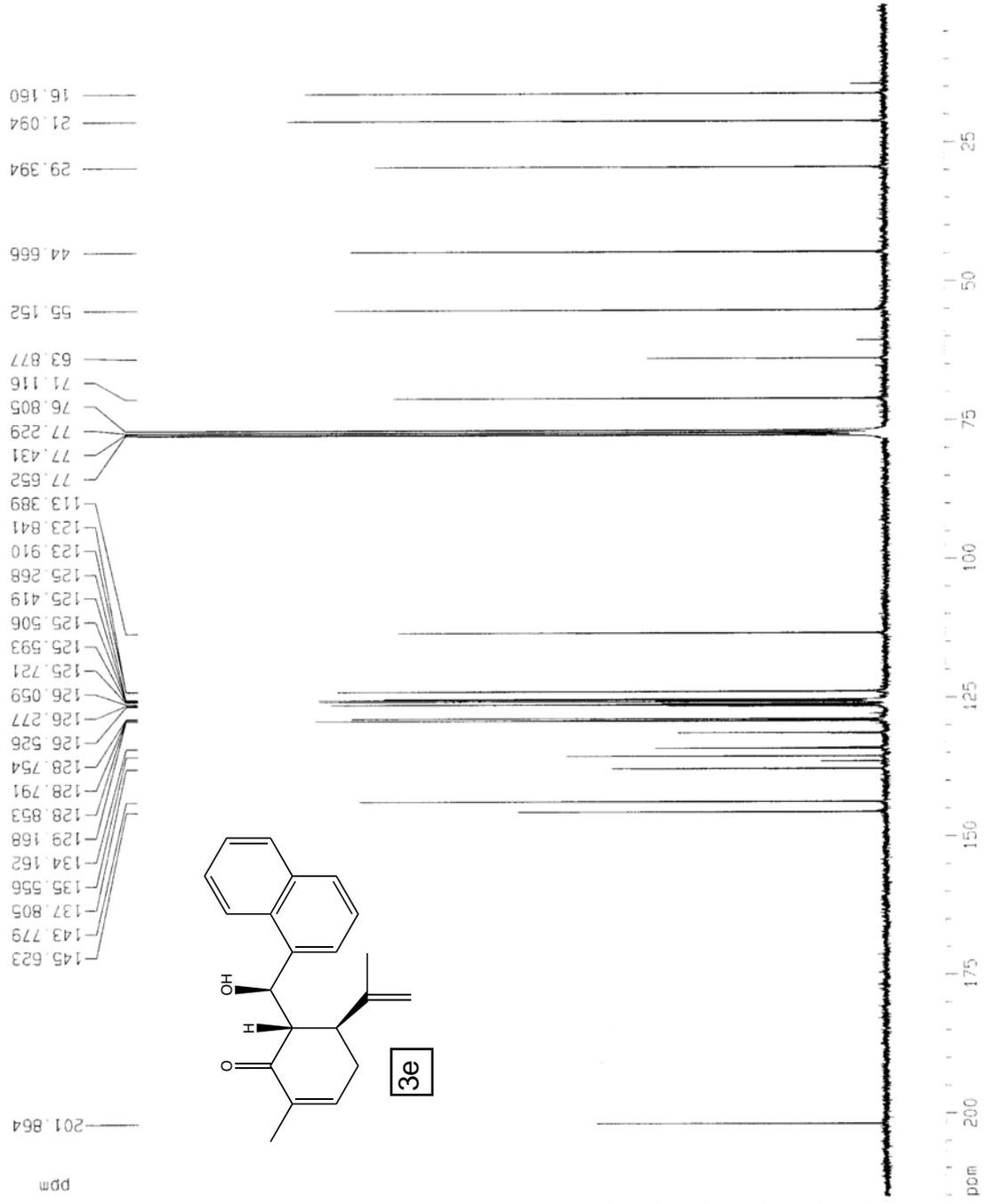
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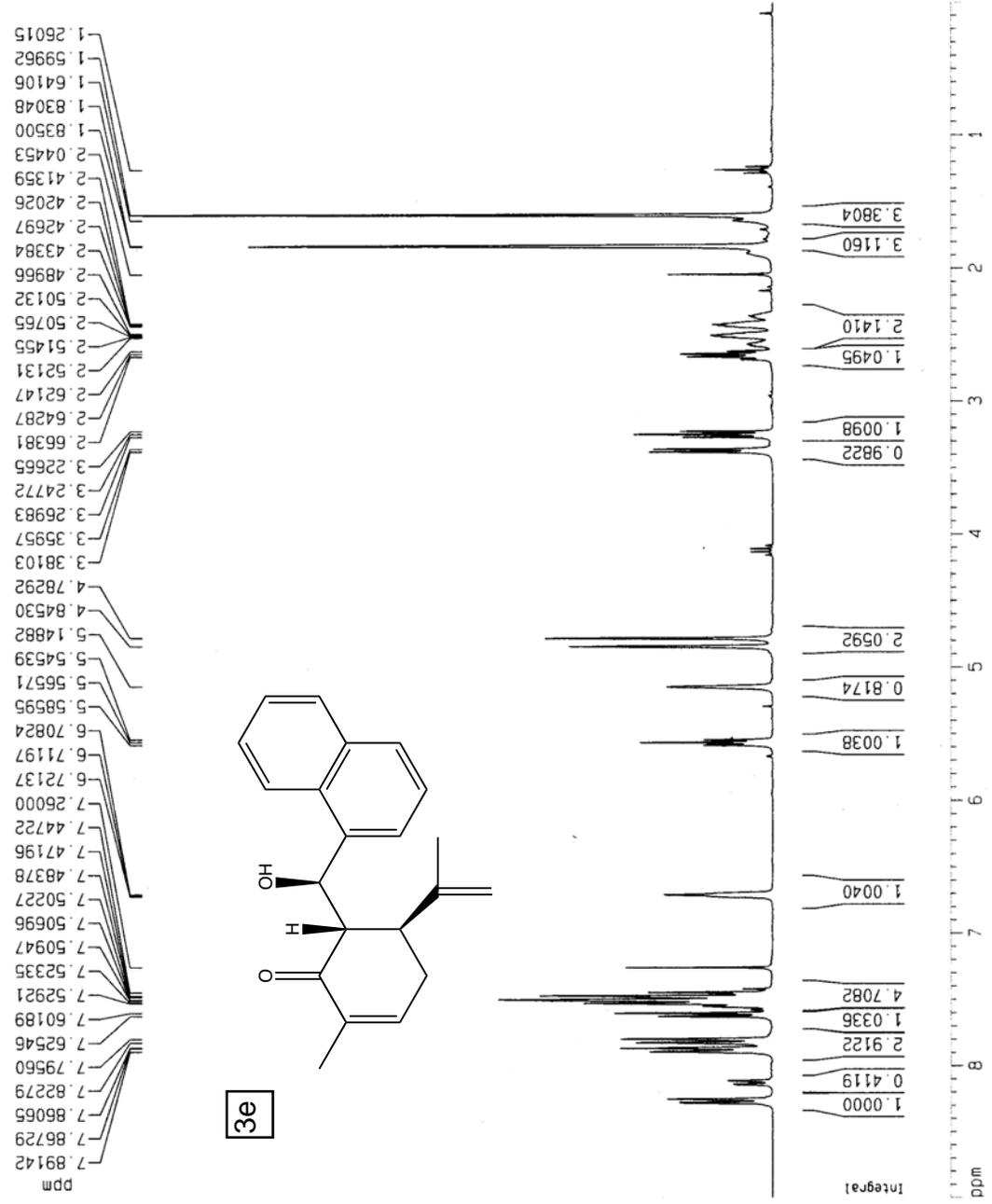
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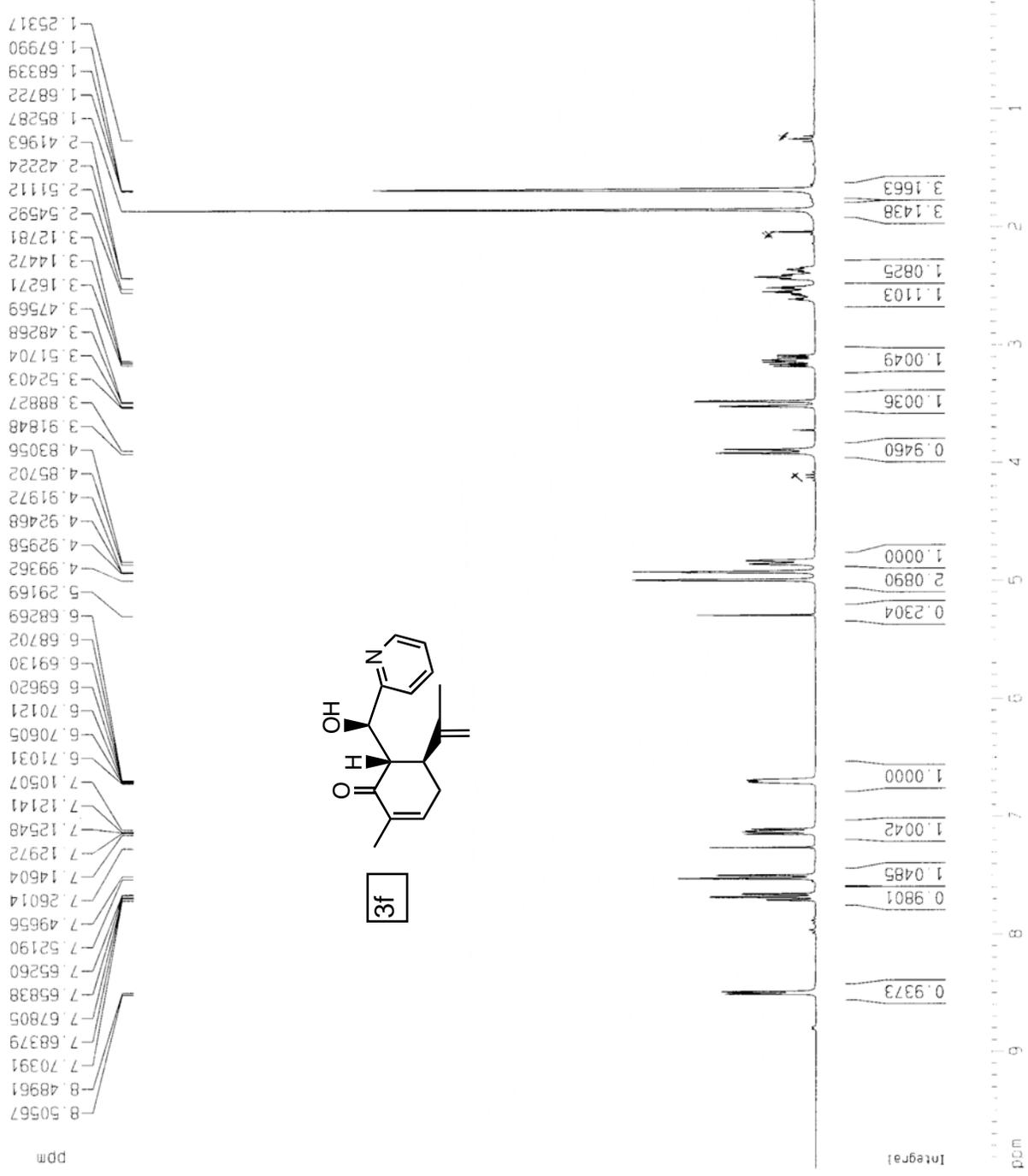
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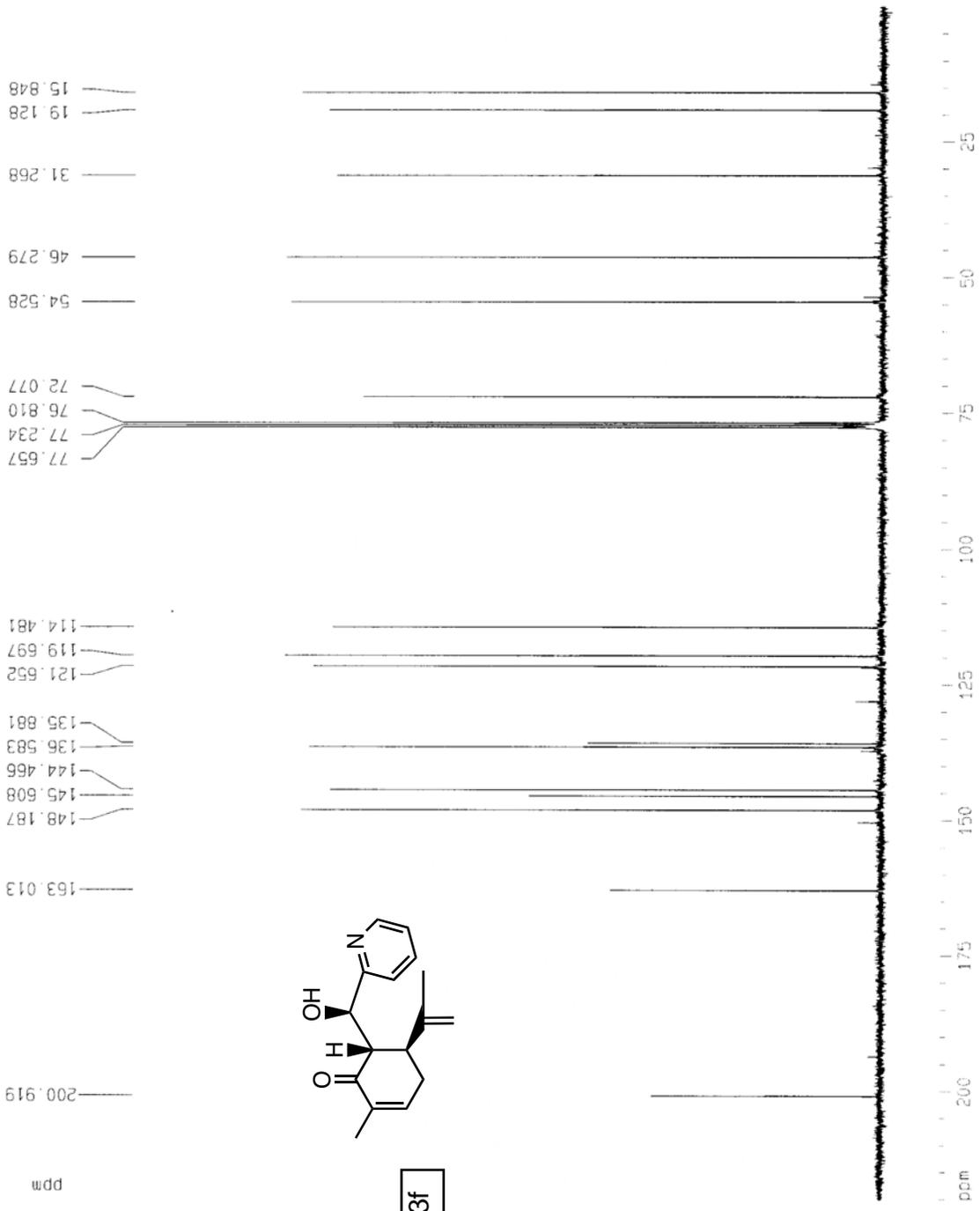
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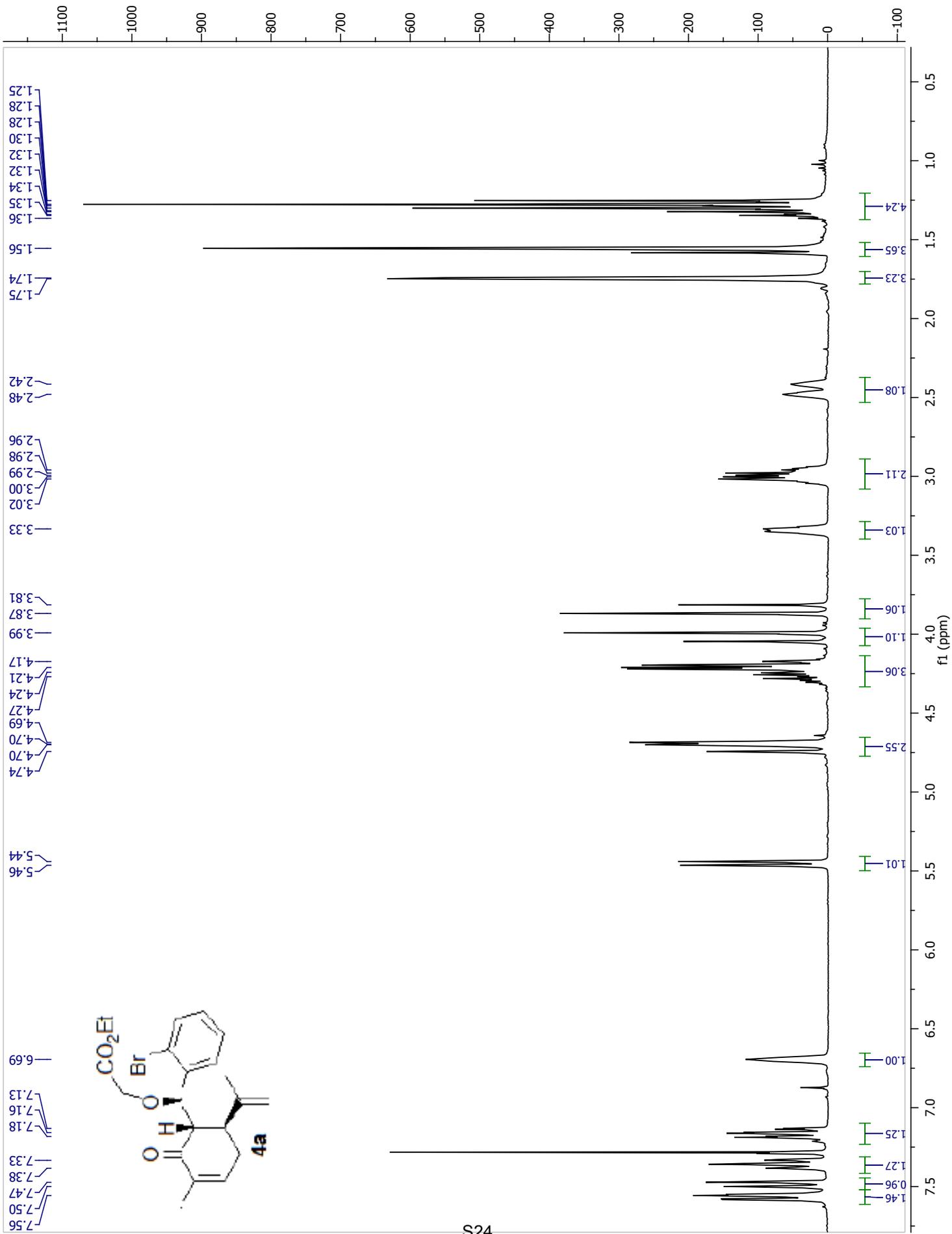
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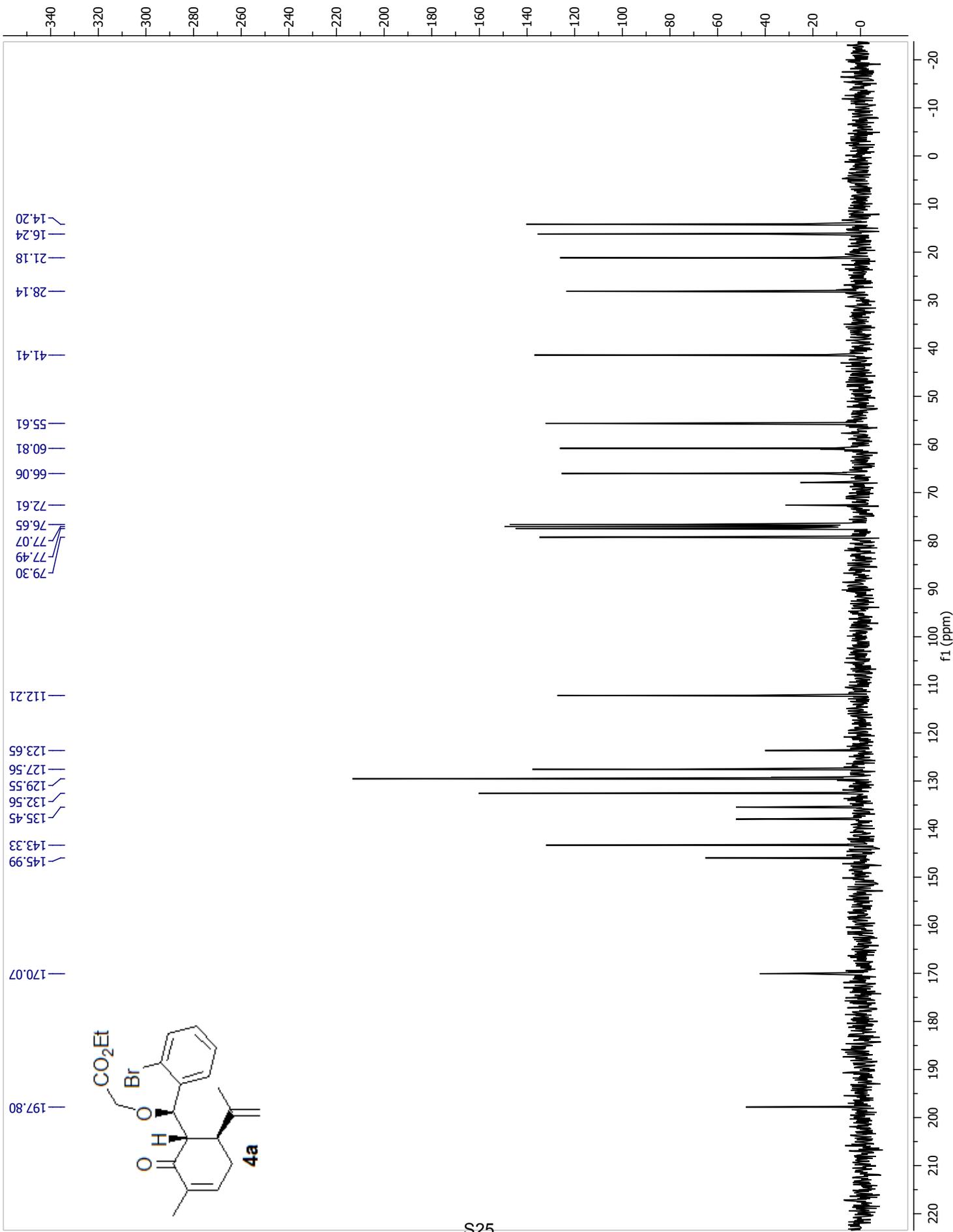
10 NMR plot parameters

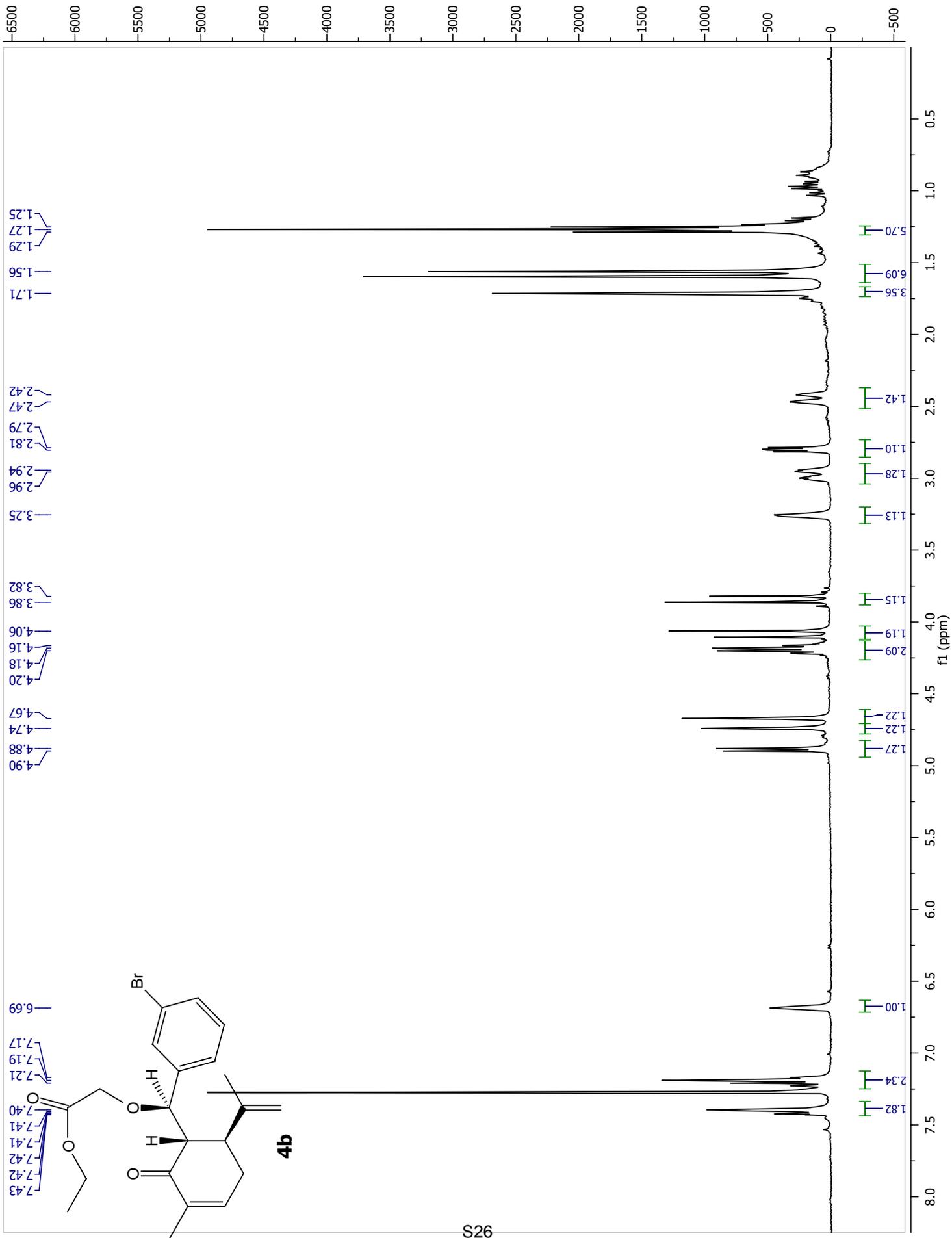
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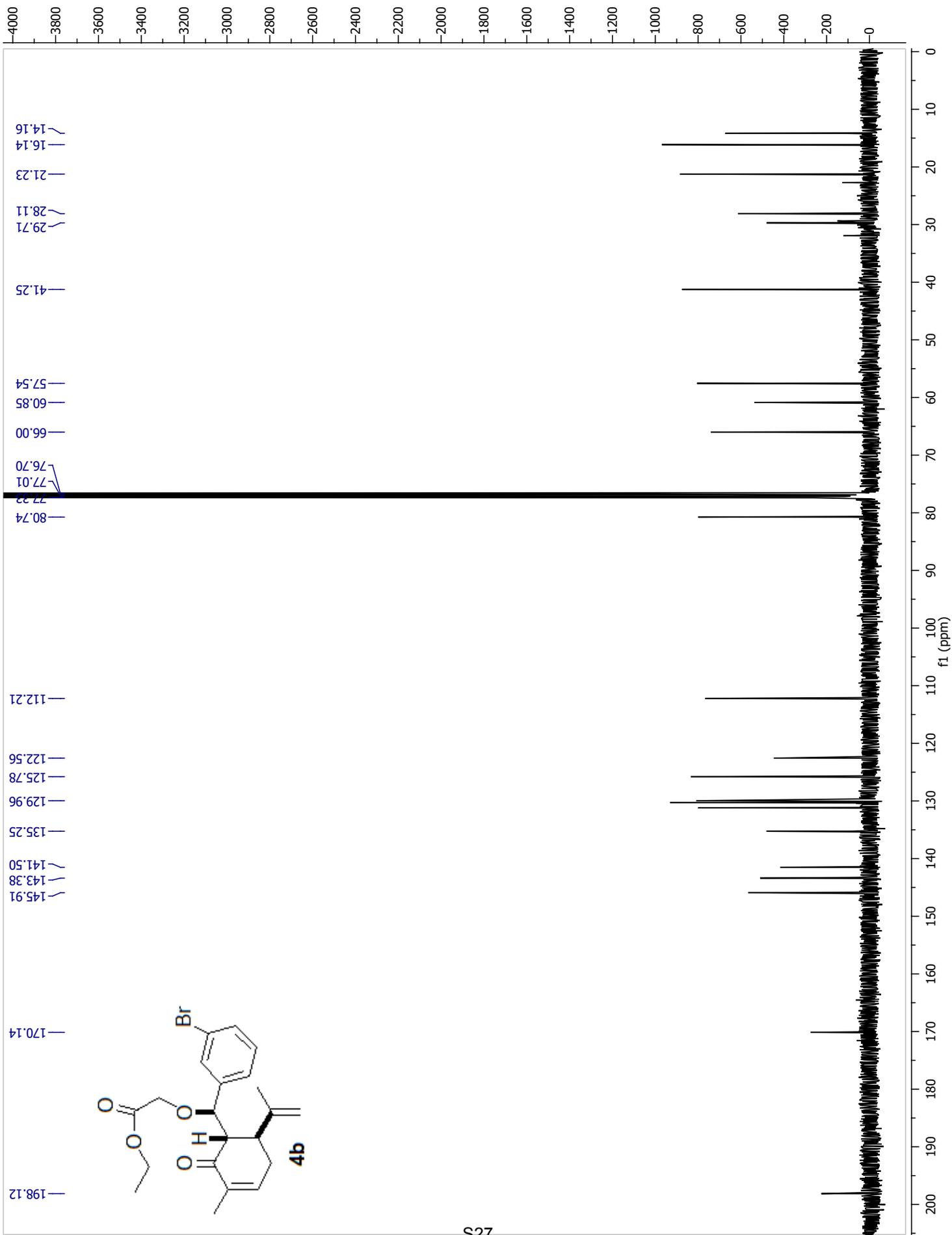


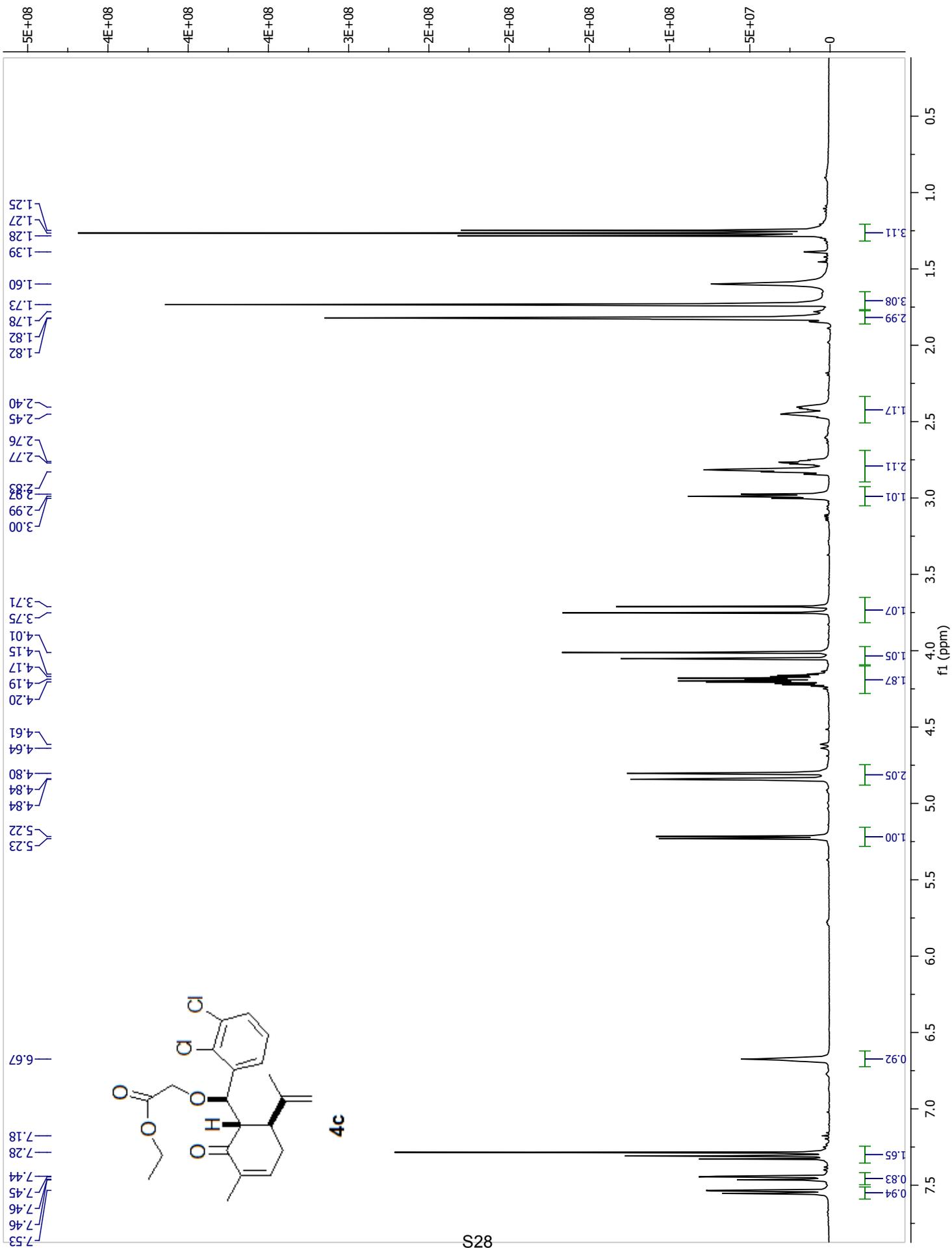
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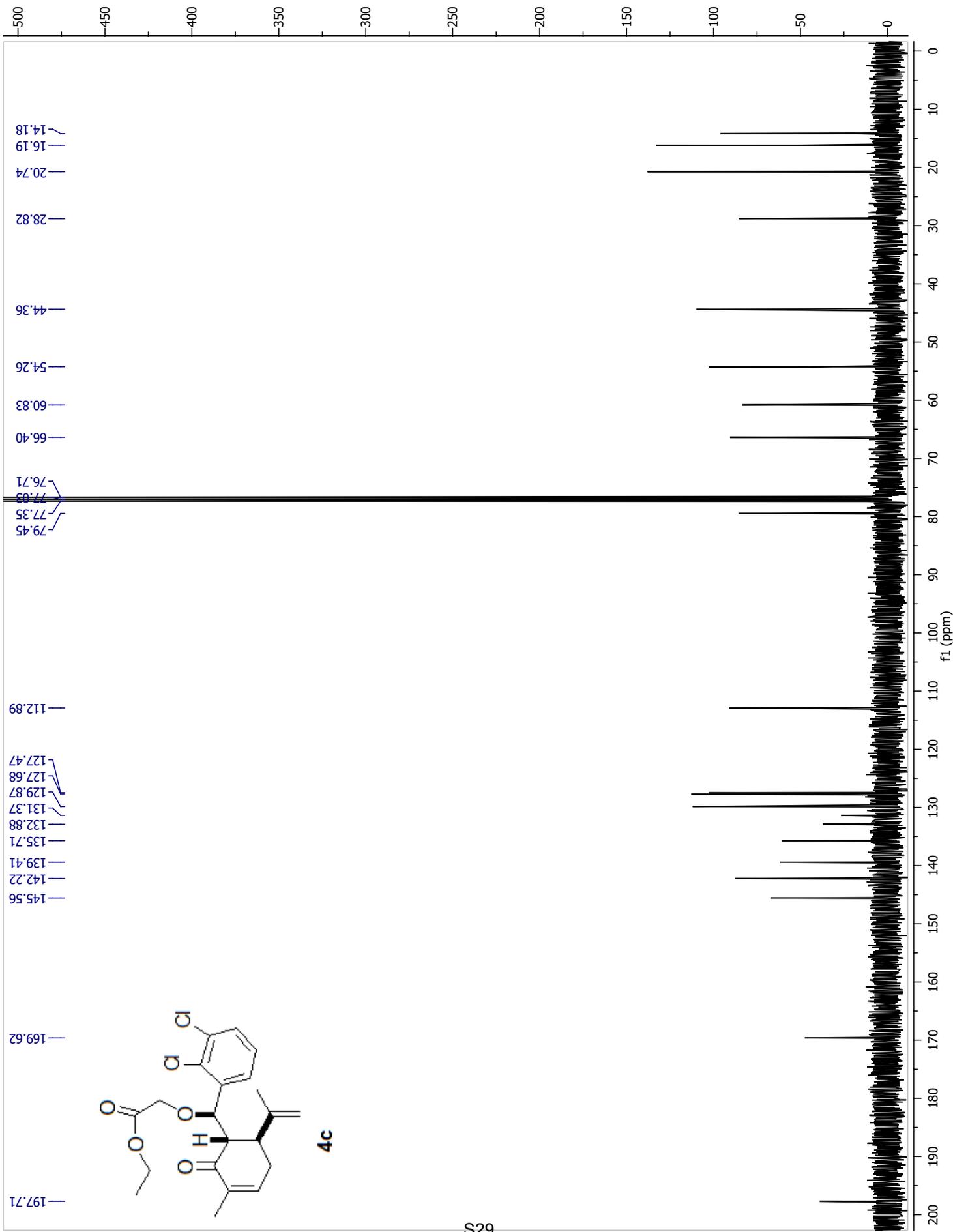


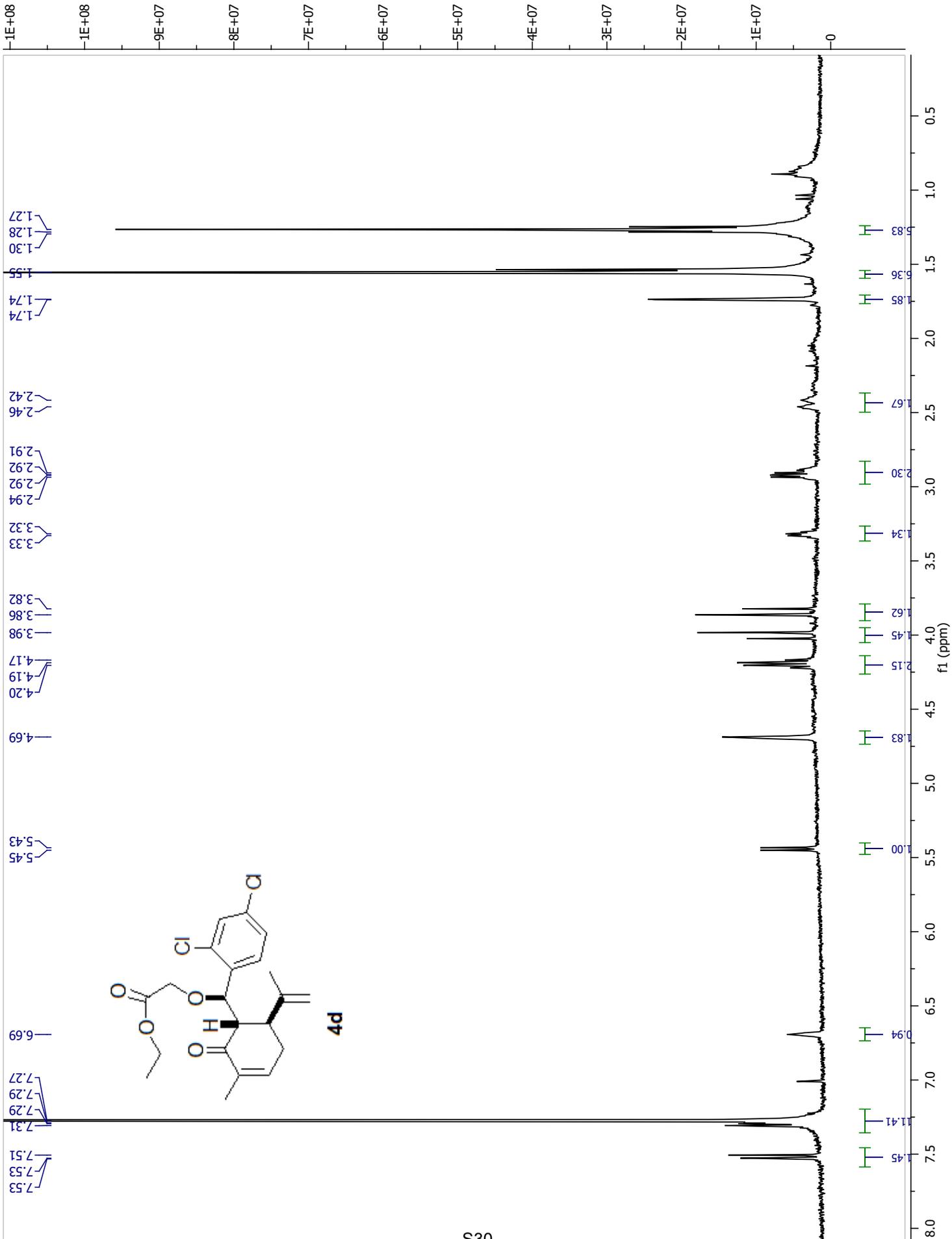


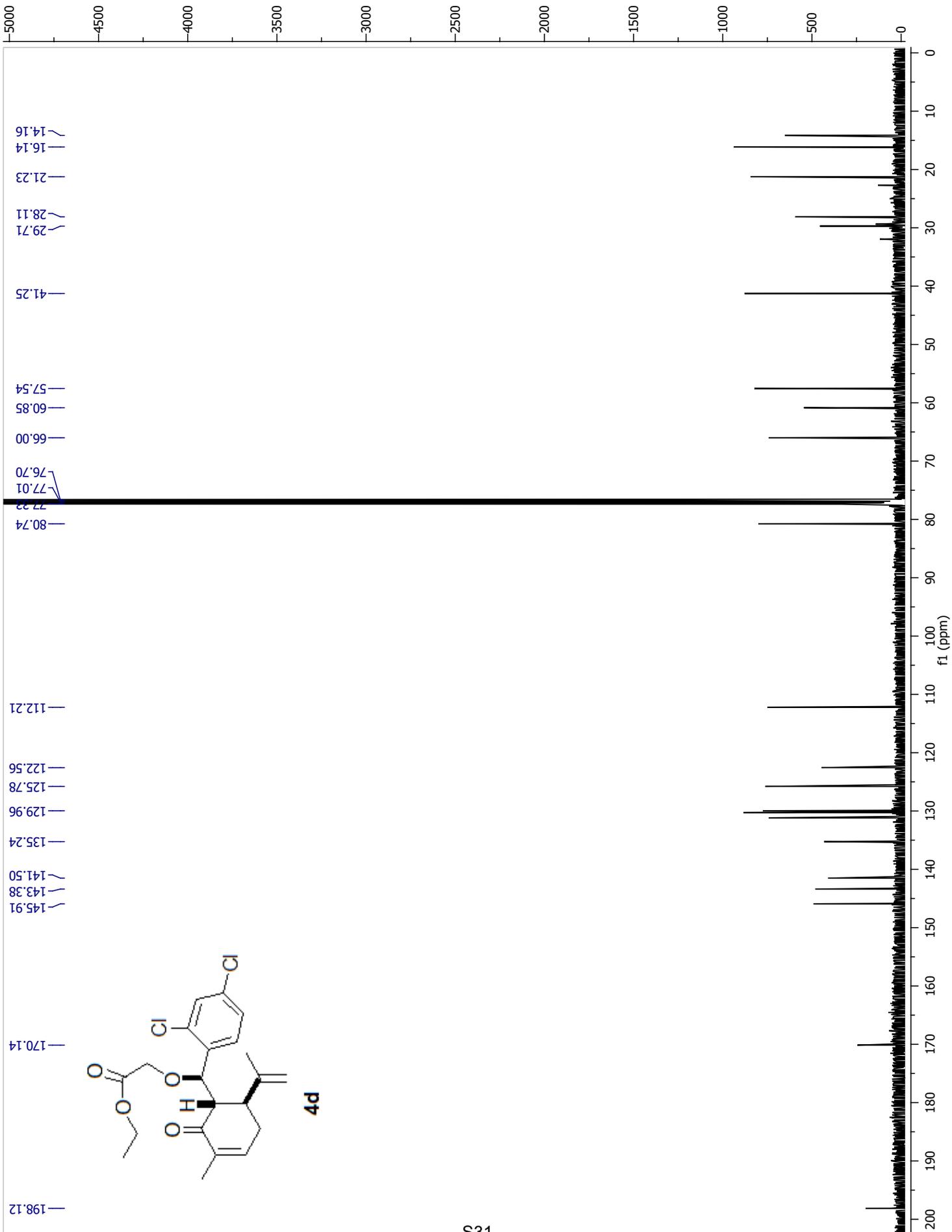












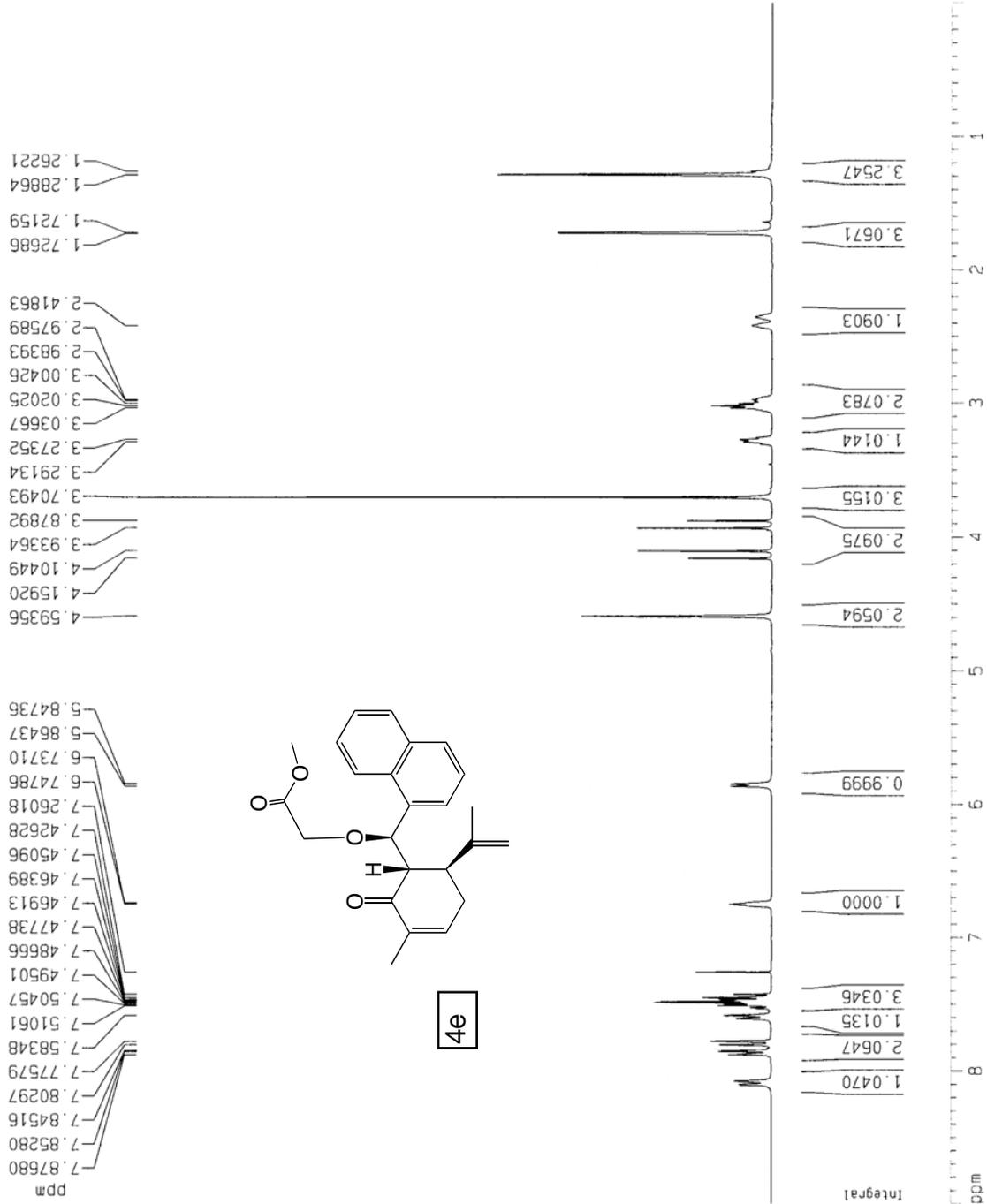
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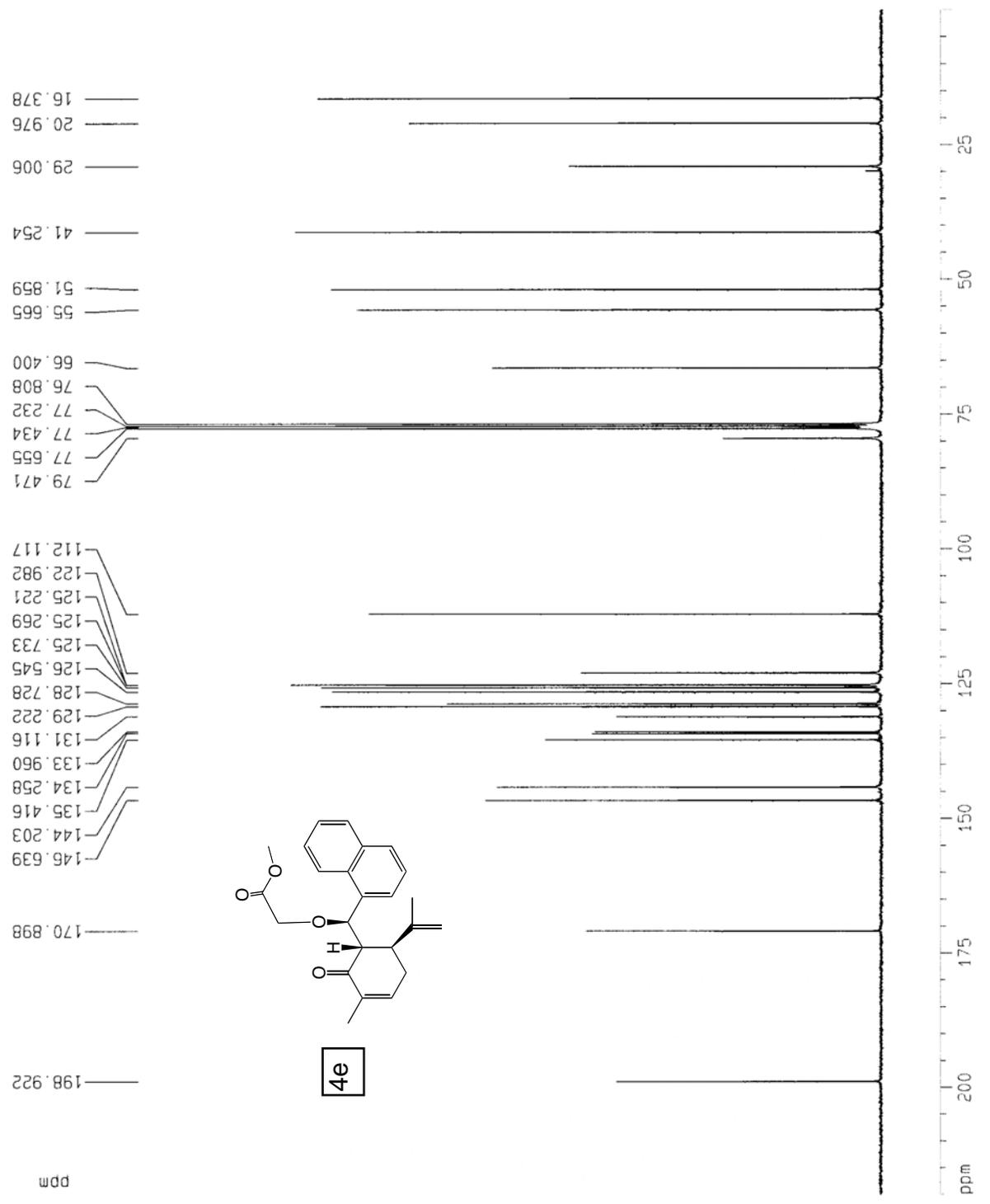
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 D11 0.03000000 sec  
 DELTA 1.89999998 sec  
 MCREST 0.00000000 sec  
 MCMRK 0.01500000 sec

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.38 usec  
 PL1 -2.80 dB  
 SF01 75.4760505 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -1.00 dB  
 PL12 15.33 dB  
 PL13 16.00 dB  
 SF02 300.1312005 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4677359 MHz  
 WDM EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

10 NMR plot parameters  
 CX 20.00 cm  
 CY 10.00 cm  
 F1P 220.000 ppm  
 F1 16602.90 Hz  
 F2P 0.000 ppm  
 F2 0.00 Hz  
 PPMCM 11.00000 ppm/cm  
 HZCM 830.14508 Hz/cm





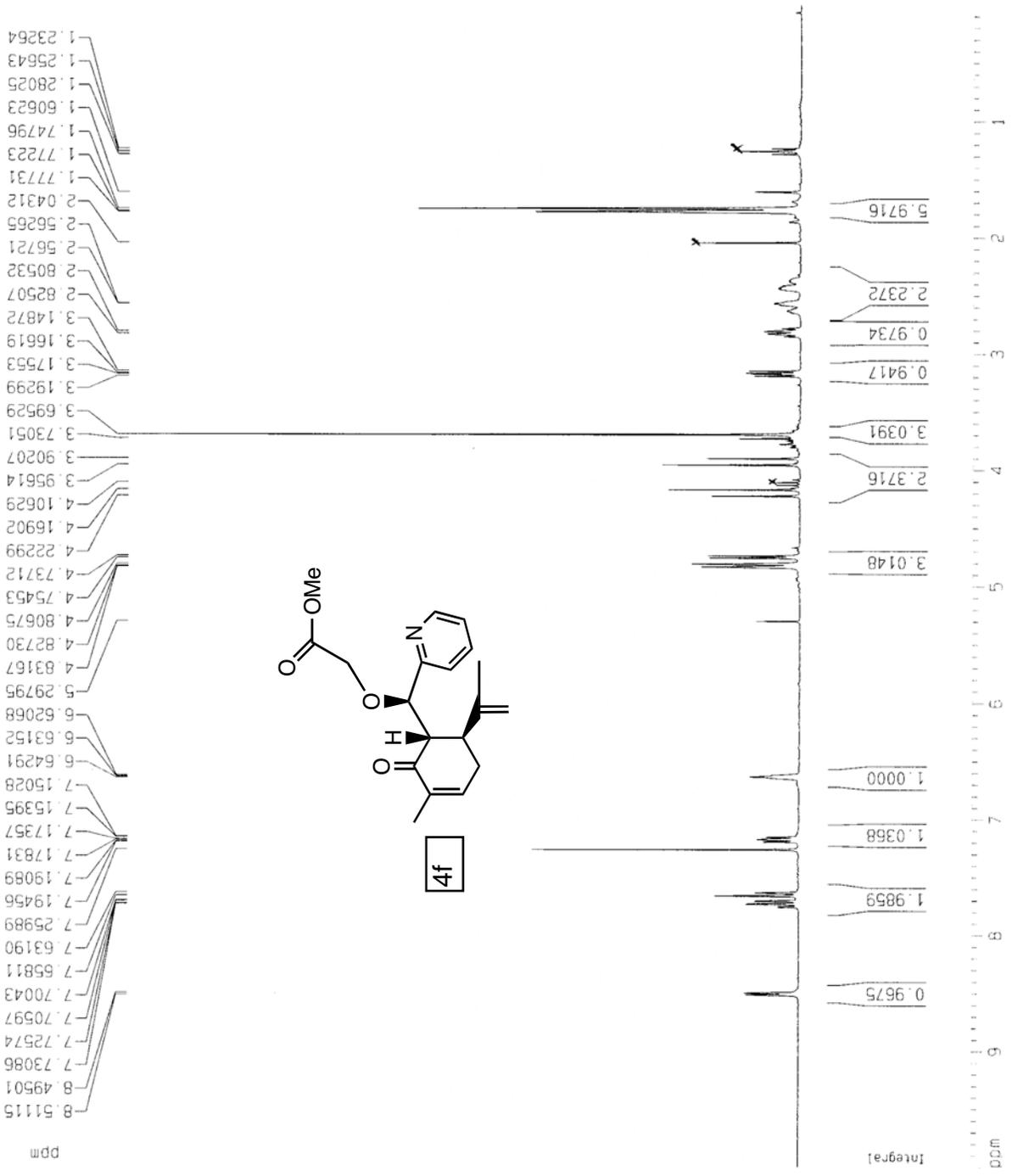
Current Data Parameters  
NAME km-1286  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20071202  
Time 12.00  
INSTRUM spect  
PROBHD 5 mm PAB80 BB-  
PULPROG zg30  
TD 17964  
SOLVENT COC13  
NS 16  
DS 2  
SWH 4495.403 Hz  
FIDRES 0.250022 Hz  
AQ 1.9998708 sec  
RG 362  
DM 111.200 usec  
DE 6.00 usec  
TE 295.5 K  
D1 1.00000000 sec  
MCREST 0.00000000 sec  
MCWK 0.01500000 sec

==== CHANNEL f1 =====  
NUC1 1H  
P1 12.20 usec  
PL1 -0.60 dB  
SF01 300.1321009 MHz

F2 - Processing parameters  
SI 32788  
SF 300.1300059 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

ID NMR plot parameters  
CX 20.00 cm  
CY 12.00 cm  
F1P 10.000 ppm  
F1 3001.30 Hz  
F2P 0.000 ppm  
F2 0.00 Hz  
PPMCM 0.50000 ppm/cm  
HZCM 150.06500 Hz/cm



Current Data Parameters  
 NAME km-1286  
 EXPNO 2  
 PROCNO 1

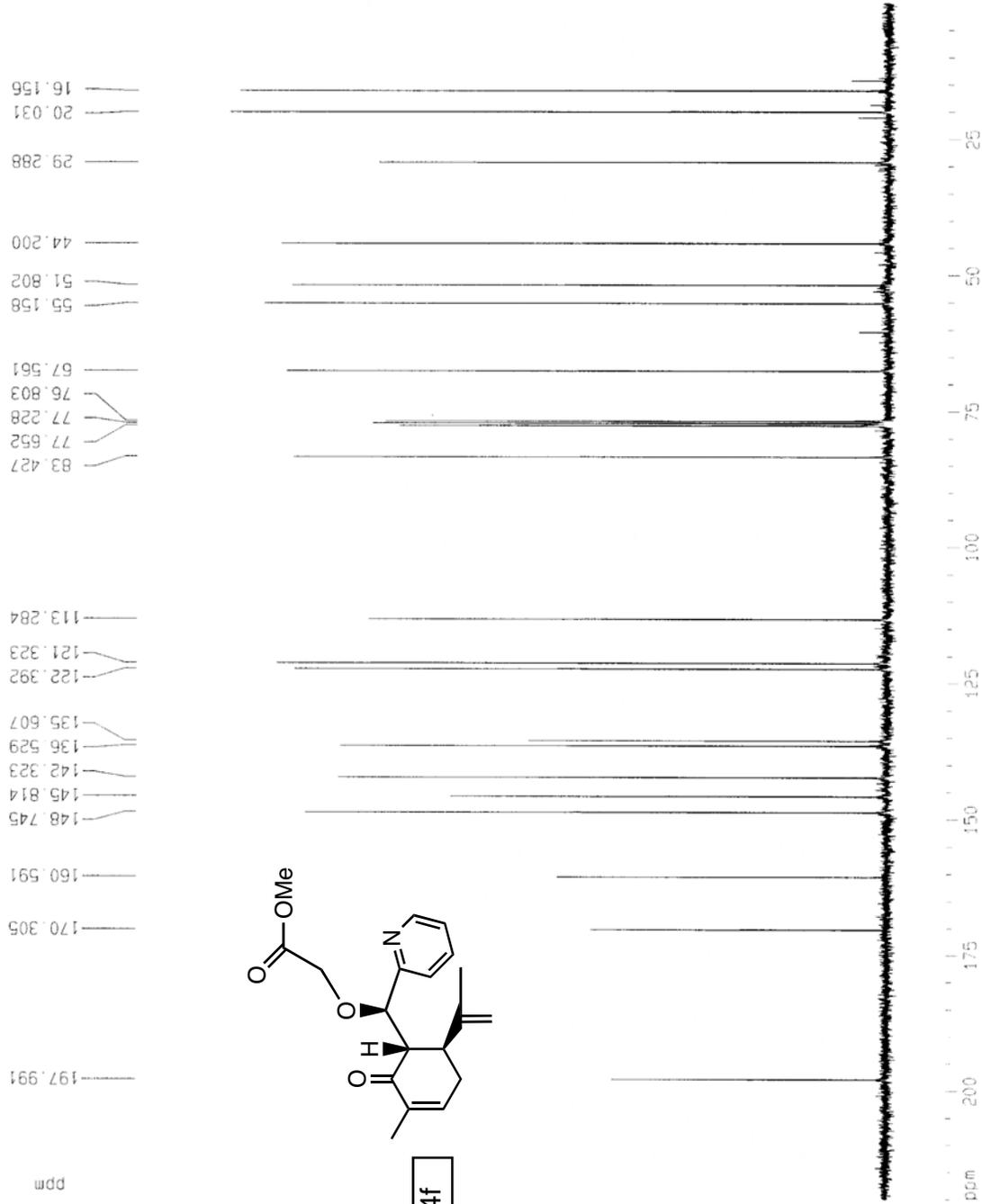
F2 - Acquisition Parameters  
 Date\_ 20071203  
 Time\_ 11:18  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zgpg30  
 TD 32372  
 SOLVENT CDCl3  
 NS 400  
 DS 4  
 SWH 18115.941 Hz  
 FIDRES 0.559618 Hz  
 AQ 0.8935172 sec  
 RG 23170.5  
 DM 27.600 usec  
 DE 6.00 usec  
 TE 297.0 K  
 D1 2.0000000 sec  
 D11 0.0300000 sec  
 DELTA 1.8999998 sec  
 WREST 0.0000000 sec  
 WCRK 0.01500000 sec

\*\*\*\*\* CHANNEL f1 \*\*\*\*\*  
 NUC1 13C  
 P1 8.38 usec  
 PL1 -2.80 dB  
 SF01 75.4752953 MHz

\*\*\*\*\* CHANNEL f2 \*\*\*\*\*  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -0.60 dB  
 PL12 15.73 dB  
 PL13 16.00 dB  
 SF02 300.1312005 MHz

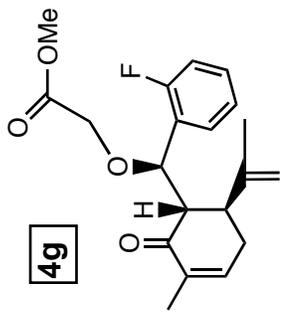
F2 - Processing parameters  
 SI 32768  
 SF 75.4677415 MHz  
 MDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

1D NMR plot parameters  
 CX 20.00 cm  
 CY 11.00 cm  
 F1 220.118 ppm  
 F2 15611.81 Hz  
 ZP -0.000 ppm  
 Z 11.00590 ppm/cm  
 PPMCM 830.55045 Hz/cm  
 HZCM



ppm

7.44273  
7.43694  
7.41806  
7.41210  
7.27318  
7.26010  
7.24720  
7.18012  
7.17717  
7.15512  
7.15226  
6.99782  
6.99470  
6.97041  
6.96399  
6.95985  
6.93618  
6.66153  
5.29838  
5.27042  
5.24518  
4.70105  
4.66250  
4.10737  
4.05226  
3.90304  
3.84791  
3.77130  
3.73132  
3.72057  
3.32755  
3.31859  
3.30957  
2.95767  
2.94813  
2.93878  
2.91437  
2.90148  
2.88922  
2.87629  
2.46752  
2.40234  
1.72733  
1.70581  
1.70090  
1.58691  
1.56251



F2 - Acquisition Parameters

Date\_ 20071002  
Time 8.39  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zg30  
TD 17984  
SOLVENT CDCl3  
NS 16  
DS 2  
SWH 4496.403 Hz  
FIDRES 0.250022 Hz  
AQ 1.9998708 sec  
RG 322.5  
DW 111.200 usec  
DE 6.00 usec  
TE 296.2 K  
D1 1.00000000 sec  
MCREST 0.00000000 sec  
MCWRR 0.01500000 sec

==== CHANNEL f1 =====

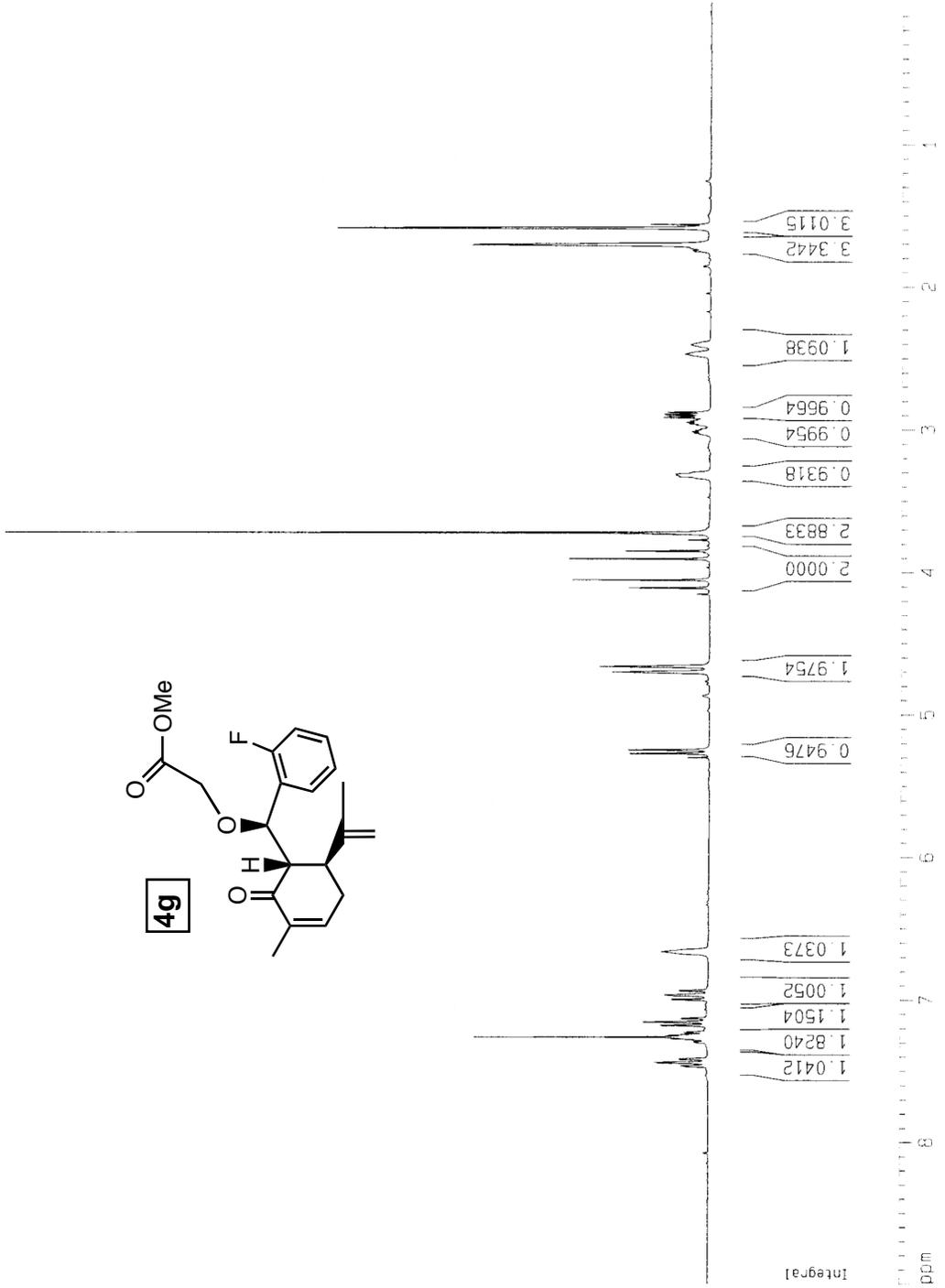
NUC1 1H  
P1 12.20 usec  
PL1 -0.60 dB  
SFO1 300.1321009 MHz

F2 - Processing parameters

SI 32768  
SF 300.1300066 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

1D NMR plot parameters

CX 20.00 cm  
CY 11.00 cm  
F1P 9.000 ppm  
F1 2701.17 Hz  
F2P 0.000 ppm  
F2 0.00 Hz  
PPMCM 0.45000 ppm/cm  
HZCM 135.05850 Hz/cm



Current Data Parameters  
 NAME km-1240  
 EXPNO 2  
 PROCNO 1

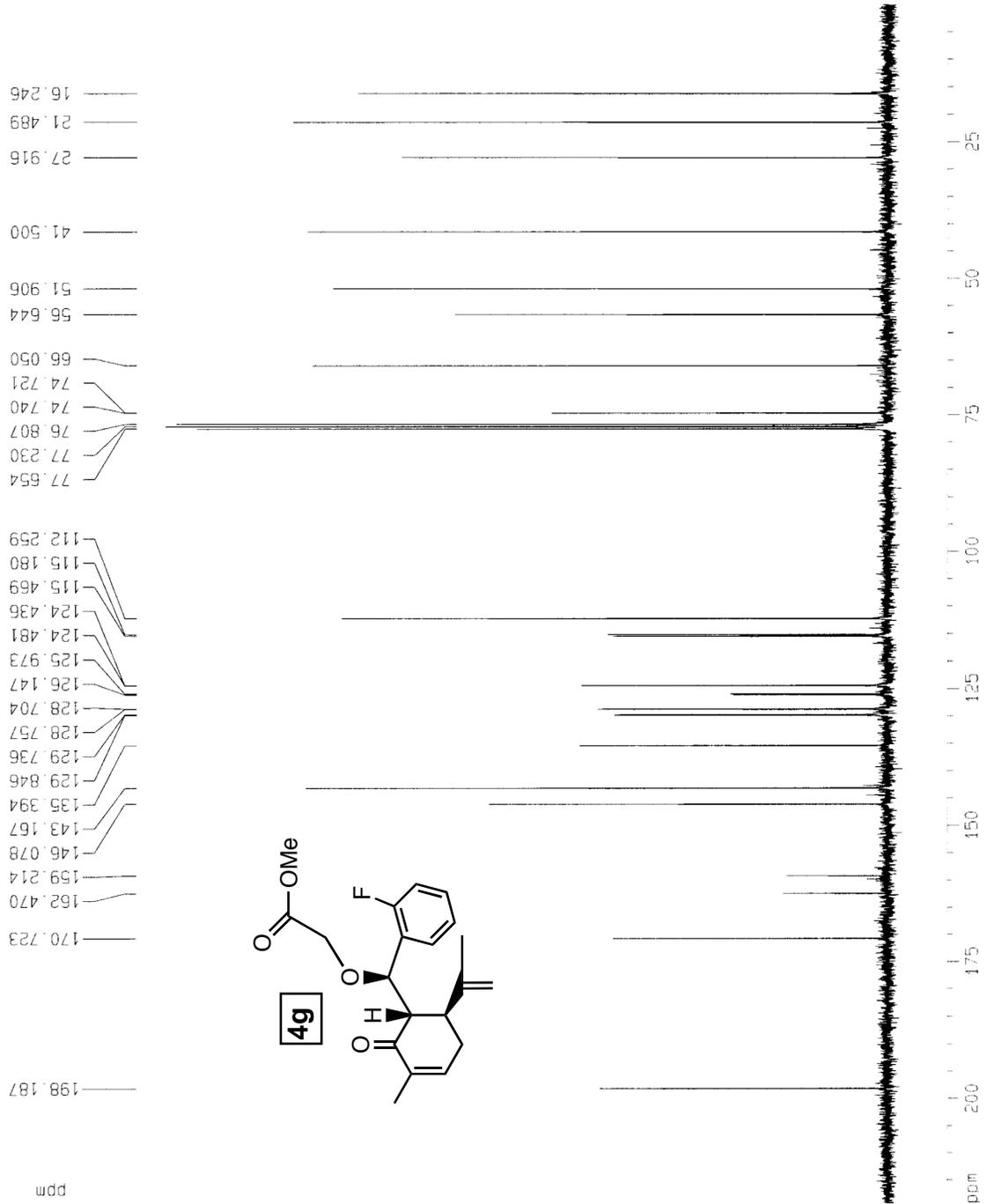
F2 - Acquisition Parameters  
 Date\_ 20071003  
 Time 13.24  
 INSTRUM spect  
 PRQBD 5 mm PABBO BB-  
 PULPROG zgpg30  
 ID 32372  
 SOLVENT CDCl3  
 NS 500  
 DS 4  
 SWH 17985.611 Hz  
 FIDRES 0.55592 Hz  
 AQ 0.899916 sec  
 RG 18390.4  
 DM 27.800 usec  
 DE 6.00 usec  
 TE 296.8 K  
 D1 2.00000000 sec  
 d11 0.03000000 sec  
 DELTA 1.8959998 sec  
 XCREST 0.00000000 sec  
 XCMARK 0.01500000 sec

\*\*\*\*\* CHANNEL f1 \*\*\*\*\*  
 NUC1 13C  
 P1 8.38 usec  
 PL1 -2.80 dB  
 SF01 75.4752953 MHz

\*\*\*\*\* CHANNEL f2 \*\*\*\*\*  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -0.60 dB  
 PL12 15.73 dB  
 PL13 16.00 dB  
 SF02 300.1312005 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4677363 MHz  
 MDM EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

1D NMR plot parameters  
 CX 20.00 cm  
 CY 10.00 cm  
 F1P 219.323 ppm  
 F1 16551.79 Hz  
 F2P -0.000 ppm  
 F2 -0.00 Hz  
 PPMCM 10.96613 ppm/cm  
 HZCM 827.58929 Hz/cm





Current Data Parameters  
 NAME Km-1354  
 EXPNO 2  
 PROCNO 1

F2 - Acquisition Parameters

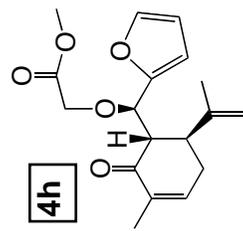
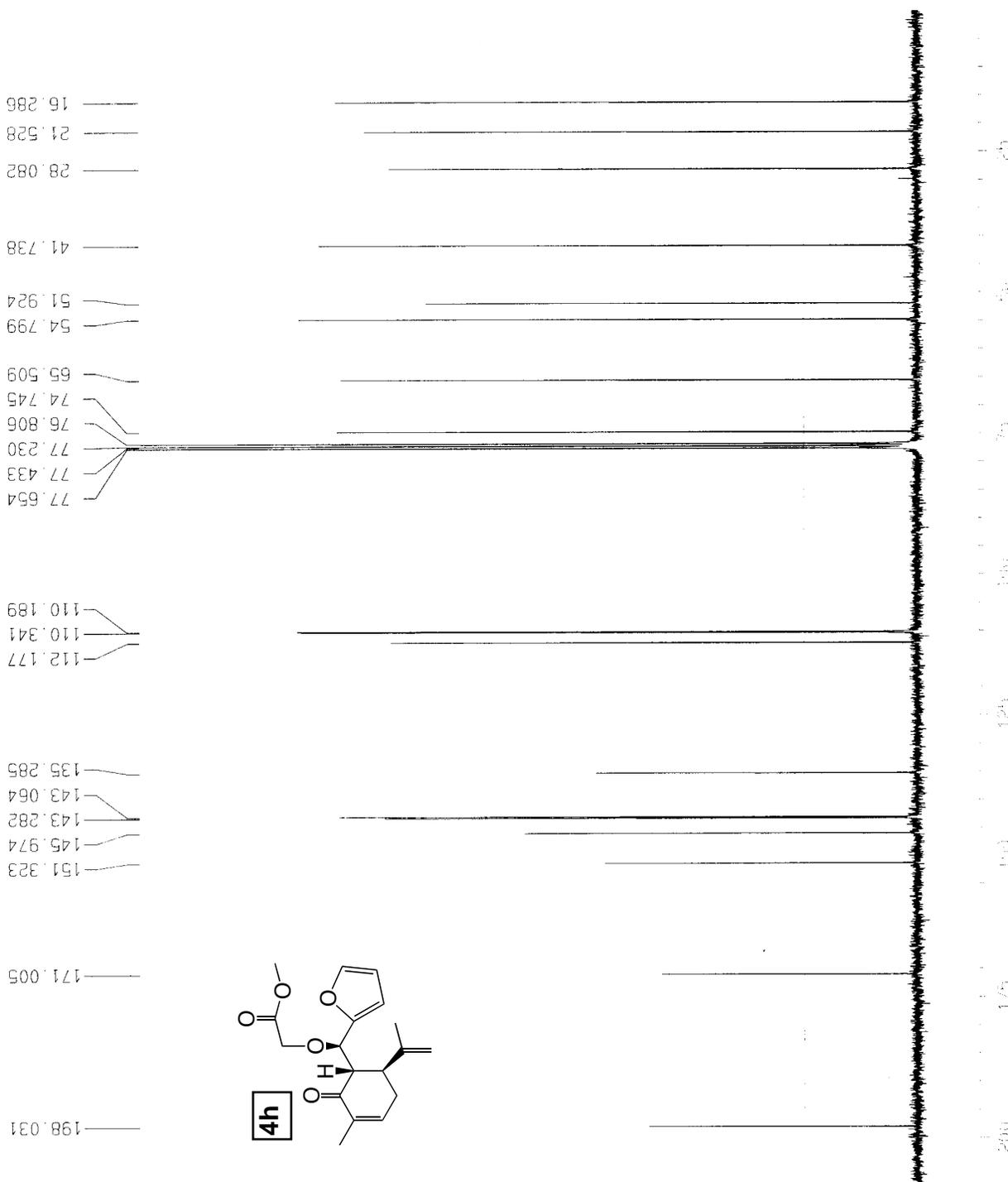
Date\_ 20080210  
 Time 9.05  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zgpg30  
 TO 36230  
 SOLVENT CDCl3  
 NS 2000  
 DS 4  
 SWH 18115.941 Hz  
 FIDRES 0.500026 Hz  
 AQ 0.9999980 sec  
 RG 23170.5  
 DW 27.600 usec  
 DE 6.00 usec  
 TE 297.0 K  
 J1 2.00000000 sec  
 J11 0.03000000 sec  
 DELTA 1.89999996 sec  
 MCREST 0.00000000 sec  
 MCLRK 0.01500000 sec

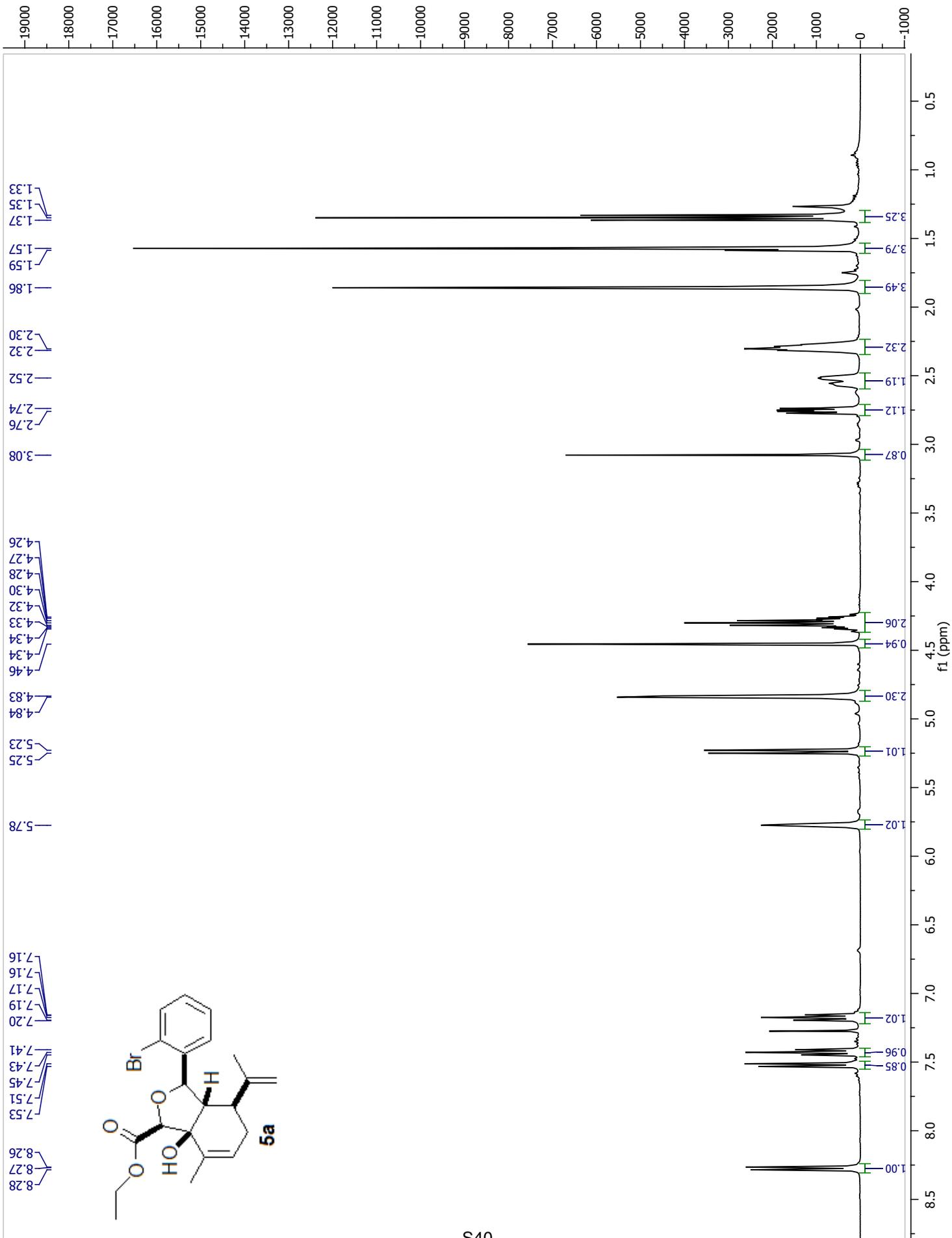
==== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.38 usec  
 PL1 -2.80 dB  
 SF01 75.4752953 MHz

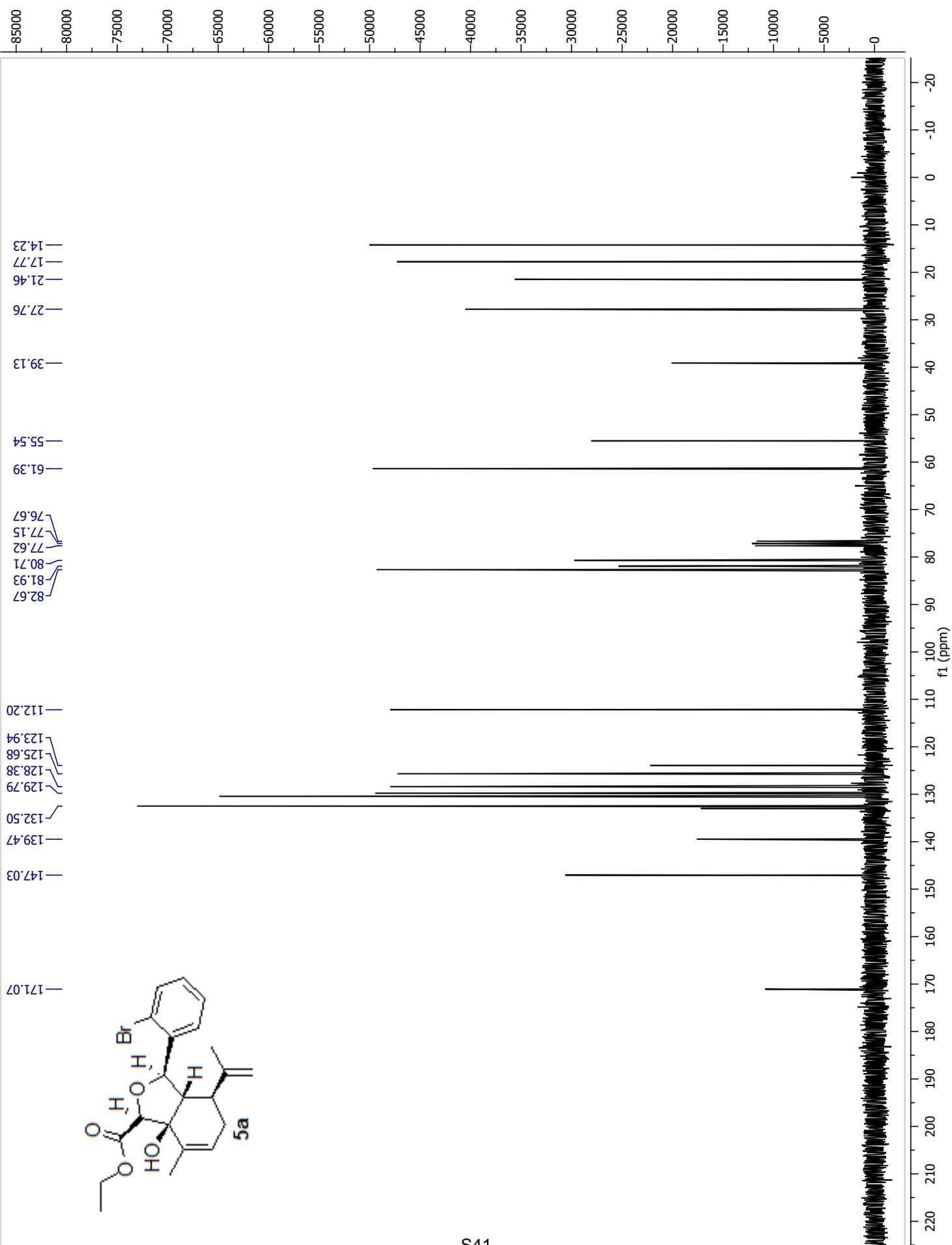
==== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -1.00 dB  
 PL12 15.33 dB  
 PL13 16.00 dB  
 SF02 300.1312005 MHz

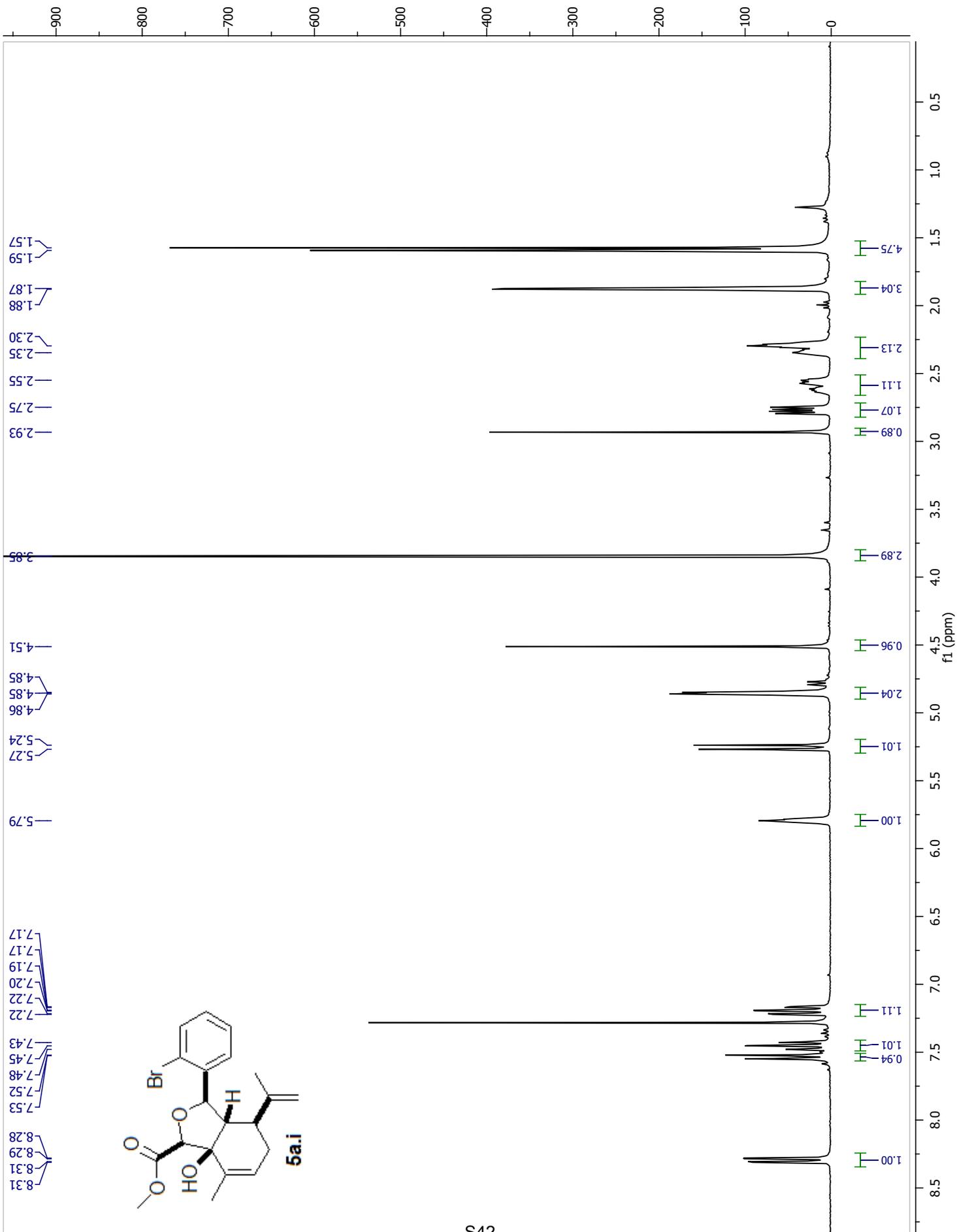
F2 - Processing parameters  
 SI 32768  
 SF 75.4677348 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

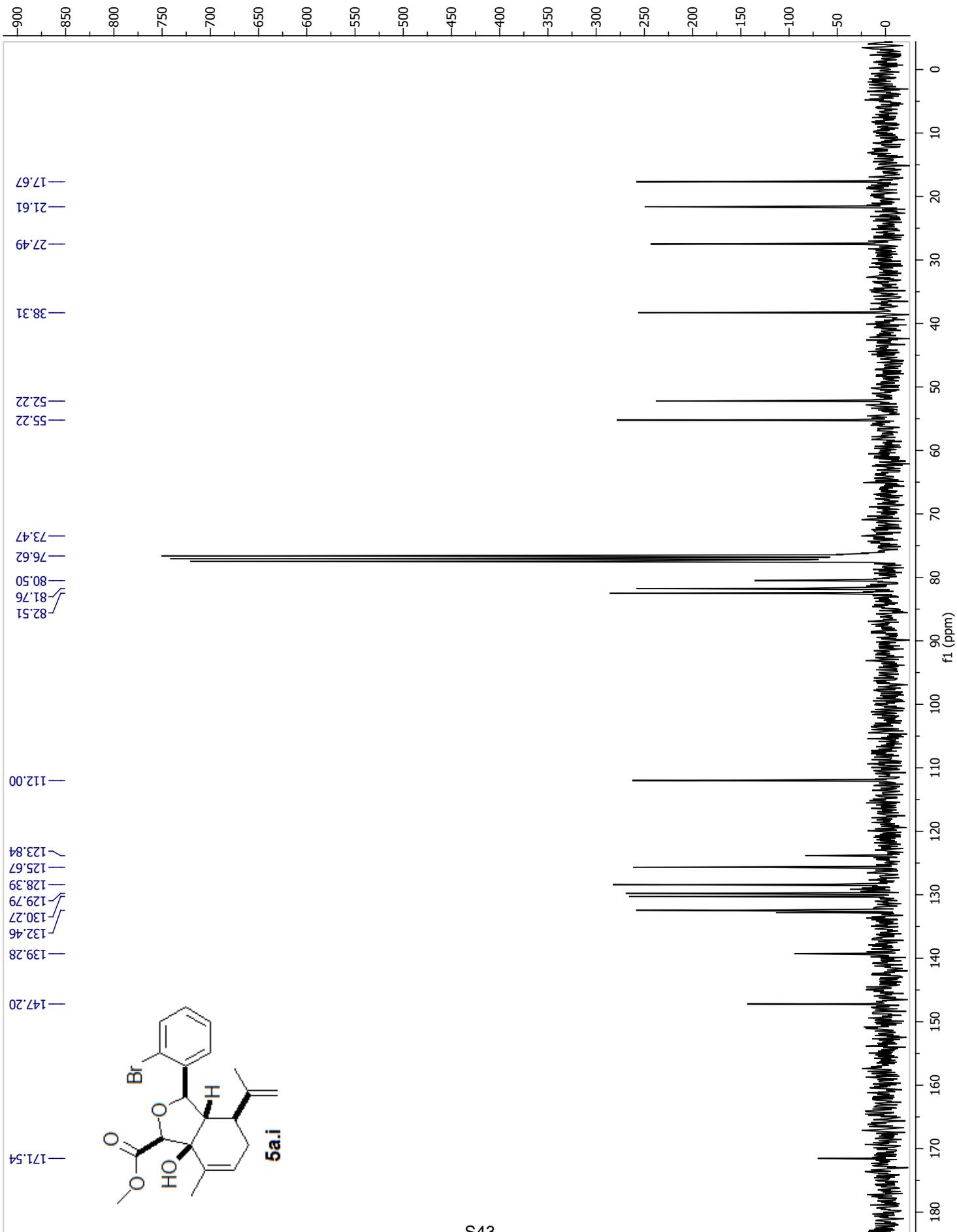
1D NMR plot parameters  
 CX 20.00 cm  
 CY 10.00 cm  
 F1p 220.206 ppm  
 F1 16618.44 Hz  
 F2p 0.000 ppm  
 F2 0.00 Hz  
 PPMCM 11.01030 ppm/cm  
 AZCM 830.92206 Hz/cm



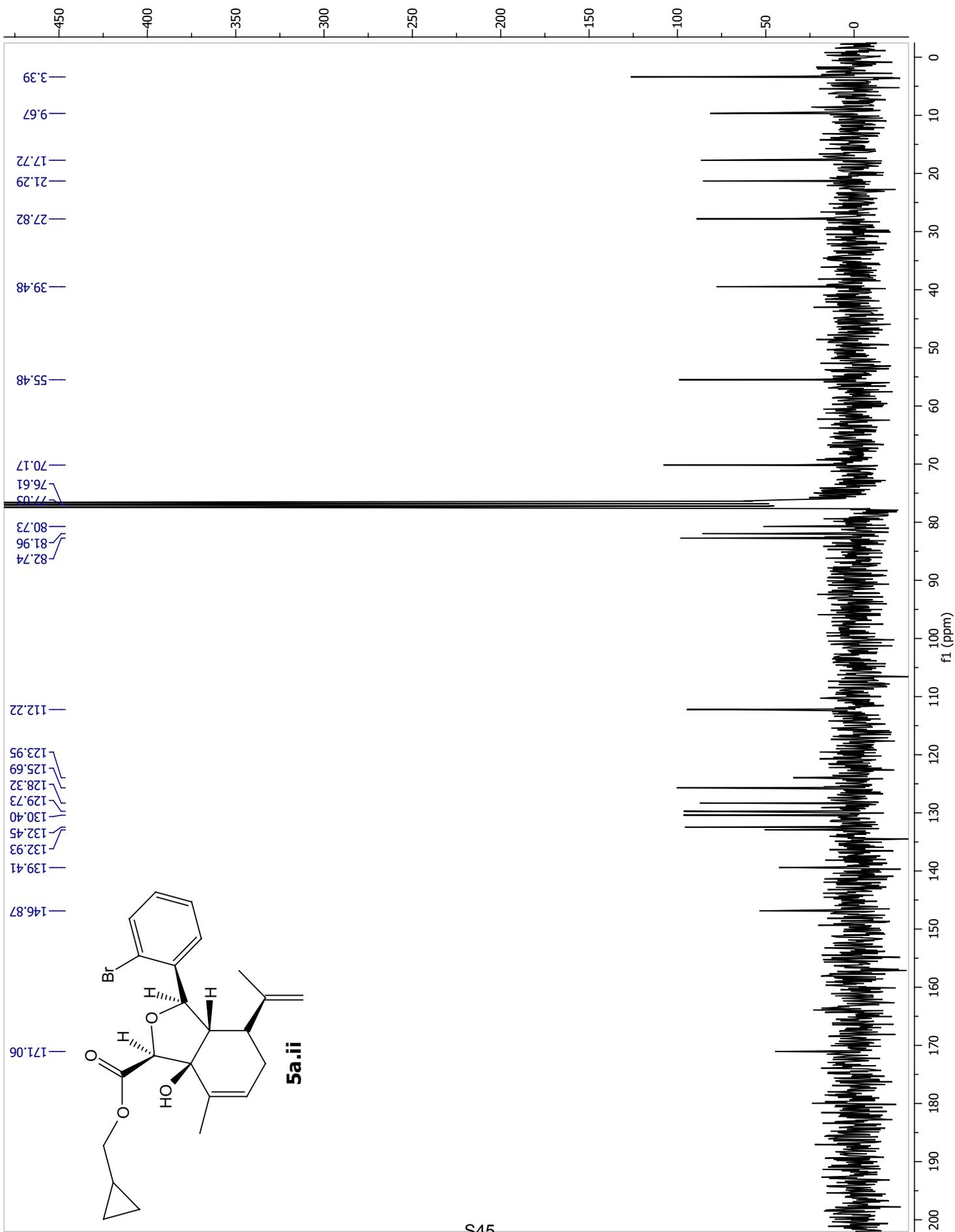


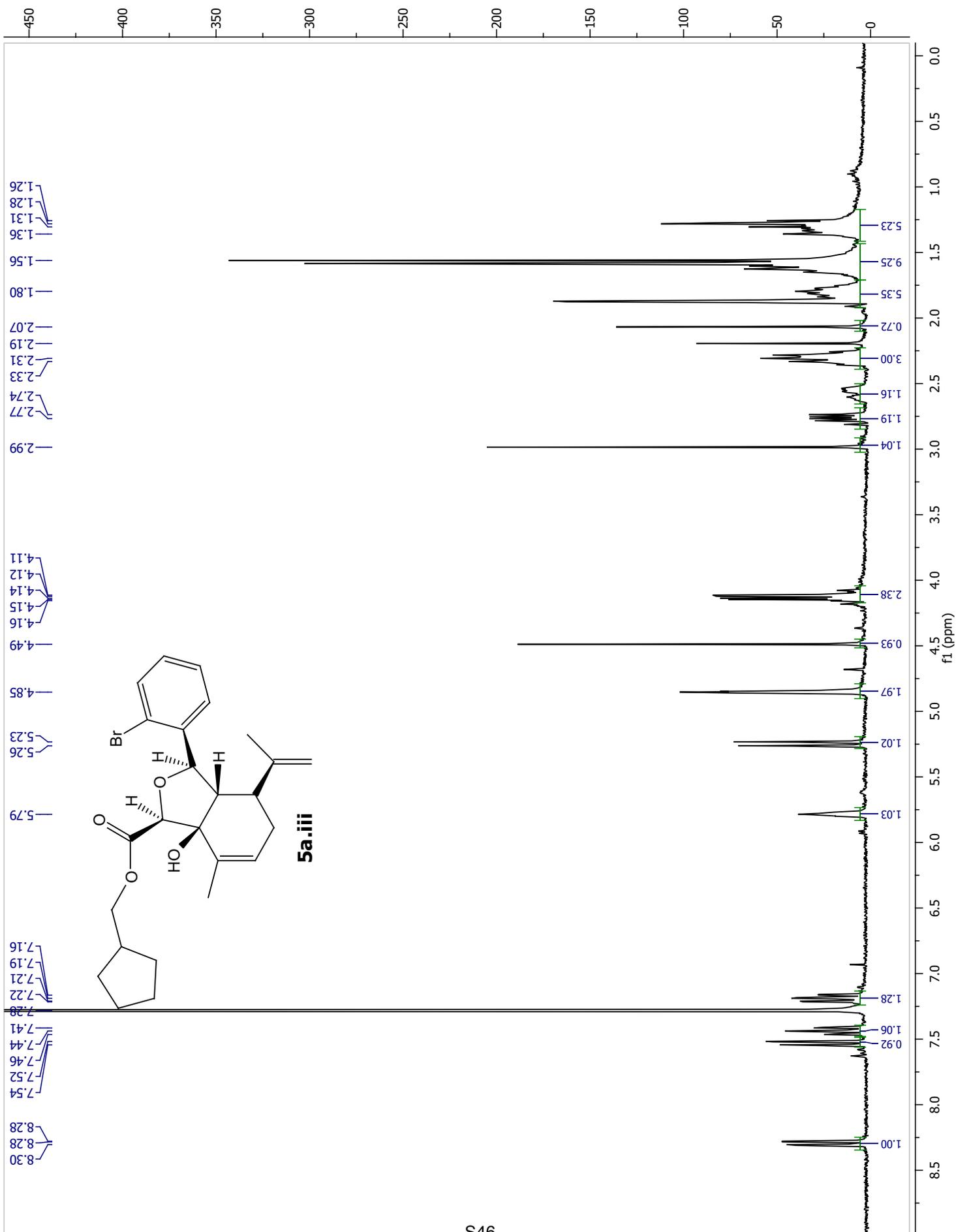


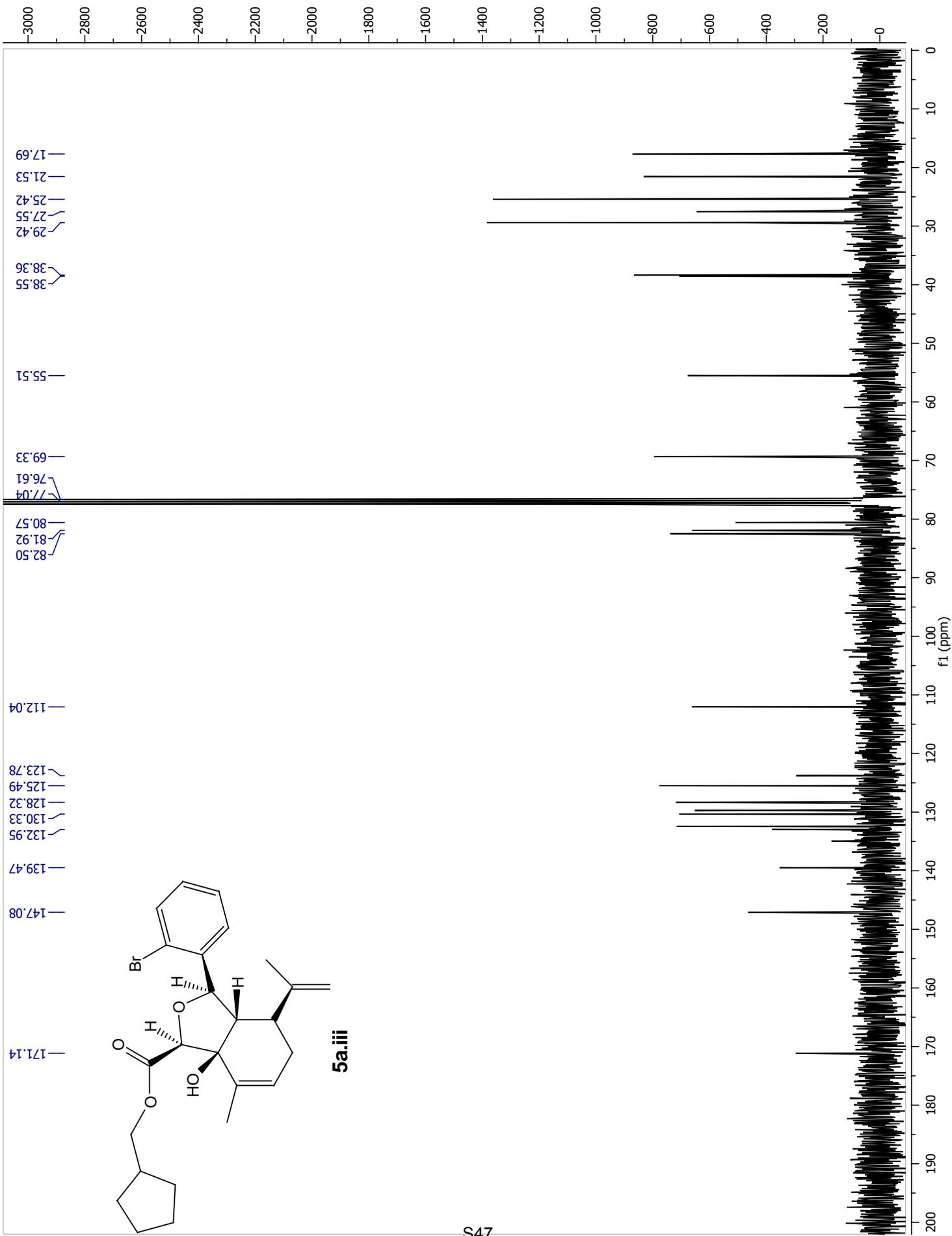


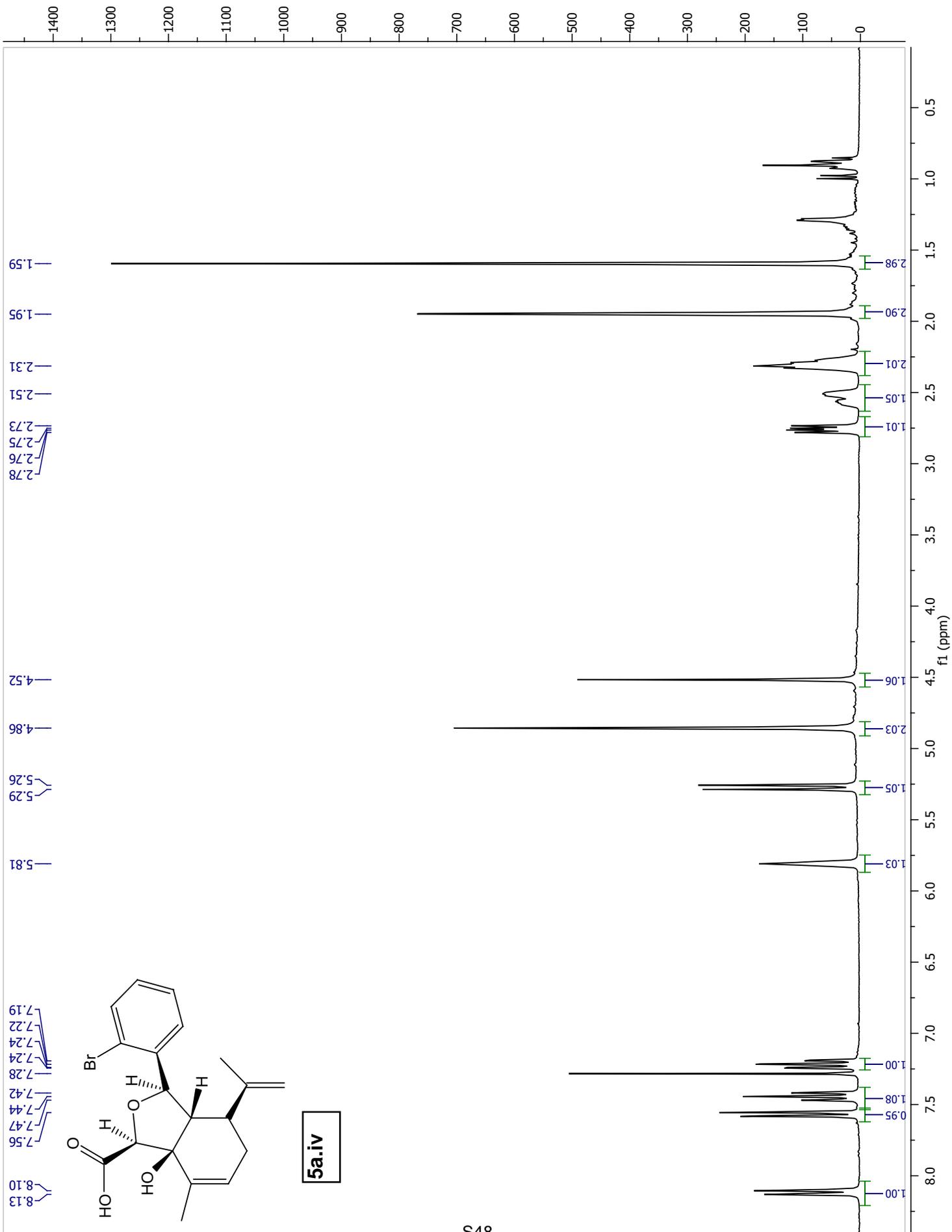


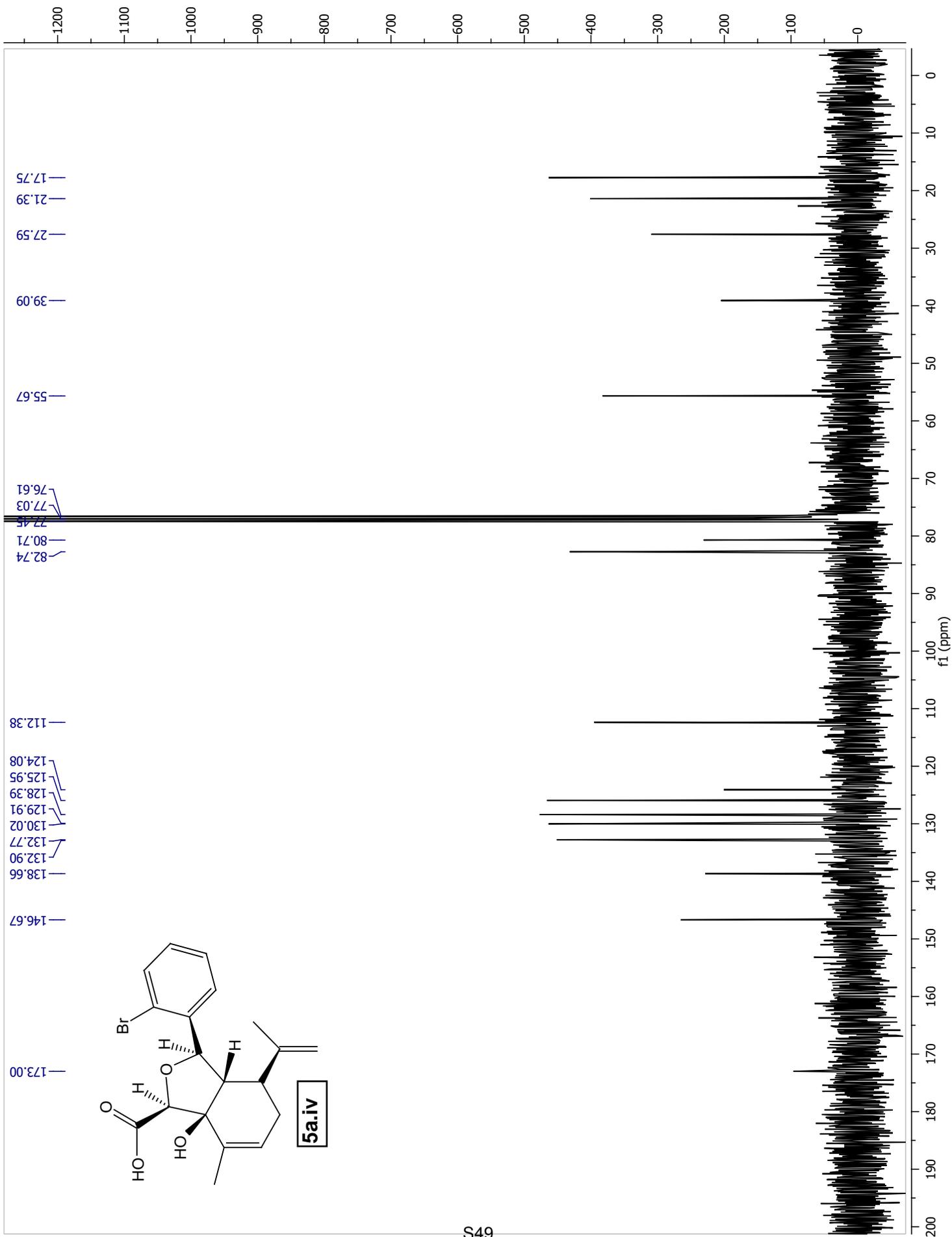




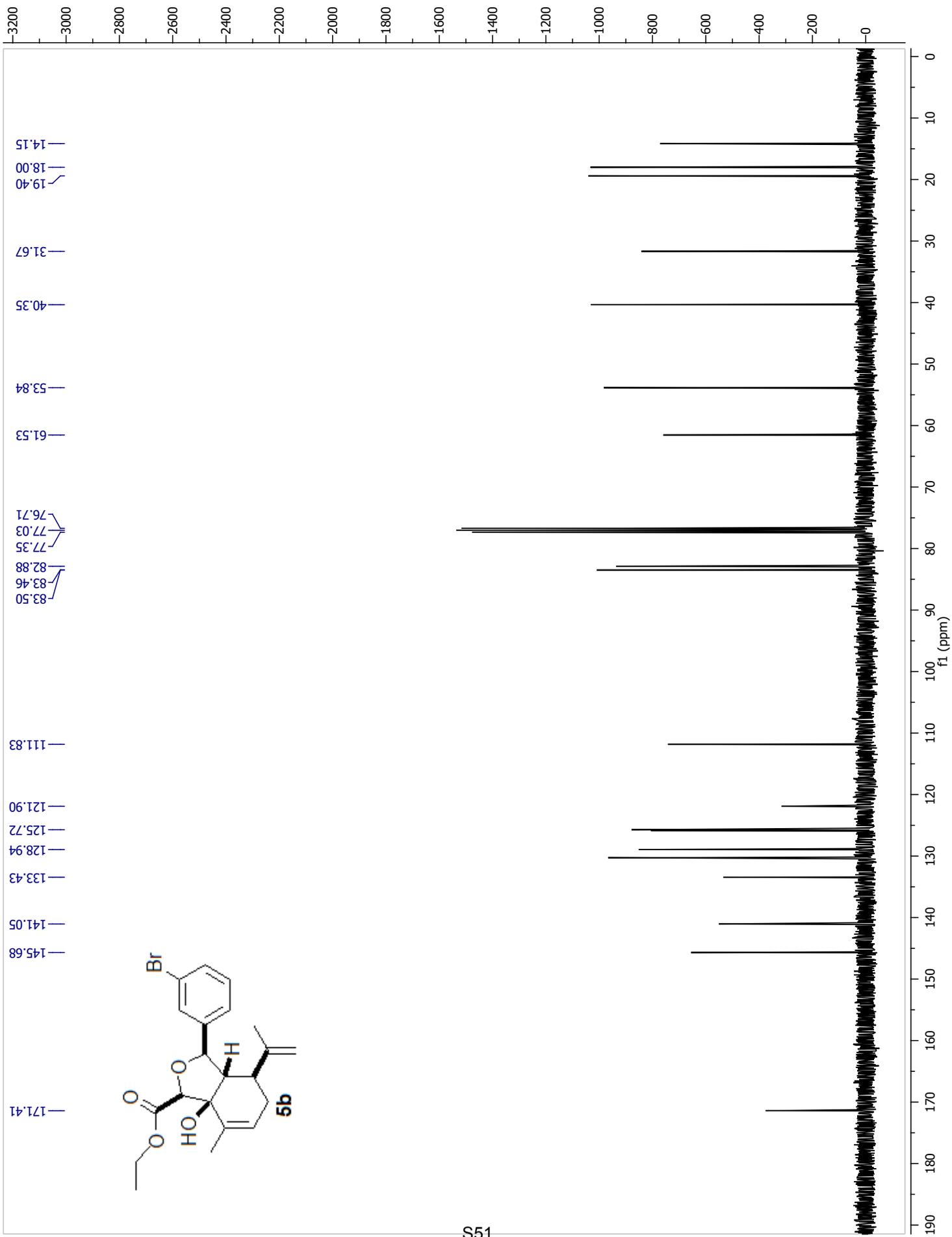


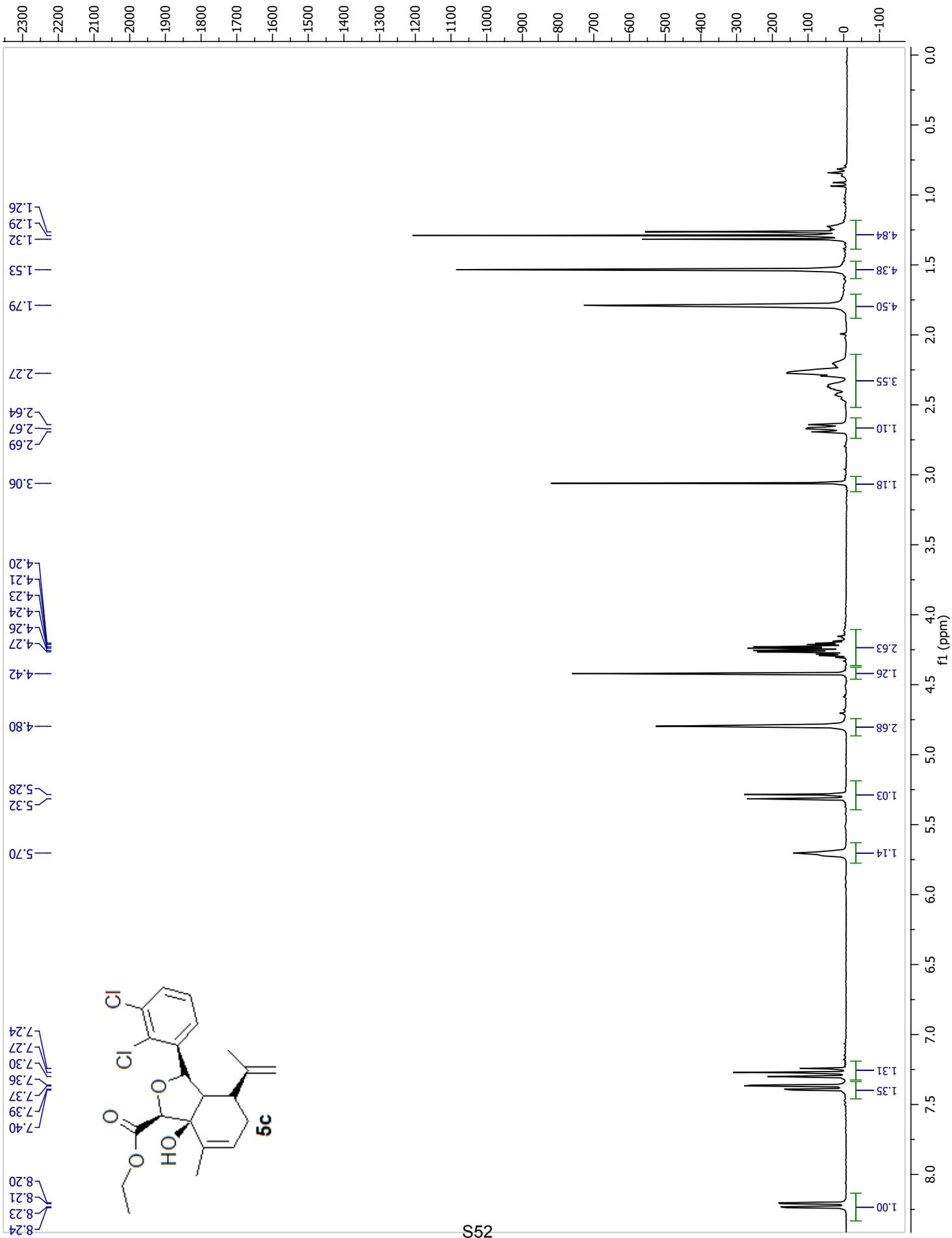


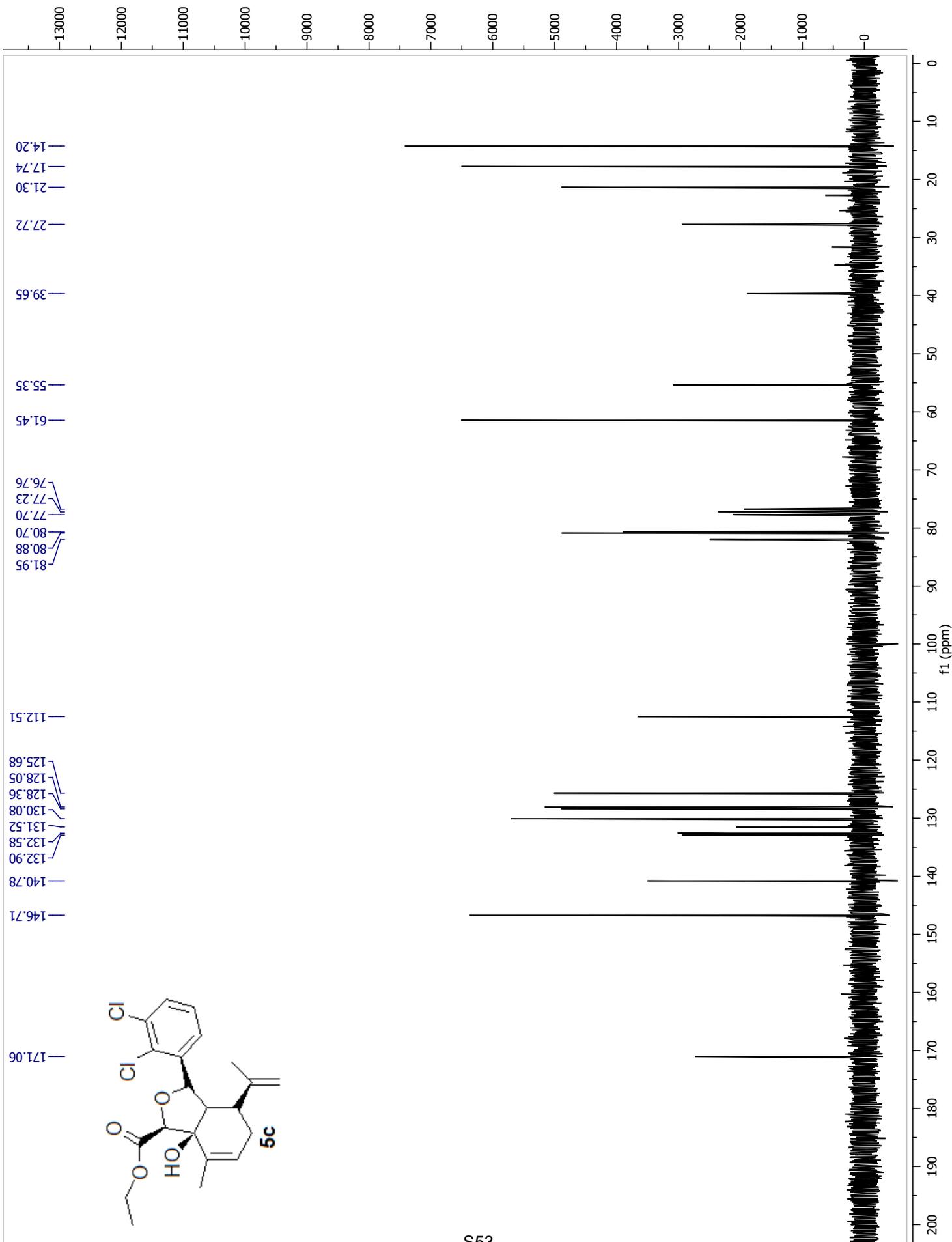


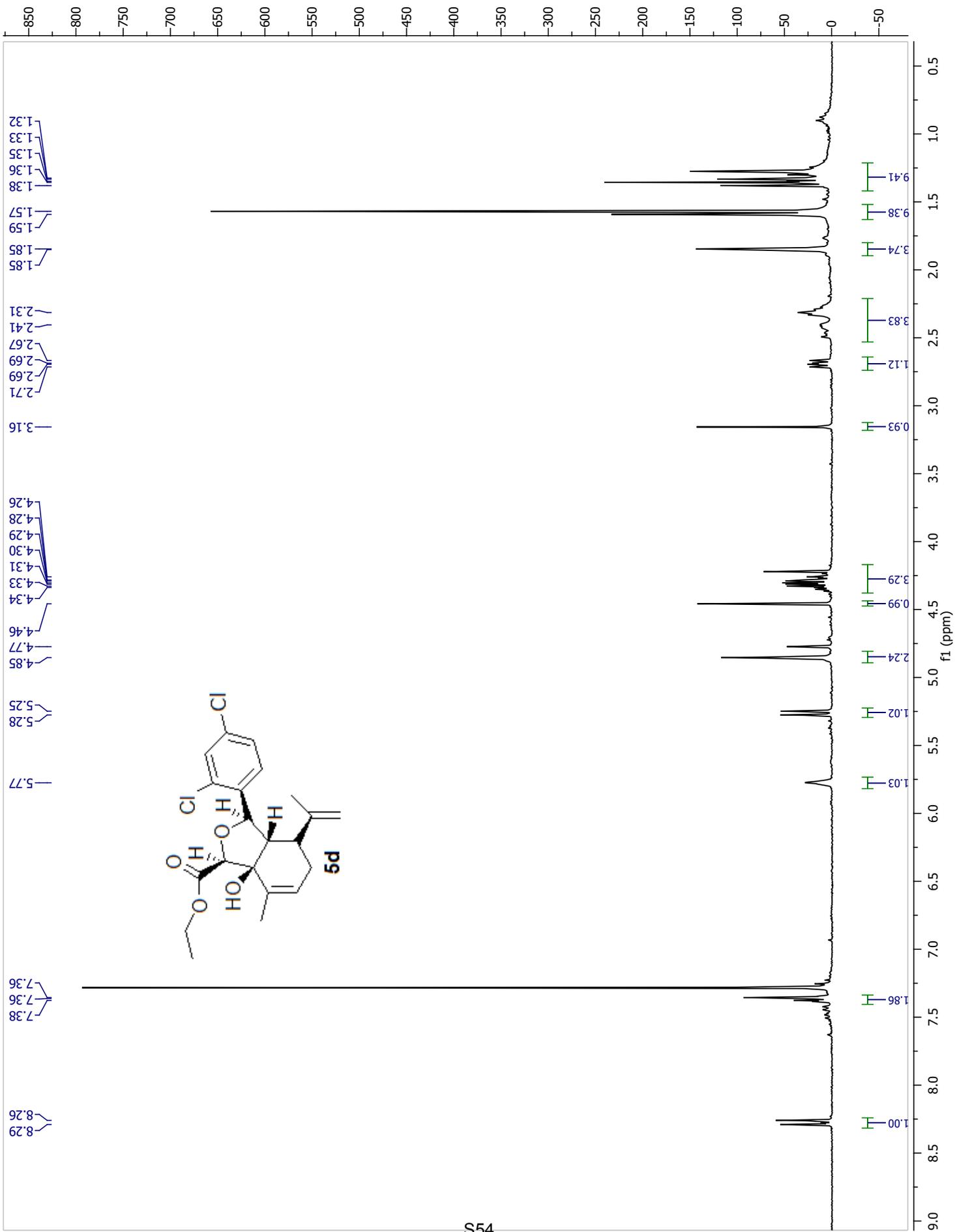


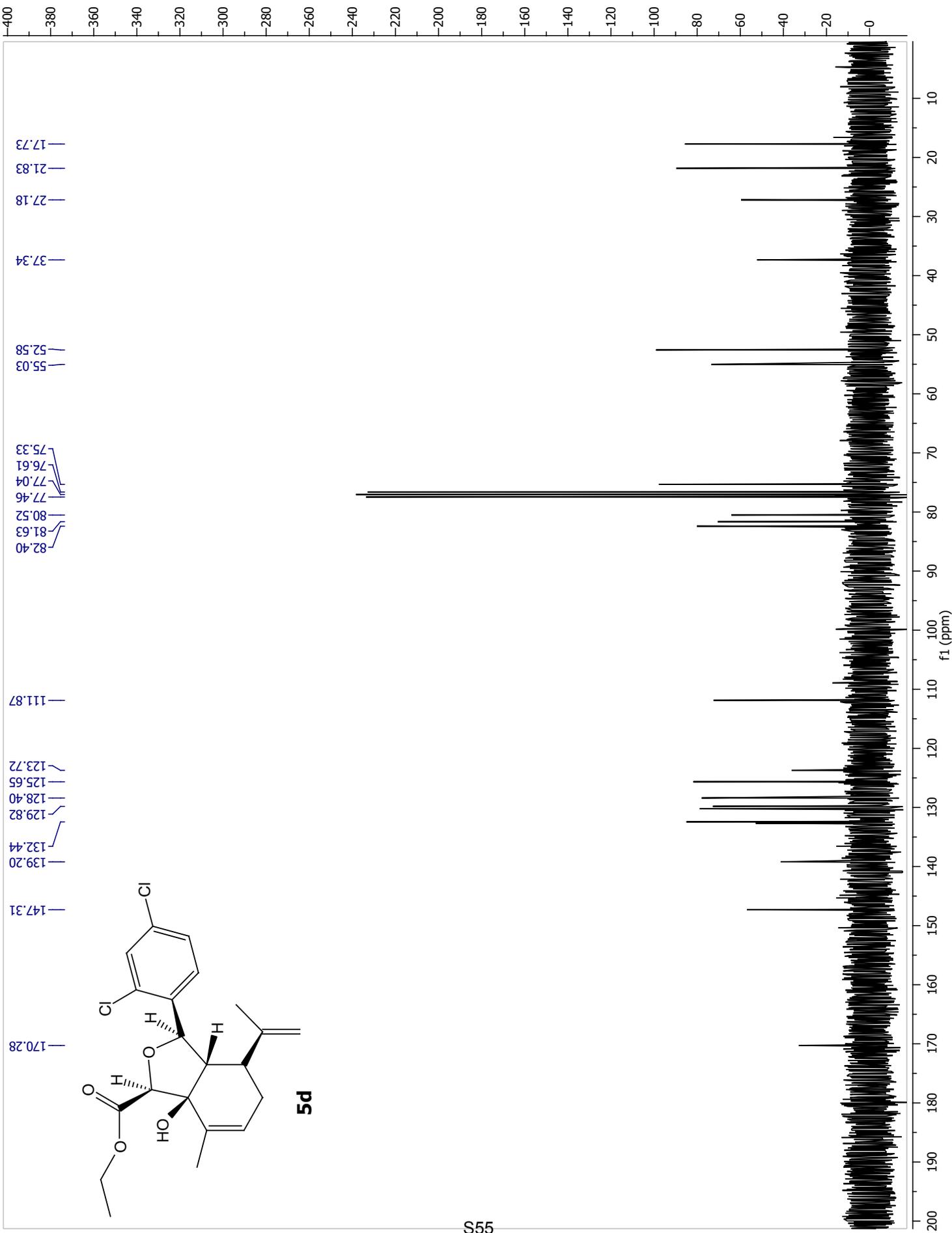












Current Data Parameters  
 NAME km-1255  
 EXPNO 11  
 PROCNO 1

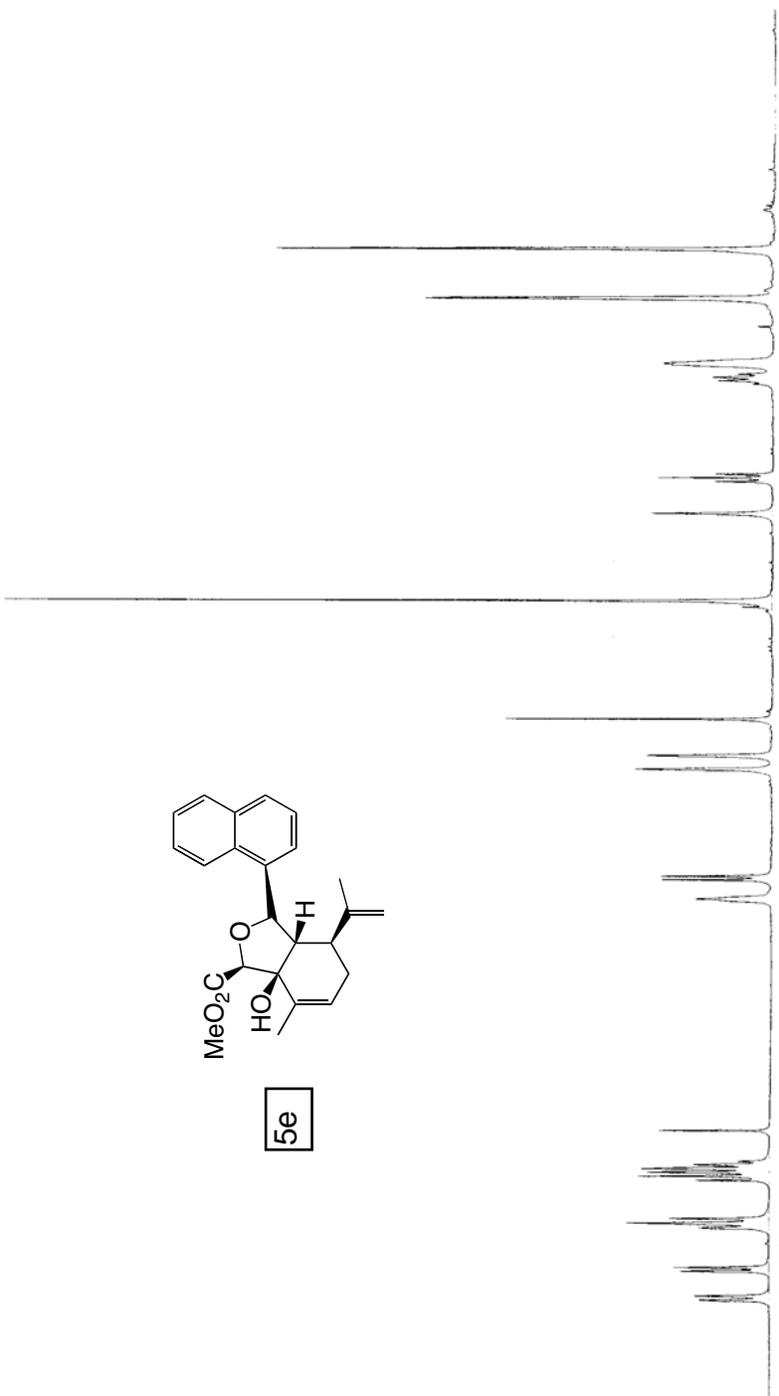
F2 - Acquisition Parameters  
 Date\_ 20080229  
 Time 11:51  
 INSTRUM spect  
 PROBHD 5 mm PABBO B2-  
 PULPROG zg30  
 TD 17984  
 SOLVENT CDCl3  
 VS 8  
 JS 2  
 SWH 4496.403 Hz  
 FIDRES 0.250022 Hz  
 40 1.9998708 sec  
 35.9  
 CW 111.200 usec  
 DE 6.00 usec  
 TE 296.8 K  
 D1 1.0000000 sec  
 VCREST 0.0000000 sec  
 VCMARK 0.0150000 sec

\*\*\*\*\* CHANNEL f1 \*\*\*\*\*  
 NUC1 1H  
 P1 12.20 usec  
 PL1 -1.00 dB  
 SF01 300.1321009 MHz

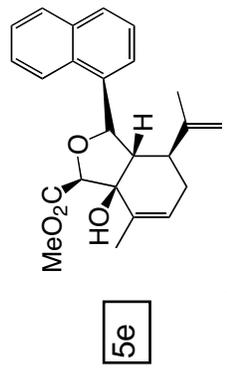
F2 - Processing parameters  
 SI 32768  
 SF 300.1300068 MHz  
 KDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

1D NMR plot parameters  
 CX 20.00 cm  
 CY 11.00 cm  
 F1 9.000 ppm  
 F2 2701.17 Hz  
 ZP 0.000 ppm  
 F2 0.00 Hz  
 ZPNCM 0.45000 ppm/cm

1.55154  
 1.87139  
 2.28859  
 2.29467  
 2.30049  
 2.36026  
 2.38236  
 2.40308  
 2.42440  
 3.00911  
 3.03306  
 3.05698  
 3.26222  
 3.82684  
 3.86531  
 4.59175  
 4.82718  
 4.91553  
 5.61196  
 5.63587  
 5.76035  
 7.25891  
 7.26081  
 7.48002  
 7.50157  
 7.50809  
 7.52896  
 7.55500  
 7.58065  
 7.82861  
 7.85793  
 7.86450  
 7.88949  
 8.14554  
 8.16950  
 8.32987  
 8.35676  
 ppm



2.9675  
 2.9460  
 1.9961  
 1.1017  
 0.9709  
 0.9109  
 3.0263  
 0.9849  
 2.0295  
 1.0175  
 1.0000  
 3.2155  
 2.1936  
 1.0119  
 0.9843  
 Integral



Current Data Parameters  
 NAME km-1255  
 EXPNO 10  
 PROCNO 1

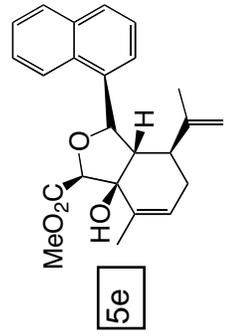
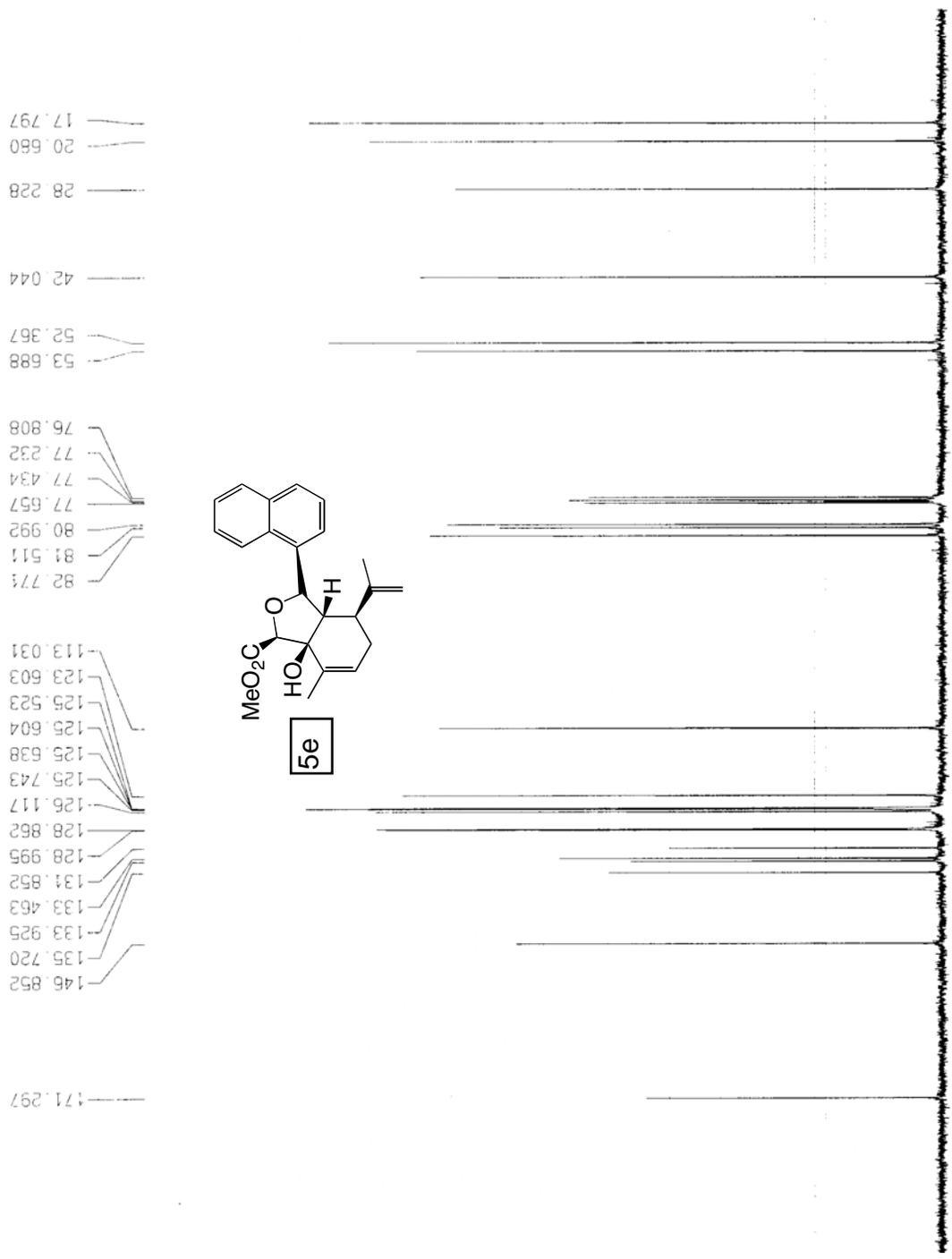
F2 - Acquisition Parameters  
 Date\_ 20080229  
 Time 11.48  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zgpg30  
 TO 32006  
 SOLVENT CDCl3  
 VS 400  
 DS 4  
 SWH 18115.941 Hz  
 FIDRES 0.555601 Hz  
 AQ 0.8999756 sec  
 RG 23170.5  
 CW 27.600 usec  
 DE 6.00 usec  
 TE 297.1 K  
 TI 2.00000000 sec  
 D1 0.03000000 sec  
 DELTA 1.89599988 sec  
 VCREST 0.00000000 sec  
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====  
 NU1 13C  
 P1 8.38 usec  
 PL1 -2.80 dB  
 SF01 75.4760505 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NU2 1H  
 PCPD2 60.00 usec  
 PL2 -1.00 dB  
 PL12 15.33 dB  
 PL13 16.00 dB  
 SF02 300.1312005 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4677447 MHz  
 KW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

1D NMR plot parameters  
 CX 20.00 cm  
 CY 10.00 cm  
 ZP 200.000 ppm  
 F1 15082.54 Hz  
 ZP 0.000 ppm  
 T2 0.16 sec  
 T2\* 16.000 sec  
 T2\*\* 15.000 sec



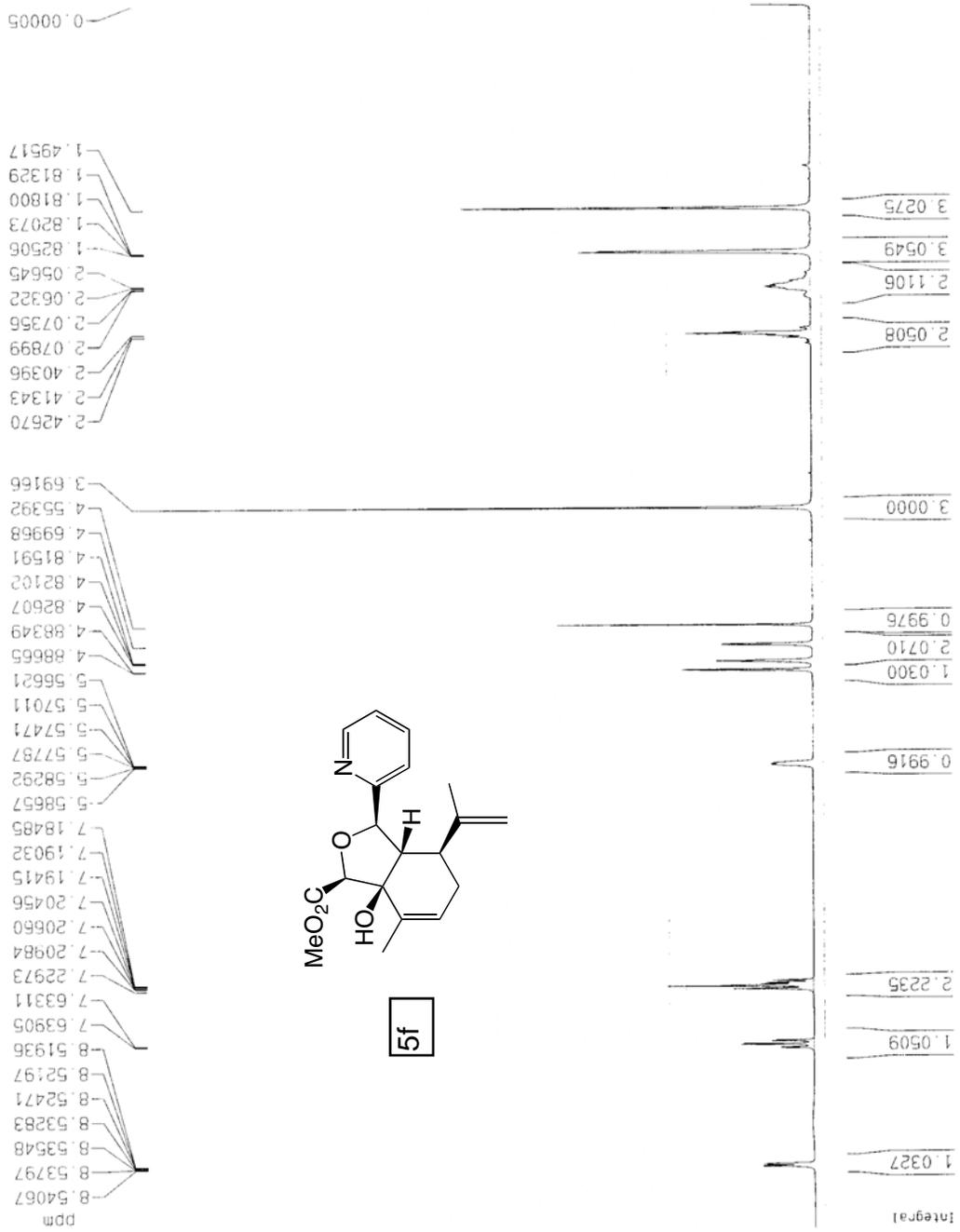
Current Data Parameters  
 NAME km-1290  
 EXPNO 7  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20080229  
 Time 17.51  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zg30  
 TD 37036  
 SOLVENT CDC13  
 NS 8  
 DS 2  
 SWH 6172.839 Hz  
 FIDRES 0.166671 Hz  
 AQ 2.999659 sec  
 RG 57  
 CW 81.000 usec  
 DE 6.00 usec  
 TE 296.2 K  
 D1 1.00000000 sec  
 MPREST 0.00000000 sec  
 MCWRR 0.01500000 sec

==== CHANNEL f1 =====  
 NUC1 1H  
 P1 12.20 usec  
 PL1 -1.00 dB  
 SF01 300.1318534 MHz

F2 - Processing parameters  
 SI 32768  
 SF 300.1300218 MHz  
 WDW EM  
 SSB 0  
 -B 0.30 Hz  
 GB 0  
 C 1.00

1D NMR plot parameters  
 CX 20.00 cm  
 CY 12.50 cm  
 F1P 9.000 ppm  
 F1 2701.17 Hz  
 F2P 0.000 ppm  
 F2 0.00 Hz  
 SPMCM 0.45000 ppm/cm



Current Data Parameters  
 NAME KB-1290  
 EXPNO 8  
 PROCNO 1

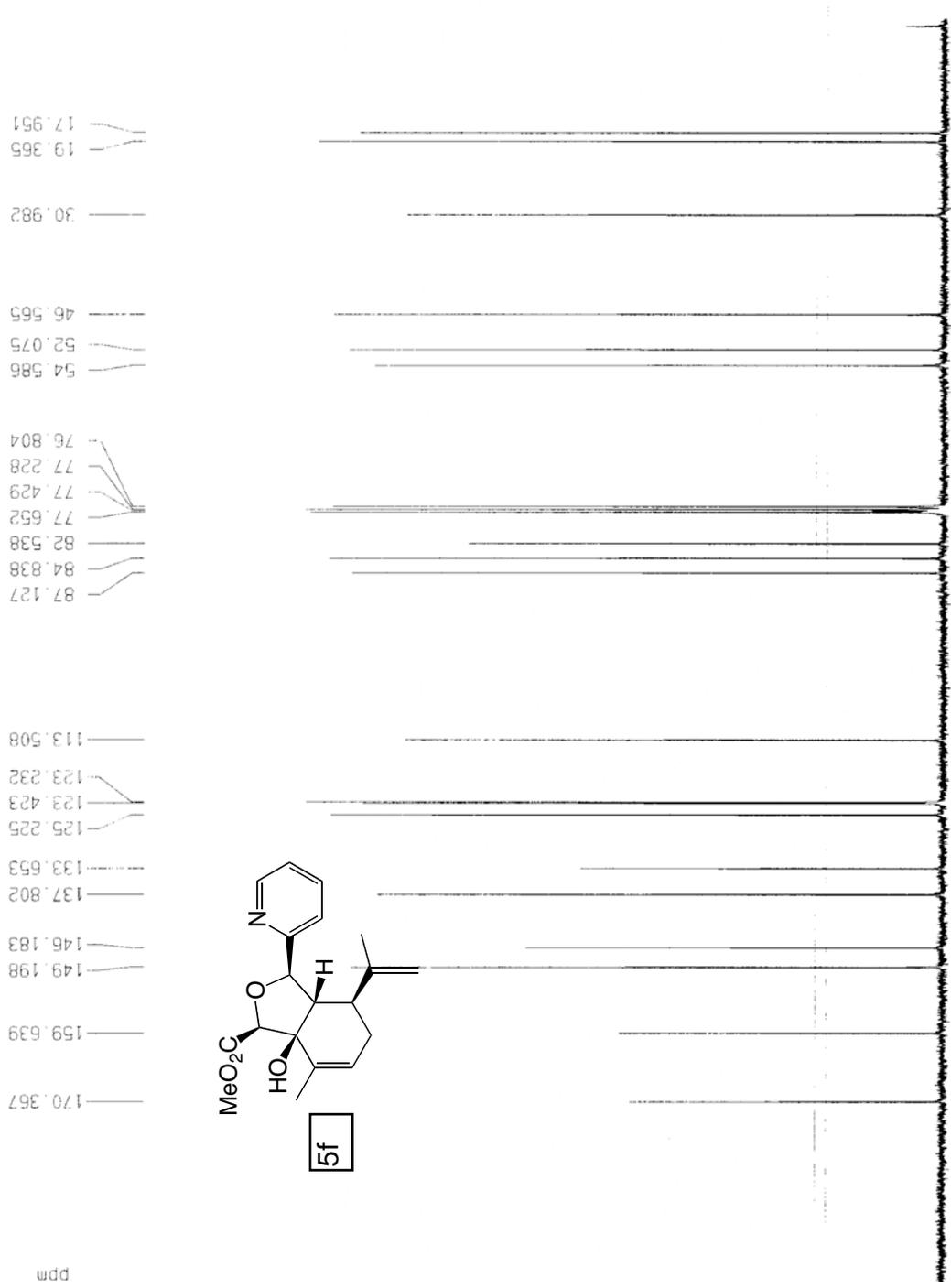
F2 - Acquisition Parameters  
 Date\_ 20080229  
 Time 18:45  
 INSTRUM spect  
 PROBO 5 mm PA6BBO BB-  
 PULPROG zgpg30  
 TD 35968  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 17985.611 Hz  
 FIDRES 0.300045 Hz  
 AQ 0.9999604 sec  
 RG 23170.5  
 KW 27.600 usec  
 DE 6.00 usec  
 TE 296.9 K  
 D1 2.0000000 sec  
 d11 0.0300000 sec  
 DELTA 1.8999999 sec  
 XRES 0.0000000 sec  
 XENRK 0.01500000 sec

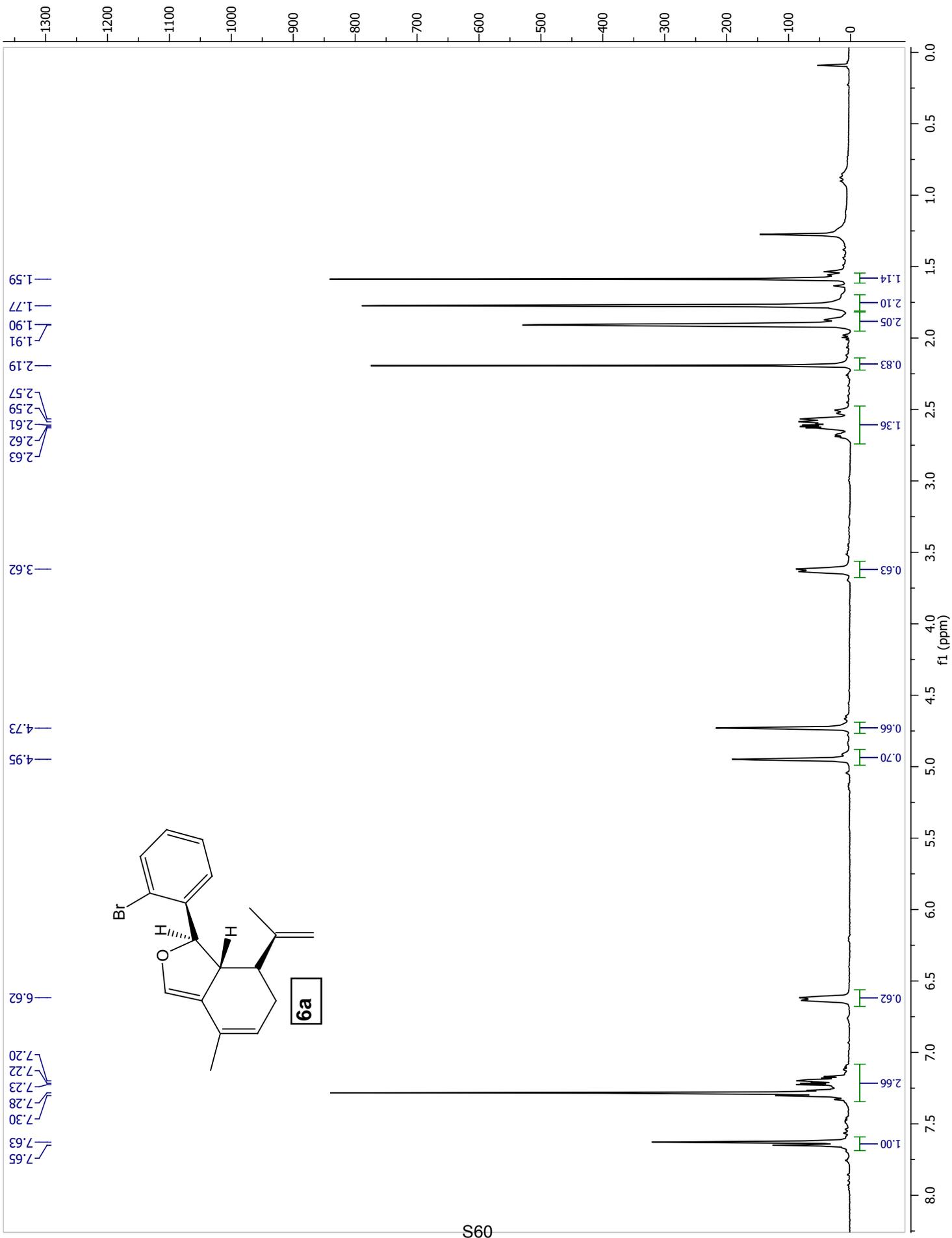
===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.38 usec  
 PL1 -2.80 dB  
 SF01 75.4760505 MHz

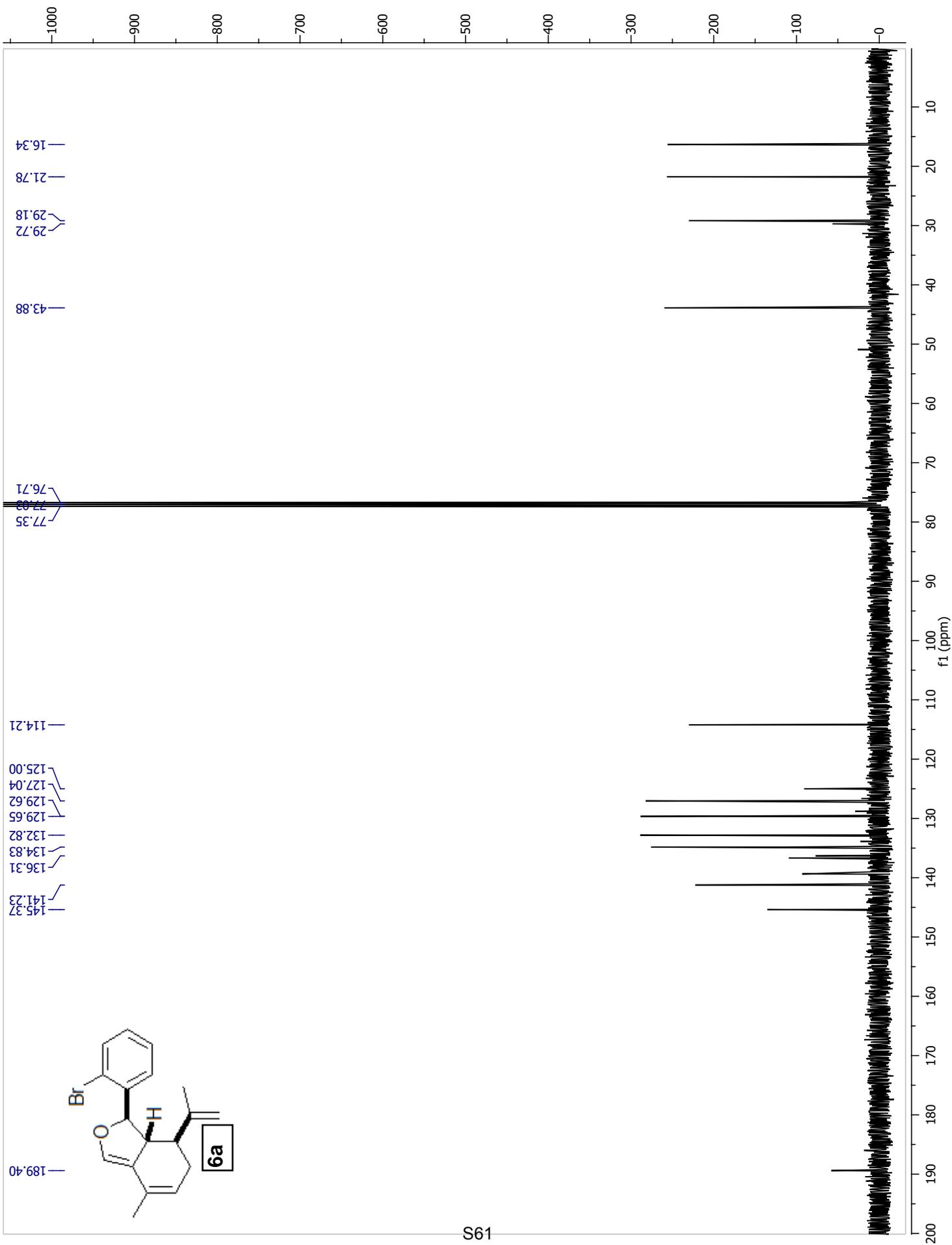
===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -1.00 dB  
 PL12 15.33 dB  
 PL13 16.00 dB  
 SF02 300.1312005 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4677395 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

ID NMR plot parameters  
 CX 20.00 cm  
 CY 10.00 cm  
 F1F 200.000 Dpp  
 F2F 255.476 MHz  
 F3F 300.131 MHz  
 GPC 1.40  
 ID 1







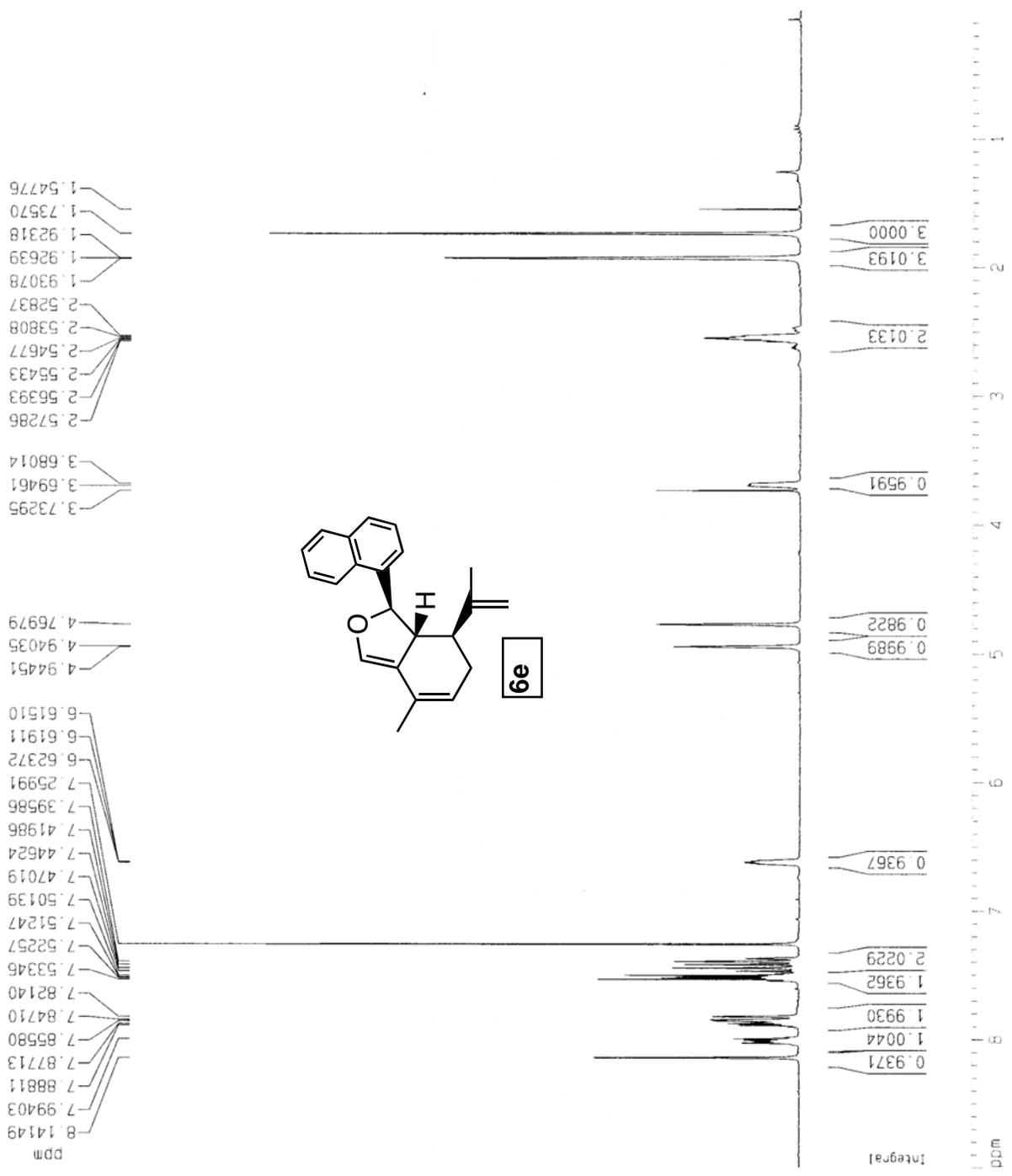
Current Data Parameters  
 NAME km-1257  
 EXPNO 9  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20071017  
 Time 9.07  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zg30  
 TD 17984  
 SOLVENT CDCl3  
 NS 32  
 DS 2  
 SWH 4496.403 Hz  
 FIDRES 0.250022 Hz  
 AQ 1.9998708 sec  
 RG 456.1  
 DM 111.200 usec  
 DE 6.00 usec  
 TE 296.2 K  
 D1 1.00000000 sec  
 MCREST 0.00000000 sec  
 MCWRRK 0.01500000 sec

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 12.20 usec  
 PL1 -0.60 dB  
 SFO1 300.1321009 MHz

F2 - Processing parameters  
 SI 32768  
 SF 300.1300068 MHz  
 WDW EM  
 SSB 0  
 -B 0.30 Hz  
 GB 0  
 PC 1.00

1D NMR plot parameters  
 CX 20.00 cm  
 CY 12.00 cm  
 F1P 9.000 ppm  
 F1 2701.17 Hz  
 F2P 0.000 ppm  
 F2 0.00 Hz  
 PPMCM 0.45000 ppm/cm  
 HZCM 135.05850 Hz/cm



Current Data Parameters  
 NAME km-1257  
 EXPNO 8  
 PROCNO 1

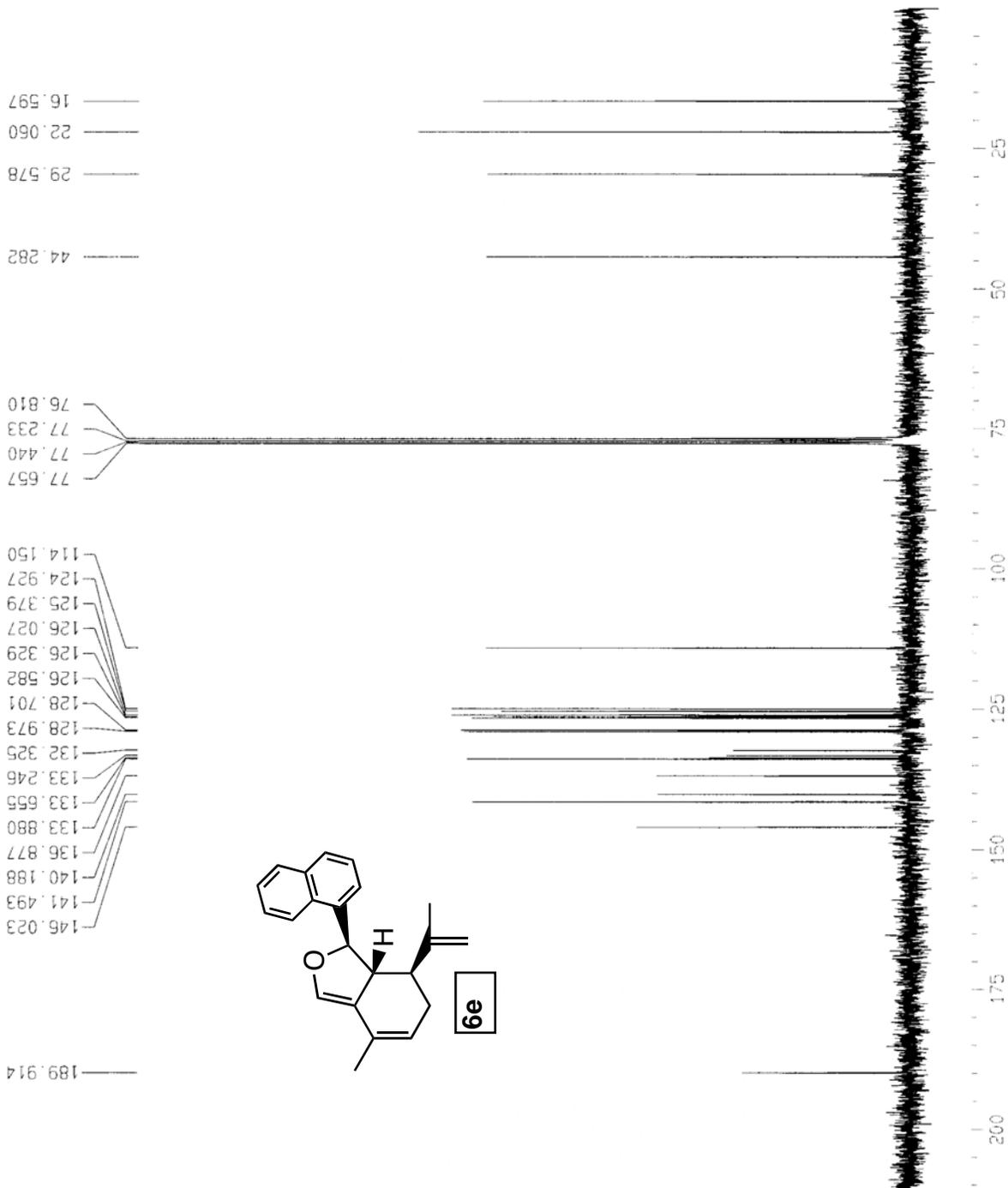
F2 - Acquisition Parameters  
 Date\_ 20071021  
 Time 11:27  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zgpg30  
 TD 35968  
 SOLVENT CDCl3  
 VS 1024  
 JS 4  
 SWH 18115.941 Hz  
 FIDRES 0.503668 Hz  
 AQ 0.9927668 sec  
 RG 26008  
 DM 27.600 usec  
 DE 6.00 usec  
 TE 297.0 K  
 D1 2.0000000 sec  
 D11 0.0300000 sec  
 DELTA 1.8999998 sec  
 MCREST 0.0000000 sec  
 MCMRK 0.0150000 sec

\*\*\*\*\* CHANNEL f1 \*\*\*\*\*  
 NUC1 13C  
 P1 8.38 usec  
 PL1 -2.80 dB  
 SFO1 75.4752953 MHz

\*\*\*\*\* CHANNEL f2 \*\*\*\*\*  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -0.60 dB  
 PL12 15.73 dB  
 PL13 16.00 dB  
 SFO2 300.1312005 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4677337 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 SC 1.40

10 NMR plot parameters  
 CX 20.00 cm  
 CY 8.00 cm  
 F1P 220.221 ppm  
 F1 16619.54 Hz  
 F2P -0.000 ppm  
 F2 -0.00 Hz  
 DPMSM 11.01103 ppm/cm  
 AZCM 830.97717 Hz/cm



```

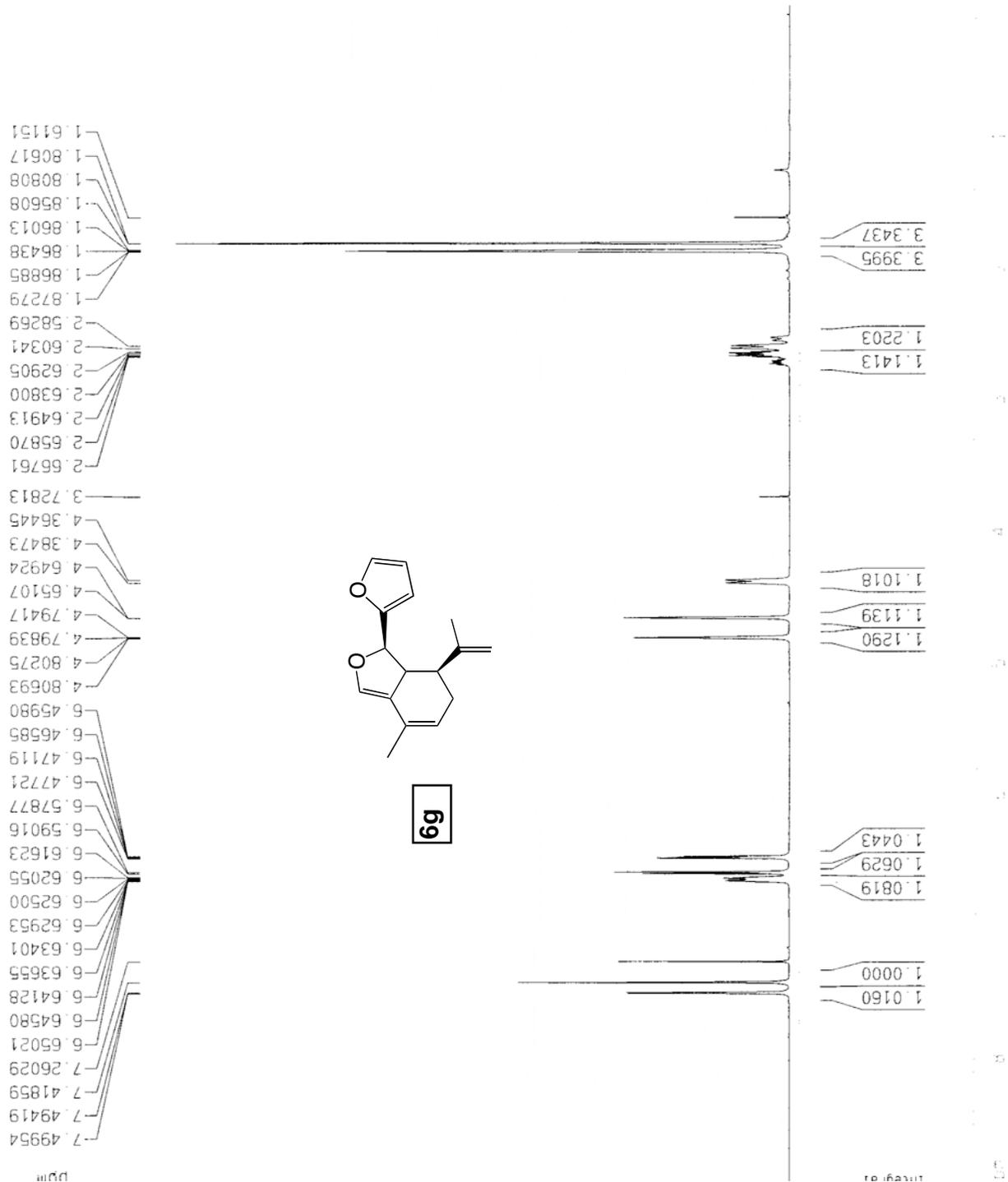
Current Data Parameters
NAME      km-1360
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20080207
Time      16:57
INSTRUM   spect
PROBHD    5 mm PABBO BBO-
PULPROG   zg30
TD         23980
SOLVENT   CDCl3
NS         16
DS         2
SWH        5995.204 Hz
FIDRES     0.250008 Hz
AQ         1.9999820 sec
RG         256
CW         83.400 usec
DE         5.00 usec
TE         296.4 K
D1         1.00000000 sec
wCREST     0.00000000 sec
wCMARK     0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         12.20 usec
PL1        -1.00 dB
SFO1       300.1318608 MHz

F2 - Processing parameters
SI         32768
SF         300.1300066 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
CY         10.00 cm
F1P        9.000 ppm
F1         2701.17 Hz
F2P        0.000 ppm
F2         0.00 Hz
SFOCM      0.45000 ppm/cm
AQCM       1.00000000 sec
    
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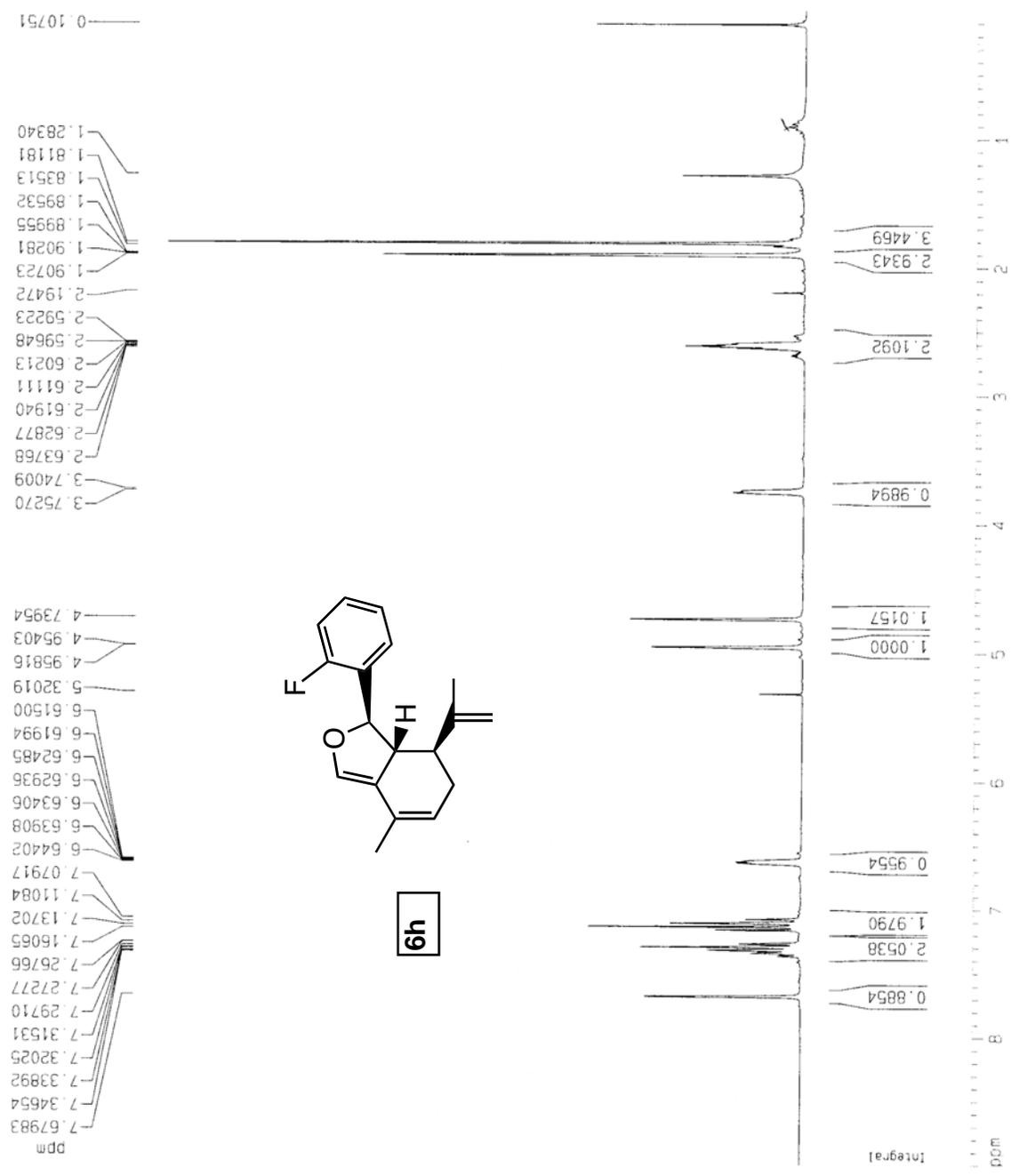
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 NAME km-1245  
 EXPNO 2  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20071006  
 Time 10.17  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zg30  
 TD 17984  
 SOLVENT COC13  
 NS 8  
 DS 2  
 SWH 4496.403 Hz  
 FIDRES 0.250022 Hz  
 AQ 1.9998708 sec  
 RG 71.8  
 DW 111.200 usec  
 DE 6.00 usec  
 TE 296.3 K  
 D1 1.0000000 sec  
 MCREST 0.0000000 sec  
 MCWRR 0.01500000 sec

\*\*\*\*\* CHANNEL f1 \*\*\*\*\*  
 NUC1 1H  
 P1 12.20 usec  
 PL1 -0.60 dB  
 SF01 300.1321009 MHz

F2 - Processing parameters  
 SI 32768  
 SF 300.1299555 MHz  
 MDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

1D NMR plot parameters  
 CX 20.00 cm  
 CY 11.00 cm  
 F1 9.000 ppm  
 F2 2701.17 Hz  
 F3 0.000 ppm  
 F4 0.00 Hz  
 SFO1 0.45000 ppm/cm  
 HZCM 135.05850 Hz/cm



```

Current Data Parameters
NAME km-1245
EXPNO 5
PROCNO 1

F2 - Acquisition Parameters
Date_ 20071005
Time 10.19
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPRG6 zgpg30
TD 32372
SOLVENT CDCl3
NS 240
DS 4
SMH 17985.611 Hz
FIDRES 0.555592 Hz
AQ 0.8959916 sec
RG 23170.5
JM 27.800 usec
JE 6.00 usec
TE 296.5 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRRK 0.01500000 sec

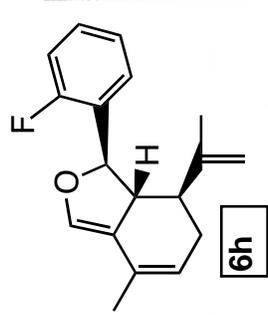
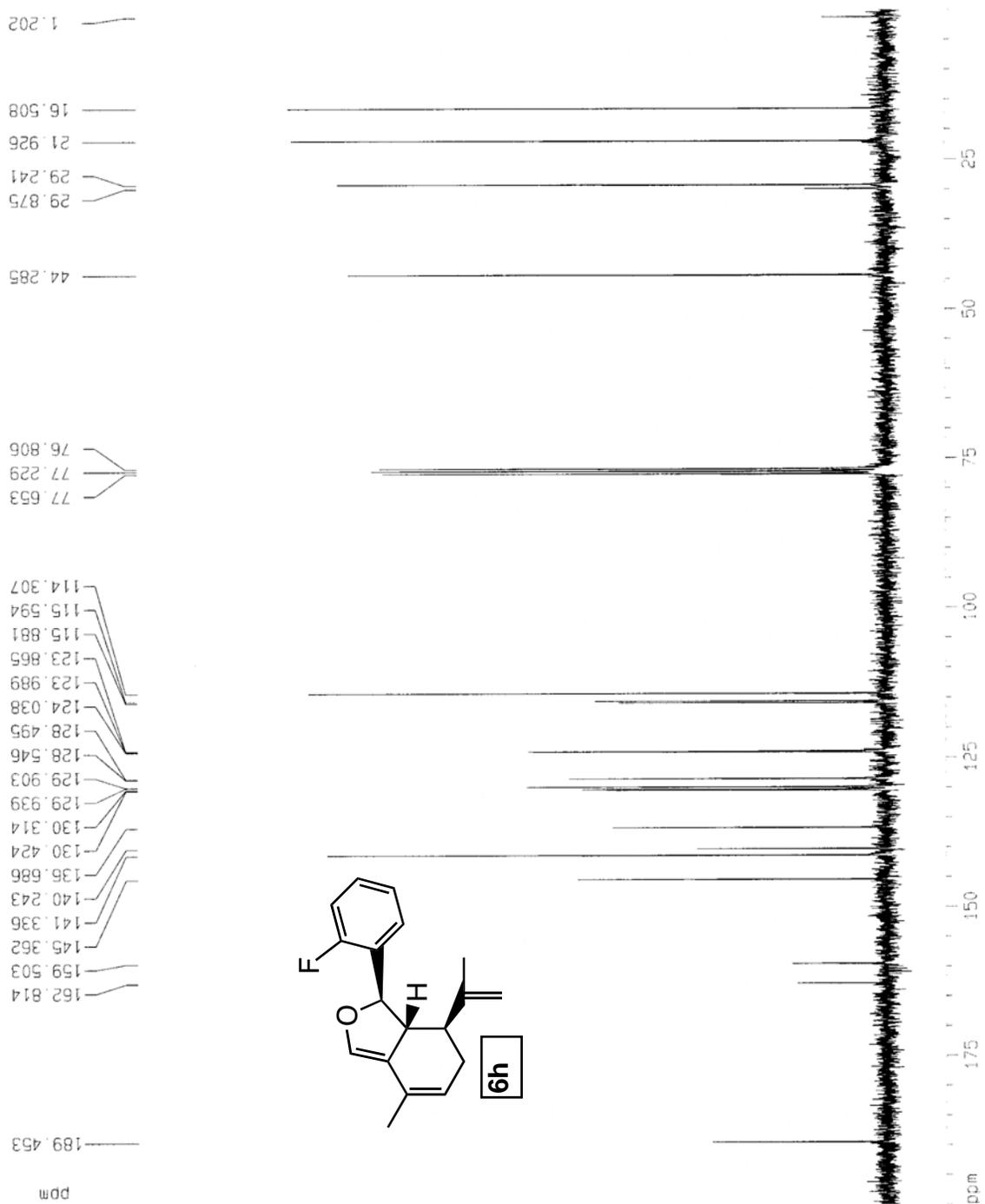
***** CHANNEL f1 *****
NUC1 13C
P1 8.38 usec
PL1 -2.80 dB
SF01 75.4752953 MHz

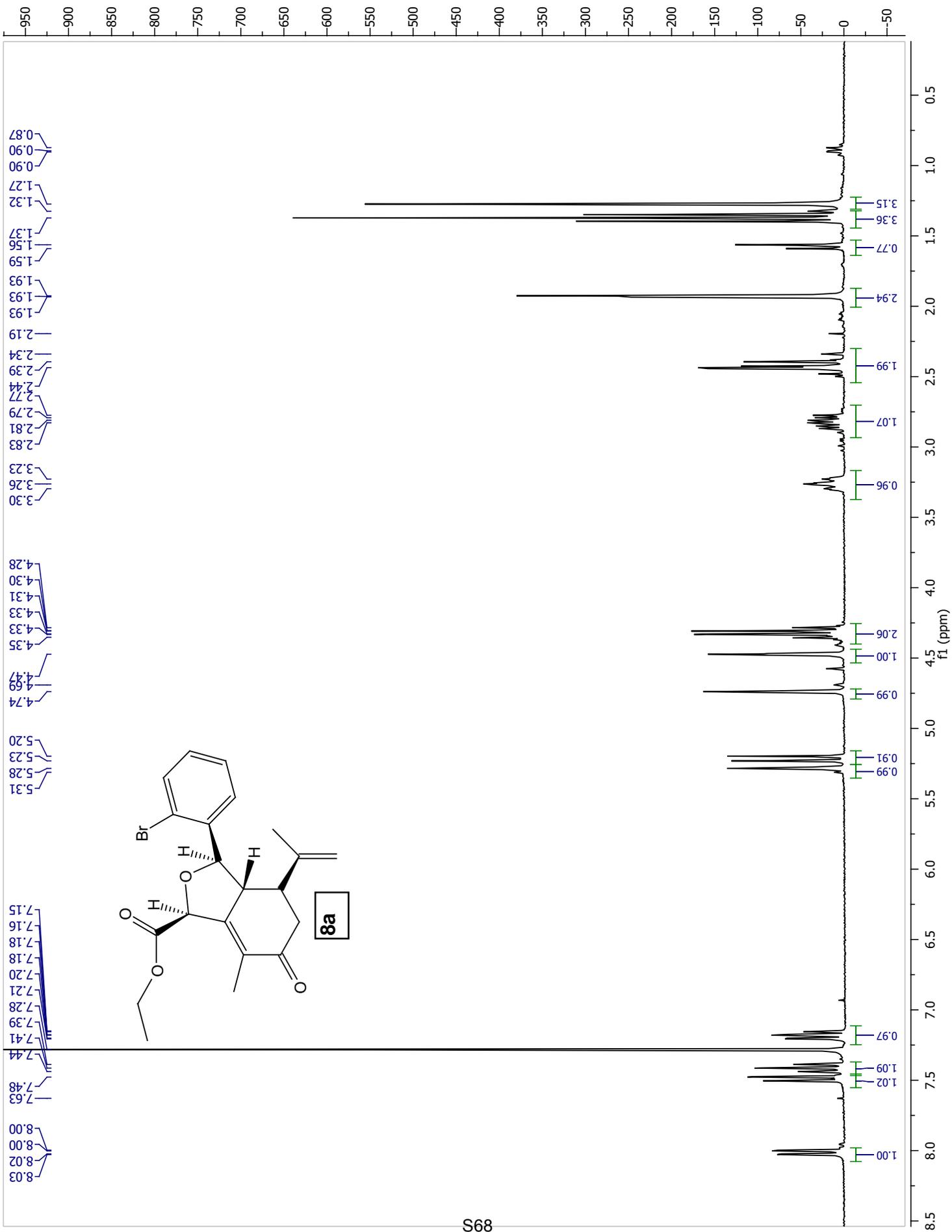
***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -0.60 dB
PL12 15.73 dB
PL13 16.00 dB
SF02 300.1312005 MHz

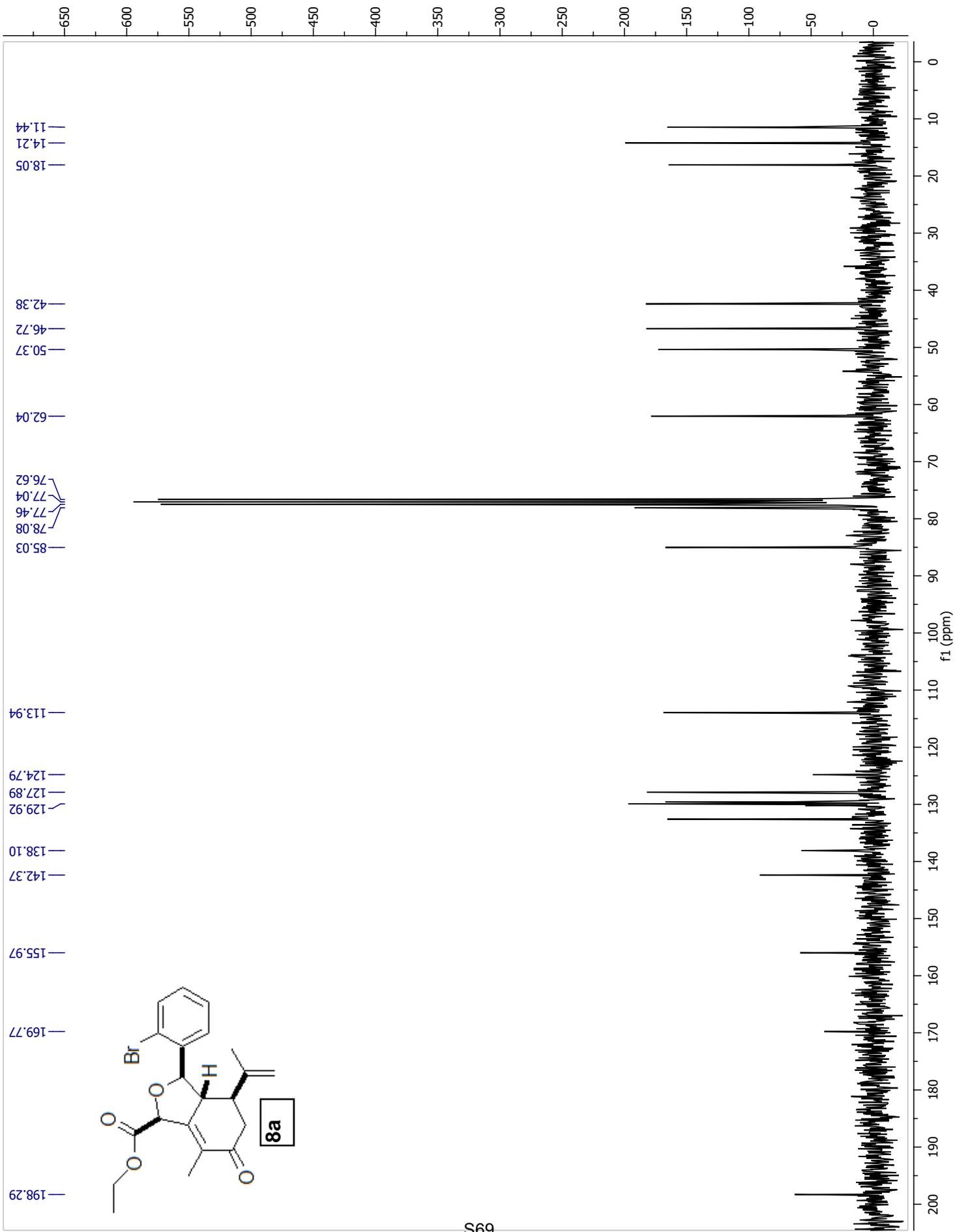
F2 - Processing parameters
SI 32768
SF 75.4677363 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
CY 10.00 cm
F1P 200.000 ppm
F1 15093.55 Hz
F2P 0.000 ppm
F2 0.00 Hz
PPMCM 10.00000 ppm/cm
HZCM 754.67737 Hz/cm

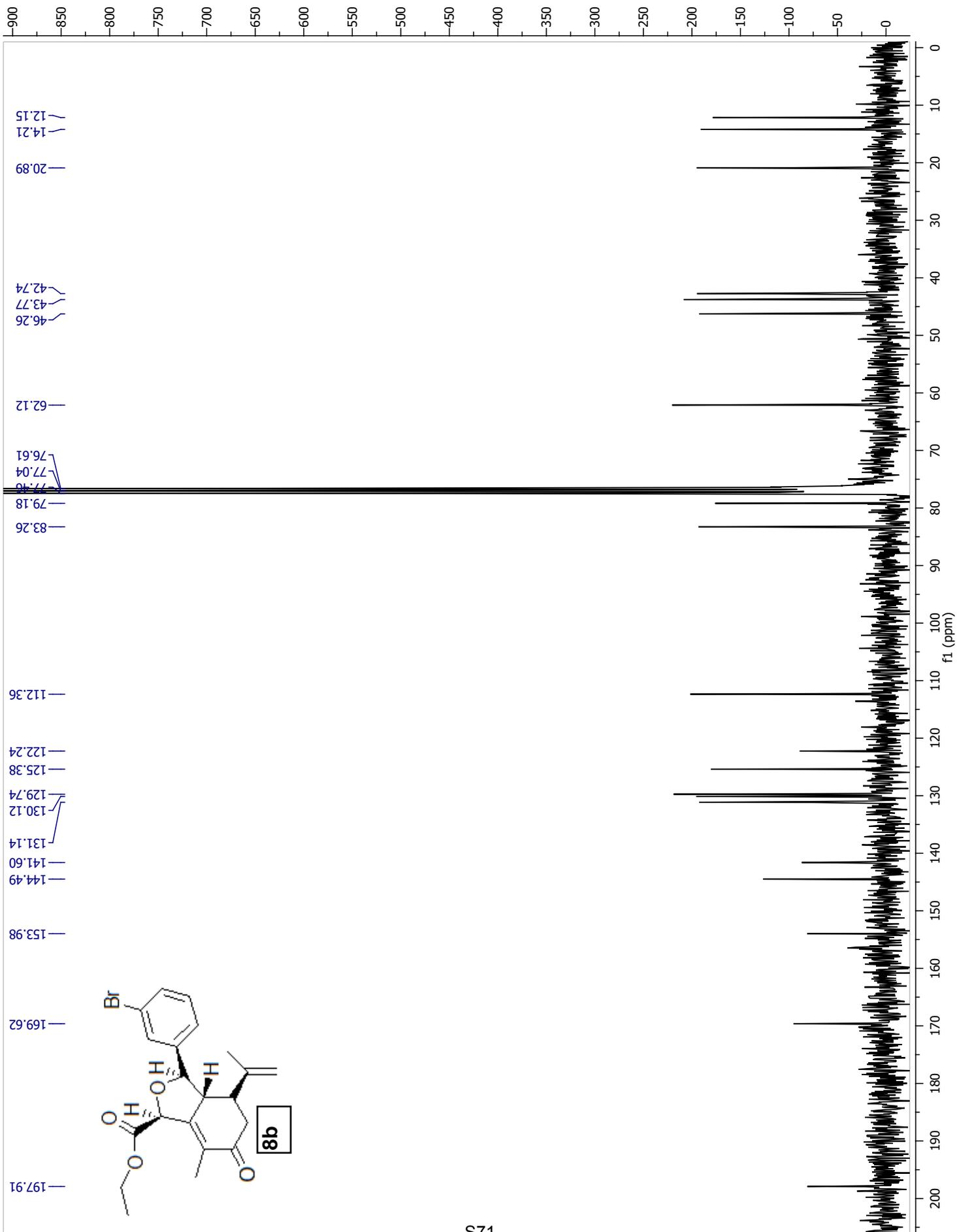
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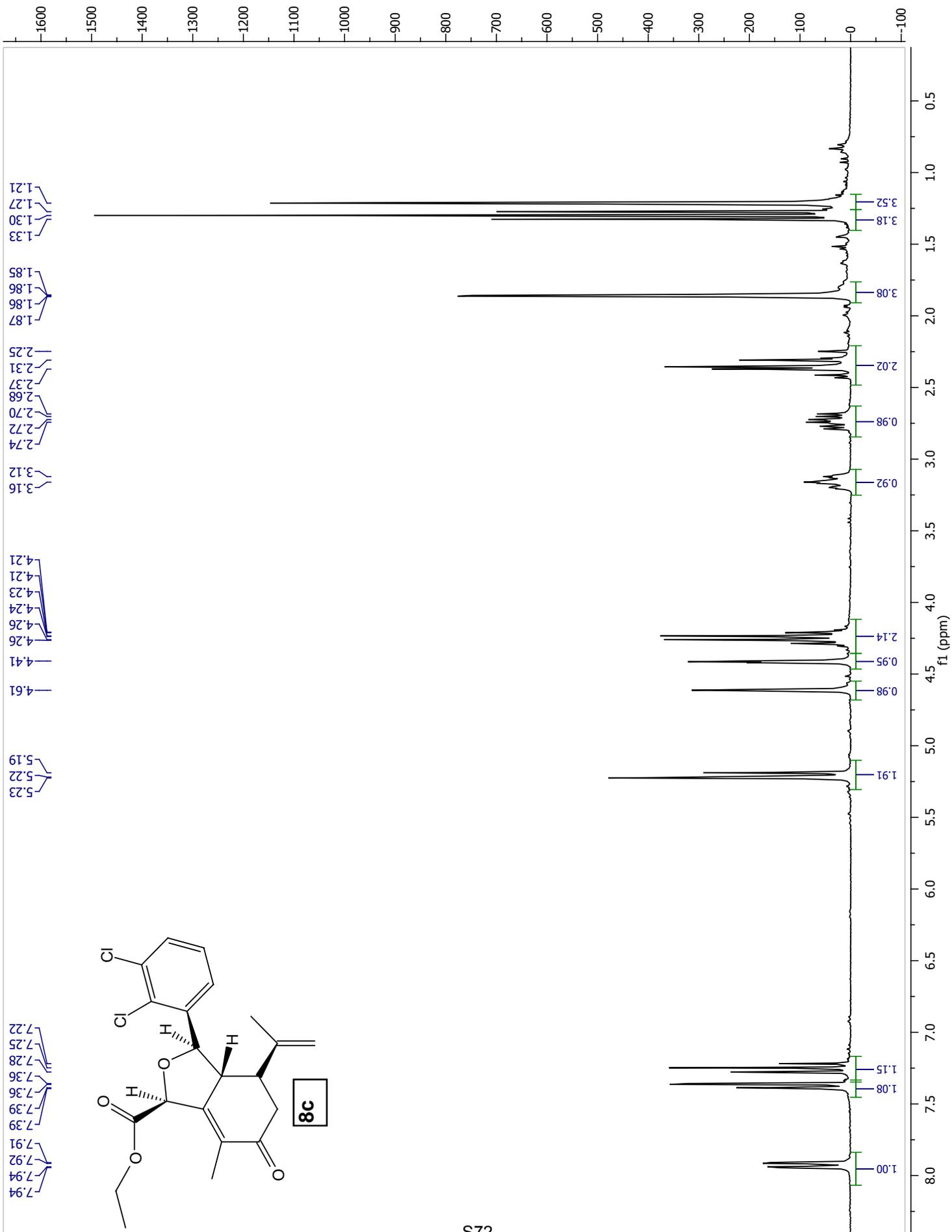


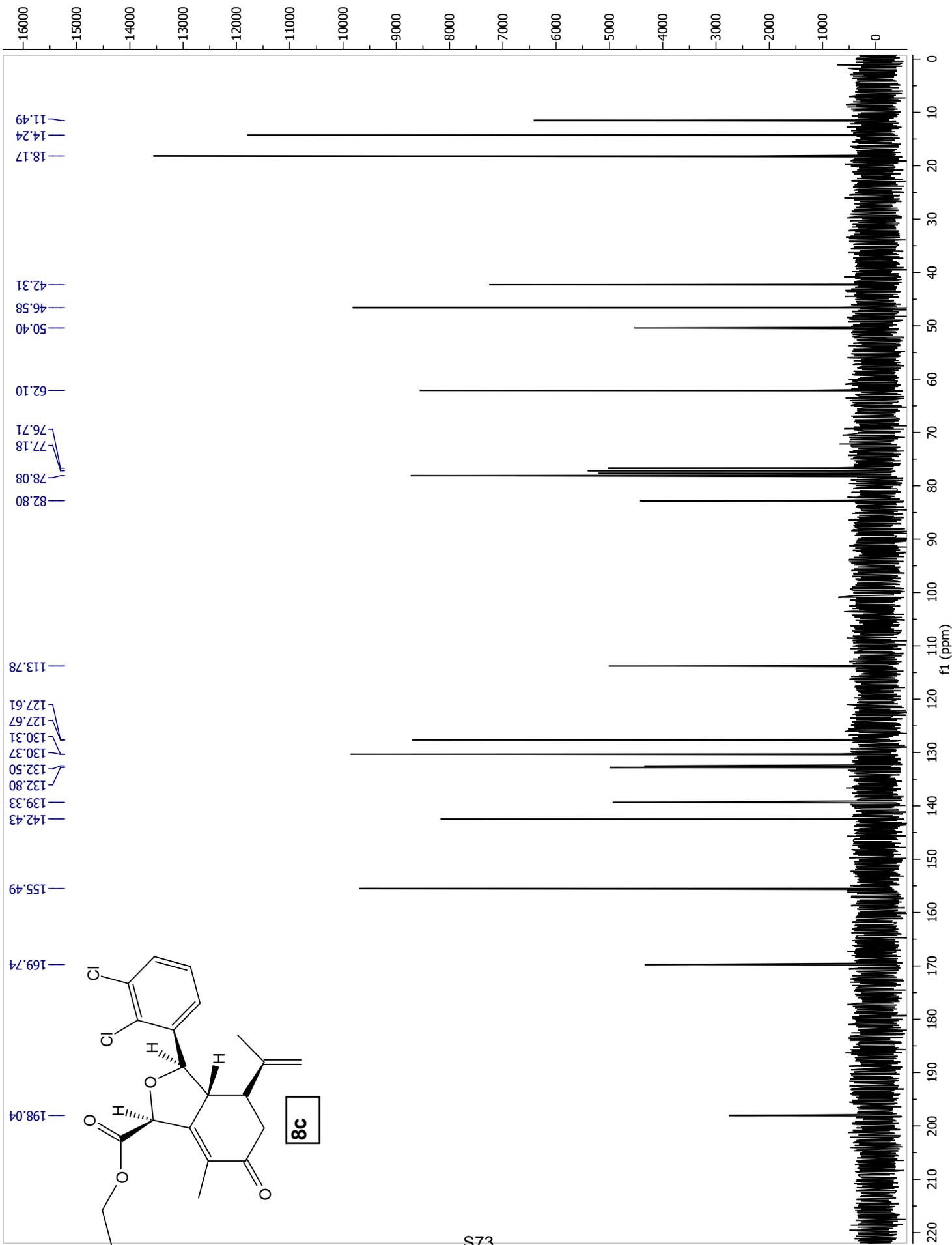


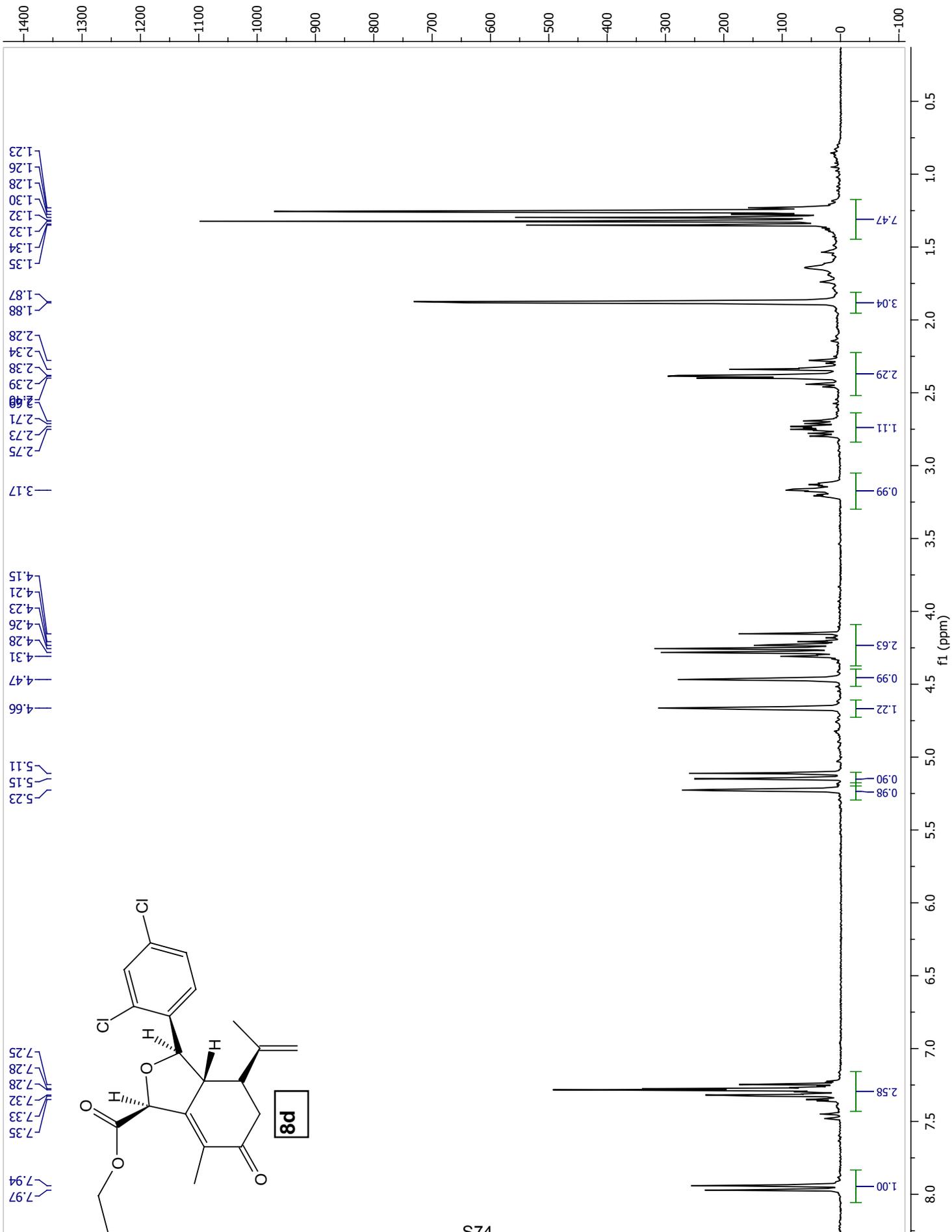


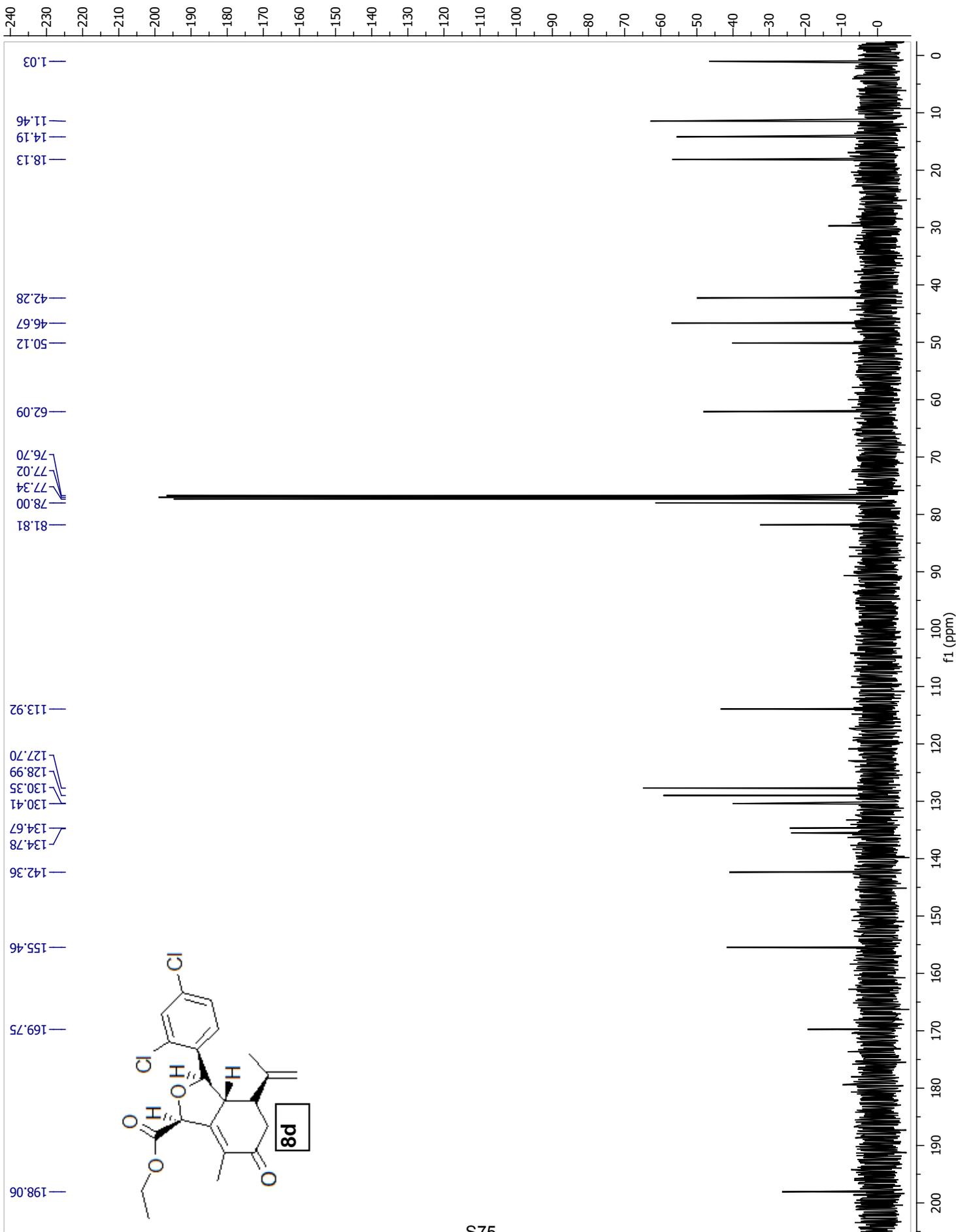




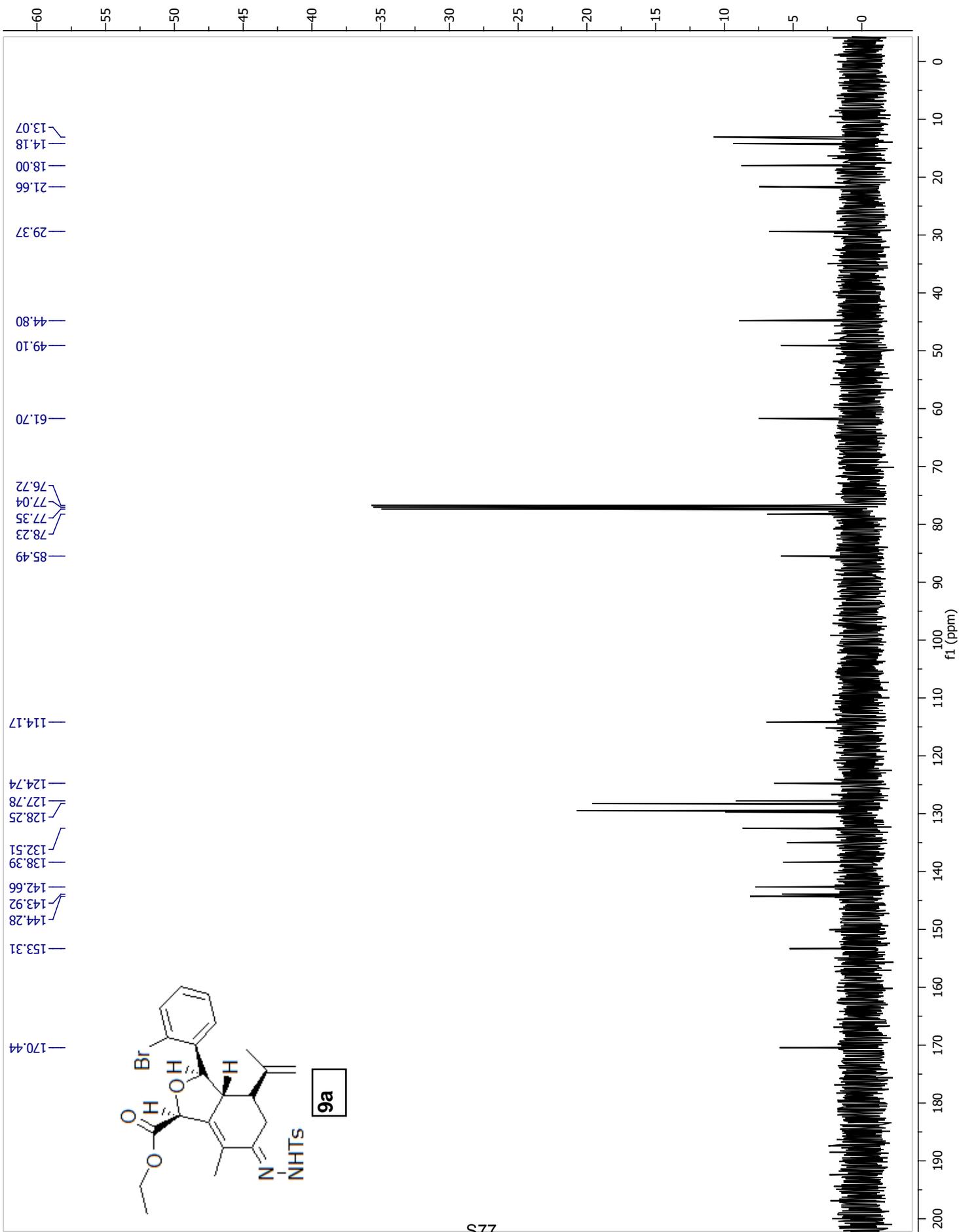




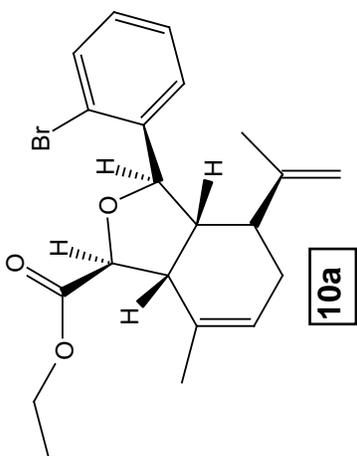




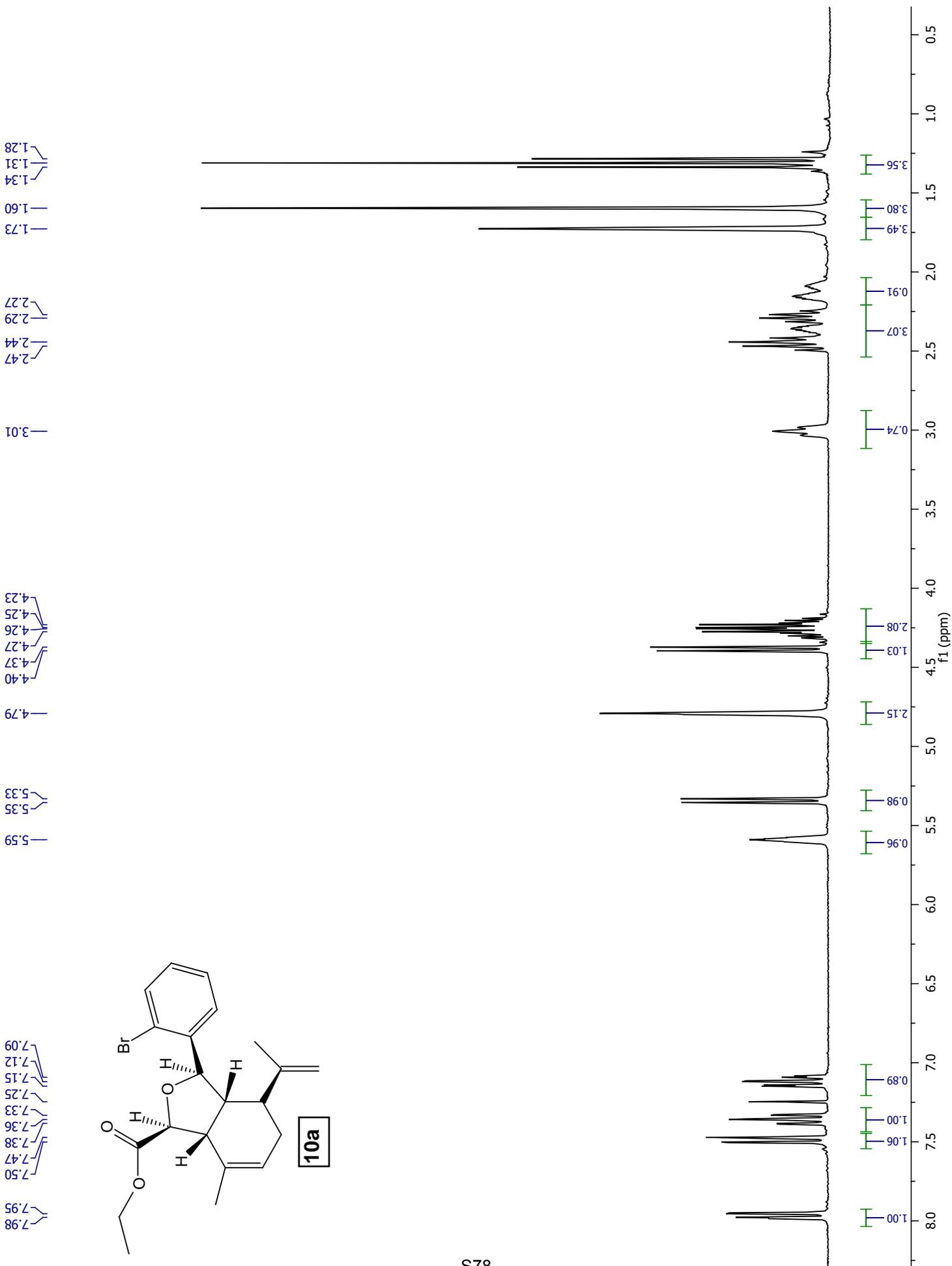


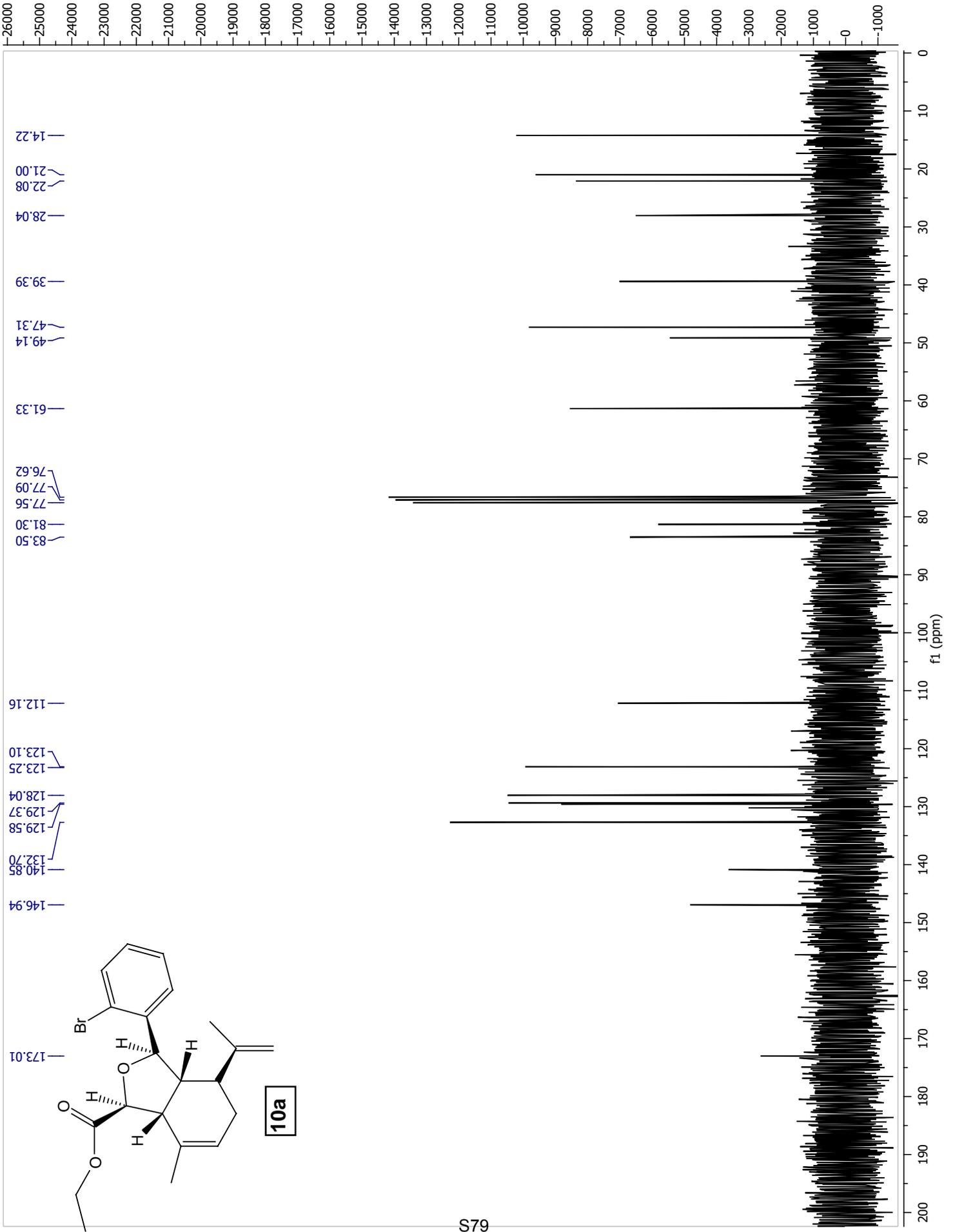


7.98  
7.95  
7.50  
7.47  
7.38  
7.36  
7.33  
7.25  
7.15  
7.12  
7.09  
5.59  
5.35  
5.33  
4.79  
4.40  
4.37  
4.27  
4.26  
4.25  
4.23  
3.01  
2.47  
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2.29  
2.27  
1.73  
1.60  
1.34  
1.31  
1.28



S78





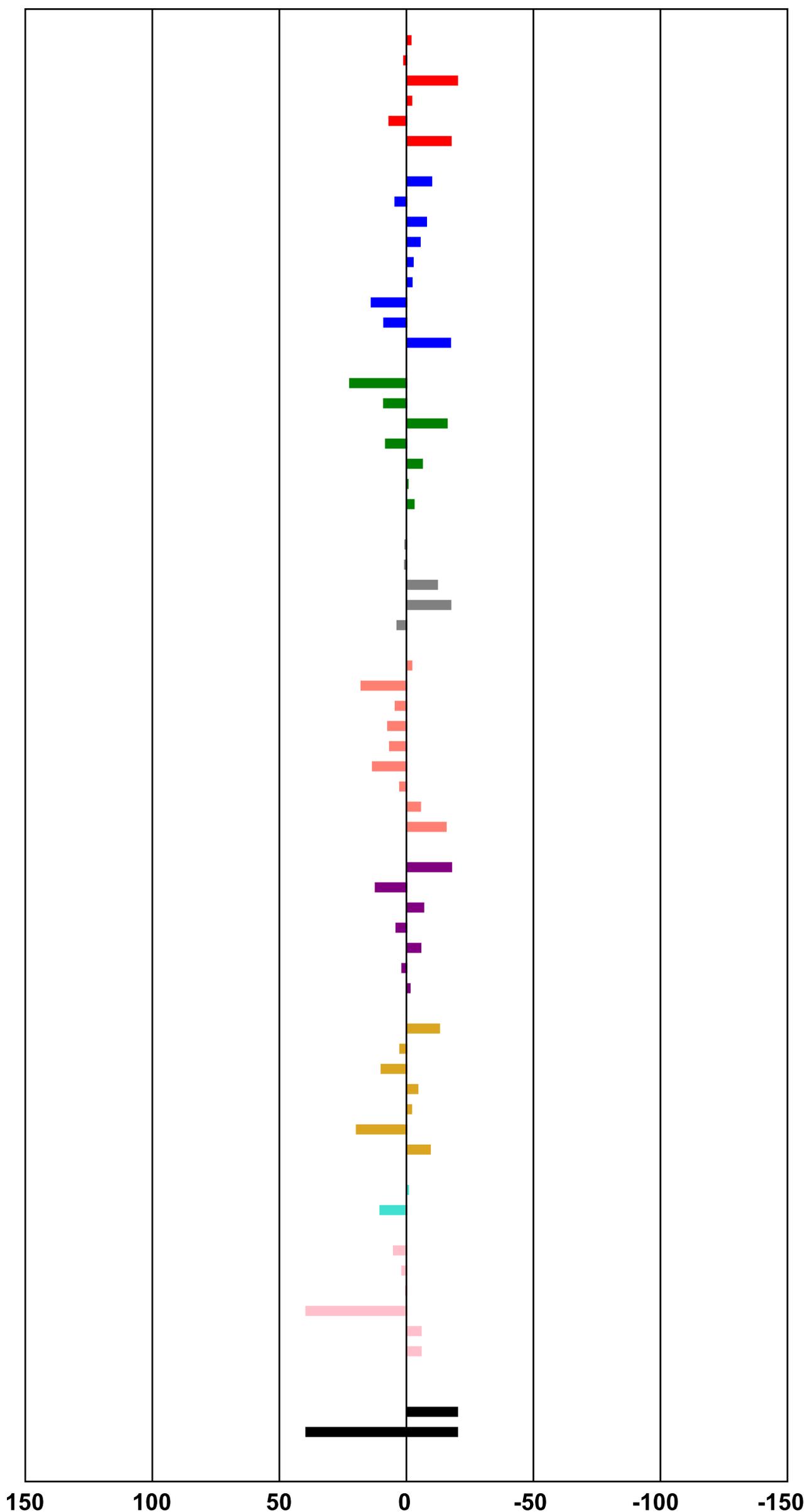
**One Dose Mean Graph**

**Panel/Cell Line**

**Growth Percent**

**Mean Growth Percent - Growth Percent**

Leukemia	
CCRF-CEM	98.31
HL-60(TB)	100.94
K-562	79.96
MOLT-4	98.02
RPMI-8226	106.77
SR	82.50
Non-Small Cell Lung Cancer	
A549/ATCC	90.16
EKVX	104.39
HOP-62	92.21
HOP-92	94.71
NCI-H226	97.47
NCI-H23	97.89
NCI-H322M	113.73
NCI-H460	108.80
NCI-H522	82.73
Colon Cancer	
COLO 205	122.22
HCC-2998	108.86
HCT-116	84.08
HCT-15	108.10
HT29	93.83
KM12	99.49
SW-620	97.10
CNS Cancer	
SF-268	100.46
SF-539	100.60
SNB-19	87.90
SNB-75	82.61
U251	103.59
Melanoma	
LOX IMVI	98.00
MALME-3M	117.74
M14	104.29
MDA-MB-435	107.24
SK-MEL-2	106.54
SK-MEL-28	113.23
SK-MEL-5	102.55
UACC-257	94.54
UACC-62	84.48
Ovarian Cancer	
IGROV1	82.34
OVCAR-3	112.11
OVCAR-4	93.31
OVCAR-5	103.98
OVCAR-8	94.44
NCI/ADR-RES	101.67
SK-OV-3	98.68
Renal Cancer	
786-0	87.09
A498	102.47
ACHN	109.85
RXF 393	95.61
SN12C	98.08
TK-10	119.56
UO-31	90.72
Prostate Cancer	
PC-3	99.33
DU-145	110.29
Breast Cancer	
MCF7	104.98
MDA-MB-231/ATCC	101.68
HS 578T	100.26
BT-549	139.44
T-47D	94.32
MDA-MB-468	94.32
Mean	100.01
Delta	20.05
Range	59.48



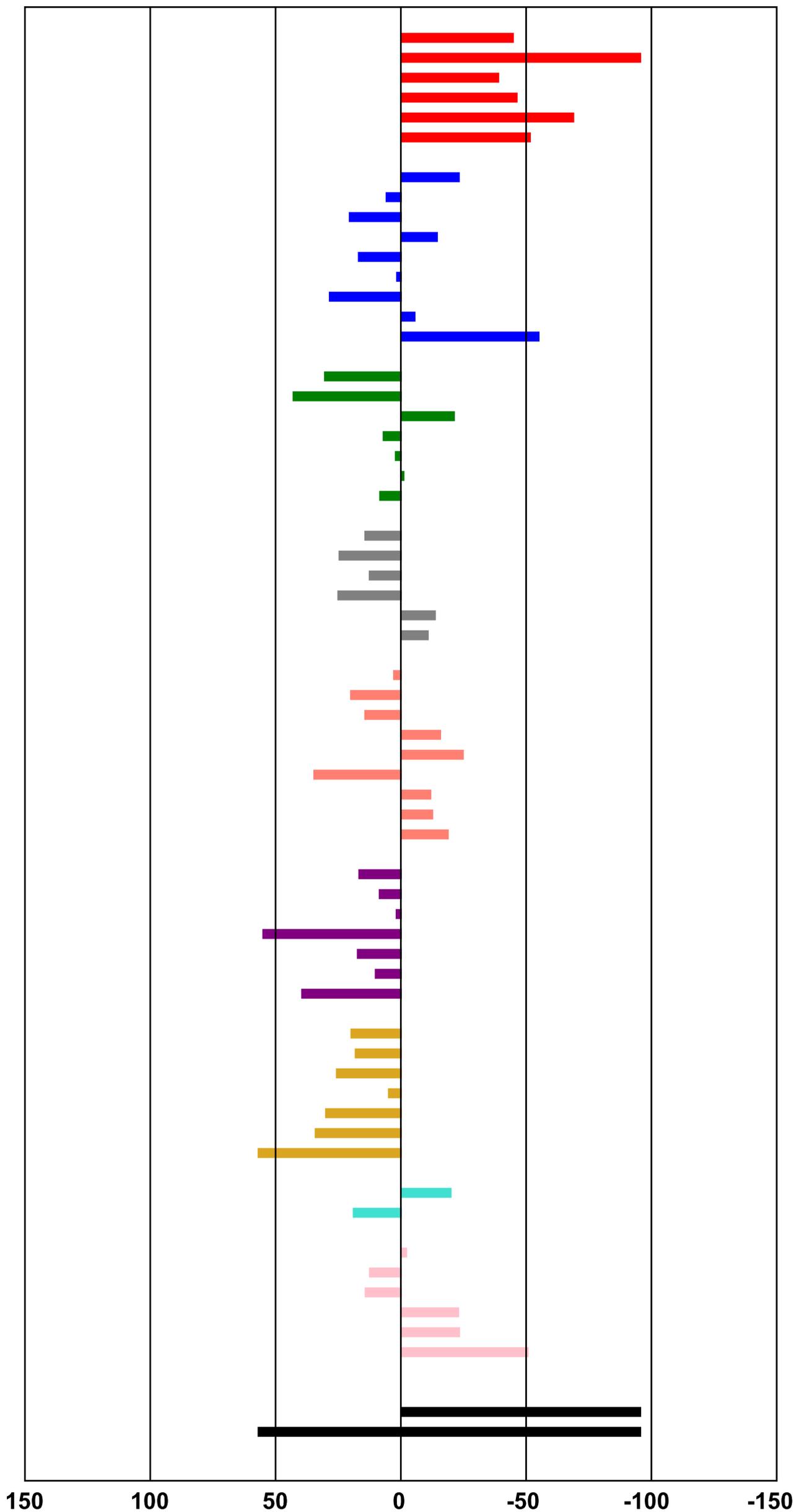
**One Dose Mean Graph**

**Panel/Cell Line**

**Growth Percent**

**Mean Growth Percent - Growth Percent**

Leukemia		
CCRF-CEM	19.48	
HL-60(TB)	-31.34	
K-562	25.32	
MOLT-4	17.98	
RPMI-8226	-4.60	
SR	12.69	
Non-Small Cell Lung Cancer		
A549/ATCC	41.04	
EKVX	69.98	
HOP-62	84.64	
HOP-92	49.80	
NCI-H226	81.04	
NCI-H23	65.80	
NCI-H322M	92.62	
NCI-H460	58.73	
NCI-H522	9.23	
Colon Cancer		
COLO 205	94.54	
HCC-2998	107.11	
HCT-116	42.99	
HCT-15	71.15	
HT29	66.30	
KM12	63.13	
SW-620	72.54	
CNS Cancer		
SF-268	78.47	
SF-295	88.75	
SF-539	76.69	
SNB-19	89.23	
SNB-75	50.59	
U251	53.44	
Melanoma		
LOX IMVI	66.98	
MALME-3M	84.13	
M14	78.49	
MDA-MB-435	48.50	
SK-MEL-2	39.43	
SK-MEL-28	98.82	
SK-MEL-5	52.40	
UACC-257	51.65	
UACC-62	45.44	
Ovarian Cancer		
IGROV1	80.84	
OVCAR-3	72.74	
OVCAR-4	65.97	
OVCAR-5	119.16	
OVCAR-8	81.47	
NCI/ADR-RES	74.31	
SK-OV-3	103.67	
Renal Cancer		
786-0	84.02	
A498	82.29	
ACHN	89.80	
RXF 393	69.05	
SN12C	94.13	
TK-10	98.26	
UO-31	121.05	
Prostate Cancer		
PC-3	44.34	
DU-145	83.07	
Breast Cancer		
MCF7	62.06	
MDA-MB-231/ATCC	76.61	
HS 578T	78.32	
BT-549	41.32	
T-47D	40.94	
MDA-MB-468	13.57	
Mean	64.24	
Delta	95.58	
Range	152.39	



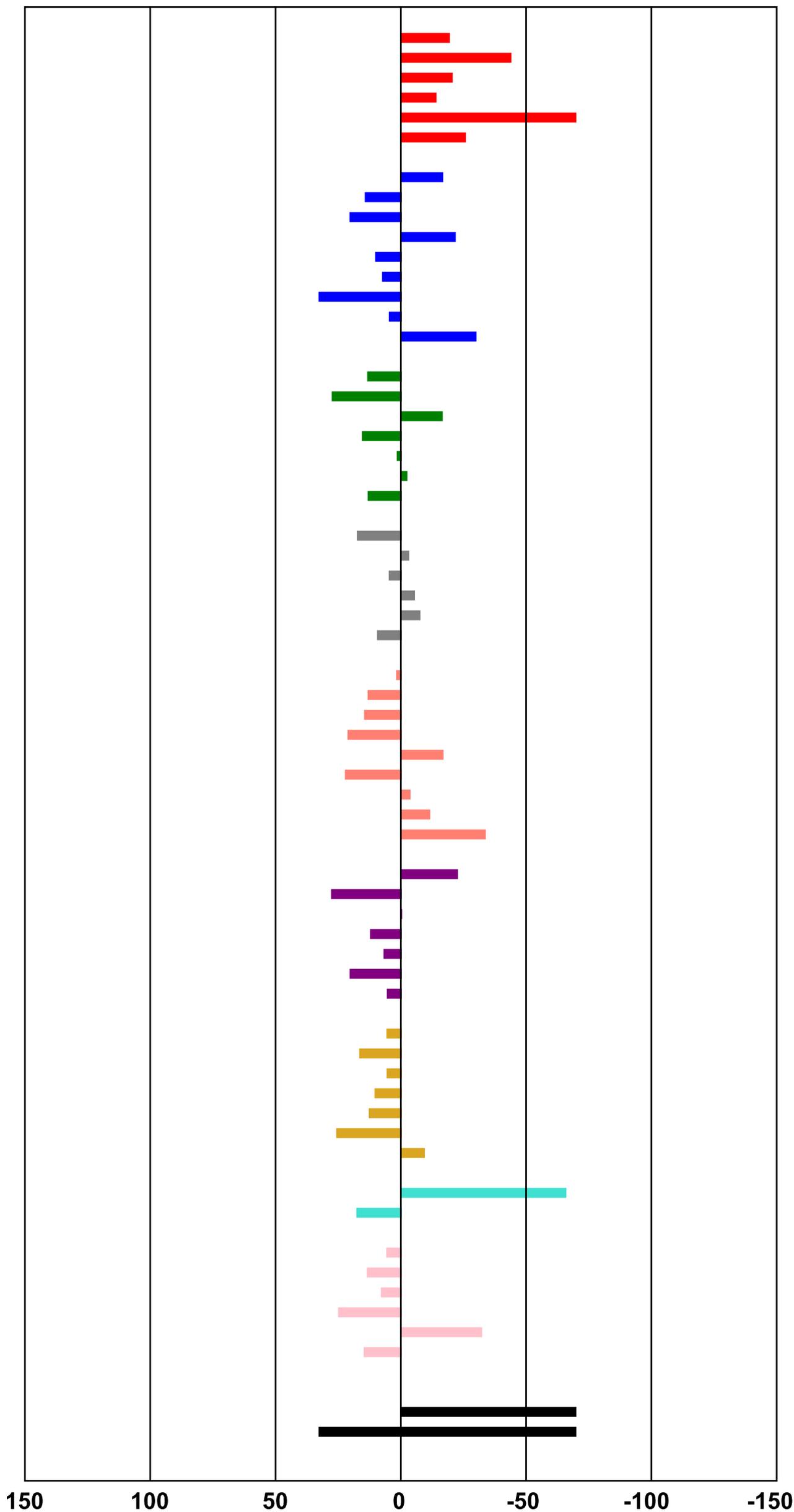
**One Dose Mean Graph**

**Panel/Cell Line**

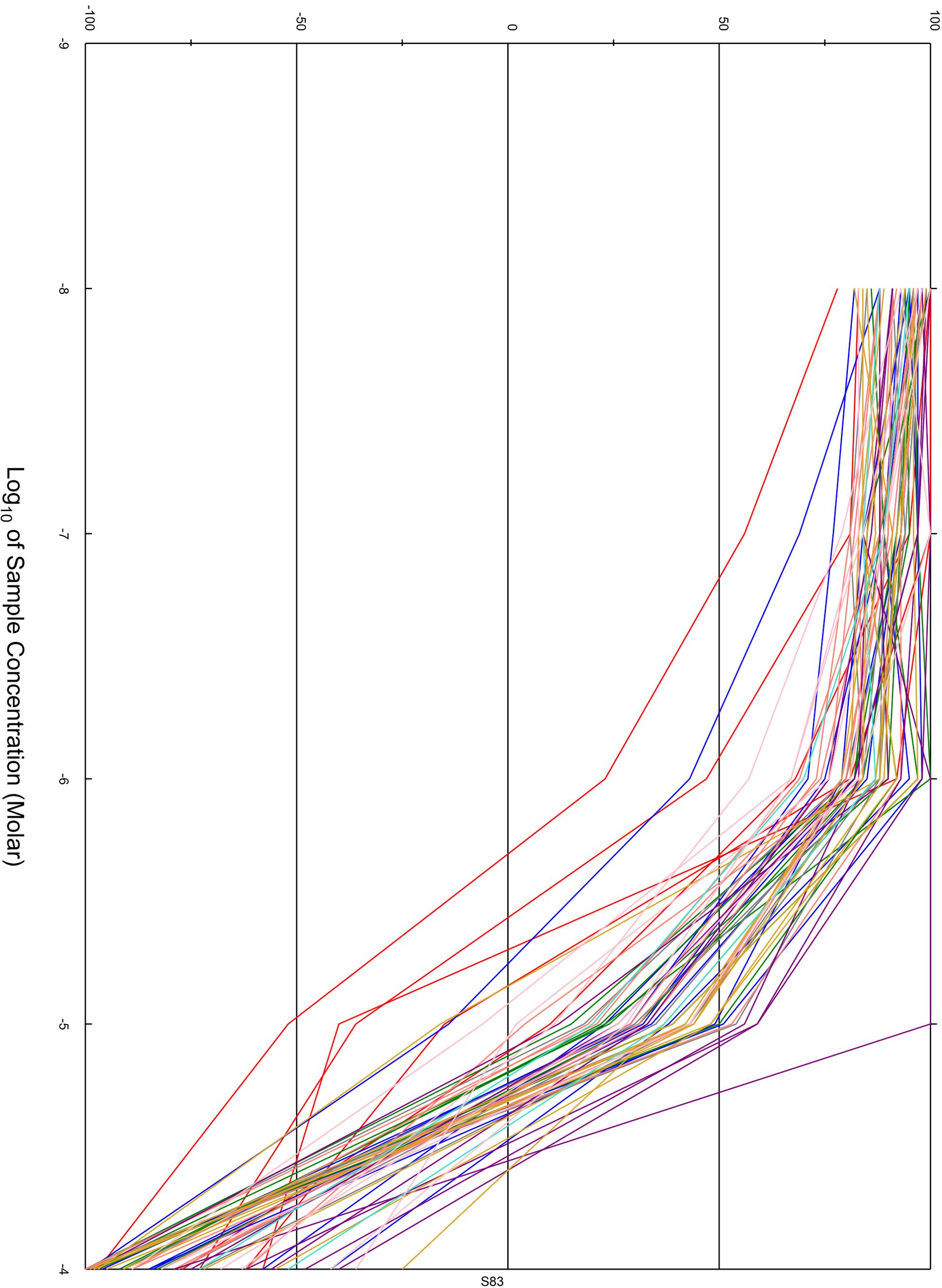
**Growth Percent**

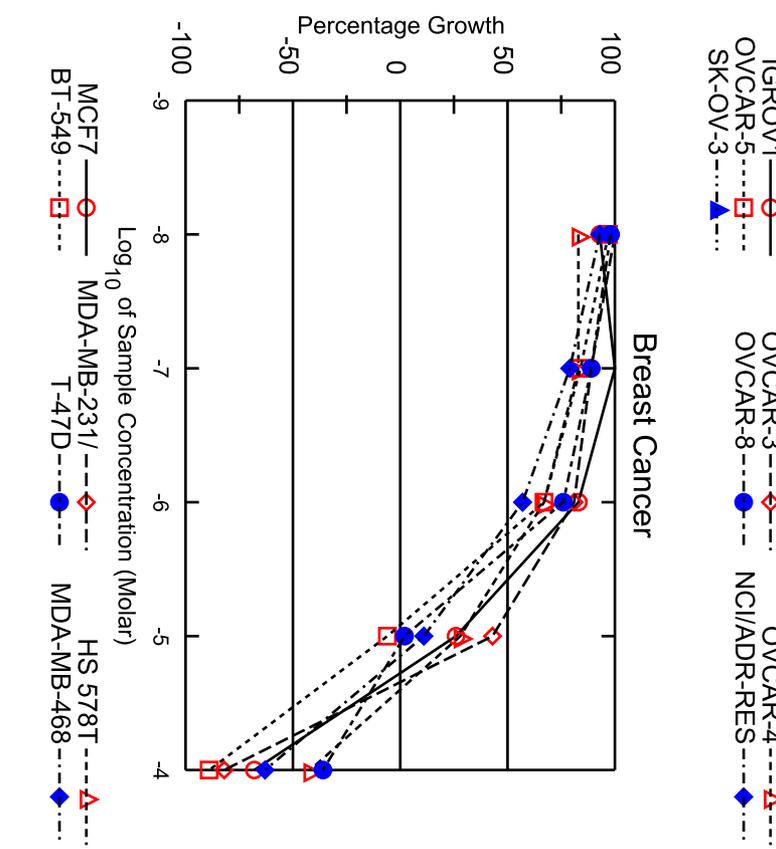
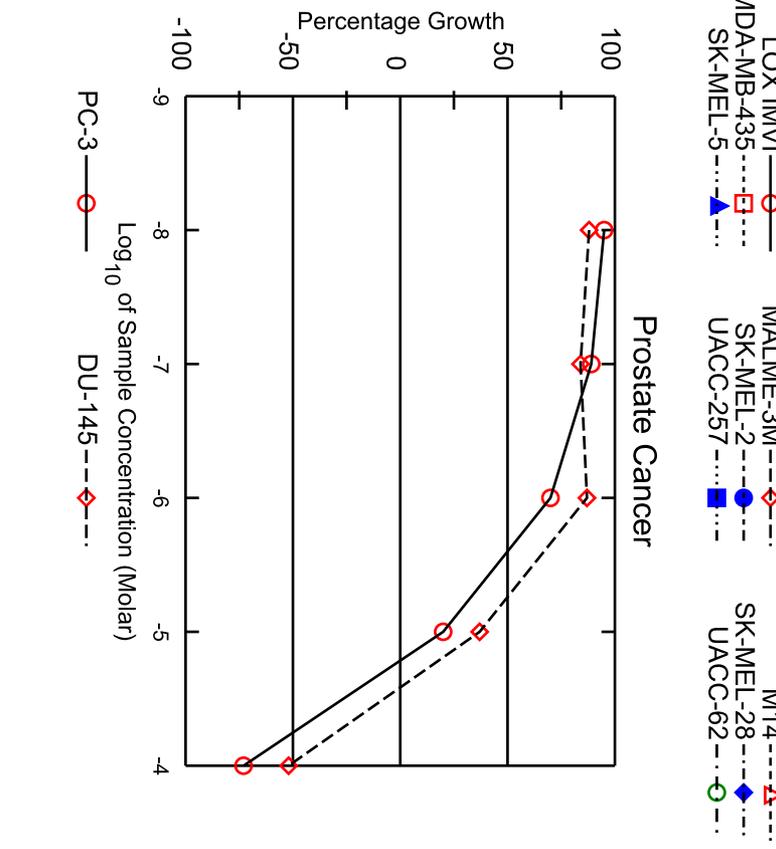
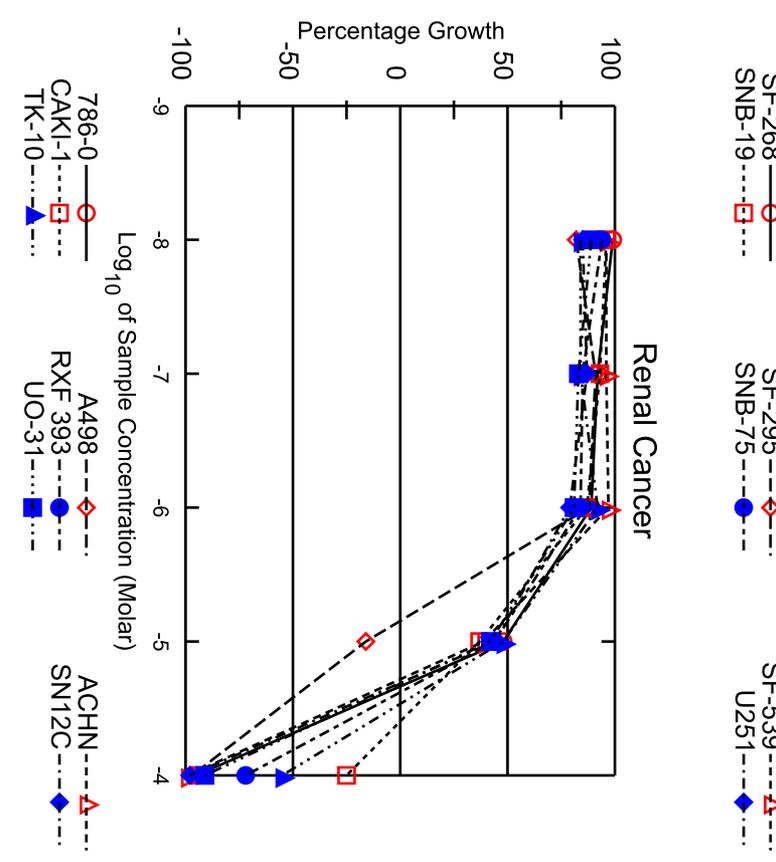
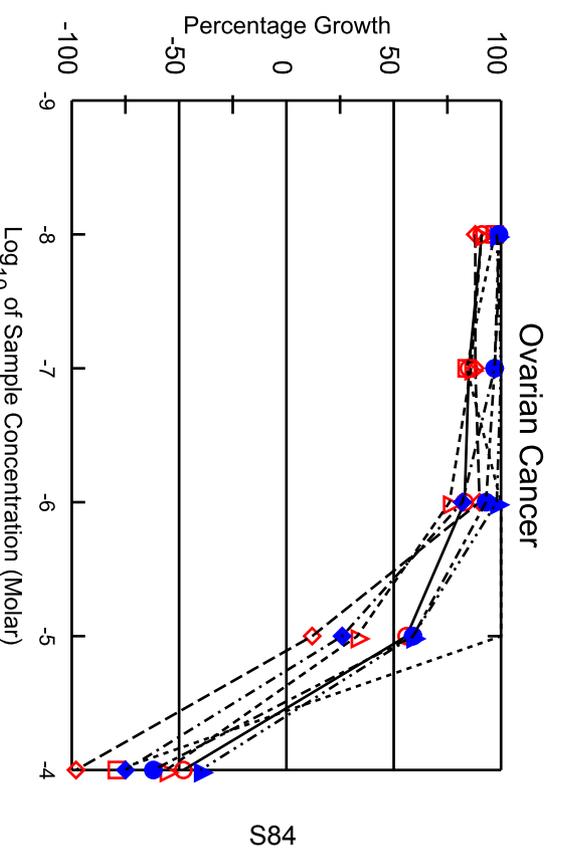
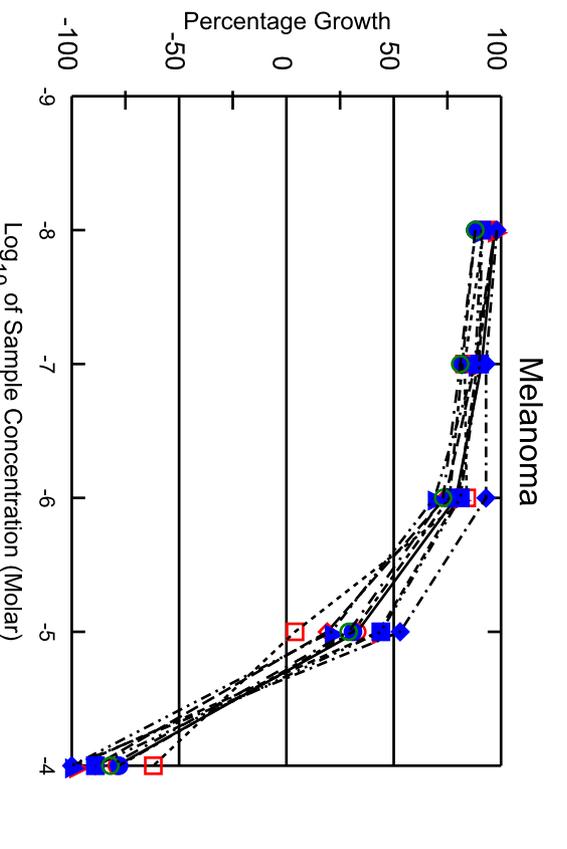
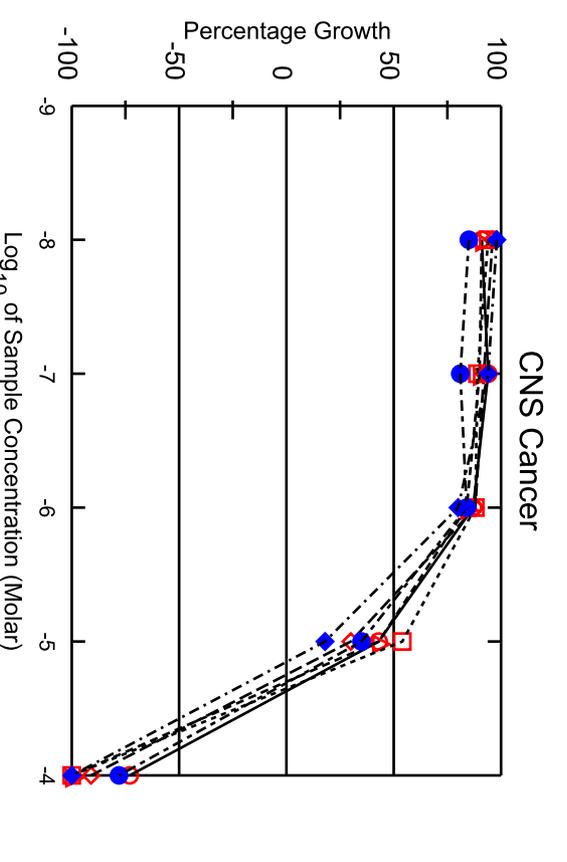
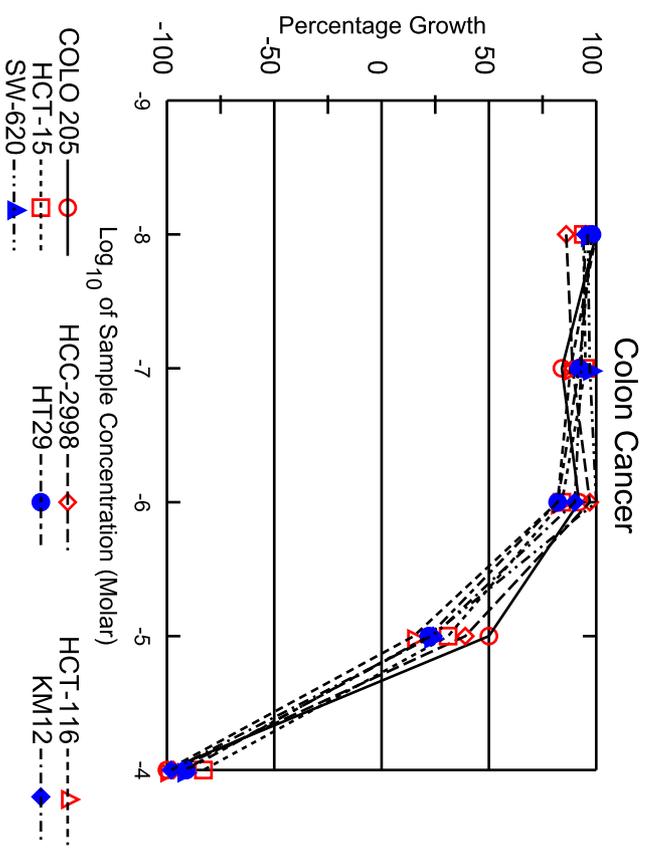
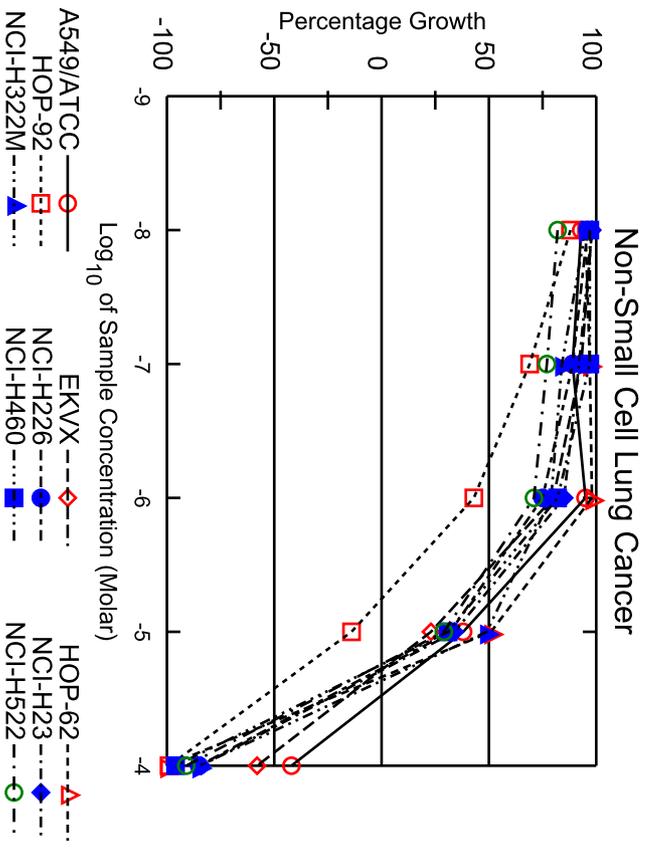
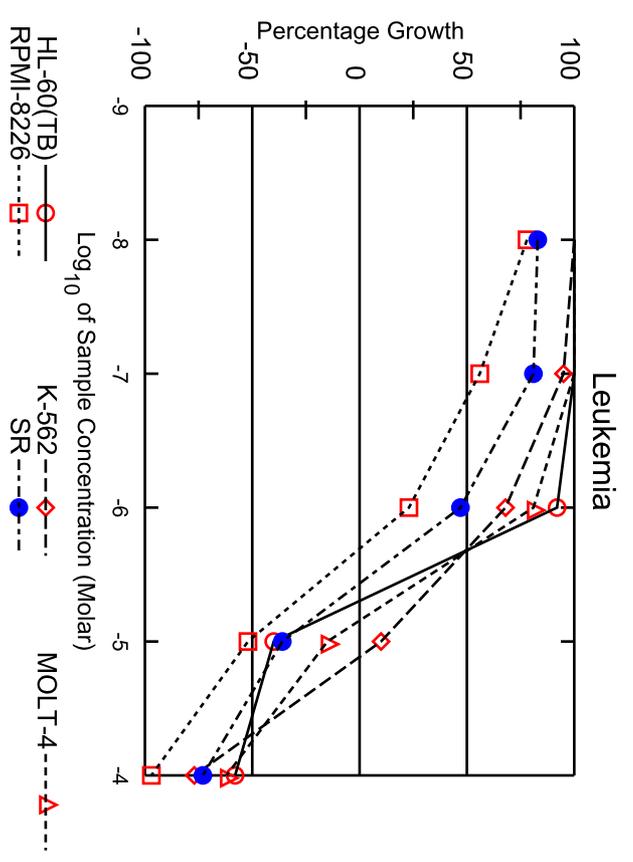
**Mean Growth Percent - Growth Percent**

Leukemia	
CCRF-CEM	68.96
HL-60(TB)	44.43
K-562	67.84
MOLT-4	74.30
RPMI-8226	18.49
SR	62.58
Non-Small Cell Lung Cancer	
A549/ATCC	71.63
EKVX	102.28
HOP-62	108.33
HOP-92	66.62
NCI-H226	98.10
NCI-H23	95.40
NCI-H322M	120.70
NCI-H460	92.68
NCI-H522	58.36
Colon Cancer	
COLO 205	101.29
HCC-2998	115.46
HCT-116	71.81
HCT-15	103.41
HT29	89.53
KM12	85.93
SW-620	101.12
CNS Cancer	
SF-268	105.37
SF-295	85.19
SF-539	92.75
SNB-19	82.90
SNB-75	80.72
U251	97.35
Melanoma	
LOX IMVI	89.75
MALME-3M	101.14
M14	102.54
MDA-MB-435	109.16
SK-MEL-2	71.47
SK-MEL-28	110.20
SK-MEL-5	84.67
UACC-257	76.82
UACC-62	54.63
Ovarian Cancer	
IGROV1	65.74
OVCAR-3	115.72
OVCAR-4	88.01
OVCAR-5	100.16
OVCAR-8	94.80
NCI/ADR-RES	108.30
SK-OV-3	93.49
Renal Cancer	
786-0	93.62
A498	104.48
ACHN	93.56
RXF 393	98.38
SN12C	100.70
TK-10	113.66
UO-31	78.96
Prostate Cancer	
PC-3	22.47
DU-145	105.62
Breast Cancer	
MCF7	93.70
MDA-MB-231/ATCC	101.47
HS 578T	95.86
BT-549	112.93
T-47D	56.11
MDA-MB-468	102.71
Mean	88.21
Delta	69.72
Range	102.21



All Cell Lines





# National Cancer Institute Developmental Therapeutics Program In-Vitro Testing Results

NSC : D - 749112 / 1	Experiment ID : 0902NS35	Test Type : 08	Units : Molar
Report Date : March 24, 2009	Test Date : February 02, 2009	QNS :	MC :
COMI : DB.10h (81264)	Stain Reagent : SRB Dual-Pass Related	SSPL : 0XCV	

Panel/Cell Line	Time Zero	Log10 Concentration											GI50	TGI	LC50	
		Ctrl	-8.0	-7.0	-6.0	-5.0	-4.0	-8.0	-7.0	-6.0	-5.0	-4.0				
<b>Leukemia</b>																
HL-60(TB)	0.986	2.104	2.149	2.153	2.017	0.589	0.415	104	104	92	-40	-58	2.08E-6	4.97E-6	3.55E-5	
K-562	0.252	1.205	1.210	1.156	0.897	0.352	0.059	101	95	68	10	-77	2.04E-6	1.32E-5	4.93E-5	
MOLT-4	0.530	1.404	1.457	1.404	1.241	0.452	0.203	106	100	81	-15	-62	2.12E-6	7.03E-6	5.62E-5	
RPMI-8226	0.854	1.741	1.548	1.348	1.056	0.409	0.028	78	56	23	-52	-97	1.48E-7	2.01E-6	9.36E-6	
SR	0.425	0.820	0.753	0.744	0.609	0.273	0.114	83	81	47	-36	-73	7.98E-7	3.67E-6	2.39E-5	
<b>Non-Small Cell Lung Cancer</b>																
A549/ATCC	0.245	1.246	1.178	1.135	1.193	0.627	0.143	93	89	95	38	-42	6.17E-6	3.01E-5	> 1.00E-4	
EKVX	0.596	1.398	1.380	1.341	1.236	0.783	0.252	98	93	80	23	-58	3.37E-6	1.94E-5	8.03E-5	
HOP-62	0.452	1.137	1.100	1.115	1.124	0.801	0.002	95	97	98	51	-100	1.01E-5	2.18E-5	4.68E-5	
HOP-92	0.824	1.239	1.188	1.112	1.004	0.712	0.011	88	69	43	-14	-99	5.52E-7	5.76E-6	2.68E-5	
NCI-H226	0.670	1.231	1.206	1.171	1.090	0.851	0.104	96	89	75	32	-85	3.82E-6	1.89E-5	5.06E-5	
NCI-H23	0.665	1.820	1.792	1.723	1.648	1.060	0.105	98	92	85	34	-84	4.88E-6	1.94E-5	5.14E-5	
NCI-H322M	0.621	1.739	1.686	1.557	1.507	1.173	0.091	95	84	79	49	-85	9.50E-6	2.32E-5	5.46E-5	
NCI-H460	0.261	2.111	2.057	2.061	1.784	0.832	0.011	97	97	82	31	-96	4.24E-6	1.75E-5	4.34E-5	
NCI-H522	0.396	1.269	1.111	1.069	1.013	0.649	0.036	82	77	71	29	-91	3.13E-6	1.74E-5	4.55E-5	
<b>Colon Cancer</b>																
COLO 205	0.280	0.968	0.978	0.859	0.910	0.621	-0.011	101	84	92	50	-100	9.77E-6	2.15E-5	4.63E-5	
HCC-2998	0.683	1.954	1.779	1.818	1.915	1.176	0.024	86	89	97	39	-97	6.40E-6	1.93E-5	4.53E-5	
HCT-116	0.251	1.689	1.718	1.524	1.424	0.472	-0.014	102	88	82	15	-100	3.00E-6	1.36E-5	3.69E-5	
HCT-15	0.376	2.159	2.057	2.072	1.878	0.927	0.063	94	95	84	31	-83	4.38E-6	1.86E-5	5.10E-5	
HT29	0.149	0.997	0.976	0.928	0.845	0.334	0.013	98	92	82	22	-91	3.41E-6	1.56E-5	4.32E-5	
KM12	0.466	2.085	2.011	1.964	1.921	0.853	0.010	95	93	90	24	-98	4.02E-6	1.57E-5	4.04E-5	
SW-620	0.291	1.511	1.462	1.472	1.520	0.572	0.025	96	97	101	23	-92	4.49E-6	1.59E-5	4.34E-5	
<b>CNS Cancer</b>																
SF-268	0.362	1.187	1.115	1.136	1.077	0.718	0.096	91	94	87	43	-73	6.95E-6	2.34E-5	6.29E-5	
SF-295	1.371	3.235	3.161	3.095	3.020	1.921	0.124	96	92	88	30	-91	4.49E-6	1.76E-5	4.57E-5	
SF-539	0.743	2.275	2.135	2.118	2.026	1.406	-0.001	91	90	84	43	-100	6.82E-6	2.00E-5	4.48E-5	
SNB-19	0.642	1.499	1.445	1.403	1.398	1.107	-0.007	94	89	88	54	-100	1.07E-5	2.25E-5	4.74E-5	
SNB-75	0.585	1.212	1.119	1.092	1.111	0.805	0.132	85	81	84	35	-78	4.94E-6	2.05E-5	5.69E-5	
U251	0.235	1.379	1.353	1.309	1.153	0.442	-0.001	98	94	80	18	-100	3.06E-6	1.42E-5	3.77E-5	
<b>Melanoma</b>																
LOX IMVI	0.270	1.964	1.911	1.807	1.603	0.835	0.058	97	91	79	33	-79	4.30E-6	1.98E-5	5.54E-5	
MALME-3M	0.708	1.590	1.562	1.485	1.365	0.879	0.080	97	88	74	19	-89	2.78E-6	1.51E-5	4.38E-5	
M14	0.256	0.915	0.897	0.846	0.785	0.540	0.005	97	89	80	43	-98	6.51E-6	2.02E-5	4.56E-5	
MDA-MB-435	0.472	1.904	1.787	1.665	1.676	0.524	0.181	92	83	84	4	-62	2.65E-6	1.14E-5	6.63E-5	
SK-MEL-2	0.507	0.967	0.913	0.886	0.856	0.649	0.111	88	82	76	31	-78	3.74E-6	1.92E-5	5.52E-5	
SK-MEL-28	0.479	1.173	1.157	1.121	1.122	0.847	-0.001	98	93	93	53	-100	1.05E-5	2.22E-5	4.71E-5	
SK-MEL-5	0.327	1.972	1.816	1.779	1.461	0.673	-0.015	91	88	69	21	-100	2.48E-6	1.49E-5	3.86E-5	
UACC-257	0.602	1.345	1.280	1.272	1.201	0.926	0.067	91	90	81	44	-89	6.71E-6	2.13E-5	5.09E-5	
UACC-62	0.265	0.930	0.852	0.804	0.748	0.459	0.048	88	81	73	29	-82	3.31E-6	1.83E-5	5.15E-5	
<b>Ovarian Cancer</b>																
IGROV1	0.319	1.316	1.225	1.164	1.144	0.874	0.167	91	85	83	56	-48	1.13E-5	3.46E-5	> 1.00E-4	
OVCAR-3	0.361	0.987	0.914	0.912	0.925	0.434	0.007	88	88	90	12	-98	3.24E-6	1.28E-5	3.65E-5	
OVCAR-4	0.397	1.403	1.310	1.266	1.165	0.733	0.177	91	86	76	33	-56	4.10E-6	2.37E-5	8.66E-5	
OVCAR-5	0.478	0.903	0.891	0.837	0.905	1.015	0.101	97	84	100	126	-79	2.36E-5	4.13E-5	7.23E-5	
OVCAR-8	0.302	1.242	1.233	1.210	1.181	0.858	0.114	99	97	93	59	-62	1.19E-5	3.06E-5	7.90E-5	
NCI/ADR-RES	0.556	1.619	1.667	1.589	1.423	0.829	0.137	105	97	82	26	-75	3.67E-6	1.79E-5	5.61E-5	
SK-OV-3	0.621	1.326	1.309	1.323	1.314	1.037	0.372	98	100	98	59	-40	1.23E-5	3.93E-5	> 1.00E-4	
<b>Renal Cancer</b>																
786-0	0.594	2.154	2.139	2.032	1.980	1.336	0.027	99	92	89	48	-95	8.73E-6	2.15E-5	4.81E-5	
A498	1.150	1.457	1.402	1.430	1.422	0.970	0.035	82	91	88	-16	-97	2.34E-6	7.07E-6	2.65E-5	
ACHN	0.334	1.372	1.332	1.333	1.344	0.734	0.002	96	96	97	39	-99	6.39E-6	1.90E-5	4.38E-5	
CAKI-1	0.864	2.632	2.565	2.515	2.399	1.520	0.651	96	93	87	37	-25	5.50E-6	3.98E-5	> 1.00E-4	
RXF 393	0.398	1.056	1.016	0.960	0.954	0.686	0.112	94	85	84	44	-72	7.00E-6	2.39E-5	6.47E-5	
SN12C	0.544	1.869	1.668	1.703	1.593	1.123	0.012	85	87	79	44	-98	6.63E-6	2.03E-5	4.59E-5	
TK-10	0.599	1.222	1.123	1.124	1.170	0.897	0.268	84	84	92	48	-55	8.93E-6	2.91E-5	8.89E-5	
UO-31	0.460	1.368	1.264	1.217	1.193	0.845	0.041	89	83	81	42	-91	6.32E-6	2.08E-5	4.92E-5	
<b>Prostate Cancer</b>																
PC-3	0.422	1.262	1.218	1.166	1.006	0.592	0.116	95	89	70	20	-73	2.49E-6	1.65E-5	5.72E-5	
DU-145	0.297	1.210	1.104	1.061	1.089	0.637	0.142	88	84	87	37	-52	5.53E-6	2.61E-5	9.45E-5	
<b>Breast Cancer</b>																
MCF7	0.265	1.401	1.317	1.441	1.209	0.557	0.084	93	103	83	26	-68	3.77E-6	1.88E-5	6.39E-5	
MDA-MB-231/ATCC	0.489	1.367	1.378	1.271	1.204	0.871	0.090	101	89	81	43	-82	6.73E-6	2.23E-5	5.59E-5	
HS 578T	0.689	1.300	1.194	1.194	1.099	0.858	0.397	83	83	67	28	-42	2.72E-6	2.48E-5	> 1.00E-4	
BT-549	1.527	2.435	2.405	2.293	2.136	1.442	0.166	97	84	67	-6	-89	1.72E-6	8.37E-6	3.40E-5	
T-47D	0.705	1.402	1.389	1.325	1.236	0.717	0.452	98	89	76	2	-36	2.24E-6	1.11E-5	> 1.00E-4	
MDA-MB-468	0.404	1.289	1.227	1.106	0.911	0.502	0.148	93	79	57	11	-63	1.43E-6	1.41E-5	6.61E-5	

Mean Graphs

Report Date :March 24, 2009

Test Date :February 02, 2009

