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Figure S1 Srf-deficient newborn epidermis is grossly defective and yet cells within the tissue still display typical signs of adhesion. Ultrastructural analysis of WT (a) and Srf-cKO (b) epidermis. Arrows indicate vacuoles, prevalent by newborn, which could be mistaken for adhesive gaps at the light microscopic

level. Morphology shows grossly disorganized epidermis. (c) Boxed area in (b) is shown at higher magnification. (d-e) Boxed areas in (c) are shown at higher magnification. Kf, keratin filaments; Des, desmosome; Der, dermis; BL, basal layer; SP, spinous layer; Gr, granular layer; Sc, stratum corneum. Bars=10µm



Figure S2 Elevated proliferation and apoptosis in *Srf*-cKO epidermis is evident beginning at E17.5. (a-f) Immunofluorescence of frozen backskin sections (10 µm) from E16.5-E18.5 embryos labeled with: (a) Ki67, indicative of S and M-phase cells and β 4 integrin, a hemidesmosomal

component (arrows denote suprabasal mitoses); and(b) active caspase3 and β 4 integrin. (c) Quantifications of phospho-histone H3 (pHH3)-positive cells, reflective of early mitosis. *indicates statistically significant differences between WT and cK0. E17.5: p=0.012; E18.5: p=0.037.



Figure S3 Signs of inflammation in the skin are not appreciable until late after *Srf* ablation during epidermal embryogenesis. (a) Wound-induced inflammatory response in P21 CD1 mouse (positive control). 24h after wounding, backskin sections were frozen, sectioned (10 μ m) and processed for immunofluorescence with CD45, Mac1 and CD3 antibodies. (b-c) Unwounded E16.5 and newborn WT and *Srf-cKO*

embryos were frozen, sectioned and processed for immunofluorescence as in (a). Dotted lines denote dermo-epidermal boundary. Bar=20 μ m. (d-e) Quantifications of the CD45 and Mac1 positive immune cells in the skin of E16.5, E17.5 and newborn mice. * indicates statistically significant differences between WT and cK0: CD45, p=0.014; Mac1, p=0.007.



Figure S4 Normal distribution of adhesion proteins in newborn *Srf*-cKO skin. Backskin sections from newborn mice were frozen, sectioned (10 μ m) and processed for immunofluorescence with α - and β -catenin, (α cat, β cat) P-cadherin (P-cad), desmoplakin (DP), Laminin 5 α , and β 4 and β 1 integrins (β 4, β 1). Note overall normal distribution of these proteins. Arrows denote suprabasal laminin 5 α and β 4 integrin. Dotted lines denote dermo-epidermal boundaries. Bar=20 μ m.



a FACS E16.5 basal cells

Figure S5 Most early transcriptional changes in basal cells depleted of Srf involve regulators of actomyosin and not cell cycle, inflammation, apicobasal polarity, or Notch signaling genes. (a) Fluorescence activated cell sorting (FACS) profile depicting populations of E16.5 basal cells used for microarray analyses. Embryos were from *K14-Cre X Rosa26YFP fl/fl* (WT) and *K14-Cre* X *Rosa26YFP fl/fl* X *Srf fl/fl* (cKO) strains. Basal cells active for Cre recombinase expressed YFP and were enriched for surface α 6 integrin (green). (b) Verification of the array results by semiquantitative RT-PCR of mRNAs from FACS-purified E16.5 basal progenitors. n=2. See Table 1 and Fig. 5a for additional details.

F-actin β4 integrin



Figure S6 Alterations the actin cytoskeleton during the development of *Srf-cKO* embryos. backskin sections from E16.5, E17.5 and E18.5 mice were frozen, sectioned (10 μ m) and processed for immunofluorescence. Phalloidin staining to mark F-actin (red) and β 4 integrin (green). Note the progressive loss of F-actin staining. Bar=20 μ m



Figure S7 Normal recruitment of myosin-IIA at the midzone of late mitotic cells deficient for Srf. WT and *Srf*-cKO E16.5 embryos were subjected to whole-mount fluorescence microscopy for F-actin (phalloidin), myosin-IIA (MIIA), and

DAPI. Shown are planar images through the basal layer. White arrows indicate late mitotic cells, enriched in MIIA in WT but not *Srf*-cKO. Yellow arrows indicate mid-zone enriched for MIIA in both WT and cKO samples. Bar=10µm.



Figure S8 LGN localization is sensitive to perturbations in actin dynamics and myosin motor activity. Live E14.5 CD1 mouse embryos were placed directly into culture medium and treated with DMSO, latrunculin (Lat 2.5μ M and 0.25μ M), jasplakinolide (Jsp 0.5μ M or blebistatin (BS, 50µm) for 1 hour at 37°C. Embryos were then frozen in OCT, sectioned (10µm) and processed for fluorescence microscopy: (a) F-actin (grayscale, phalloidin staining), Par3 and pericentrin (PC) (red). For the DMSO and latrunculin treatments, see Fig. 5 and main text. (b) Phosphohistone H3 (pHH3)(green), LGN (red) and DAPI depicting representative examples of immunolocalization. Dotted line denotes dermo-epidermal boundary. Bars=20 μ m. (c) Quantifications of the experimental analyses, showing percentages of early mitotic cells (pHH3+) in which LGN can be detected by immunolabeling. * indicates p<0.05. DMSO vs Lat 2.5 μ m p=0.035 DMSP vs Jsp 0.5 μ m p=0.029.(d) Percentages of early mitotic cells (pHH3+) in which LGN exhibited an apical crescent. * indicates p<0.05. DMSO vs Lat 0.25 μ m p=0.008, DMSO vs Jsp 0.5 μ m p=0.004; DMSO vs BS 50 μ m p=0.003.







Figure 6e



Figure S9 Full scans

Table S1. Summary of Microarray Analysis of decrease in GeneExpression in E16.5Srf cKO vs WT Basal Cells.

The listed probe sets were down regulated (fold change >1.5 and p<0.05) in the two independent experiment.

Probe Set	Gene Symbol	Gene Title	Mean fold change
1427682 a at	Egr2	early growth response 2	9.09
1427683 at	Egr2	early growth response 2	8.12
1442067 at			6.73
1417065 at	Ear1	early growth response 1	5.05
1436329 at	Ear3	early growth response 3	3.88
1449141 at	Fblim1	filamin binding LIM protein 1	3.69
		histocompatibility 2 K1 K	0.00
1425336 x at	H2-K1	region	3.22
1439183 at	Acer1	alkaline ceramidase 1	3,155
1450645 at	Mt4	metallothionein 4	3 14
1450981 at	Cnn2	calponin 2	3 14
1100001_ut	01112	prostaglandin-endoperoxide	0.11
1436448 a at	Ptas1	synthase 1	2.96
1449204 at	Gib5	gap junction protein beta 5	2.62
AFFX-b-			
ActinMur/M12481 5 at	Actb	actin, beta	2.41
		short chain	
		dehydrogenase/reductase	
1457025 at	Sdr16c6	family 16C, member 6	2.39
—		fatty acid binding protein 4,	
1451263_a_at	Fabp4	adipocyte	2.38
		lymphocyte antigen 6	
1416930_at	Ly6d	complex, locus D	2.38
AFFX-b-			
ActinMur/M12481_M_a			
t	Actb	actin, beta	2.32
1453568_at	Dapl1	death associated protein-like 1	2.30
1460330_at	Anxa3	annexin A3	2.30
1422587_at	Tmem45a	transmembrane protein 45a	2.22
1424701_at	Pcdh20	protocadherin 20	2.16
		chloride channel calcium	
1437578_at	Clca2	activated 2	2.15
1417942_at	Lypd3	Ly6/Plaur domain containing 3	2.10
		elongation of very long chain	
		fatty acids (FEN1/Elo2,	
1451308_at	Elovl4	SUR4/Elo3, yea	2.08
1417928_at	Pdlim4	PDZ and LIM domain 4	2.08
1452679_at	Tubb2b	tubulin, beta 2B	1.99
		cysteine and glycine-rich	
1425810_a_at	Csrp1	protein 1	1.94
		RIKEN cDNA 1110020C03	
	1110020C03Rik	gene /// similar to CG4329-PA,	
1429808_at	///	isoform A	1.94
AFFX-b-	•		
ActinMur/M12481_3_at	Actb	actin, beta	1.94
1425811_a_at	Csrp1	cysteine and glycine-rich	1.93

1425811_a_at	Csrp1	cysteine and glycine-rich protein 1	1.93
1417408 at	F3	coagulation factor III	1.90
1456539 at			1.90
1427038 at	Penk	preproenkephalin	1.90
1436722 a at	Actb	actin, beta	1.88
		elongation of very long chain	
		fatty acids (FEN1/Elo2.	
1424306 at	Elovl4	SUR4/Elo3, yea	1.86
—		thioesterase superfamily	
1431211 s at	Them5	member 5	1.83
		N-acetylneuraminate pyruvate	
1424265_at	Npl	lyase	1.83
1427910_at	Cst6	cystatin E/M	1.81
—		Rho GTPase activating protein	
1424842_a_at	Arhgap24	24	1.81
1455519_at	Dsg1b	desmoglein 1 beta	1.80
1426677_at	Flna	filamin, alpha	1.75
1415779_s_at	Actg1	actin, gamma, cytoplasmic 1	1.74
	-	proprotein convertase	
1448312_at	Pcsk2	subtilisin/kexin type 2	1.69
		insulin-like growth factor	
1423062_at	lgfbp3	binding protein 3	1.69
1437591_a_at	Wdr1	WD repeat domain 1	1.68
1419734_at	Actb	actin, beta	1.68
1444254_at	Tns4	tensin 4	1.63
1431429_a_at	Arl4a	ADP-ribosylation factor-like 4A	1.63
1459898_at	Sbsn	suprabasin	1.63
1423054_at	Wdr1	WD repeat domain 1	1.63
1450851_at	Wdr1	WD repeat domain 1	1.62
1448318_at	Plin2	perilipin 2	1.57
		serine (or cysteine) peptidase	
		inhibitor, clade B (ovalbumin),	
1429297_at	Serpinb12	member	1.57
1427256_at	Vcan	versican	1.57
		RIKEN cDNA 2200001115	
1437019_at	2200001I15Rik	gene	1.52
1419602_at	Hoxa2	homeo box A2	1.52

Table S2. Summary of Microarray Analysis of Increase in GeneExpression in E16.5Srf cKO vs WT Basal Cells.

Srf cKO vs WT Basal Cells. The listed probe sets were up regulated (fold change >1.5 and p<0.05) in the two independent experiment.

		eukaryotic translation initiation factor 2,	
1420491 at	Eif2s1	subunit 1 alpha	3.64
1418930 at	Cxcl10	chemokine (C-X-C motif) ligand 10	3.29
1442886 at			3.23
1442700 at	Pde4b	phosphodiesterase 4B, cAMP specific	2.33
1439780 at	Rpl7l1	ribosomal protein L7-like 1	2.23
_	120000311		
	0Rik ///		
	1200015M		
	12Rik ///		
	1200016E2		
	4Rik ///	RIKEN cDNA 1200003110 gene ///	
	A130040M	RIKEN cDNA 1200015M12 gene ///	
	12Rik ///	RIKEN cDNA 1200016E24 gene ///	
	F430024C	RIKEN cDNA A130040M12 gene ///	
1427932 s at	06Rik	RIKEN cDNA F430024C06 gene	2 14
1127002_0_ut	1200016E2		2.11
	4Rik ///		
	3930401B1		
	9Rik ///		
	A130040M	RIKEN cDNA 1200016E24 gene ///	
	12Rik ///	RIKEN cDNA 3930401B19 gene ///	
	F430024C	RIKEN cDNA A130040M12 gene ///	
1453238 s at	06Rik	RIKEN cDNA F430024C06 gene	1 94
1100200_0_4	o or the	acyl-Coenzyme A dehydrogenase long-	
1448987 at	Acadl	chain	1.83
in locol_at		LIM and senescent cell antigen-like	
1418230 a at	Lims1	domains 1	1.81
1419721 at	Niacr1	niacin receptor 1	1.80
	D17H6S56		
1417821 at	E-5	DNA segment, Chr 17, human D6S56E 5	1.80
	_ •	cholinergic receptor, nicotinic, alpha	
1440681 at	Chrna7	polypeptide 7	1 75
1439200 x at			1.70
1100200_A_at		cysteine rich transmembrane BMP regulator	
1426951 at	Crim1	1 (chordin like)	1.69
	•	growth arrest and DNA-damage-inducible	
1450971 at	Gadd45b	45 beta	1.69
1416104 at	Mpdu1	mannose-P-dolichol utilization defect 1	1.68
1436983 at	Crebbp	CREB binding protein	1.62
1460713 at	BC048355	cDNA sequence BC048355	1.62
1452406 x at	Erdr1	ervthroid differentiation regulator 1	1.62
1416124 at	Ccnd2	cvclin D2	1.57
1450780 s at	Hmga2	high mobility aroup AT-hook 2	1.57
1422851 at	Hmga2	high mobility group AT-hook 2	1.52
1429660 s at	Smc2	structural maintenance of chromosomes 2	1.52
1418283 at	Cldn4	claudin 4	1.52
1455956 x at	Ccnd2	cyclin D2	1.52

Gene	Forward primer	Reverse primer
HPRT	gatcagtcaacgggggacataaa	cttgcgctcatcttaggctttgt
Ppib	gtgagcgcttcccagatgaga	tgccggagtcgacaatgatg
Srf	gttgcccgccaccatcat	cgggcggatcattcactctt
itga6	ctggacacccgcgaggacaac	tcaaccggccatcgcagaaact
Krt14	cgccgccctggtgtgg	atctggcggttggtggaggtca
Krt5	gttgaacgccgctgacct	cttcggaaggacacactggac
Notch1	caaactggcctgggtggggacat	aaaaggccagaaagagctgccctgag
Jag1	aagggaacagactgagctatatgactta	atttattgccaggaacaacacatcaaag
Pard6b	gggacctgccgcctataaataat	acacgggccggaagtcct
Pard6g	caagcctgggaagtttgaagattt	tgcggcatgctgatgttga
Prkcc	cagcgacagagaaaacttcctgaa	tcccgccatcatctcaaacata
Numa1	gtcaggcccccttggagact	agcgggccagagactgagtg
Gpsm2	tctgctgcaaagagatccaaaca	tcatgggcaggtacaaaaagtcc
Myh9	cctgccataagggaacctaatcac	gcgctctggtgcctctccta
Fos	agagcgccccatccttacg	ggtgggctgccaaaataaactc
junb	acacaggcgcatctctgaagc	cggctccggaccagcata
Egr2	ggccgtagacaaaatcccagtaac	gaatttgcccatgtaagtgaaggtc
Egr1	atgtgggtggtaaggtggtcacta	aaccggcccagcaagaca
Nab2	gatccggaagtacagcgtcatcta	acttgtcgggacagtgagaagagtt
Gadd45a	tgccgggaaagtcgctacat	tttctcgcagcttccttcttcag
Gadd45b	tacgaggcggccaaactgat	tgatacccggacgatgtcaatg
Cnn2	aatgggcttcctgtttcttcatct	tcgtgggaaagcaaacttagtcc
Actb	cggccaggtcatcactattgg	aggggccggactcatcgta
Actg1	cccaaagctaacagagagaagatgacg	gtggtaaagctgtagccccgttca
Flna	gattggggaggagacggtgat	tttgctggctaccctgaggatag
Wdr1	tggagcggggcgtctcta	aatccgctgggtgcatacttg

Table S3. Semi-Quantitative RT-PCR Primers table.