

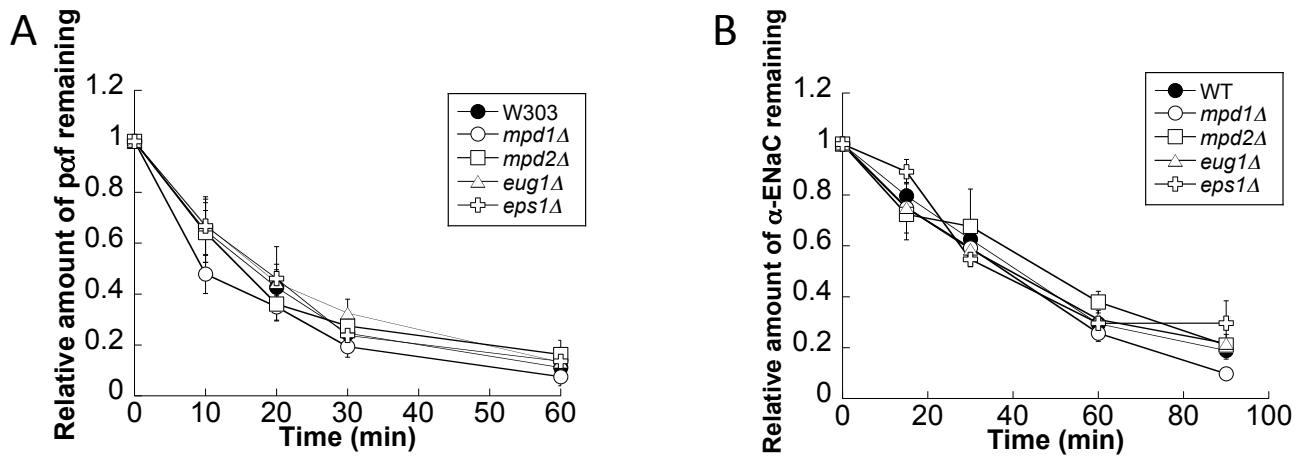
## Supplemental Table 1 Yeast Strains

Strain	Genotype	Source
W303	<i>MAT<math>\alpha</math>, ade2-1, can1-100, his3-11,15, leu2-3,112, trp1-1, ura3-1</i>	This lab
<i>mpd1<math>\Delta</math></i>	<i>MAT<math>\alpha</math>, ade2-1, can1-100, his3-11,15, leu2-3,112, trp1-1, ura3-1, mpd1::KANMX</i>	This study
<i>mpd2<math>\Delta</math></i>	<i>MAT<math>\alpha</math>, ade2-1, can1-100, his3-11,15, leu2-3,112, trp1-1, ura3-1, mpd2::KANMX</i>	This study
<i>eug1<math>\Delta</math></i>	<i>MAT<math>\alpha</math>, ade2-1, can1-100, his3-11,15, leu2-3,112, trp1-1, ura3-1, eug1::KANMX</i>	This study
<i>eps1<math>\Delta</math></i>	<i>MAT<math>\alpha</math>, ade2-1, can1-100, his3-11,15, leu2-3,112, trp1-1, ura3-1, eps1::KANMX</i>	This study
M4492	<i>MAT<math>\alpha</math> pdi1::HIS3 <math>\Delta</math>eps1 <math>\Delta</math>eug1 <math>\Delta</math>mpd1 <math>\Delta</math>mpd2::G418 ura3 trp1 his3 [pBH1800 (MPD1 CEN TRP1)]</i>	Norgaard et al., 2001
SRH01	<i>MAT<math>\alpha</math> pdi1::HIS3 <math>\Delta</math>eps1 <math>\Delta</math>eug1 <math>\Delta</math>mpd1 <math>\Delta</math>mpd2::G418 ura3 trp1 his3 [pSG01]</i>	This study
<i>pdi1<math>\Delta</math>[PDI1<sub>CGHC-CGHC</sub>]</i>	<i>MAT<math>\alpha</math>, ade2-1, can1-100, his3-11,15, leu2-3,112, trp1-1, ura3-1, pdi1::HIS3, [pBH1464]</i>	Luz and Lennarz, 1998
<i>pdi1<math>\Delta</math>[PDI1<sub>SGHS-CGHC</sub>]</i>	<i>MAT<math>\alpha</math>, ade2-1, can1-100, his3-11,15, leu2-3,112, trp1-1, ura3-1, pdi1::HIS3, [pBH1852]</i>	Luz and Lennarz, 1998
<i>pdi1<math>\Delta</math>[PDI1<sub>CGHC-SGHS</sub>]</i>	<i>MAT<math>\alpha</math>, ade2-1, can1-100, his3-11,15, leu2-3,112, trp1-1, ura3-1, pdi1::HIS3, [pBH1630]</i>	Luz and Lennarz, 1998
<i>pdi1<math>\Delta</math>[PDI1<sub>222-302<math>\Delta</math>]</sub></i>	<i>MAT<math>\alpha</math>, ade2-1, can1-100, his3-11,15, leu2-3,112, trp1-1, ura3-1, pdi1::HIS3, [pRS-<math>\Delta</math>222-302]</i>	Gillece et al., 1999
M4130	<i>MAT<math>\alpha</math>, pdi1::HIS3, ade2, can1, ura3, leu2, trp1, his3, [pCT37]</i>	Norgaard et al., 2001
<i>ire1<math>\Delta</math></i>	<i>Mat<math>\alpha</math>, lys2, his3, leu2, ura3, ire1::KANMX</i>	This lab
SEY6210	<i>MAT<math>\alpha</math>, ura3-52, leu2-3,112, trp1-<math>\Delta</math>901, his3-<math>\Delta</math>200, lys2-801, suc2-<math>\Delta</math>9</i>	Nakatsukasa et al., 2001
<i>htm1<math>\Delta</math></i>	<i>MAT<math>\alpha</math>, ura3-52, leu2-3,112, trp1-<math>\Delta</math>901, his3-<math>\Delta</math>200, lys2-801, suc2-<math>\Delta</math>9, htm1::HIS3</i>	Nakatsukasa et al., 2001
W303	<i>MAT<math>\alpha</math>, ade2-1, can1-100, his3-11,15, leu2-3,112, trp1-1, ura3-1</i>	Gauss et al., 2011
KKY415	<i>MAT<math>\alpha</math>, pdi1-1, his3-11::HIS3-UPRE LacZ, trp1-1, his3-11,15, ura3-1, can1-100, ade2-1, leu2-3,112</i>	Gauss et al., 2011

## Supplemental Table 2. Plasmids

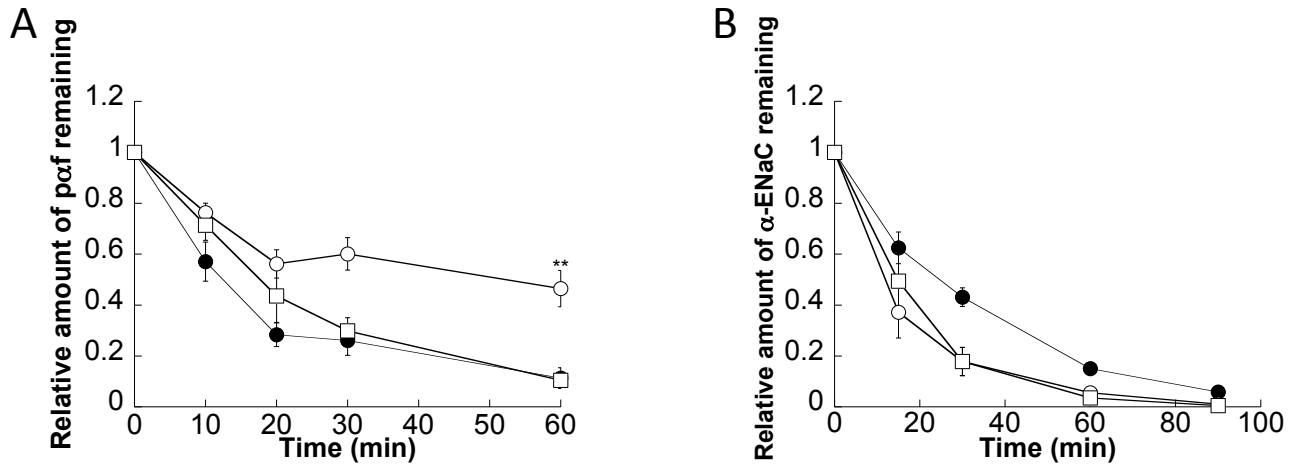
Plasmid Name	Notes	Selectable Marker	Reference
pRS316CPY*-3HA	CPY* expression	<i>URA3</i>	Bhamidipati <i>et al.</i> , 2005
pSLW1-B29	ApoB29 expression	<i>URA3</i>	Hrizo <i>et al.</i> , 2007
pBH1800	<i>CEN MPD1</i>	<i>TRP1</i>	Norgaard <i>et al.</i> , 2001
pBH1464	<i>CEN PDI1</i> <sub>CGHC-CGHC</sub>	<i>TRP1</i>	Holst <i>et al.</i> , 1997
pBH1852	<i>CEN PDI1</i> <sub>SGHS-CGHC</sub>	<i>TRP1</i>	Holst <i>et al.</i> , 1997
pBH1630	<i>CEN PDI1</i> <sub>CGHC-SGHS</sub>	<i>TRP1</i>	Holst <i>et al.</i> , 1997
pRS-Δ222-302	<i>CEN PDI1</i> <sub>222-302Δ</sub>	<i>TRP1</i>	Gillece <i>et al.</i> , 1999
pCT37	Galactose inducible <i>PDI1</i>	<i>URA3</i>	Norgaard <i>et al.</i> , 2001
pFA6a-KanMX6	KanMX	<i>AMP<sup>R</sup></i>	Longtine <i>et al.</i> , 1998
pSG01	<i>CEN PDI1</i> <sub>CGHC-CGHC</sub>	<i>LEU2</i>	This Study
pRS426GPD ENaC-HA	ENaC alpha subunit expression	<i>URA3</i>	Buck <i>et al.</i> , 2010
pRS426MET25 ENaC-HA	Methionine repressible ENaC alpha subunit expression	<i>URA3</i>	This Study
pSM36-ppafΔG-HA	paf expression	<i>URA3</i>	Kim <i>et al.</i> 2005
pKK223	PrA*-Ab expression	<i>LEU2</i>	Kanehara <i>et al.</i> 2010
pJJB20	Vector control corresponding to ApoB29	<i>URA3</i>	Hrizo <i>et al.</i> , 2007
pRS316	Vector control corresponding to CPY*	<i>URA3</i>	Bhamidipati <i>et al.</i> , 2005
pcDNA3.1	Vector control corresponding to PDI, ERp57, and ERp72	Neomycin <sup>R</sup>	Invitrogen, Iowa City, IA
pcDNA3.1-hPDI	Human PDI expression	Neomycin <sup>R</sup>	This Study
pcDNA3.1-hERp57	Human ERp57 expression	Neomycin <sup>R</sup>	This Study
pcDNA3.1-hERp72	Human ERp72 expression	Neomycin <sup>R</sup>	This Study

## Supplemental Figure 1



**Supplemental Figure 1. The non-essential PDIs do not contribute to the ERAD of paf or  $\alpha$ -ENaC.** Cycloheximide chase reactions were performed as described in the Materials and Methods in wild type (●), *mpd1* $\Delta$  (○), *mpd2* $\Delta$  (□), *eug1* $\Delta$  (△), and *eps1* $\Delta$  (⊠) yeast strains expressing paf (A) from the pSM36-ppaf $\Delta$ G-HA plasmid or  $\alpha$ -ENaC (B) from the pRS426GPD ENaC-HA plasmid. Chase reactions were performed at 30°C, and lysates were immunoblotted with anti-HA antibody. Anti-G6PD antiserum was used as a loading control. Data represent the means of 4-6 experiments,  $\pm$  SEM. The lack of visible error bars indicates that the SEM is less than the size of the symbol.

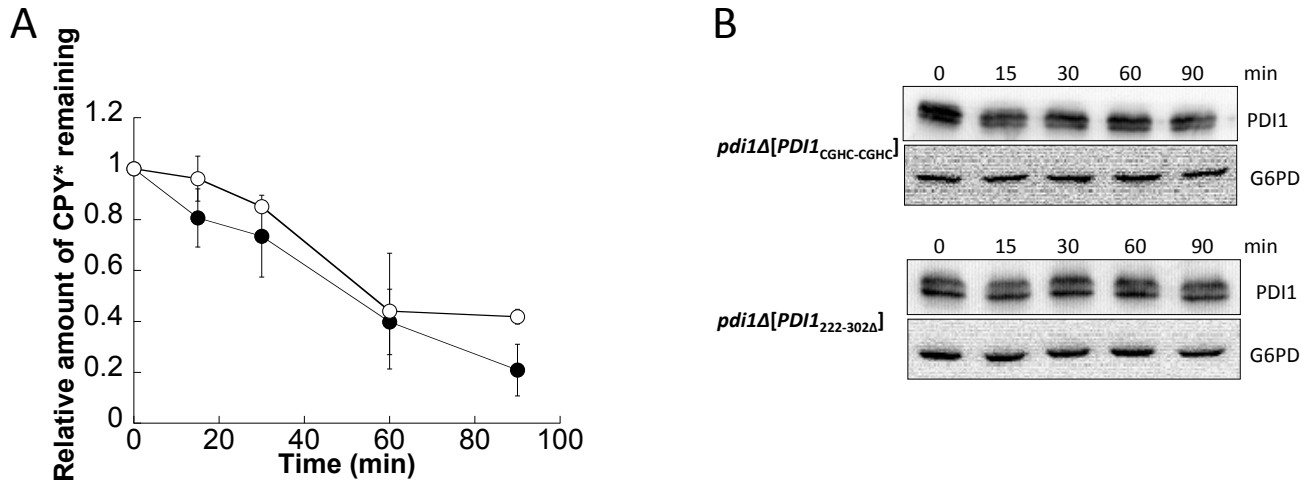
## Supplemental Figure 2



### Supplemental Figure 2. Pdi1 is necessary for the degradation of paf, but not $\alpha$ -ENaC.

Cycloheximide chase reactions were performed as described in the Materials and Methods in wild type (●), M4492 (○), or SRH01 (□) yeast strains expressing paf (A) from the pSM36-ppaf $\Delta$ G-HA plasmid or  $\alpha$ -ENaC (B) from the pRS426MET25 ENaC-HA plasmid. For expression of  $\alpha$ -ENaC, strains were grown overnight at 26°C in selective medium supplemented with 2mM methionine to repress the expression of  $\alpha$ -ENaC. The cells were then harvested and resuspended in selective medium without methionine for 90 min to induce the expression of  $\alpha$ -ENaC. Chase reactions were subsequently performed at 30°C, and lysates were immunoblotted with anti-HA antibody. Anti-G6PD antiserum was used as a loading control. Data represent the means of 4-6 experiments,  $\pm$  SEM. The lack of visible error bars indicates that the SEM is less than the size of the symbol. Where indicated (\*\*),  $p < 0.01$ .

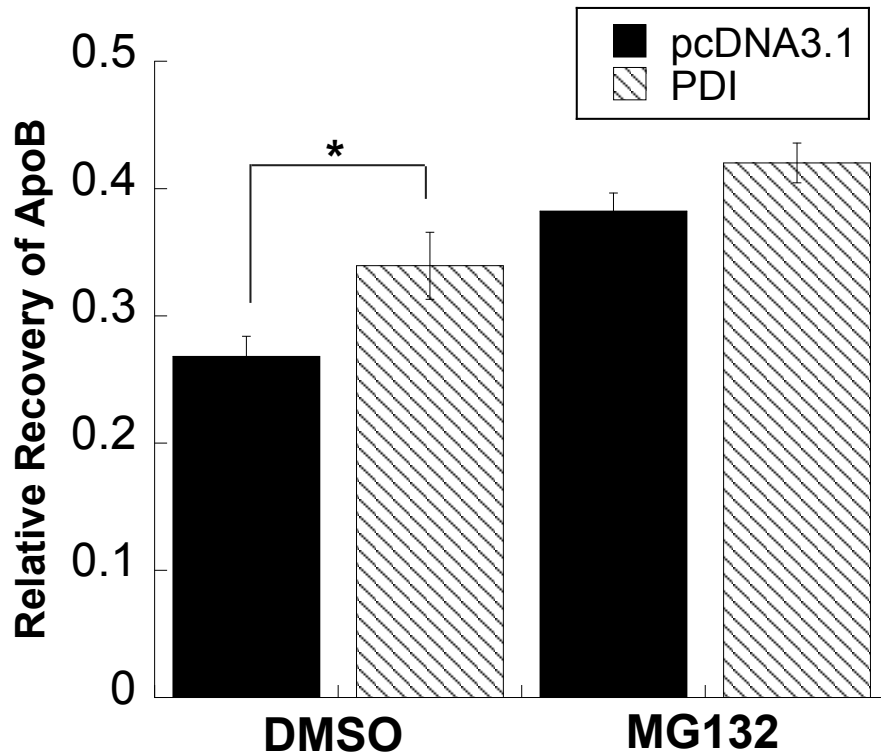
## Supplemental Figure 3



### Supplemental Figure 3. The chaperone activity of Pdi1 is not necessary for the ERAD of CPY\*.

Cycloheximide chase reactions were performed as described in the Materials and Methods in *pdi1Δ* [*PDI1*<sub>CGHC-CGHC</sub>] (●) and *pdi1Δ* [*PDI1*<sub>222-302Δ</sub>] (○) yeast strains expressing CPY\* (A). Chase reactions were performed at 30°C, and lysates were immunoblotted with anti-HA antibody. Anti-G6PD antiserum was used as a loading control. Data represent the means of 4-6 experiments, ± SEM. The lack of visible error bars indicates that the SEM is less than the size of the symbol. Lysates were also immunoblotted with anti-Pdi1 antiserum and representative images are shown (B).

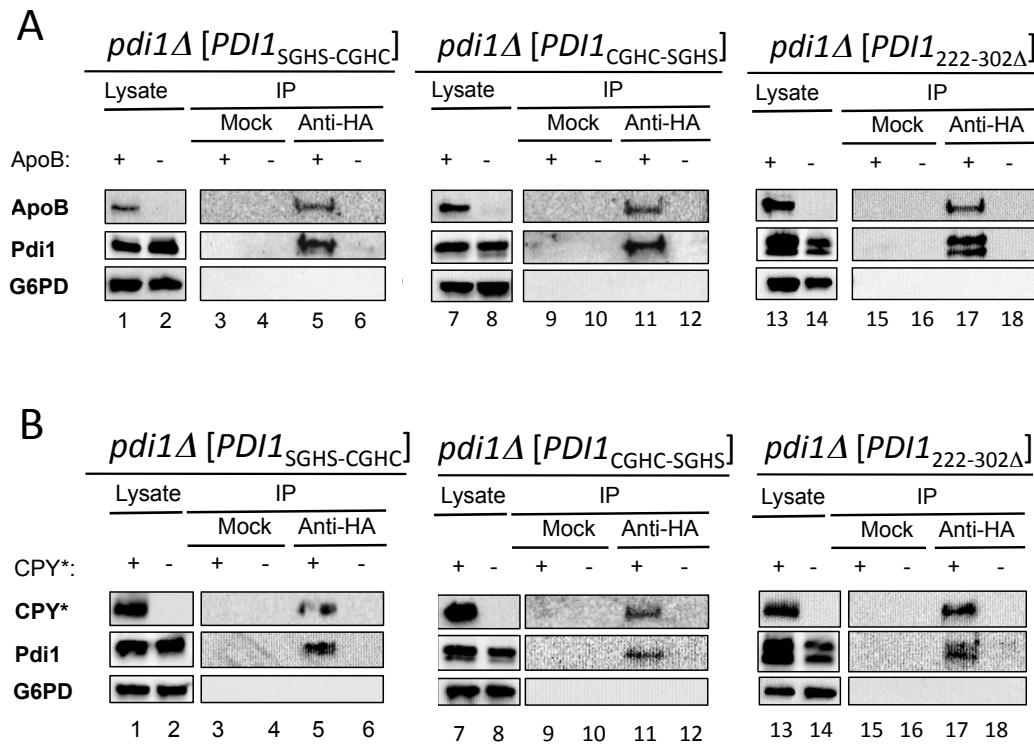
## Supplemental Figure 4



### **Supplemental Figure 4. PDI promotes ApoB secretion independent of ERAD activity.**

Following a metabolic labeling reaction in the presence of either DMSO or the proteasome inhibitor MG132, as indicated, a 90 min chase was performed as described in the Materials and Methods in McArdle-RH7777 cells transfected with a vector control (pcDNA3.1 lacking an insert), or containing the PDI gene in pcDNA3.1. The “Relative Recovery of ApoB” indicates the amount of ApoB-precipitable material recovered from cell lysates and secreted into the medium at the completion of the chase divided by the amount of ApoB-precipitable material recovered after 30 min of chase. Data represent the means of 3 independent experiments,  $\pm$  SEM. Where indicated (\*)  $p < 0.05$ .

## Supplemental Figure 5



### Supplemental Figure 5. Mutant forms of Pdi1 are able to co-precipitate with ApoB29 and CPY\*.

Native immunoprecipitation reactions were performed using anti-HA resin, or unconjugated Sepharose ("Mock"), using lysates from *pdi1Δ* yeast strains expressing active site mutant (lanes 1-6 and 7-12) or chaperone mutant (lanes 13-18) forms of *PDI1* on a plasmid. As indicated, each strain also expressed (A) ApoB29 ("ApoB"), (B) CPY\* ("CPY\*"), or harbored an empty vector control ("-"). A total of 1% of the input for the precipitation was also examined ("Lysate"). After precipitation and SDS-PAGE, the indicated proteins were examined by immunoblot analysis. The doublet observed for Pdi1 in some panels is due to differential glycosylation.