

The three-dimensional structure of nylon hydrolase and its mechanism of nylon-6 hydrolysis

SUPPLEMENTARY DATA ONLINE

Legend for supplementary figures

Fig. S1. Comparison of amino acid sequences of NylC from *Arthrobacter* plasmid pOAD2 (NylC_{p2}), *Agromyces* (NylC_A) and *Kocuria* (NylC_K). Amino acid residues are shown as *one-letter* codes. Fifteen residues differed between NylC_{p2} and NylC_K, of which the five residues in *green* boxes are altered in both NylC_K and NylC_A. Ten residues in *blue* boxes are unique to NylC_K.

Fig. S2. Stereoview of the NylC_A heterodimer shown as a ribbon diagram. The α -subunit and β -subunit in molecule A are colored in *dark green* and *light green*, respectively. The distance (32.6 Å) between Pro260 and Thr267 at C _{α} is shown.

Fig. S3. Stereoviews of overall structure of NylC_A. Superimposition of the NylC_A structure (*green*) with L-aminopeptidase D-Ala-esterase/amidase from *Ochrobactrum anthropi* (DmpA: PDB ID code, 1B65; *magenta*) (A), β -peptidyl amino peptidase from *Sphingosinicella xenopeptidilytica* (BapA: PDB ID code, 3N33; *orange*) (B), and ornithine acetyltransferase from *Streptomyces clavuligerus* (PDB ID code, 1VZ6; *yellow*) (C). Positions of N-terminus of α -subunit (α -N term), C-terminus of α -subunit (α -C term), N-terminus of β -subunit (β -N term) and C-terminus of β -subunit (β -C term) in NylC_A are shown.

Fig. S4. Multiple three-dimensional alignment of NylC_A and typical N-tn hydrolases. A multiple three-dimensional alignment was performed using secondary structure matching (SSM) of the following proteins (42): DmpA, L-aminopeptidase D-Ala-esterase/amidase (PDB ID code, 1B65); BapA, β -peptidyl amino peptidase (PDB ID code, 3N33); and OAT, ornithine acetyltransferase (PDB ID code, 1VZ6). The secondary structures are shown as *green* (helix) or *orange* (β -strand) letters. The numbering of DmpA, BapA and OAT is the same as the numbering registered in the protein data bank. Helices and β -strands of NylC_A shown in Fig. 2A are illustrated at the top with *green* cylinders (helix) and *orange* arrows (β -strand). The catalytic residue (nucleophile) is in yellow. Amino acid residues affecting the thermostability of NylC_A (Asp36, Ser111, Gly122, Tyr130, Ala137 and Met225) are marked with asterisks. Because the residues of the regions of 261-266 of NylC_A were not able to fit in the electron density map due to poor electron density distribution, they were excluded from the three-dimensional alignment.

Fig. S5. CD spectra at far UV (200-250 nm). A. CD measurements were carried out using a J-720WI spectropolarimeter (Jasco, Japan) at 25 °C. B. A temperature shift from 25°C to 75°C (at 1°C min⁻¹) was performed. CD measurements were conducted at 75°C. A cuvette with a path length of 1 mm was used for all far UV CD measurements. The results are expressed as the mean residue molar ellipticity, $[\theta]$, and defined as $[\theta] = 100 (\theta_{\text{obs}} - \theta_{\text{back}}) l^{-1} c^{-1}$, where θ_{obs} is the observed ellipticity in degrees, θ_{back} is the observed ellipticity in degrees in the absence of enzyme (background), c is the molar concentration of the residue, and l is the length of the light path (in centimeters). The protein concentration used in all far UV CD measurements was 0.1 mg ml⁻¹.

Table S1. Sequences of primers used for the construction of NylC mutants.

Mutation	Oligonucleotide	5'	Sequence	3'
G111S	FEG111S		GGAGCC <u>AGCT</u> ACGGGCTCGAGGCGGGC	
	REG111S		CCCGTA <u>GCT</u> GGCTCCGCCGGCCAGGCA	
D122G	FED122G		GGGGTGAGC <u>GGC</u> GCGCTCCTGGAACGCCTC	
	RED122G		TTCCAGGAGCGC <u>GCC</u> GCTCACCCCGGCGCC	
H130Y	FEH130Y		CTCGAGT <u>TAT</u> CGCACCGGCTTCGCCGAGCTC	
	REH130Y		GGTGCG <u>ATA</u> CTCGAGGCGTTCCAGGAGCGC	
L137A	FEL137A		GCCGAG <u>GCC</u> CAGCTGGTGTCGTCGGCG	
	REL137A		CAGCT <u>GGC</u> CCTCGGCCGAAGCCGGTGCG	
V225M	FV225M		GTGATC <u>ATG</u> ACCGCGCGGGCACGGTGGTG	
	RV225M		GCGGT <u>CAT</u> GATCACACCGACCGGGTTCGG	
D36A	FD36A		GGCAACT <u>TCC</u> ACGATCAGCGCGATCGTC	
	RD36A		GATCGT <u>GGA</u> GTTGCCGGCCTCGGTGACGGG	
E263Q	FE263Q		GGCAACT <u>TGC</u> ACGATCAGCGCGATCGTC	
	RE263Q		GATCGT <u>GCA</u> GTTGCCGGCCTCGGTGACGGG	

Underlined bases indicate the mutated sites.

Table S2. The root mean square deviation of backbone structure among subunit molecules of NylC_A.

	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O
A		0.3	0.29	0.24	0.26	0.22	0.2	0.27	0.23	0.29	0.14	0.24	0.19	0.27	0.15
B	0.3		0.37	.3	0.31	0.28	0.25	0.31	0.34	0.38	0.33	0.29	0.34	0.37	0.32
C	0.29	0.37		0.2	0.19	0.18	0.38	0.22	0.31	0.29	0.26	0.23	0.21	0.23	0.28
D	0.24	0.3	0.2		0.15	0.19	0.33	0.15	0.26	0.27	0.24	0.17	0.19	0.2	0.22
E	0.26	0.31	0.19	0.15		0.18	0.34	0.2	0.27	0.3	0.2	0.21	0.23	0.24	0.25
F	0.22	0.28	0.18	0.19	0.18		0.28	0.23	0.27	0.29	0.24	0.22	0.21	0.24	0.24
G	0.2	0.25	0.38	0.33	0.34	0.28		0.33	0.27	0.37	0.24	0.31	0.3	0.35	0.25
H	0.27	0.31	0.22	0.15	0.2	0.23	0.33		0.27	0.26	0.25	0.17	0.23	0.25	0.24
I	0.23	0.3	0.31	0.26	0.27	0.27	0.27	0.27		0.36	0.24	0.26	0.26	0.3	0.26
J	0.29	0.38	0.29	0.27	0.3	0.29	0.37	0.26	0.36		0.27	0.2	0.26	0.28	0.27
K	0.14	0.33	0.26	0.24	0.26	0.24	0.24	0.25	0.24	0.27		0.23	0.18	0.26	0.17
L	0.24	0.29	0.23	0.17	0.21	0.22	0.31	0.17	0.2	0.2	0.23		0.21	0.22	0.23
M	0.19	0.34	0.21	0.19	0.23	0.21	0.3	0.23	0.26	0.26	0.18	0.21		0.13	0.17
N	0.27	0.37	0.23	0.2	0.24	0.24	0.35	0.25	0.3	0.28	0.26	0.22	0.13		0.24
O	0.15	0.32	0.28	0.22	0.25	0.24	0.25	0.24	0.26	0.27	0.17	0.23	0.17	0.24	

The root mean square deviations (rmsd) (Å) between the fifteen molecules (A - O) at C_α were calculated by secondary structure matching (SSM).

Table S3. Search for proteins with analogous folds using the program DALI

PDB ID	Z	RMSD	LALI	NRES	%ID	PDB description
1B65	26.3	2.8	259	363	20	D-Aminopeptidase
2DRH	25.9	2.7	256	354	21	Hypothetical D-aminopeptidase (361aa long)
3N5I	25.4	3.1	264	371	19	β -Peptidyl aminopeptidase
3N33	25.2	3.6	267	367	18	β -Peptidyl aminopeptidase
3N2W	25.2	3.6	266	367	18	β -Peptidyl aminopeptidase
1VZ8	14.3	3.4	204	376	14	Ornithine acetyl-transferase
1VZ6	14.3	3.4	204	376	14	Ornithine acetyl-transferase
1VZ7	14.1	3.4	204	376	14	Ornithine acetyl-transferase
3IT4	7.3	2.7	136	193	9	Arginine biosynthesis bifunctional protein ArgJ

Z, Z-score (strength of structural similarity in S.D. above expected); RMSD, positional root mean square deviation of superimposed C α atoms in angstroms; LALI, total number of equivalenced residues; NRES, Number of amino acids in the protein; %ID, percentage of sequence identity.

Table S4. Peptides used for analysis of substrate specificity of NylC_{p2}.

Dipeptide	Gly-Gly	Gly-L-Ala	Gly-L-Leu	
	Gly-L-Phe	Gly-L-Pro	Gly-L-Val	
	L-Ala-L-Ala	D,L-Ala-Gly	D,L-Ala-D,L-Asn	
	L-Ala-L-Asp	L-Ala-L-His	D,L-Ala-D,L-Leu	
	L-Ala-L-Val	L-Ala-L-Ile	L-Ala-L-Phe	
	L-Ala-L-Pro	L-Ala-L-Thr	L-Ala-L-Trp	
	L-Ala-L-Tyr	L-Ala-L-Glu	L-Ala-L-Lys	
	L-Ala-L-Met	L-Ser-Gly	L-Ser-L-Ala	
	L-Val-Gly	L-Val-L-Ala	L-Leu-L-Ala	
	L-Leu-Gly	L-Leu-L-Tyr	L-Leu-L-Leu	
	L-Pro-Gly	L-Pro-L-Ala	L-Phe-Gly	
	L-Phe-L-Ala	L-Trp-L-Ala	L-Trp-Gly	
	L-Tyr-Gly	L-Tyr-L-Ala	L-Asp-Gly	
	L-Glu-L-Ala	L-Glu-L-Val	L-Met-Gly	
	L-Met-L-Ala	L-His-Gly	L-His-L-Ala	
	L-His-L-Leu	L-Lys-Gly		
	Tripeptide	Gly-Gly-Gly	Gly-Gly-L-Ala	
		Gly-Gly-L-His	Gly-Gly-L-Phe	
		Gly-L-Pro-L-Ala	Gly-L-Leu-L-Tyr	
		Gly-D,L-Leu-D,L-Ala	Gly-L-Ala-L-Ala	
L-Ala-L-Ala-L-Ala		D,L-Ala-Gly-Gly		
D,L-Ala-D,L-Leu-Gly		L-Leu-Gly-Gly		
Gly-L-Phe-L-Ala		L-Glu-L-Val-L-Phe		
L-Ser-L-Gln-Gly				
Tetrapeptide	Gly-Gly-Gly-Gly			
Pentapeptide	Gly-Gly-Gly-Gly-Gly			
	L-Ala-L-Ala-L-Ala-L-Ala-L-Ala			
Hexapeptide	Gly-Gly-Gly-Gly-Gly-Gly			

NylC_{p2} has no detectable activity with the 66 peptides shown in this table (ref. 14, 15).

Positions of amino acid sequence

Enzymes

36 41 50 60 62 111 122 130 137 225 230 231 257 263 354

NyIC_{p2}

D A M I A G D H L V T V V E G

NyIC_A

D A M I A S G Y A M T V V E G

NyIC_K

A V T V S S G Y A M G I L Q A

Fig. S1

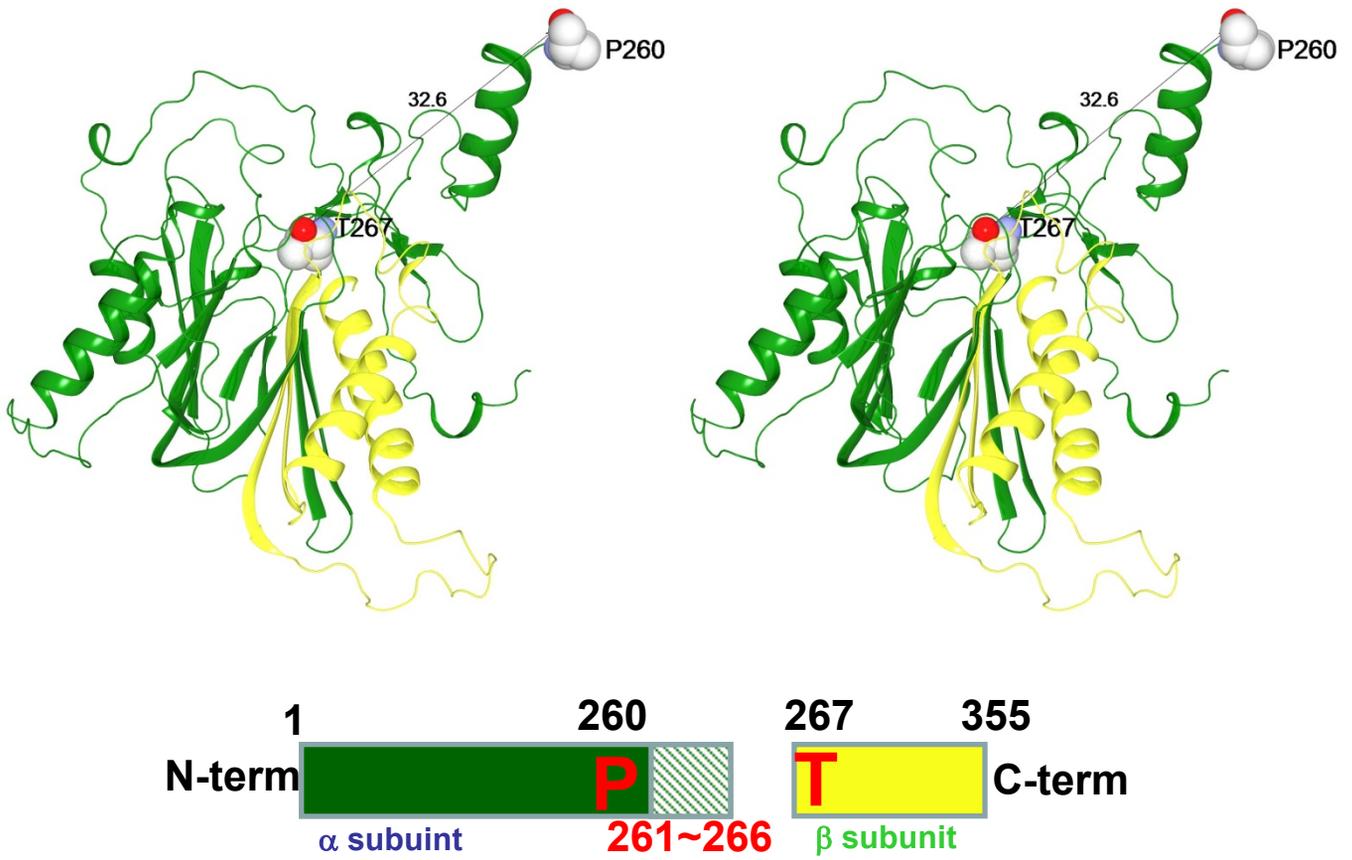


Fig. S2

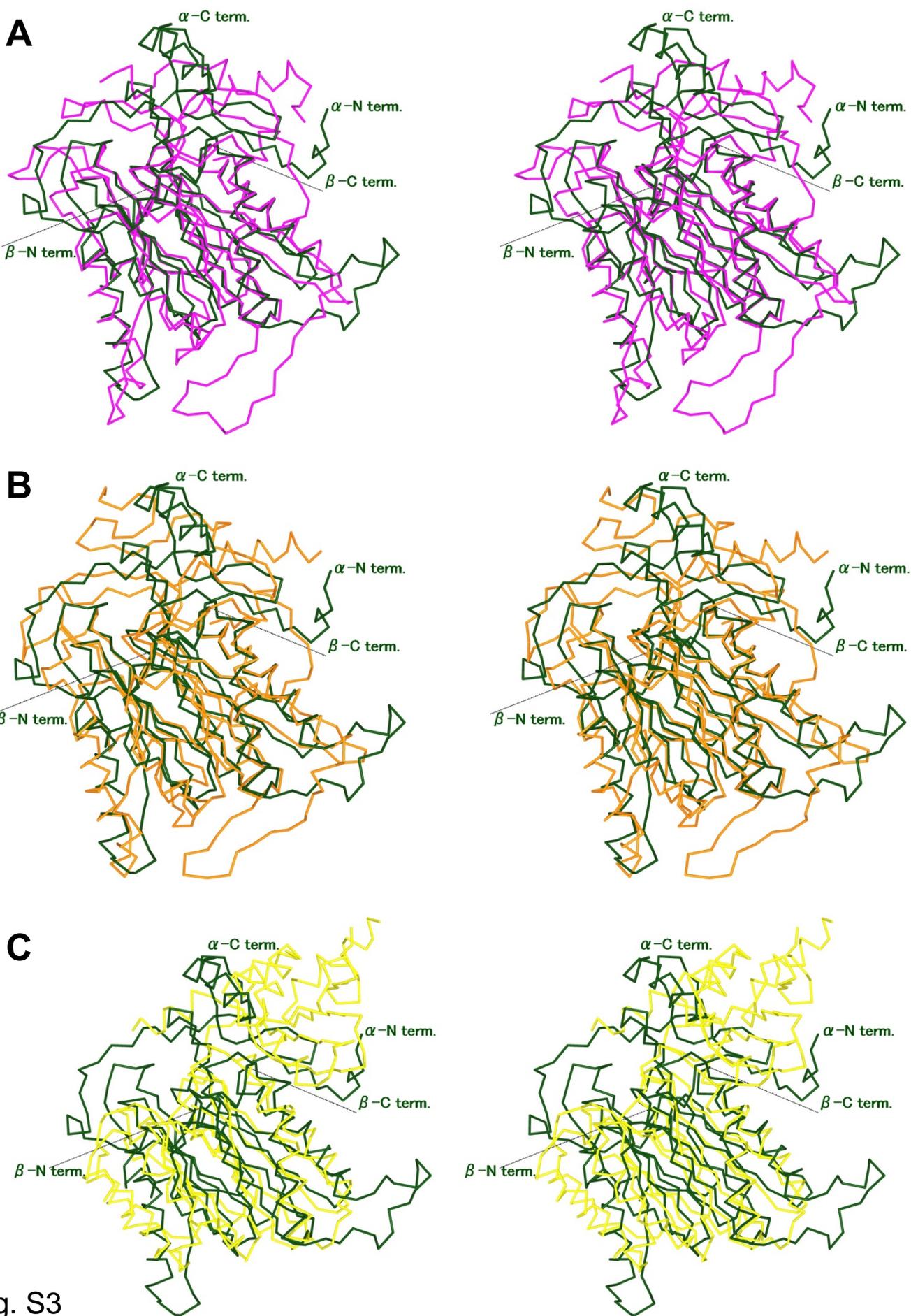


Fig. S3

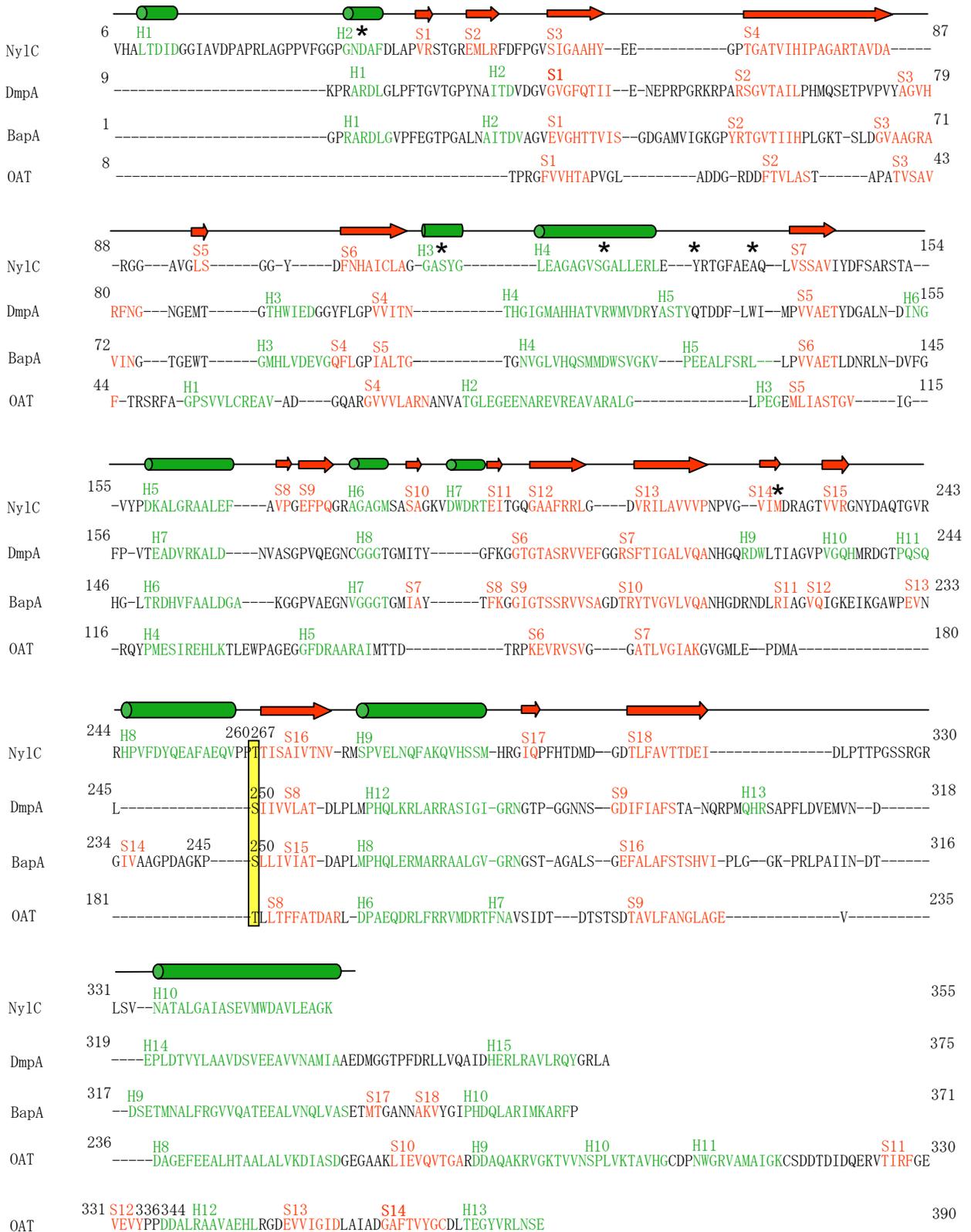


Fig. S4

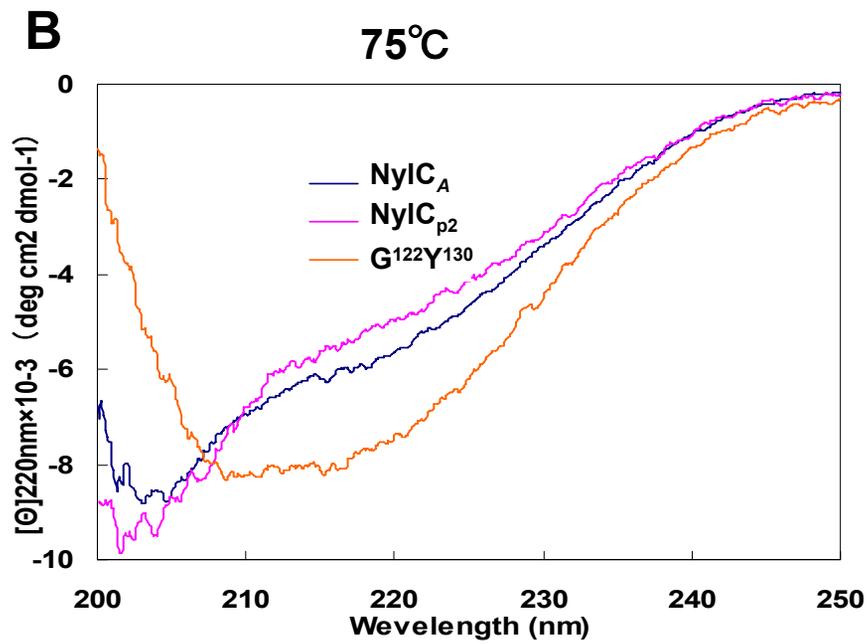
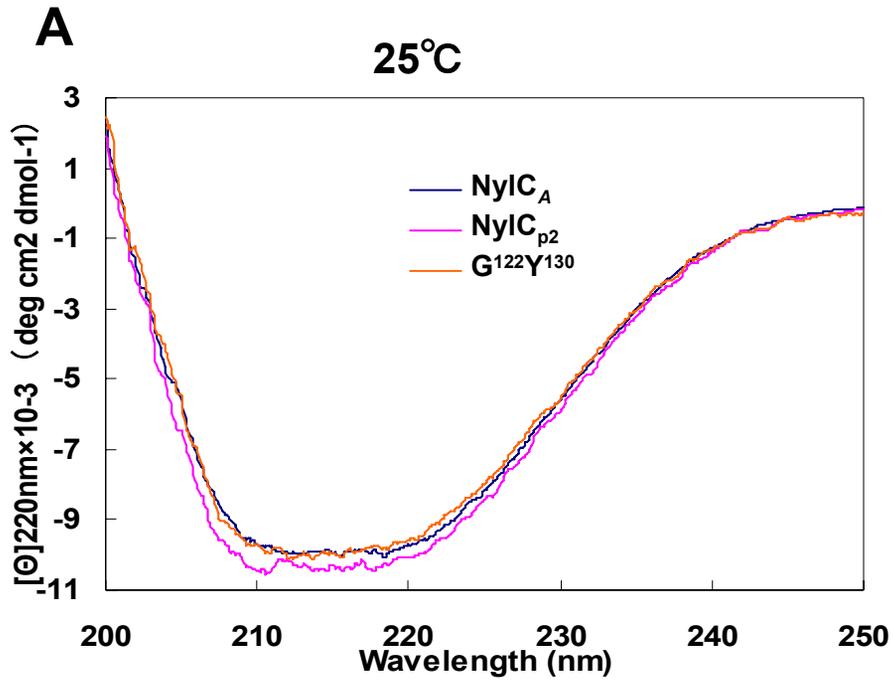


Fig. S5