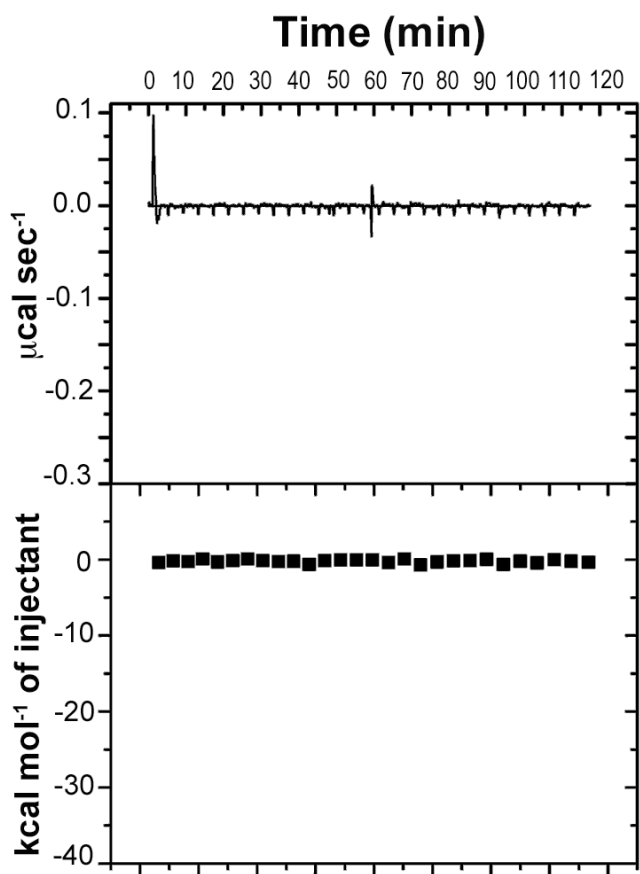
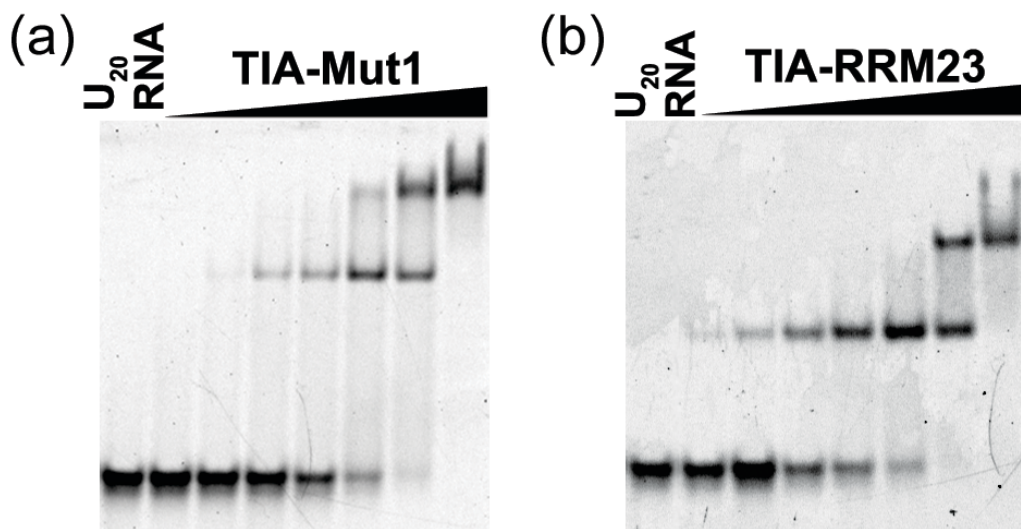


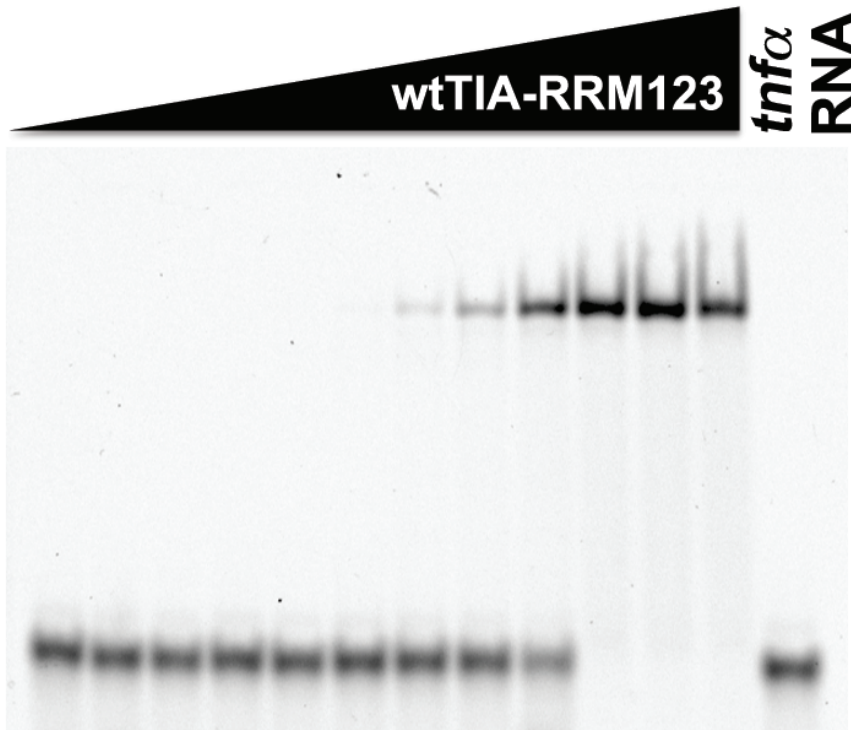
Supplementary Figure S1. Titration of wtTIA-RRM123 (70 μM) into buffer.



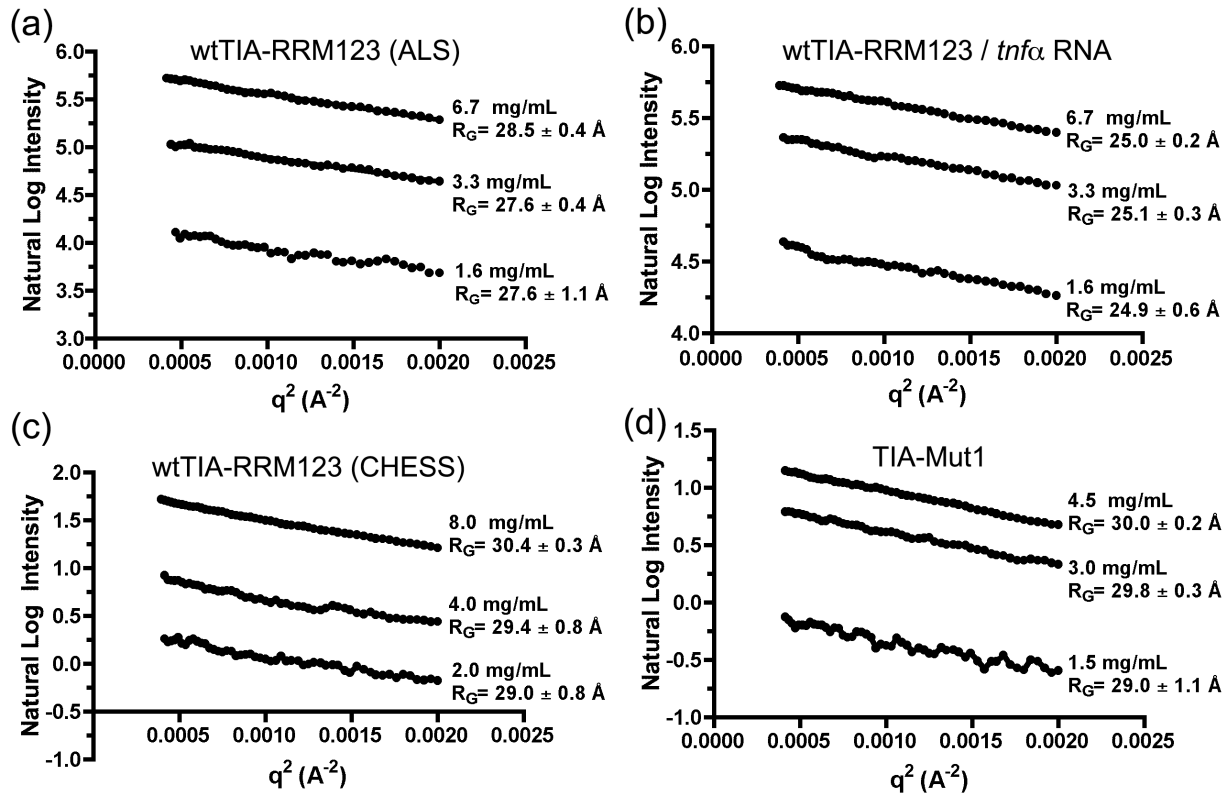
Supplementary Figure S2. Qualitative electrophoretic mobility shift assays confirm formation of two complexes during titration of (a) TIA-Mut1 or (b) TIA-RRM23 into fluorescein-labeled U₂₀ RNA (2.5 μM). The final protein concentrations were 0.3, 0.5, 0.9, 1.4, 2.2, 3.5 and 5.6 μM for TIA-Mut1, or 0.3, 0.5, 0.8, 1.3, 2.0, 3.3, and 5.3 μM for TIA-RRM23.



Supplementary Figure S3. Qualitative electrophoretic mobility shift assay demonstrating a single complex between wtTIA-RRM123 and the *tnf α* RNA oligonucleotide (5'-UUAUUUAUUUA-3') used for SAXS data collection. The fluorescein-labeled RNA (2.5 μ M final concentration) was detected by emission at 520 nm. The final protein concentrations were 0.002, 0.005, 0.013, 0.03, 0.07, 0.16, 0.37, 0.86, 2.0, 4.7, 10.9, and 25.3 μ M.



Supplementary Figure S4. Guinier plots of (a) wtTIA-RRM123 (Advanced Light Source), (b) wtTIA-RRM123 / *tnf- α* RNA, (c) wtTIA-RRM123 (Cornell High Energy Synchrotron Source), and (d) TIA-Mut1 are linear with R_G values among the concentration series that match within measurement error.



Supplementary Figure S5. Kratky plots (a) or pairwise distance distribution functions (b) of wtTIA-RRM123 SAXS data collected either at the Advanced Light Source (ALS) or Cornell High Energy Synchrotron Source (CHESS).

